



Review

# Choline, Other Methyl-Donors and Epigenetics

Steven H. Zeisel

UNC Nutrition Research Institute, Departments of Nutrition and Pediatrics, University of North Carolina at Chapel Hill, 500 Laureate Drive, Kannapolis, NC 28081, USA; [steven\\_zeisel@unc.edu](mailto:steven_zeisel@unc.edu); Tel.: +1-704-250-5006

Received: 6 March 2017; Accepted: 19 April 2017; Published: 29 April 2017

**Abstract:** Choline dietary intake varies such that many people do not achieve adequate intakes. Diet intake of choline can modulate methylation because, via betaine homocysteine methyltransferase (BHMT), this nutrient (and its metabolite, betaine) regulate the concentrations of *S*-adenosylhomocysteine and *S*-adenosylmethionine. Some of the epigenetic mechanisms that modify gene expression without modifying the genetic code depend on the methylation of DNA or of histones; and diet availability of choline and other methyl-group donors influences both of these methylations. Examples of methyl-donor mediated epigenetic effects include the changes in coat color and body weight in offspring when pregnant *agouti* mice are fed high choline, high methyl diets; the changes in tail kinking in offspring when pregnant *Axin(Fu)* mice are fed high choline, high methyl diets; the changes in *Cdkn3* methylation and altered brain development that occurs in offspring when pregnant *rodents* are fed low choline diets. When choline metabolism is disrupted by deleting the gene *Bhmt*, DNA methylation is affected (especially in a region of chromosome 13), expression of specific genes is suppressed, and liver cancers develop. Better understanding of how nutrients such as choline and methyl-donors influence epigenetic programs has importance for our understanding of not only developmental abnormalities but also for understanding the origins of chronic diseases.

**Keywords:** choline; epigenetics; DNA methylation; brain development; liver cancer

## 1. Introduction

Our genetic code is the same in almost every cell in our body, yet all tissues do not express every gene to the same extent. For the most part, this difference between tissues occurs because there are epigenetic mechanisms that modify gene expression without modifying the genetic code [1–3]. This is fortunate because it permits retuning of gene expression to achieve some degree of adaptation to the environment, including the nutrition environment. The field of epigenetics is relatively new, and to date, epigenetic mechanisms involve covalent modifications to DNA (the addition of a methyl group to carbon five in cytosine [2] or the hydroxymethylation of carbon five in cytosine [4,5]); covalent modifications to histone proteins (methylation [6,7], acetylation [8], biotinylation [9] of lysine in histone tails); as well as gene expression/translation modification by microRNAs and other noncoding RNAs [10] and by nucleosome positioning/remodeling [11]. Genome-wide epigenetic status is established in early life with different cell types developing different epigenetic programs that are usually maintained throughout later life. Thus, gene expression retuning in early life can affect metabolism and organ function in later life.

## 2. Nutrient-Responsive Epigenetic Mechanisms

There are “nutrient responsive” epigenetic mechanisms that change genes’ expression, thereby modulating metabolic pathways [3,12–23]. Studies on dietary choline and other methyl-group donors provide some of the best examples of nutrient responsive epigenetic mechanisms. DNA and histone methylases are directly influenced by the availability of methyl-groups derived from diet (from

choline/betaine, methyl-folate or methionine) because these are the precursors that are converted to the universal methyl-donor *S*-adenosylmethionine [24–28], needed for the methylation of cytosine in DNA or lysine in histones (epigenetic marks). That diet could modify DNA cytosine methylation was appreciated more than 40 years ago when it was observed that feeding rats a diet very low in choline and methionine resulted in the decreased methylation of cytosines in DNA of liver [29] which was correlated with changes in the expression of a variety of hepatic genes [30] and was linked to the spontaneous development of liver cancers on this diet [29,31,32]. Later, methylation indicator-mouse models were developed in which changes in DNA methylation during *in utero* development resulted in easily visible and stable phenotypes in mouse offspring. The mouse *agouti* gene regulates body weight, risk for diabetes and also the production of black or yellow pigment in hair. The *agouti* wildtype *A* mouse has a brown coat and is lean and not diabetic; the *agouti* mutant *A<sup>vy</sup>* has a solid yellow coat and a marked prevalence of obesity and diabetes but this phenotype can vary considerably from one mouse carrying this mutation to another, even among siblings of a single litter, because the expression of the *agouti* gene is epigenetically regulated [18]. Feeding pregnant *A<sup>vy</sup>/A* mice a diet high in choline, methionine, betaine, vitamin B12 and folic acid resulted in more offspring born that were brown and thin rather than possessing the normal yellow and fat phenotype for the *A<sup>vy</sup>/A* mouse. These phenotypes were predicted by the methylation status of *A<sup>vy</sup>* (methylation of cytosines in the *A<sup>vy</sup>* gene acts to suppress gene expression) [18,33,34]. There was a critical period in early life during which the epigenetic marks were formed (in this case, the period before birth), and after this time, restoration of a control diet did not reverse them. Mice with a different methylation indicator-gene (*Axin(Fu)*) develop kinks in their tails. When pregnant *Axin(Fu)* mice were fed diets high in choline, methionine, betaine and vitamin B12, cytosines in the gene *Axin(Fu)* were hypermethylated, gene expression was suppressed and offspring had tails with fewer kinks [16]. Again, there was a critical *in utero* time-window during which the epigenetic marks were established [16]. Other examples of choline-methyl diet modification of epigenetics include the following: insulin-like growth factor II (*IGF2*), which is an important growth-stimulating factor and metabolic regulator; and the expression of *IGF2*, which is negatively regulated by a long noncoding RNA made by the *H19* gene [35–38]. *H19* expression, in turn, is regulated by the methylation of a region shared by both genes called *Igf2DMR2*. When pregnant rats were fed choline-deficient diets, *Igf2DMR2* in fetal liver was hypermethylated, *H19* was inhibited and *IGF2* expression increased [39]. In people, maternal choline intake modulated epigenetic marks in the placenta; women with higher intake of choline had higher placental promoter methylation in the corticotrophin releasing hormone (*CRH*) and glucocorticoid receptor (*NR3C1*) genes and this was associated with lower placental expression of the corticotrophin-releasing hormone [40]. Though much of the epigenome is established during early life, some epigenetic marks are modifiable in later life. For example, methylation levels of specific CpG sites from *Srebf2*, *Agnat3*, *Esr1* and *Fasn* promoter regions in liver were changed when rats were fed a high choline (high methyl) diet, and these effects were gender dependent [41,42]. Why some epigenetic marks are fixed in early life and others are plastic is not understood.

### 3. Choline, Epigenetics and Fetal Development

The dietary choline-mediated effects on epigenetic marks are not limited to the genes discussed above; feeding pregnant mice a diet low in choline decreased cytosine methylation in the gene cyclin dependent kinase 3 (*Cdkn3*), resulting in increased expression of this gene that inhibits cell cycling [43]. These diet-mediated epigenetic changes in *Cdkn3* were associated with decreased proliferation of neural progenitor cells in the fetal brain [43,44]. This is especially interesting because maternal dietary choline is essential for normal brain development.

There is extra demand for choline during pregnancy and this increases the choline requirements of the mother and often exceeds the capacity for endogenous production of choline (in the form of phosphatidylcholine) in the liver [45]. Estrogen can induce the gene *PEMT* encoding the enzyme catalyzing the production of phosphatidylcholine in liver, and this means that young women are

somewhat less dependent on dietary intake of choline than are men [46,47]. Unfortunately, almost half of women have a gene variant that blocks estrogen-induction of the capacity to make their own choline [46,47]; these women may be especially sensitive to dietary choline variations during pregnancy. The placenta and mammary gland deliver large amounts of choline from mother to the fetus and infant, respectively [48,49]. This results in plasma and tissue choline concentrations that are much higher in the fetus and young infant than they are in adults [50,51].

Maternal supply of choline to the fetus and infant is important for brain development. In rodents, decreased supply of choline during a critical window of brain development (in the rodent, days 11–17 of gestation) results in decreased neurogenesis in the brain cortex [52] and hippocampus [44,53]. Lower maternal choline intake results in smaller fetal brains, with markedly diminished numbers of fetal progenitor cells in the germinal layers of both the cortex and hippocampus [52]. Also, with lower maternal choline intake, there is a loss of the normal layering of the developing fetal cortex because neural progenitor cells differentiate too early, forming layer 6 of the cortex, and leave too few remaining progenitors to form the five other later-born layers of the brain [52]. These six different layers connect to different neural pathways in the brain, and when they are not formed properly, these pathways are disrupted. One mechanism for this effect of choline is mediated by changes in the epidermal growth factor (EGF) signaling (required for normal timing of differentiation) in the developing cortex. Low choline availability to the fetal brain results in decreased production of the receptor for EGF [52]. Similarly, low maternal choline intake results in premature differentiation of neural progenitors in the fetal hippocampus [54–57]. This area of the brain is important for memory function. If pregnant rats are fed a diet supplemented with choline, their offspring perform better in tests of visuospatial and auditory memory [58–60] and this improvement lasts for their lifetime. Conversely, when pregnant rats are fed diets low in choline, their offspring have worse visuospatial and auditory memory [61]. Babies born from women who eat more choline during pregnancy perform better at 7 years of age in visuospatial memory testing [62], suggesting that the effects of choline in rodents may also apply to people. However, one observational study [63] reported that there was no association between cord blood choline concentrations and scores on standard intelligence tests in 5 year olds.

In addition to influencing brain neurogenesis, fetuses from pregnant mice fed a low choline diet have fewer small blood vessels in their hippocampi when compared to fetuses from mothers fed a normal or high choline diet [64]. Endothelial progenitor cells in brains of the low choline group, responding to increased expression of *Vegfc* and *Angpt2*, differentiate prematurely and stop dividing, with resulting reductions in blood vessel formation [64]. The increased expression of *Vegfc* and *Angpt2* is associated with the decreased methylation of cytosines within these two genes [64].

Thus, choline has an important role in regulating the development of the brain, and epigenetic mechanisms that likely contribute to the underlying pathways for these effects.

#### 4. Choline, Epigenetics and Liver Cancer

In addition to its role in brain development, choline and other methyl-donors also have a role in carcinogenesis. Rodents fed low choline, low methyl diets develop liver cancers [65–68]. A meta-analysis of 11 studies in people calculated that diets low in choline increased the overall relative risk for developing cancer [69] with the largest reported effects found for lung (30% increase; also see [70]), nasopharyngeal (58% increase; also see [70]) and breast cancer (60% increase; also see [71]). An increment in diet intake of 100 mg/day of choline and betaine (a metabolite derived from choline) helped reduce cancer incidence by 11% [69].

It is possible that these effects of choline are mediated by epigenetic mechanisms [72]. Choline-methyl-deficient diets result in decreased hepatic concentrations of *S*-adenosylmethionine, and increased concentrations of *S*-adenosylhomocysteine in the livers of male rats and mice [73]. Low *S*-adenosylmethionine combined with high *S*-adenosylhomocysteine concentrations act to inhibit methyltransferases' activity [74]. *S*-adenosylhomocysteine has high affinity for DNA methyltransferases (DNMTs) and competes for binding of *S*-adenosylmethionine, thereby

inhibiting DNMT enzymatic activity [75]. Choline-methyl-deficient diets result in decreased DNA methylation [30,72] at many sites including the oncogene *c-myc* [76], but also in increased methylation at other cytosines (including the hypermethylation of several tumor-suppressor genes such as *p53*, *p16<sup>INK4a</sup>*, *PtprO*, *Cdh1*, and *Cx26*) [72,77–79]. Histone methylation is also affected by methyl-deficient diets. Feeding a choline-methyl-deficient diet results in decreased hepatic histone H3 lysine 9 trimethylation (H3K9me3) and histone H4 lysine 20 trimethylation (H4K20me3) [72]. In addition, several histone methyltransferases are down-regulated (*Suv39h1*, *Prdm2/Riz1*, and *Suv420h2*) [72]. Finally, down-regulation of microRNAs (miR-122 and miR-29), and up-regulation of miR-34a, miR-155, and miR-221 occur in methyl-deficient liver [72]. These microRNAs act to modify the expression and translation of specific gene products.

When a key gene (*Bhmt*; betaine homocysteine methyltransferase) in choline metabolism is deleted in mice, these mice develop hepatocarcinomas [80]. The betaine-dependent methylation of homocysteine (by BHMT) is important for maintaining concentrations of *S*-adenosylmethionine, and for removing *S*-adenosylhomocysteine in liver and for maintaining normal patterns of DNA methylation [80]. Deleting *Bhmt* causes mostly loss of methylation at 63 differentially methylated cytosines. Of these, 33 (mostly hypomethylated) are located in one locus spanning 15.5 Mb on chromosome 13 (from 93.5 to 109 Mb (mm9), which includes the *Bhmt* gene located at chr13: 94,386,846–94,407,713) [80]. It is possible that the sensitivity to DNA methylation perturbations that we observe in chromosome 13 is a regulatory mechanism meant to sense methyl-metabolism, as not only *Bhmt* but other 1-carbon metabolism genes are located there: dimethylglycine dehydrogenase (*Dmgdh*) at chr.13: 94,444,391, methionine synthase reductase (*Mtrr*) at chr.13: 68,699,657 and 5-methyltetrahydrofolate-homocysteine methyltransferase (*Mtr*; also in chromosome 13 but outside the locus at chr.13: 12,279,086) [80]. The altered DNA methylome in *Bhmt*-null mice is associated with changes in gene expression in liver; of 18 differentially expressed genes, five are located on chromosome 13, with four out of the five falling within the chromosome 13 hypomethylated block: *Arsb*, *F2rl2*, *Iqgap2*, and *Enc1* [80]. *Iqgap2* belongs to a family of scaffolding proteins, which mediate Rho GTPase and Ca<sup>2+</sup>/calmodulin signaling in regulating multiple cellular processes, such as cell adhesion, motility, and exocytosis [81]; *Iqgap2*-null mice develop spontaneous hepatocellular carcinomas [82]. *F2rl2* is a member of the protease-activated receptor-3 (PAR-3) family and is involved in homeostasis, adhesion, proliferation, and migration [83,84]; *F2rl2* binds to signal transduction proteins and transcription factors involved in carcinogenesis [85,86].

Thus, choline and methyl metabolism has an important role in regulating carcinogenesis, and epigenetic mechanisms likely contribute to the underlying pathways for these effects.

## 5. Choline Intake Is Marginal in Many People

Choline is a required nutrient and in 1998, an Adequate Intake (AI) and a Tolerable Upper Limit (UL) for choline was established [87]. In 2016, the US Food and Drug Administration (FDA) set a Recommended Daily Intake (RDI) for choline based on the AIs as part of the new Nutrition Facts label for packaged foods (published in the Federal Register on May 27, 2016; FDA-2012-N-1210-0875, Federal Register Number:2016-11867). The US AI/RDI varies by age and gender, but is 550 mg/day in adult men and 425 mg/day in adult women (450 mg/day in pregnant and 550 mg/day lactating women). The European Food Safety Authority recently established slightly different recommendations for Adequate Intake levels for choline (European Food Safety Authority, Parma, Italy, 17 August 2016).

The US Department of Agriculture maintains a database of the choline content of foods (<https://www.ars.usda.gov/northeast-area/beltsville-md/beltsville-human-nutrition-research-center/nutrient-data-laboratory/docs/usda-database-for-the-choline-content-of-common-foods-release-2-2008/>); foods such as eggs and meats contain more choline than do most plant sources. There is a wide variation in choline intake in the diet; the US National Health and Nutrition Examination Survey (NHANES 2009–2012) reports that only 11% of adult Americans achieve the Adequate Intake level of choline, with mean intake being 300 mg/day (10%ile being 200 mg/day; 90%ile being

500 mg/day) [88]. Similar ranges of intake were reported in the Framingham Offspring Study [89], the Atherosclerosis Risk in Communities study [90,91] and the Nurse's Health Study [92]. Intake of choline is even lower in low-income countries such as Jamaica and The Gambia [93,94].

## 6. Conclusions

Choline is an essential nutrient with functional relevance in a wide array of biological pathways including epigenetic modulation of gene expression. Timing of nutrient availability is important in determining epigenetic outcomes; changes in availability during early life potentially having a greater effect than changes occurring later in life. The Developmental Origins of Health and Disease (DOHaD) hypothesis [95] is that diet in early life influences developmental plasticity and alters susceptibility to adult cardiovascular disease, type 2 diabetes, obesity [96], neuropsychiatric diseases [97], immune/inflammatory diseases [98] and cancer [99]. Epigenetic mechanisms are likely a major mechanism of DOHaD [100]. Thus, better understanding of how nutrients such as choline and methyl-donors influence epigenetic programs has importance for our understanding of not only developmental abnormalities but also for understanding the origins of chronic diseases.

**Acknowledgments:** This work was supported by a grant from the NIH (DK56350).

**Conflicts of Interest:** Steven Zeisel has an equity interest in Nutrigen Biosciences, Metabolon and in SNPitty. He is on the scientific advisory board for SNPitty and Metabolon. He receives consulting fees from Enzymotec, Committee for Essential Nutrients, and from Williams and Connelly. Zeisel has research funding from Nestle and Balchem. Nutrigen, Balchem, Committee for Essential Nutrients and Enzymotec have choline-related products.

## References

1. Deichmann, U. Epigenetics: The origins and evolution of a fashionable topic. *Dev. Biol.* **2016**, *416*, 249–254. [[CrossRef](#)] [[PubMed](#)]
2. Fuks, F. DNA methylation and histone modifications: Teaming up to silence genes. *Curr. Opin. Genet. Dev.* **2005**, *15*, 490–495. [[CrossRef](#)] [[PubMed](#)]
3. Zeisel, S.H. Epigenetic mechanisms for nutrition determinants of later health outcomes. *Am. J. Clin. Nutr.* **2009**, *89*, 1488S–1493S. [[CrossRef](#)] [[PubMed](#)]
4. Tahiliani, M.; Koh, K.P.; Shen, Y.; Pastor, W.A.; Bandukwala, H.; Brudno, Y.; Agarwal, S.; Iyer, L.M.; Liu, D.R.; Aravind, L.; et al. Conversion of 5-methylcytosine to 5-hydroxymethylcytosine in mammalian DNA by MLL partner TET1. *Science* **2009**, *324*, 930–935. [[CrossRef](#)] [[PubMed](#)]
5. Yin, R.; Mao, S.Q.; Zhao, B.; Chong, Z.; Yang, Y.; Zhao, C.; Zhang, D.; Huang, H.; Gao, J.; Li, Z.; et al. Ascorbic acid enhances Tet-mediated 5-methylcytosine oxidation and promotes DNA demethylation in mammals. *J. Am. Chem. Soc.* **2013**, *135*, 10396–10403. [[CrossRef](#)] [[PubMed](#)]
6. Brownell, J.E.; Zhou, J.; Ranalli, T.; Kobayashi, R.; Edmondson, D.G.; Roth, S.Y.; Allis, C.D. Tetrahymena histone acetyltransferase A: A homolog to yeast Gcn5p linking histone acetylation to gene activation. *Cell* **1996**, *84*, 843–851. [[CrossRef](#)]
7. Taunton, J.; Hassig, C.A.; Schreiber, S.L. A mammalian histone deacetylase related to the yeast transcriptional regulator Rpd3p. *Science* **1996**, *272*, 408–411. [[CrossRef](#)] [[PubMed](#)]
8. Berger, S.L. Histone modifications in transcriptional regulation. *Curr. Opin. Genet. Dev.* **2002**, *12*, 142–148. [[CrossRef](#)]
9. Chew, Y.C.; West, J.T.; Kratzer, S.J.; Ilvarsonn, A.M.; Eissenberg, J.C.; Dave, B.J.; Klinkebiel, D.; Christman, J.K.; Zempleni, J. Biotinylation of histones represses transposable elements in human and mouse cells and cell lines and in drosophila melanogaster. *J. Nutr.* **2008**, *138*, 2316–2322. [[CrossRef](#)] [[PubMed](#)]
10. Fabian, M.R.; Sonenberg, N.; Filipowicz, W. Regulation of mRNA translation and stability by microRNAs. *Annu. Rev. Biochem.* **2010**, *79*, 351–379. [[CrossRef](#)] [[PubMed](#)]
11. Ekici, M.; Schmitz, F.; Hohl, M.; Seigel, G.M.; Thiel, G. Chromatin structure and expression of synapsin I and synaptophysin in retinal precursor cells. *Neurochem. Int.* **2008**, *53*, 165–172. [[CrossRef](#)] [[PubMed](#)]
12. Hayakawa, K.; Hirosawa, M.; Tabei, Y.; Arai, D.; Tanaka, S.; Murakami, N.; Yagi, S.; Shiota, K. Epigenetic switching by the metabolism-sensing factors in the generation of orexin neurons from mouse embryonic stem cells. *J. Biol. Chem.* **2013**, *288*, 17099–17110. [[CrossRef](#)] [[PubMed](#)]

13. Shin, H.J.; Kim, H.; Oh, S.; Lee, J.G.; Kee, M.; Ko, H.J.; Kweon, M.N.; Won, K.J.; Baek, S.H. AMPK-SKP2-CARM1 signalling cascade in transcriptional regulation of autophagy. *Nature* **2016**, *534*, 553–557. [[CrossRef](#)] [[PubMed](#)]
14. Fullgrabe, J.; Klionsky, D.J.; Joseph, B. The return of the nucleus: Transcriptional and epigenetic control of autophagy. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 65–74. [[CrossRef](#)] [[PubMed](#)]
15. Dolinoy, D.C.; Weidman, J.R.; Waterland, R.A.; Jirtle, R.L. Maternal genistein alters coat color and protects avy mouse offspring from obesity by modifying the fetal epigenome. *Environ. Health Perspect.* **2006**, *114*, 567–572. [[CrossRef](#)] [[PubMed](#)]
16. Waterland, R.A.; Dolinoy, D.C.; Lin, J.R.; Smith, C.A.; Shi, X.; Tahiliani, K.G. Maternal methyl supplements increase offspring DNA methylation at Axin Fused. *Genesis* **2006**, *44*, 401–406. [[CrossRef](#)] [[PubMed](#)]
17. Waterland, R.A.; Travisano, M.; Tahiliani, K.G. Diet-induced hypermethylation at agouti viable yellow is not inherited transgenerationally through the female. *FASEB J.* **2007**, *21*, 3380–3385. [[CrossRef](#)] [[PubMed](#)]
18. Wolff, G.L.; Kodell, R.L.; Moore, S.R.; Cooney, C.A. Maternal epigenetics and methyl supplements affect agouti gene expression in Avy/a mice. *FASEB J.* **1998**, *12*, 949–957. [[PubMed](#)]
19. Mehedint, M.G.; Niculescu, M.D.; Craciunescu, C.N.; Zeisel, S.H. Choline deficiency alters global histone methylation and epigenetic marking at the Re1 site of the calbindin 1 gene. *FASEB J.* **2010**, *24*, 184–195. [[CrossRef](#)] [[PubMed](#)]
20. Fryer, A.A.; Nafee, T.M.; Ismail, K.M.; Carroll, W.D.; Emes, R.D.; Farrell, W.E. Line-1 DNA methylation is inversely correlated with cord plasma homocysteine in man: A preliminary study. *Epigenetics* **2009**, *4*, 394–398. [[CrossRef](#)] [[PubMed](#)]
21. Hsu, A.; Wong, C.P.; Yu, Z.; Williams, D.E.; Dashwood, R.H.; Ho, E. Promoter de-methylation of cyclin D2 by sulforaphane in prostate cancer cells. *Clin. Epigenetics* **2011**. [[CrossRef](#)] [[PubMed](#)]
22. Lee, H.S.; Barraza-Villarreal, A.; Biessy, C.; Duarte-Salles, T.; Sly, P.D.; Ramakrishnan, U.; Rivera, J.; Herceg, Z.; Romieu, I. Dietary supplementation with polyunsaturated fatty acid during pregnancy modulates DNA methylation at IGF2/H19 imprinted genes and growth of infants. *Physiol. Genom.* **2014**, *46*, 851–857. [[CrossRef](#)] [[PubMed](#)]
23. Oommen, A.M.; Griffin, J.B.; Sarath, G.; Zempleni, J. Roles for nutrients in epigenetic events. *J. Nutr. Biochem.* **2005**, *16*, 74–77. [[CrossRef](#)] [[PubMed](#)]
24. Zeisel, S.H. Choline: Critical role during fetal development and dietary requirements in adults. *Annu. Rev. Nutr.* **2006**, *26*, 229–250. [[CrossRef](#)] [[PubMed](#)]
25. Salbaum, J.M.; Kappen, C. Genetic and epigenomic footprints of folate. *Prog. Mol. Biol. Transl. Sci.* **2012**, *108*, 129–158. [[PubMed](#)]
26. Kok, D.E.; Dhonukshe-Rutten, R.A.; Lute, C.; Heil, S.G.; Uitterlinden, A.G.; van der Velde, N.; van Meurs, J.B.; van Schoor, N.M.; Hooiveld, G.J.; de Groot, L.C.; et al. The effects of long-term daily folic acid and vitamin b12 supplementation on genome-wide DNA methylation in elderly subjects. *Clin. Epigenetics* **2015**. [[CrossRef](#)] [[PubMed](#)]
27. Nilsson, E.; Matte, A.; Perfilyev, A.; de Mello, V.D.; Kakela, P.; Pihlajamaki, J.; Ling, C. Epigenetic alterations in human liver from subjects with type 2 diabetes in parallel with reduced folate levels. *J. Clin. Endocrinol. Metab.* **2015**, *100*, E1491–E1501. [[CrossRef](#)] [[PubMed](#)]
28. Ly, A.; Hoyt, L.; Crowell, J.; Kim, Y.I. Folate and DNA methylation. *Antioxid. Redox. Signal* **2012**, *17*, 302–326. [[CrossRef](#)] [[PubMed](#)]
29. Wilson, M.J.; Shivapurkar, N.; Poirier, L.A. Hypomethylation of hepatic nuclear DNA in rats fed with a carcinogenic methyl-deficient diet. *Biochem. J.* **1984**, *218*, 987–990. [[CrossRef](#)] [[PubMed](#)]
30. Wainfan, E.; Poirier, L.A. Methyl groups in carcinogenesis: Effects on DNA methylation and gene expression. *Cancer Res.* **1992**, *52*, 2071s–2077s. [[PubMed](#)]
31. Ghoshal, A.K.; Farber, E. The induction of liver cancer by dietary deficiency of choline and methionine without added carcinogens. *Carcinogenesis* **1984**, *5*, 1367–1370. [[CrossRef](#)] [[PubMed](#)]
32. Newberne, P.M.; Rogers, A.E. Labile methyl groups and the promotion of cancer. *Ann. Rev. Nutr.* **1986**, *6*, 407–432. [[CrossRef](#)] [[PubMed](#)]
33. Waterland, R.A.; Jirtle, R.L. Transposable elements: Targets for early nutritional effects on epigenetic gene regulation. *Mol. Cell Biol.* **2003**, *23*, 5293–5300. [[CrossRef](#)] [[PubMed](#)]
34. Cooney, C.A.; Dave, A.A.; Wolff, G.L. Maternal methyl supplements in mice affect epigenetic variation and DNA methylation of offspring. *J. Nutr.* **2002**, *132*, 2393S–2400S. [[PubMed](#)]

35. Bartolomei, M.S.; Zemel, S.; Tilghman, S.M. Parental imprinting of the mouse H19 gene. *Nature* **1991**, *351*, 153–155. [[CrossRef](#)] [[PubMed](#)]
36. DeChiara, T.M.; Robertson, E.J.; Efstratiadis, A. Parental imprinting of the mouse insulin-like growth factor II gene. *Cell* **1991**, *64*, 849–859. [[CrossRef](#)]
37. Bell, A.C.; Felsenfeld, G. Methylation of a CTCF-dependent boundary controls imprinted expression of the Igf2 gene. *Nature* **2000**, *405*, 482–485. [[PubMed](#)]
38. Claycombe, K.J.; Uthus, E.O.; Roemmich, J.N.; Johnson, L.K.; Johnson, W.T. Prenatal low-protein and postnatal high-fat diets induce rapid adipose tissue growth by inducing Igf2 expression in sprague dawley rat offspring. *J. Nutr.* **2013**, *143*, 1533–1539. [[CrossRef](#)] [[PubMed](#)]
39. Kovacheva, V.P.; Mellott, T.J.; Davison, J.M.; Wagner, N.; Lopez-Coviella, I.; Schnitzler, A.C.; Blusztajn, J.K. Gestational choline deficiency causes global and Igf2 gene DNA hypermethylation by up-regulation of Dnmt1 expression. *J. Biol. Chem.* **2007**, *282*, 31777–31788. [[CrossRef](#)] [[PubMed](#)]
40. Jiang, X.; Yan, J.; West, A.A.; Perry, C.A.; Malysheva, O.V.; Devapatla, S.; Pressman, E.; Vermeylen, F.; Caudill, M.A. Maternal choline intake alters the epigenetic state of fetal cortisol-regulating genes in humans. *FASEB J.* **2012**, *26*, 3563–3574. [[CrossRef](#)] [[PubMed](#)]
41. Cordero, P.; Champion, J.; Milagro, F.I.; Martinez, J.A. Transcriptomic and epigenetic changes in early liver steatosis associated to obesity: Effect of dietary methyl donor supplementation. *Mol. Genet. Met.* **2013**, *110*, 388–395. [[CrossRef](#)] [[PubMed](#)]
42. Cordero, P.; Gomez-Uriz, A.M.; Champion, J.; Milagro, F.I.; Martinez, J.A. Dietary supplementation with methyl donors reduces fatty liver and modifies the fatty acid synthase DNA methylation profile in rats fed an obesogenic diet. *Genes Nutr.* **2013**, *8*, 105–113. [[CrossRef](#)] [[PubMed](#)]
43. Niculescu, M.D.; Craciunescu, C.N.; Zeisel, S.H. Dietary choline deficiency alters global and gene-specific DNA methylation in the developing hippocampus of mouse fetal brains. *Faseb. J.* **2006**, *20*, 43–49. [[CrossRef](#)] [[PubMed](#)]
44. Craciunescu, C.N.; Albright, C.D.; Mar, M.H.; Song, J.; Zeisel, S.H. Choline availability during embryonic development alters progenitor cell mitosis in developing mouse hippocampus. *J. Nutr.* **2003**, *133*, 3614–3618. [[PubMed](#)]
45. Zeisel, S.H.; Mar, M.-H.; Zhou, Z.-W.; da Costa, K.-A. Pregnancy and lactation are associated with diminished concentrations of choline and its metabolites in rat liver. *J. Nutr.* **1995**, *125*, 3049–3054. [[PubMed](#)]
46. Resseguie, M.; Song, J.; Niculescu, M.D.; da Costa, K.A.; Randall, T.A.; Zeisel, S.H. Phosphatidylethanolamine N-methyltransferase (PEMT) gene expression is induced by estrogen in human and mouse primary hepatocytes. *Faseb. J.* **2007**, *21*, 2622–2632. [[CrossRef](#)] [[PubMed](#)]
47. Resseguie, M.E.; da Costa, K.A.; Galanko, J.A.; Patel, M.; Davis, I.J.; Zeisel, S.H. Aberrant estrogen regulation of PEMT results in choline deficiency-associated liver dysfunction. *J. Biol. Chem.* **2011**, *286*, 1649–1658. [[CrossRef](#)] [[PubMed](#)]
48. Sweiry, J.H.; Yudilevich, D.L. Characterization of choline transport at maternal and fetal interfaces of the perfused Guinea-pig placenta. *J. Physiol.* **1985**, *366*, 251–266. [[CrossRef](#)] [[PubMed](#)]
49. Fischer, L.M.; da Costa, K.A.; Galanko, J.; Sha, W.; Stephenson, B.; Vick, J.; Zeisel, S.H. Choline intake and genetic polymorphisms influence choline metabolite concentrations in human breast milk and plasma. *Am J. Clin. Nutr.* **2010**, *92*, 336–346. [[CrossRef](#)] [[PubMed](#)]
50. Zeisel, S.H.; Epstein, M.F.; Wurtman, R.J. Elevated choline concentration in neonatal plasma. *Life Sci.* **1980**, *26*, 1827–1831. [[CrossRef](#)]
51. Ilcol, Y.O.; Ozbek, R.; Hamurtekin, E.; Ulus, I.H. Choline status in newborns, infants, children, breast-feeding women, breast-fed infants and human breast milk. *J. Nutr. Biochem.* **2005**, *16*, 489–499. [[CrossRef](#)] [[PubMed](#)]
52. Wang, Y.; Surzenko, N.; Friday, W.B.; Zeisel, S.H. Maternal dietary intake of choline in mice regulates development of the cerebral cortex in the offspring. *FASEB J.* **2016**, *30*, 1566–1578. [[CrossRef](#)] [[PubMed](#)]
53. Albright, C.D.; Tsai, A.Y.; Friedrich, C.B.; Mar, M.H.; Zeisel, S.H. Choline availability alters embryonic development of the hippocampus and septum in the rat. *Brain Res. Dev. Brain Res.* **1999**, *113*, 13–20. [[CrossRef](#)]
54. Albright, C.D.; Mar, M.H.; Craciunescu, C.N.; Song, J.; Zeisel, S.H. Maternal dietary choline availability alters the balance of netrin-1 and DCC neuronal migration proteins in fetal mouse brain hippocampus. *Brain Res. Dev. Brain Res.* **2005**, *159*, 149–154. [[CrossRef](#)] [[PubMed](#)]

55. Albright, C.D.; Mar, M.H.; Friedrich, C.B.; Brown, E.C.; Zeisel, S.H. Maternal choline availability alters the localization of p15Ink4B and p27Kip1 cyclin-dependent kinase inhibitors in the developing fetal rat brain hippocampus. *Dev. Neurosci.* **2001**, *23*, 100–106. [[CrossRef](#)] [[PubMed](#)]
56. Albright, C.D.; Siwek, D.F.; Craciunescu, C.N.; Mar, M.H.; Kowall, N.W.; Williams, C.L.; Zeisel, S.H. Choline availability during embryonic development alters the localization of calretinin in developing and aging mouse hippocampus. *Nutr. Neurosci.* **2003**, *6*, 129–134. [[CrossRef](#)] [[PubMed](#)]
57. Albright, C.D.; Tsai, A.Y.; Mar, M.-H.; Zeisel, S.H. Choline availability modulates the expression of TGF $\beta$ 1 and cytoskeletal proteins in the hippocampus of developing rat brain. *Neurochem. Res.* **1998**, *23*, 751–758. [[CrossRef](#)] [[PubMed](#)]
58. Meck, W.H.; Williams, C.L.; Cermak, J.M.; Blusztajn, J.K. Developmental periods of choline sensitivity provide an ontogenetic mechanism for regulating memory capacity and age-related dementia. *Front. Integr. Neurosci.* **2007**. [[CrossRef](#)] [[PubMed](#)]
59. Meck, W.H.; Williams, C.L. Metabolic imprinting of choline by its availability during gestation: Implications for memory and attentional processing across the lifespan. *Neurosci. Biobehav. Rev.* **2003**, *27*, 385–399. [[CrossRef](#)]
60. Meck, W.H.; Smith, R.A.; Williams, C.L. Pre- and postnatal choline supplementation produces long-term facilitation of spatial memory. *Dev. Psychobiol.* **1988**, *21*, 339–353. [[CrossRef](#)] [[PubMed](#)]
61. Meck, W.H.; Williams, C.L. Choline supplementation during prenatal development reduces proactive interference in spatial memory. *Brain Res. Dev. Brain Res.* **1999**, *118*, 51–59. [[CrossRef](#)]
62. Boeke, C.E.; Gillman, M.W.; Hughes, M.D.; Rifas-Shiman, S.L.; Villamor, E.; Oken, E. Choline intake during pregnancy and child cognition at age 7 years. *Am. J. Epidemiol.* **2013**, *177*, 1338–1347. [[CrossRef](#)] [[PubMed](#)]
63. Signore, C.; Ueland, P.M.; Troendle, J.; Mills, J.L. Choline concentrations in human maternal and cord blood and intelligence at 5 y of age. *Am. J. Clin. Nutr.* **2008**, *87*, 896–902. [[PubMed](#)]
64. Mehedint, M.G.; Craciunescu, C.N.; Zeisel, S.H. Maternal dietary choline deficiency alters angiogenesis in fetal mouse hippocampus. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 12834–12839. [[CrossRef](#)] [[PubMed](#)]
65. da Costa, K.; Cochary, E.F.; Blusztajn, J.K.; Garner, S.C.; Zeisel, S.H. Accumulation of 1,2-sn-diradylglycerol with increased membrane-associated protein kinase C may be the mechanism for spontaneous hepatocarcinogenesis in choline deficient rats. *J. Biol. Chem.* **1993**, *268*, 2100–2105. [[PubMed](#)]
66. da Costa, K.A.; Garner, S.C.; Chang, J.; Zeisel, S.H. Effects of prolonged (1 year) choline deficiency and subsequent refeeding of choline on 1,2-sn-diradylglycerol, fatty acids and protein kinase C in rat liver. *Carcinogenesis* **1995**, *16*, 327–334. [[CrossRef](#)] [[PubMed](#)]
67. Rogers, A.E.; Zeisel, S.H.; Groopman, J. Diet and carcinogenesis. *Carcinogenesis* **1993**, *14*, 2205–2217. [[CrossRef](#)] [[PubMed](#)]
68. Zeisel, S.H.; da Costa, K.A.; Albright, C.D.; Shin, O.H. Choline and hepatocarcinogenesis in the rat. *Adv. Exp. Med. Biol.* **1995**, *375*, 65–74. [[PubMed](#)]
69. Sun, S.; Li, X.; Ren, A.; Du, M.; Du, H.; Shu, Y.; Zhu, L.; Wang, W. Choline and betaine consumption lowers cancer risk: A meta-analysis of epidemiologic studies. *Sci. Rep.* **2016**, *6*, 35547. [[CrossRef](#)] [[PubMed](#)]
70. Ying, J.; Rahbar, M.H.; Hallman, D.M.; Hernandez, L.M.; Spitz, M.R.; Forman, M.R.; Gorlova, O.Y. Associations between dietary intake of choline and betaine and lung cancer risk. *PLoS ONE* **2013**. [[CrossRef](#)] [[PubMed](#)]
71. Zhang, C.X.; Pan, M.X.; Li, B.; Wang, L.; Mo, X.F.; Chen, Y.M.; Lin, F.Y.; Ho, S.C. Choline and betaine intake is inversely associated with breast cancer risk: A two-stage case-control study in china. *Cancer Sci.* **2013**, *104*, 250–258. [[CrossRef](#)] [[PubMed](#)]
72. Pogribny, I.P.; James, S.J.; Beland, F.A. Molecular alterations in hepatocarcinogenesis induced by dietary methyl deficiency. *Mol. Nutr. Food Res.* **2012**, *56*, 116–125. [[CrossRef](#)] [[PubMed](#)]
73. Shivapurkar, N.; Poirier, L.A. Tissue levels of S-adenosylmethionine and S-adenosylhomocysteine in rats fed methyl-deficient, amino acid-defined diets for one to five weeks. *Carcinogenesis* **1983**, *4*, 1051–1057. [[CrossRef](#)] [[PubMed](#)]
74. Tehlivets, O.; Malanovic, N.; Visram, M.; Pavkov-Keller, T.; Keller, W. S-adenosyl-L-homocysteine hydrolase and methylation disorders: Yeast as a model system. *Biochim. Biophys. Acta* **2013**, *1832*, 204–215. [[CrossRef](#)] [[PubMed](#)]



75. Lin, N.; Qin, S.; Luo, S.; Cui, S.; Huang, G.; Zhang, X. Homocysteine induces cytotoxicity and proliferation inhibition in neural stem cells via DNA methylation in vitro. *FEBS J.* **2014**, *281*, 2088–2096. [[CrossRef](#)] [[PubMed](#)]
76. Tsujiuchi, T.; Tsutsumi, M.; Sasaki, Y.; Takahama, M.; Konishi, Y. Hypomethylation of CpG sites and c-myc gene overexpression in hepatocellular carcinomas, but not hyperplastic nodules, induced by a choline-deficient L-amino acid-defined diet in rats. *Jpn. J. Cancer Res.* **1999**, *90*, 909–913. [[CrossRef](#)] [[PubMed](#)]
77. Tryndyak, V.P.; Han, T.; Muskhelishvili, L.; Fuscoe, J.C.; Ross, S.A.; Beland, F.A.; Pogribny, I.P. Coupling global methylation and gene expression profiles reveal key pathophysiological events in liver injury induced by a methyl-deficient diet. *Mol. Nutr. Food Res.* **2011**, *55*, 411–418. [[CrossRef](#)] [[PubMed](#)]
78. Motiwala, T.; Ghoshal, K.; Das, A.; Majumder, S.; Weichenhan, D.; Wu, Y.Z.; Holman, K.; James, S.J.; Jacob, S.T.; Plass, C. Suppression of the protein tyrosine phosphatase receptor type O gene (PTPRO) by methylation in hepatocellular carcinomas. *Oncogene* **2003**, *22*, 6319–6331. [[CrossRef](#)] [[PubMed](#)]
79. Tsujiuchi, T.; Shimizu, K.; Itsuzaki, Y.; Onishi, M.; Sugata, E.; Fujii, H.; Honoki, K. CpG site hypermethylation of E-cadherin and Connexin26 genes in hepatocellular carcinomas induced by a choline-deficient L-amino acid-defined diet in rats. *Mol. Carcinog.* **2007**, *46*, 269–274. [[CrossRef](#)] [[PubMed](#)]
80. Lupu, D.S.; Orozco, L.D.; Wang, Y.; Cullen, J.M.; Pellegrini, M.; Zeisel, S.H. Altered methylation of specific DNA loci in the liver of Bhmt-null mice results in repression of Iqgap2 and F2rl2 and is associated with development of preneoplastic foci. *FASEB J.* **2017**. [[CrossRef](#)] [[PubMed](#)]
81. Briggs, M.W.; Sacks, D.B. Iqgap proteins are integral components of cytoskeletal regulation. *EMBO Rep.* **2003**, *4*, 571–574. [[CrossRef](#)] [[PubMed](#)]
82. Gnatenko, D.V.; Xu, X.; Zhu, W.; Schmidt, V.A. Transcript profiling identifies iqgap2(-/-) mouse as a model for advanced human hepatocellular carcinoma. *PLoS ONE* **2013**. [[CrossRef](#)] [[PubMed](#)]
83. Guo, X.; Wang, M.; Zhao, Y.; Wang, X.; Shen, M.; Zhu, F.; Shi, C.; Xu, M.; Li, X.; Peng, F.; et al. Par3 regulates invasion of pancreatic cancer cells via interaction with Tiam1. *Clin. Exp. Med.* **2015**. [[CrossRef](#)]
84. Segal, L.; Katz, L.S.; Lupu-Meiri, M.; Shapira, H.; Sandbank, J.; Gershengorn, M.C.; Oron, Y. Proteinase-activated receptors differentially modulate in vitro invasion of human pancreatic adenocarcinoma PANC-1 cells in correlation with changes in the expression of CDC42 protein. *Pancreas* **2014**, *43*, 103–108. [[CrossRef](#)] [[PubMed](#)]
85. Warner, D.R.; Pisano, M.M.; Roberts, E.A.; Greene, R.M. Identification of three novel Smad binding proteins involved in cell polarity. *FEBS Lett.* **2003**, *539*, 167–173. [[CrossRef](#)]
86. Odom, D.T.; Zizlsperger, N.; Benjamin Gordon, D.; Bell, G.W.; Rinaldi, N.J.; Murray, H.L.; Volkert, T.L.; Schreiber, J.; Alexander Rolfe, P.; Gifford, D.K.; et al. Control of pancreas and liver gene expression by HNF transcription factors. *Science* **2004**, *303*, 1378–1381. [[CrossRef](#)] [[PubMed](#)]
87. Institute of Medicine; National Academy of Sciences USA. Choline. In *Dietary reference intakes for folate, thiamin, riboflavin, niacin, vitamin b12, panthothenic acid, biotin, and choline*; National Academy Press: Washington, DC, USA, 1998.
88. Wallace, T.C.; Fulgoni, V.L., 3rd. Assessment of total choline intakes in the United States. *J. Am. Coll. Nutr.* **2016**, *35*, 108–112. [[CrossRef](#)] [[PubMed](#)]
89. Cho, E.; Zeisel, S.H.; Jacques, P.; Selhub, J.; Dougherty, L.; Colditz, G.A.; Willett, W.C. Dietary choline and betaine assessed by food-frequency questionnaire in relation to plasma total homocysteine concentration in the Framingham Offspring study. *Am. J. Clin. Nutr.* **2006**, *83*, 905–911. [[PubMed](#)]
90. Bidulescu, A.; Chambless, L.E.; Siega-Riz, A.M.; Zeisel, S.H.; Heiss, G. Usual choline and betaine dietary intake and incident coronary heart disease: The Atherosclerosis Risk in Communities (ARIC) study. *BMC Cardiovasc. Disord.* **2007**. [[CrossRef](#)] [[PubMed](#)]
91. Bidulescu, A.; Chambless, L.E.; Siega-Riz, A.M.; Zeisel, S.H.; Heiss, G. Repeatability and measurement error in the assessment of choline and betaine dietary intake: The Atherosclerosis Risk in Communities (ARIC) study. *Nutr. J.* **2009**. [[CrossRef](#)] [[PubMed](#)]
92. Cho, E.; Willett, W.C.; Colditz, G.A.; Fuchs, C.S.; Wu, K.; Chan, A.T.; Zeisel, S.H.; Giovannucci, E.L. Dietary choline and betaine and the risk of distal colorectal adenoma in women. *J. Natl. Cancer Inst.* **2007**, *99*, 1224–1231. [[CrossRef](#)] [[PubMed](#)]
93. Dominguez-Salas, P.; Moore, S.E.; Cole, D.; da Costa, K.A.; Cox, S.E.; Dyer, R.A.; Fulford, A.J.; Innis, S.M.; Waterland, R.A.; Zeisel, S.H.; et al. DNA methylation potential: Dietary intake and blood concentrations of one-carbon metabolites and cofactors in rural African women. *Am. J. Clin. Nutr.* **2013**. [[CrossRef](#)] [[PubMed](#)]

94. Gossell-Williams, M.; Fletcher, H.; McFarlane-Anderson, N.; Jacob, A.; Patel, J.; Zeisel, S. Dietary intake of choline and plasma choline concentrations in pregnant women in Jamaica. *West Indian Med. J.* **2005**, *54*, 355–359. [[CrossRef](#)] [[PubMed](#)]
95. Mochizuki, K.; Hariya, N.; Honma, K.; Goda, T. Relationship between epigenetic regulation, dietary habits, and the developmental origins of health and disease theory. *Congenit. Anom.* **2017**. [[CrossRef](#)] [[PubMed](#)]
96. Barker, D.J. Fetal programming of coronary heart disease. *Trends Endocrinol. Metab.* **2002**, *13*, 364–368. [[CrossRef](#)]
97. Faa, G.; Manchia, M.; Pintus, R.; Gerosa, C.; Marcialis, M.A.; Fanos, V. Fetal programming of neuropsychiatric disorders. *Birth Defects Res. C Embryo Today* **2016**, *108*, 207–223. [[CrossRef](#)] [[PubMed](#)]
98. Chen, T.; Liu, H.X.; Yan, H.Y.; Wu, D.M.; Ping, J. Developmental origins of inflammatory and immune diseases. *Mol. Hum. Reprod.* **2016**, *22*, 858–865. [[CrossRef](#)] [[PubMed](#)]
99. Walker, C.L.; Ho, S.M. Developmental reprogramming of cancer susceptibility. *Nat. Rev. Cancer* **2012**, *12*, 479–486. [[CrossRef](#)] [[PubMed](#)]
100. Ong, M.L.; Lin, X.; Holbrook, J.D. Measuring epigenetics as the mediator of gene/environment interactions in DOHaD. *J. Dev. Orig. Health Dis.* **2015**, *6*, 10–16. [[CrossRef](#)] [[PubMed](#)]



© 2017 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).