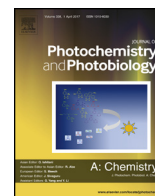




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Reactive nitrogen/oxygen species production by nitro/nitrosyl supramolecular ruthenium porphyrin complexes



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ABSTRACT

This manuscript reports on new nitro/nitrosyl Ru-based complexes, which were synthesized with the purpose of using them as precursors to obtain supramolecular ruthenium porphyrin species ($\{TPyP[Ru(NO_2)(5,5'\text{-Mebipy})_2]_4\}(PF_6)_4$) and ($\{TPyP[Ru(NO)(5,5'\text{-Mebipy})_2]_4\}(PF_6)_{12}$). The photochemical and photophysical properties of these porphyrin species were investigated. Results show that the complex containing nitrite is able to produce NO by homolytic O–NO cleavage ($\Phi_{NO}^{PPh} = 0.05$) while the $\{TPyP[Ru(NO)(5,5'\text{-Mebipy})_2]_4\}(PF_6)_{12}$ does it by direct labilization ($\Phi_{NO}^{PPh} = 0.53$) of the Ru–NO bond. Furthermore, a triplet quantum yield of 0.09 and 0.27 was observed for complexes containing nitrite and nitric oxide, respectively. The reactive oxygen species quantum yield for the complex $\{TPyP[Ru(NO)(5,5'\text{-Mebipy})_2]_4\}(PF_6)_{12}$ (0.78) is consistent with the sum of quantum yields NO release (0.53) and triplet state (0.27), which suggests that both processes participate in the formation of the reactive species. Our results show that combining these characteristics, NO production and triplet states, on the same platform could induce a synergic effect, leading to a considerable improvement in the photodynamic action of these complexes.

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1. Introduction

Photochemotherapy is a clinical treatment method, whereby cancer cells can be destroyed combining light and a photosensitizer drug (PS). Photodynamic Therapy (PDT) is one of the most known techniques that combine visible or near-infrared light, a PS and a suitable amount of molecular oxygen [1–3]. Nowadays, photodynamic action is accepted as one of the various methods

available to treat different kinds of cancer and other clinical applications [4–9].

Currently, many studies are being carried out aiming to develop more efficient PS molecules [10–13], as well as to discover more efficient photodynamic mechanisms [13–16]. Among the various strategies reported in the literature, ruthenium complexes have received a great deal of attention as potential PDT agents [17–22].

Typically, photodynamic action can be described as two distinct mechanisms: Type I mechanism, which is based on the production of free radicals, such as $\cdot OH$, $HO_2\cdot$, $O_2\cdot$ and type II mechanism, involving energy transfer to the molecular oxygen. Both processes can induce damage to membranes, DNA and other biomolecules, which can lead to cellular death by necrosis or apoptosis [23,24]. A third mechanism has been reported for ruthenium-NO complexes [25,26], which involves the photorelease of the NO radical, which

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can also be important for hypoxic regions. Combining these mechanisms on the same platform, which could be able to induce a synergic effect resulting in a considerable improvement of photodynamic action, is strongly desirable.

Furthermore, nitric oxide (NO) has been described as an anti-tumor agent [27,28], which, combined with reactive oxygen species, (ROS) produces a synergistic photodynamic action between ROS and reactive nitrogen species (RNOS) [25,26,29,30]. It was shown that using UV–vis irradiation on a NO-compound leads to tumor destruction [29]. Considering the role of NO in cancer therapy and the high triplet quantum yield of porphyrins and phthalocyanines, we decided, rationally, to design supramolecular ruthenium porphyrin complexes that could have these desirable properties. Considering this, nitrosyl ruthenium complexes were synthesized, which were able to photo-release NO and also produce triplet states [25,26,29,30]. Thus, in this work, the synthesis and characterization of two new supramolecular ruthenium porphyrin complexes, {TPyP[Ru(NO)(5,5'-Mebipy)₂]₄} (PF₆)₁₂ and {TPyP[Ru(NO₂)(5,5'-Mebipy)₂]₄} (PF₆)₄, are presented, and the purpose is to study their photodynamic properties.

2. Materials and methods

2.1. Chemicals for the synthesis

Solvents were purified using standard methods. All chemicals used were of reagent grade or comparable purity. RuCl₃·3H₂O, 5,5'-dimethyl-2,2'-bipyridine (5,5'-Mebipy), 5,10,15,20-Tetra(4-pyridyl)porphyrin (TPyP), sodium nitrite and hexafluorophosphoric acid were used as received from Aldrich.

2.2. Synthesis

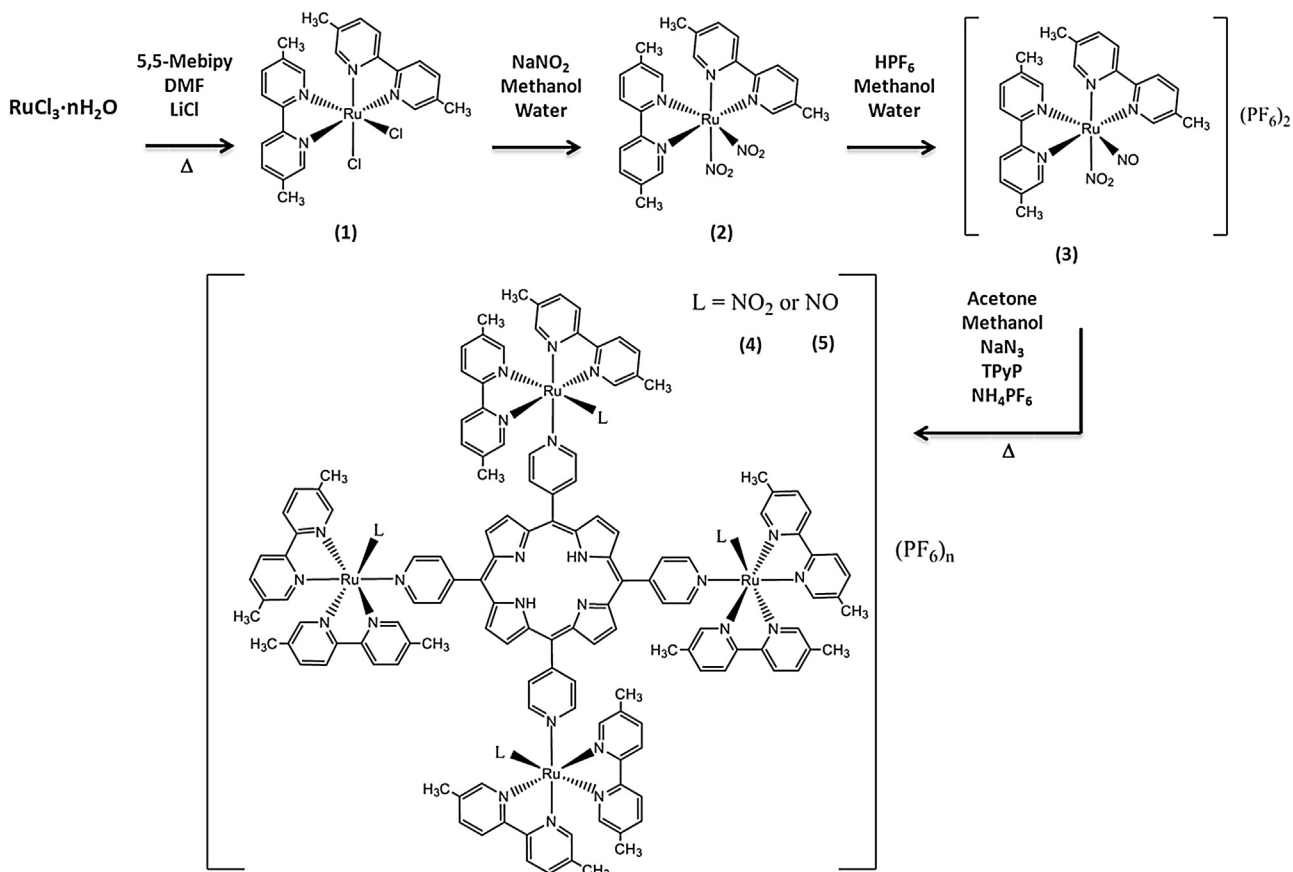
cis-[RuCl₂(5,5'-Mebipy)₂] (**1**), *cis*-[Ru(NO₂)₂(5,5'-Mebipy)₂] (**2**), and *cis*-[Ru(NO)(NO₂)(5,5'-Mebipy)₂](PF₆)₂ (**3**) were developed as precursors to coordinate the porphyrin by the peripheral pyridine rings, leading to the formation of TPyP{[Ru(NO₂)(5,5'-Mebipy)]₄}(PF₆)₄ (**4**) and TPyP{[Ru(NO)(5,5'-Mebipy)]₄}(PF₆)₁₂ (**5**) species. Complexes (**1–3**) are analogues described in the literature for 2,2-bipyridine [31,32]. The synthetic route for these syntheses is illustrated in Scheme 1.



In a Schlenk flask with 10 mL of degassed dimethylformamide, 1.0 g (3.8 mmol) of RuCl₃·3H₂O, 1.2 g (6.5 mmol) of 5,5'-Mebipy and 1.1 g (25.9 mmol) of lithium chloride were added, as described in the literature [31]. The reaction was stirred and heated for 8 h at 130 °C. At the end of the reaction time, it was cooled down and 50 mL of cold acetone were added. The flask was left in the fridge for 1 h. The solution was filtered and the dark solid was washed in ice water and ether. Yield 91.4%. Elemental analysis (%) calc. for C₂₄H₂₈Cl₂N₄O₂Ru: C 50.60, H 4.90, N 9.72. Found: C 50.86, H 4.70 and N 10.07. Conductivity (dichloromethane): 1.44 ohm⁻¹ cm² mol⁻¹, T = 298 K (neutral electrolyte).

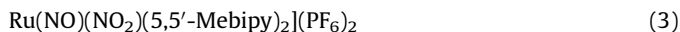


In a Schlenk flask, 0.30 g (0.5 mmol) of *cis*-[RuCl₂(5,5'-Mebipy)₂] was suspended in 50 mL of water, boiled for 15 min and stirred. The deep red solution was cooled down to room temperature and filtered and 0.90 g (7.8 mmol) of sodium nitrite was added. The solution was then refluxed for 90 min. The flask was cooled for 1 h, then filtered and washed with water and ether (3 × 5 mL) [32].

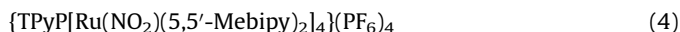


Scheme 1. Representation of the route for the synthesis of complexes (1)–(5).

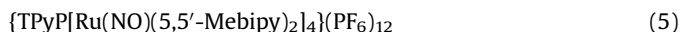
Yield 94.0%. Elemental analysis (%) calc. for $C_{24}H_{24}N_6O_4Ru$: C 51.33, H 4.31, N 14.97. Found: C 50.95, H 4.49 and N 15.39. Conductivity (dichloromethane): $1.80 \text{ ohm}^{-1} \text{ cm}^2 \text{ mol}^{-1}$, $T = 298 \text{ K}$ (neutral electrolyte).



A sample of 0.20 g (0.4 mmol) of *cis*-[Ru(NO)₂(5,5'-Mebipy)₂] was suspended in 10 mL of degassed methanol and was stirred. Then 2 mL of concentrated HPF₆ were added dropwise. After 15 min, the orange solid was converted into a yellow solid, which was filtered off and washed in methanol and ether (3 × 5 mL) [32]. Yield 94.0%. Elemental analysis (%) calc. for $C_{24}H_{24}F_{12}N_6O_3P_2Ru$: C 34.50, H 2.90, N 10.06. Found: C 34.38, H 2.87 and N 9.95. Conductivity (methanol): $165.7 \text{ ohm}^{-1} \text{ cm}^2 \text{ mol}^{-1}$, $T = 298 \text{ K}$ (electrolyte 2:1).



In a Schlenk flask, 0.30 g (0.3 mmol) of *cis*-[Ru(NO)(NO₂)(5,5'-Mebipy)₂](PF₆)₂ were suspended in 20 mL of acetone and stirred vigorously. An equimolar amount of sodium azide (NaN₃) (0.023 g, 0.36 mmol) was dissolved in 5.0 mL of methanol and was added slowly, dropwise to the above solution. After 10 min, 0.05 g, (0.08 mmol) of TPyP, previously dissolved in 20 mL of chloroform, was added and the flask was wrapped in aluminum foil and heated (60 °C). After 24 h of reaction, its volume was reduced to approximately 1 mL and NH₄PF₆ (0.5 g, 3.0 mmol), solubilized in 1 mL of methanol, was added. Then ethyl ether was added to precipitate a brown solid. The flask remained in the refrigerator for about 1 h. The formed solid was filtered off and washed several times in water to remove remaining NH₄PF₆ and with ether (3 × 5 mL). Yield: 90.0%. Elemental analysis (%) calc. for $C_{136}H_{122}F_{24}N_{28}O_8P_4Ru_4$: C 49.99, H 3.98 and N 12.00. Found: C 49.08, H 3.65, N 12.12.



A mass of {TPyP[Ru(NO₂)(5,5'-Mebipy)₂]₄}(PF₆)₄ (0.2 g, 0.06 mmol) was dissolved in 20 mL of acetonitrile. 1 mL of HPF₆, diluted in 5 mL of methanol was added to this solution and stirred for 2 h. After that, the volume of the solution was reduced to about 2 mL and 20 mL of cold water was added, leading to a formation of a light brown precipitate that was collected by filtration and washed several times in cold water and ethyl ether. Yield: 90.0%. Elemental analysis (%) calc. for $C_{136}H_{122}F_{72}N_{28}O_4P_{12}Ru_4$: C 37.10, H 2.82 and N 9.00. Found: C 36.87, H 2.67, N 9.12.

2.3. Instrumentation

Elemental analyses were performed in a Fisons EA 1108 model (Thermo Scientific, Waltham, Massachusetts). The FTIR spectra of the complexes were recorded using CsI pellets in the 4000–200 cm^{-1} region in a FT MB-102 instrument (Bomen–Michelson). The UV–vis spectra of the complex was recorded in CH₂Cl₂ for complex (1), in acetonitrile for complexes (2)–(3) and DMSO for complexes (4) and (5) in a Hewlett Packard diode array – 8452A. Cyclic voltammetry (CV) measurements of the complexes were performed in an electrochemical analyzer BAS model 100 B (Bioanalytical Systems, West Lafayette, Indiana). These experiments were carried out at room temperature in previously degassed CH₂Cl₂ for complex (1), acetonitrile for complexes (2)–(3) and DMF for complex (4)–(5), containing 0.1 mol L^{-1} Bu₄N⁺ClO₄[−] (PTBA) (FlukaPurum, St. Louis, MO) as a supporting electrolyte. A one-compartment cell was used with both working and auxiliary electrodes, which were stationary Pt foils, while the reference electrode was Ag/AgCl, 0.1 mol L^{-1} PTBA. Under such conditions, the ferrocene is oxidized at 0.43 V (Fc⁺/Fc). All the NMR

spectra were recorded at 298 K and measured using a 9.4 T Bruker Avance III spectrometer. The molar conductivity measurements (Λ_m) were taken in acetone, dichloromethane and methanol at 298 K using concentrations of $1.0 \times 10^{-3} \text{ M}$ of the complexes.

2.4. X-ray crystallography

Orange crystals of *cis*-[Ru(NO)₂(5,5'-Mebipy)₂] (2) and *cis*-[Ru(NO)(NO₂)(5,5'-Mebipy)₂](PF₆)₂ (3) were obtained by slow evaporation of a methanol/hexane solution (2:1) at 298 K. The data collection was performed using Mo-K α radiation ($\lambda = 0.71073 \text{ \AA}$) on an Enraf–Nonius Kappa-CCD diffractometer at 293 K. The final unit cell parameters were based on all reflections. Data collections were performed using the COLLECT program [33]. Data reduction was carried out using the Denzo-SMN and Scalepack software [34]. The structures were solved by the direct method using SHELXS-97 and refined using the software SHELXL-97 [35].

The hydrogen atoms were calculated at idealized positions using the riding model of SHELXL97 [35]. The Gaussian method was used for the absorption corrections [36]. The projection views of the structures were prepared using ORTEP-3 for Windows [37]. Hydrogen atoms were stereochemically positioned and refined with the riding model.

2.5. Photophysical characterization

The UV–vis spectra were acquired using a Beckman DU640 spectrophotometer and the fluorescence spectra were obtained with a Fluorolog-3 spectrofluorometer – Horiba/Jobin-Yvon Inc. The concentrations were monitored spectrophotometrically and all photophysical measurements were performed at room temperature. The fluorescence quantum yield (Φ_f) was obtained by comparing it with an emission standard of a known compound, as described in reference [22], which was *meso*-tetrakis(4-N-methylpyridiniumyl) porphyrin (TMPyP) in an aqueous solution at pH 6.8 ($\Phi_{f0} = 0.05$) [38]. The Φ_f values were calculated according to Eq. (1).

$$\phi_f = \phi_{f0} \frac{F_f A_0 n^2}{F_{f0} A n_0^2} \quad (1)$$

where Φ_f and Φ_{f0} are the quantum yields of the investigated compound and reference, respectively. A and A_0 are the absorbance values at the excitation wavelength of the compound and reference solutions and F_f and F_{f0} are the integrated fluorescence intensities of the compound and reference samples. The sample and the standard were both excited at the same relevant wavelength. n 's are the refractive indexes of the solvents containing the compound (n) and the reference (n_0).

The triplet quantum yields and transient absorptions were obtained through transient absorption experiments. Briefly, samples were excited by a Q-switched, pulsed Nd:YAG laser (Quantel U.S.A. (BigSky) Brilliant B; 5–6 ns full width at half-maximum (fwhm), 1 Hz, ~10 mm in diameter) tuned to 532 nm by using the appropriate second harmonic generator. A 150 W xenon arc lamp served as a probe beam and was aligned orthogonally to the laser excitation light. Detection was achieved using a monochromator (Spex 1702/04) optically coupled to a photomultiplier tube (R928, Hamamatsu). More details can be found in [11]. The quantum yields of the formation of the triplet excited state, Φ_{isc} , were calculated using the partial saturation method, Eq. (2), [39,40].

$$\Delta A(\lambda, I) = a(1 - e^{bt}) \quad (2)$$

where $a = [\varepsilon_T(\lambda) - \varepsilon_g(\lambda)]C_0L$ and $b = 2.3\Phi_{isc}\varepsilon_g(\lambda_{exc})$. The parameters ε_T and ε_g are the triplet excited state and the ground state extinction coefficients ($M^{-1}cm^{-1}$) at the wavelength of analysis while $\varepsilon_g(\lambda_{exc})$ is the ground state extinction coefficient at the excitation wavelength (532 nm), C_0 is the sample concentration ($mol\ L^{-1}$), L is the optical path of the sample (cm) and I is the intensity of excitation ($einstein\ cm^{-2}$). The absorbance changes, $\Delta A(\lambda, I)$, were acquired at 420 nm and 470 nm for different laser intensities from transient absorption experiments. The probed wavelengths correspond to the ground state bleaching of the singlet and the triplet excited states, respectively.

2.6. NO photorelease detection

Chemiluminescent NO detection was used to confirm the release of free NO during the photolysis of (4) and (5) porphyrins in dimethylsulfoxide solutions (DMSO, HPLC grade Sigma-Aldrich). Measurements were carried out in a GE Sievers NOA 280i apparatus, as described in the literature [41]. A xenon lamp XBO 75W/2 OFR – OSRAM irradiation induced NO photorelease of porphyrin solutions (3.0 mL). The irradiance at 320–850 nm was measured by Spectra-Physics 407A radiometer at 0° of incidence angle and the value used was $870 \pm 70\ mW\ cm^{-2}$. Optical absorption spectra, before and after irradiation, were monitored by a spectrophotometer. All experiments were performed in a dark room at a temperature of $295.0 \pm 1.0\ K$.

NO release during porphyrin photolysis was measured at $t = 1,000\ s$ for each sample. The yield reaction of NO release was calculated by Eq. (3), where M_{NO} and M_{PPh} are the masses of the formed NO and consumed porphyrin, respectively.

$$\Phi_{NO}^{PPh} = \frac{M_{NO}}{M_{PPh}} \quad (3)$$

2.7. Photochemical reaction

The displacement of NO from complex (5) was carried out by irradiation, at 395 nm, of a solution containing $\{TPyP[Ru(NO)(5,5'-Mebipy)]_4\}(PF_6)_{12}$ (0.02 g/2 mL of DMSO) and $[RuCl_3(dppb)(H_2O)]$ (0.015 g/2 mL CH_2Cl_2), which was used as a trap to capture the NO released from the porphyrin. The same experiment was performed in the absence of light. The photoreaction experiment was followed by the $^{31}P\{^1H\}$ NMR technique.

2.8. Determination of quantum yields for reactive oxygen species (RNOS)

Quantum yields for reactive oxygen species (Φ_{Δ}) were determined using the relative method of a chemical quencher of RNOS 1,3-diphenylisobenzofuran (DPBF) with zinc phthalocyanine as a reference [11,42]. The DPBF was used as an efficient quencher for oxygen singlet [11] and the radical ion as the anion radical superoxide [42].

Porphyrin solutions containing the quencher were prepared in the dark and irradiated in the Q-band region using a 661 nm laser. The absorbance value of solution was adjusted to ~ 0.2 at the irradiation wavelength. The disappearance of DPBF was monitored using a UV–vis spectrophotometer by absorption decays of DPBF at 417 nm. The quantum yields of ROS (Φ_{Δ}) were determined using:

$$\Phi_{\Delta} = \Phi_{\Delta}^0 \frac{R I_{abs}^0}{R^0 I_{abs}} \quad (4)$$

where R and R^0 are the rates of consumption of the DPBF in the presence of the compound under investigation and the reference, respectively. I_{abs} and I_{abs}^0 are the rates of light absorption by the sensitizer under investigation and the reference, respectively. Φ_{Δ}^0 is the quantum yield of the reference for ROS formation. In DMSO, the Φ_{Δ}^0 value for zinc phthalocyanine is 0.67 [11].

3. Results and discussion

Complexes (1–5) were obtained in good yields utilizing mild conditions. Elemental analysis and molar conductivity suggest the structures and purity of the complexes. The 1H NMR data for complexes (1) to (5) are summarized to support the information (see Figs. S1–S4 and Table S1). All compounds exhibited well-resolved characteristic 1H NMR peaks. Aromatic hydrogens of 5,5-Mebipy and pyrrolic and pyridyl protons of TPyP were found in the range of 7.0–10.0 ppm, giving an integrated signal corresponding to 78 hydrogens and singlets relative to methyl groups, ranging from 2.0 to 2.9 ppm, making a total of 48 hydrogens that are consistent with the expected structures for complexes (4) and (5). Singlets observed for the methyl group were confirmed by the COSY 1H NMR experiment. A singlet at -2.9 ppm (2H) was assigned to the inner ring protons from the TPyP. The presence of the counter-ion PF_6^- with a chemical shift at $\delta -144$ ppm was confirmed by $^{31}P\{^1H\}$ NMR spectra.

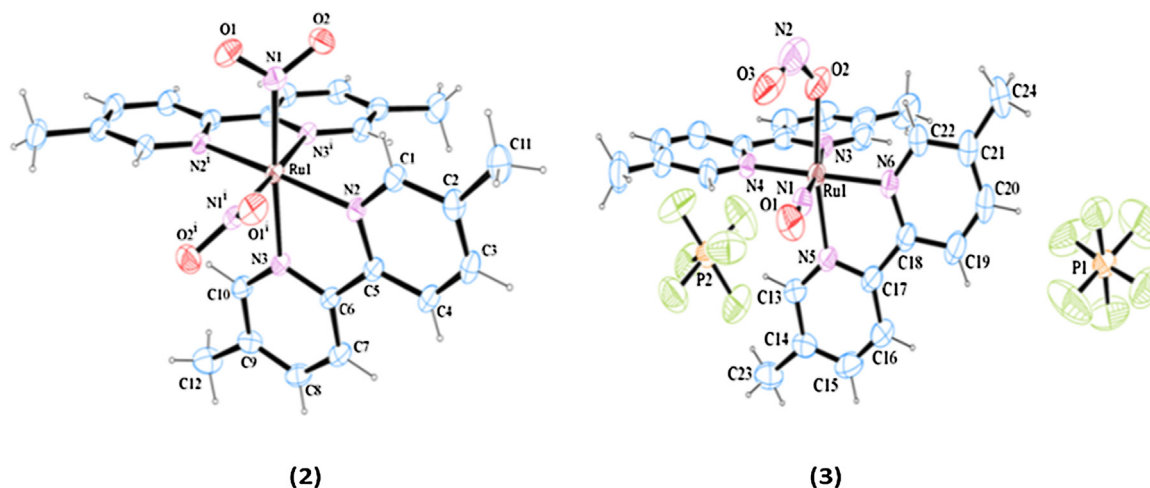


Fig. 1. ORTEP structures of *cis*- $[Ru(NO_2)_2(5,5'-Mebipy)_2] \cdot 3.5H_2O$ (2) and *cis*- $[Ru(NO)(NO_2)(5,5'-Mebipy)_2](PF_6)_2$ (3). Thermal ellipsoids with 30% probability. For (2): space group, C2/c; $a = 11.2860(9)\ \text{\AA}$, $b = 17.3225(11)\ \text{\AA}$, $c = 15.3621(12)\ \text{\AA}$, $\beta = 111.104(5)^\circ$. For (3): $P2_1/n$, $a = 9.0970(2)\ \text{\AA}$, $b = 30.4250(6)\ \text{\AA}$, $c = 11.5830(3)\ \text{\AA}$, $\beta = 98.5850(10)^\circ$.

The main IR bands for complexes (1) to (5) are summarized in Figs. S5–S9. IR measurements show that nitrosyl complexes (3) and (5) presented strong bands ranging from 1940 to 1945 cm^{-1} , which are attributed to the νNO^+ stretching [43]. Complexes (2) and (4) presented a NO_2^- group coordinated to the metal center by the oxygen, agreeing with a nitro complex with ν_{as} and ν_{s} at 820 cm^{-1} and 824 cm^{-1} , respectively. At approximately 1331/1290 cm^{-1} and 1328/1281 cm^{-1} , stretching of $\nu_{\text{as}}\text{NO}_2^-$ and $\nu_{\text{s}}\text{NO}_2^-$ for complexes (2) and (4) was also observed, which is consistent with nitro complexes [44]. On the other hand, complex (3) presented a band at 1945 cm^{-1} assigned to νNO^+ stretching and ν_{as} and ν_{s} , from NO_2^- were observed at 1453 cm^{-1} and 1061 cm^{-1} , respectively. These results are consistent with nitrite complexes and in both cases, the coordination mode was confirmed by crystallographic data (Table S2).

In the IR spectra of complex (5), ν_{s} and ν_{as} (C–H) aromatic stretching was observed in the region of 3500 cm^{-1} . Both bands, C=N and C=C, of the pyridine rings and CH_3 of 5,5'-group were observed ranging from 1630 to 1400 cm^{-1} [45]. At 827 cm^{-1} and 547 cm^{-1} , an intense band from ν_{as} and ν_{s} (P–F) was observed and attributed to the counter-ion. The low intensity band at 547 cm^{-1} was assigned to the $\nu\text{Ru-N}$ stretching [45].

The electrochemical behavior for the compounds was evaluated by cyclic voltammetry and differential pulse voltammetry experiments (Fig. S10). Complex (1) showed a reversible process attributed to the oxidation of $\text{Ru}^{\text{II}}/\text{Ru}^{\text{III}}$ where $\text{ipa}/\text{ipc}=1.0$ and $E_{1/2}$ of 291 mV, reinforced by differential pulse voltammetry. Complexes (2), (4) and (5) have the presence of a Ru-NO^+ fragment in common. Thus, the electrochemistry of these compounds is based on the nitrosyl group (1A: $\text{NO}^+ \rightarrow \text{NO}^0$, 1B: $\text{NO}^0 \rightarrow \text{NO}^-$, 2A: $\text{NO}^- \rightarrow \text{NO}^0$ and 2B: $\text{NO}^0 \rightarrow \text{NO}^+$).

Cyclic voltammogram of complex (3) showed a quasi-reversible process ($\text{ipa}/\text{ipc} = 1.11$) with E_{pa} and E_{pc} values of 821 and 736 mV, respectively. This can be attributed to the redox pair of $\text{Ru}^{\text{II}}\text{-NO}_2$ to $\text{Ru}^{\text{III}}\text{-NO}_2$ [46]. After oxidation the $[\text{Ru}^{\text{III}}\text{-(NO}_2)_2(5,5'\text{-Mebipy)}_2]^{2+}$ species has a limited stability due to the easy intermolecular disproportionation reaction, which is responsible for the formation of the nitrite and nitrosyl complex, as previously reported for the $\text{cis-}[\text{Ru}(\text{NO}_2)_2(\text{bipy})_2]$ compound [46].

3.1. X ray diffraction

Single crystals suitable for X-ray diffraction of complexes (2) and (3) were obtained by slow evaporation of methanol/hexane (4:1) solutions at room temperature. Complexes (2) and (3) crystallized into a monoclinic crystalline system and space group $C2/c$ and $P2_1/n$, respectively (Table S2). In the structure of complex (2) (Fig. 1), H_2O molecules were omitted for the sake of clarity.

Complexes (2) and (3) exhibit the expected octahedral coordination geometries: the two bidentate 5,5'-Mebipy ligands are situated in position *cis* to each other and the ligand NO_2^- in complex (2) is in the *cis* position to NO_2^- , and in complex (3), NO_2 is also situated *cis* to NO . The nitrite ion is coordinated by the oxygen atom (O2) with O2-N2-O3 bond angle of 116.7 (8) $^\circ$. The bond distances O1-N1, N2-O2, and N2-O3 (1.130 (5) 1.244 (8) and 1.147 (8) \AA) are within the range reported for other nitrite complexes [44] (Table S3).

The Ru-N3 distance in complex (3), which is *trans* to Ru-N1(NO) is 2.087 (4) \AA , while Ru-N5 (5,5'-Mebipy) *trans* to ONO is practically the same, 2.090 (4) \AA . The bond distances Ru-N1(NO) and Ru-O2 (ONO) are 1.758 (4) and 2.067 (4), respectively. The nitrosyl structure is practically in a linear coordination mode and the structural parameters, typical for Ru-NO complexes [43,44], are consistent with the IR data.

Complex (2) presents a symmetrical structure with space group $C2/c$ and consequently the distances of the atoms are identical, as

can be observed in Ru(1)-N(1) and Ru(1)-N(1)ⁱ 2.033(2) \AA and Ru(1)-N(2) and Ru(1)-N(2)ⁱ 2.078(2) \AA . The bond distances, O1-N1 and N2-O2, 1.231 (3) and 1.268 (3) \AA respectively, and O1-N1-O2 angle 117.2 (2), are in agreement with the literature on nitro complexes [45,46].

3.2. UV-vis absorbance and fluorescence emission

The electronic spectra for complex (1–3) are summarized in the supplementary material (Table S4). The electronic (black curve) and fluorescence emission (gray curve) spectra for $\text{TPyP}\{[\text{Ru}(\text{NO}_2)(5,5'\text{-Mebipy})]_4\}(\text{PF}_6)_4$ (4) and $\text{TPyP}\{[\text{Ru}(\text{NO})(5,5'\text{-Mebipy})]_4\}(\text{PF}_6)_{12}$ (5), in DMSO are presented in Fig. 2a and b, respectively. The band around 300 nm can be attributed to the $\pi \rightarrow \pi^*$ intraligand transition of unsaturation in 5,5'-Mebipy ligands, while the band at 420 nm can be assigned as the B-band of porphyrins and the bands ranging from 515 to 647 nm are the Q-bands. The molar absorption coefficients were obtained in different concentrations from 1 to 100 μM for the maximum of the main peaks, in Table 1.

The fluorescence spectra of these samples show the well-known dual emission of free base porphyrins. The fluorescence quantum yield was obtained and the results show that for complex (4) it is 0.002, while for complex (5) it is 0.003.

3.3. Triplet state formation

Nanosecond transient absorption experiments for a great variety of porphyrins have been extensively reported in the

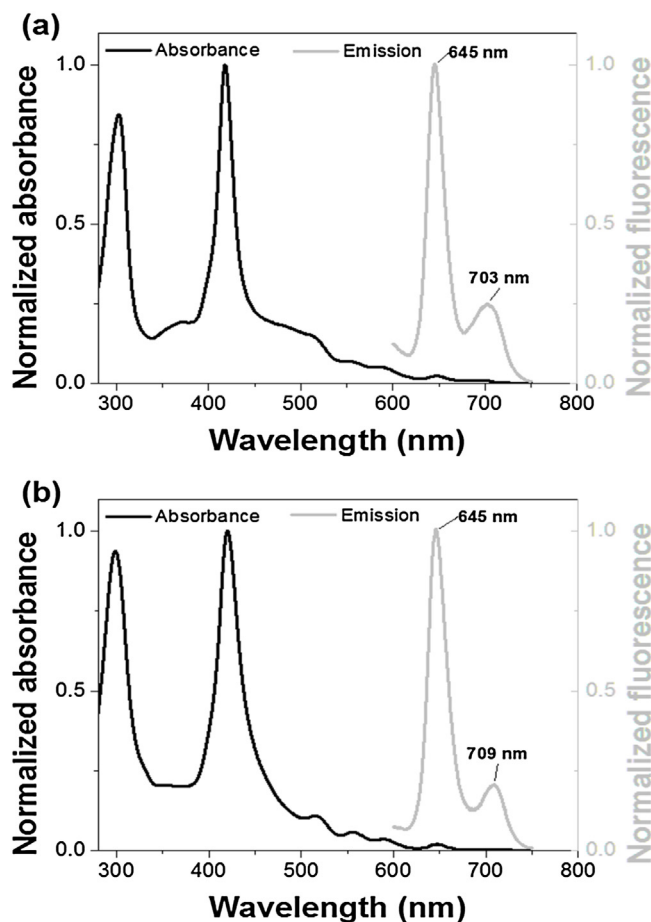


Fig. 2. Absorption spectrum in the UV/Vis region (black curve) and fluorescence emission spectrum (gray curve): (a) complex (4) and (b) complex (5) in DMSO.

Table 1

UV/Vis absorption properties obtained at room temperature for samples containing porphyrin complexes (4) and (5). The values of the extinction coefficient (ϵ) are in units of $10^5 \text{ M}^{-1} \text{ cm}^{-1}$, \pm standard deviation. The wavelengths, in nm, for the center of the observed bands are presented between parentheses.

Complex	Intraligand Band	B-Band	$Q_y(1,0)$	$Q_y(0,0)$	$Q_x(1,0)$	$Q_x(0,0)$
(4)	2.38 ± 0.05 (300)	2.96 ± 0.09 (419)	0.47 ± 0.02 (514)	0.23 ± 0.01 (545)	0.16 ± 0.04 (590)	0.07 ± 0.01 (647)
(5)	2.15 ± 0.03 (300)	2.17 ± 0.05 (420)	0.21 ± 0.05 (515)	0.11 ± 0.01 (555)	0.31 ± 0.01 (590)	0.03 ± 0.01 (647)

literature regarding the formation and dynamics of triplet states [47–52]. The transient absorption spectrum of a free base tetrapyrrolyl porphyrin shows characteristic ground state bleaching between 400 nm and 430 nm and a growth from 430 nm to 500 nm [52]. In transient absorption, a positive signal represents an increase in absorption, and generally for porphyrins, this is assigned to the triplet excited state absorption, $T_1 \rightarrow T_n$.

Transient absorption of porphyrin complexes (4) and (5) depicted very distinct triplet excited state behavior although they differ slightly in their molecular structure (see Fig. 3). Both compounds exhibit tri-exponential kinetics for the triplet excited state relaxation, as shown in Table 2. The origin of each time constant will not be further discussed since it is not the main aim of the current research. The most relevant aspect here is to show that both compounds have a long lived triplet excited state and their calculated triplet quantum yield is 0.09 and 0.27 for complexes (4) and (5), respectively.

3.4. NO photorelease from nitrosyl ruthenium porphyrins complexes

The nitrosyl ruthenium porphyrin complexes, when dissolved in DMSO are stable in the dark, but upon irradiation with visible light, the solution of (4) and (5) release nitric oxide (NO) as demonstrated by spectral modifications (Fig. 4) and NO^\bullet chemiluminescent signal (Fig. 5).

The decrease in the absorption of the band at 420 nm (Fig. 4) can be attributed to the consumption of the initial complex, while the band centered at 300 nm is associated to the intraligand $\pi \rightarrow \pi^*$ transitions in 5,5'-Mebipy ligands [48]. The porphyrin transition bands in the region 330–550 nm in the UV-vis spectra are sufficiently intense to mask the metal ligand charge transfer

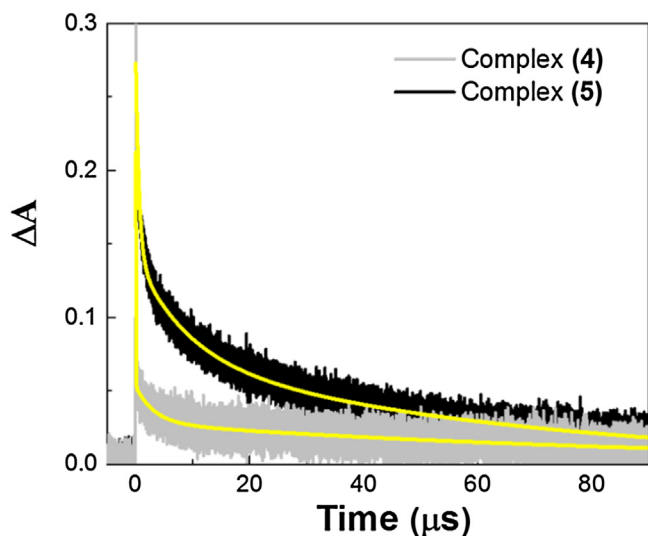


Fig. 3. Absorption changes monitored at 460 nm after pulsed light excitation ($\lambda_{\text{exc}} = 532 \text{ nm}$ and 1 mJ/pulse) of complex (4), gray curve and complex (5), black curve, in DMSO solution. The yellow line corresponds to a tri-exponential function. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Table 2

Depopulation time constants and formation quantum yield of triplet state for {TPyP[Ru(NO₂)(5,5'-Mebipy)₄](PF₆)₄} (4) and (b) {TPyP[Ru(NO)(5,5'-Mebipy)₄](PF₆)₁₂} (5) in DMSO.

Compound	τ_1^a	τ_2^a	τ_3^a	Φ_T
(4)	2.8×10^{-8} (94%)	3.2×10^{-6} (3%)	9.5×10^{-5} (3%)	0.09
(5)	5.8×10^{-7} (50%)	8.9×10^{-6} (24%)	6.4×10^{-5} (26%)	0.27

^a The numbers between parenthesis represent the contribution of the time constant to the overall kinetics.

(MLCT) involving $d_\pi(\text{Ru}^{\text{II}}) \rightarrow \pi^*(5,5'\text{-Mebipy})$ and $d_\pi(\text{Ru}^{\text{II}}) \rightarrow \pi^*(\text{NO}^+)$ in the case of complex (5) or $d_\pi(\text{Ru}^{\text{III}}) \rightarrow \pi^*(5,5'\text{-Mebipy})$ and $d_\pi(\text{Ru}^{\text{III}}) \rightarrow \pi^*(\text{ONO}^-)$ for complex (4) (Figs. S11–S13).

Thus, the spectral changes are associated with nitrosyl ligand reduction ($\text{NO}^+ \rightarrow \text{NO}^0$) in the case of complex (5) and {TPyP[Ru^{III}(solvent)(5,5'-Mebipy)₄](PF₆)₁₂} formation, as suggested by controlled reduction potential (electrolysis), performed in CH_3CN ,

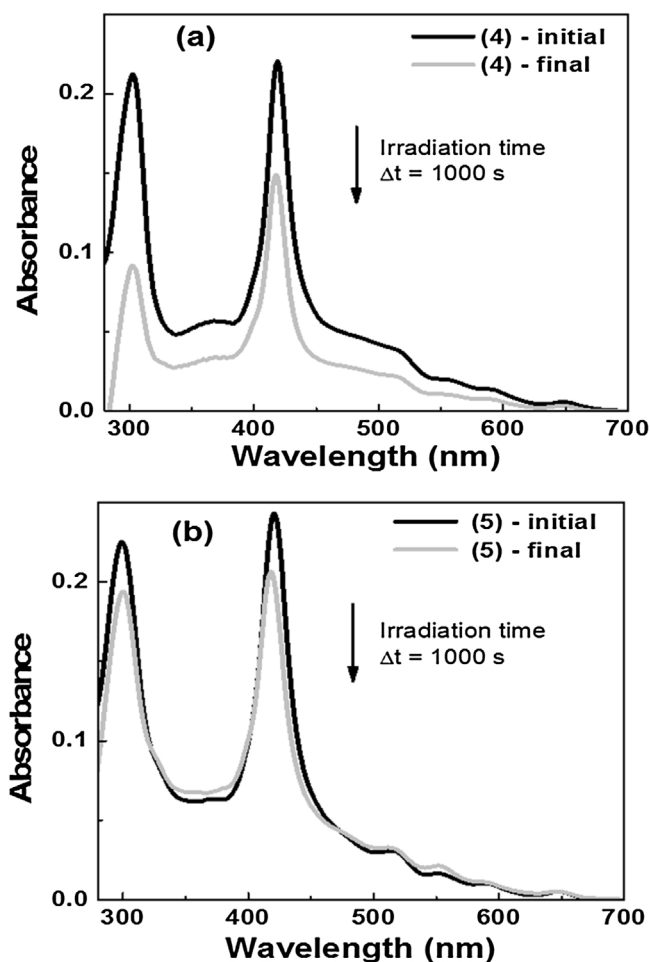


Fig. 4. Absorption spectra modifications for porphyrin solutions upon irradiation with visible light during $\Delta t = 1000 \text{ s}$. (a) {TPyP[Ru(NO₂)(5,5'-Mebipy)₄](PF₆)₄} (4) and (b) {TPyP[Ru(NO)(5,5'-Mebipy)₄](PF₆)₁₂} (5).

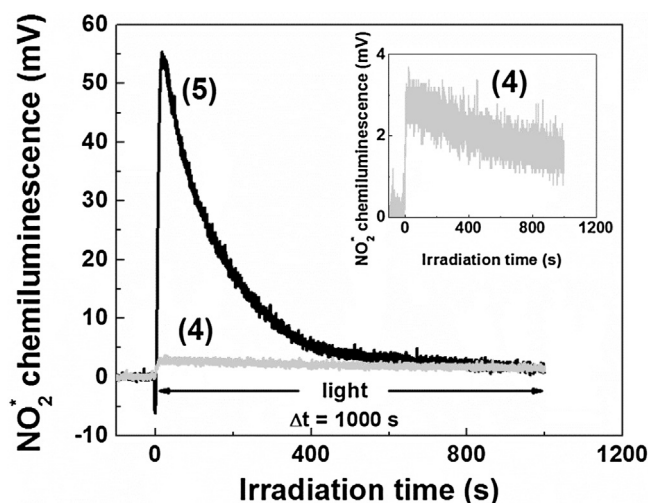


Fig. 5. Chemiluminescent signal from photolysis of (4) and (5) compounds irradiated with visible light during $\Delta t = 1000$ s. The inset shows the rescaled (4) curve.

0.1 M PTBA (Fig. S10e). The changes in the complex (4) spectrum were associated with nitrite ligand oxidation ($\text{ONO}^- \rightarrow \text{NO}^0$) and subsequent formation of metal–oxo species. In this case, there was no metal–solvent coordination since the ruthenium of complex (4) has an octahedral structure ($\text{TPyP}[\text{Ru}^{\text{IV}}=\text{O}]$) (Fig. S10d). NO photolabilization from the coordinated ruthenium complex has been demonstrated by a direct labilization of the Ru–NO bond, in our case complex (5), and by homolytic O–NO cleavage resulting in a metal–oxo species, in this study as complex (4) [53–55].

The detection of NO was directly registered by the chemiluminescence emitted from the NO_2 excited state (NO_2^*). The amount of NO released (M_{NO} , in moles) from the porphyrin complexes during photolysis was calculated taking the ratio of the area under the decay curve (Fig. 5) by the proportionality constant $B = 7.9 \pm 0.2 \times 10^{12} \text{ mV s mol}^{-1}$. Constant B was obtained from the calibration curve as previously reported [41] (Fig. S14). Additionally, the amount of porphyrin consumed (M_{PPh} , in moles) is directly determined from the optical absorption spectra at the Soret band, acquired immediately before and after photolysis (see Fig. 5), according to Eq. (5).

$$M_{\text{PPh}} = \frac{(A_{\text{initial}} - A_{\text{final}}) \cdot \text{volume}}{\varepsilon_{\text{Soret-band}}} \quad (5)$$

Compared to the initial amount of porphyrins in the reaction flask, the amount of NO released represents a reaction yield of 0.53 ± 0.03 mol of NO per mol of (5) and 0.05 ± 0.01 mol of NO per mol of (4). These results are summarized in Table 3.

The NO quantum yield is highly dependent on the structure of nitrosyl porphyrin ruthenium complexes, $\text{TPyP}[\text{Ru}^{\text{II}}-\text{N}=\text{O}]$ or $\text{TPyP}[\text{Ru}^{\text{III}}-\text{O}-\text{N}=\text{O}]$. The lower reaction photorelease yield of free NO for complex (4) could be explained in terms of the recombination reaction between NO^+ and $\text{TPyP}[\text{Ru}^{\text{IV}}=\text{O}]$, as

Table 3

Amounts of NO released (M_{NO}) and porphyrin consumed (M_{PPh}) during the photolysis of $\{\text{TPyP}[\text{Ru}(\text{NO}_2)(5,5'\text{-Mebipy})_4](\text{PF}_6)_4$ (4) and $\{\text{TPyP}[\text{Ru}(\text{NO})(5,5'\text{-Mebipy})_4](\text{PF}_6)_{12}$ (5) porphyrins. The reaction yield of NO release ($\Phi_{\text{NO}}^{\text{PPh}}$) was determined for an irradiation time of $\Delta t = 1,000$ s.

Complexes	$M_{\text{PPh}} (\times 10^{-12} \text{ moles})$	$M_{\text{NO}} (\times 10^{-12} \text{ moles})$	$\Phi_{\text{NO}}^{\text{PPh}}$
(4)	730 ± 10	40 ± 10	0.05 ± 0.03
(5)	570 ± 10	300 ± 10	0.53 ± 0.03

suggested in Ref. [53]. It is supported by a predominantly ($> 50\%$) short triplet lifetime and lower triplet quantum yield, $\tau_1 = 2.8 \times 10^{-8} \text{ s}$ and $\Phi_T = 0.09$, respectively (Table 2). These data indicate that NO release is one of the processes involved in the dissipation of the triplet state energy, since the intersystem crossing process to the complex (4) ground state is lower.

NO photorelease from complex (5) was monitored by the photoreactivity of nitrosyl (NO^0) with $[\text{Ru}^{\text{III}}\text{Cl}_3(\text{dppb})(\text{H}_2\text{O})]$, which in turn was reduced to $[\text{Ru}^{\text{II}}(\text{NO})\text{Cl}_3(\text{dppb})]$ and is confirmed by ^{31}P { ^1H } NMR data. The $[\text{Ru}^{\text{II}}(\text{NO})\text{Cl}_3(\text{dppb})]$ $^{31}\text{P}\{^1\text{H}\}$ NMR spectrum shows a chemical shift at 14.6 ppm and 11.1 ppm, whose phosphorus atoms of dppb are *trans*-dppb-NO and *trans*-dppb-Cl, respectively (Fig. S15) [56]. On the other hand, in the absence of light, no chemical shift and consequently no reduction of Ru^{III} complex was detected, indicating an absence of NO^0 photorelease.

Additionally, another mechanism should also be involved in the NO production from similar nitrosyl ruthenium complexes. It has been reported that under air atmosphere and upon ultraviolet irradiation, photoinduced electron transfer could be present in the photoredox transformations of a nitrosyl phthalocyanine ruthenium complex, resulting in superoxide anion (O_2^-) formation [57].

This hypothesis is corroborated by the fact that the estimated Q-band energy for $\{\text{TPyP}[\text{Ru}(\text{NO})]\}$ is 2.17 eV, and the ground state $\{\text{TPyP}[\text{Ru}(\text{NO})]^+/\{\text{TPyP}[\text{Ru}(\text{NO})]^0$ reduction potential is +1.17 V. Thus, the estimated $\Delta E_{0,0}$ gives the $\{\text{TPyP}[\text{Ru}(\text{NO})]^+/\{\text{TPyP}[\text{Ru}(\text{NO})]^0$ potential as -1.00 eV for the singlet state, which implies that $\{\text{TPyP}[\text{Ru}(\text{NO})]^*$ can easily reduce molecular oxygen to superoxide anion, since molecular oxygen has a one-electron reduction potential of -0.33 eV [57,58]. This mechanism could also be present for complex (4), $\{\text{TPyP}[\text{Ru}(\text{NO}_2)(5,5'\text{-Mebipy})_4](\text{PF}_6)_4$, $\Delta E_{0,0} = -1.55$ eV. Moreover, it is well known that superoxide can react quickly with nitric oxide producing peroxyxynitrite (ONOO^-) and could also be in the photochemical reaction [59].

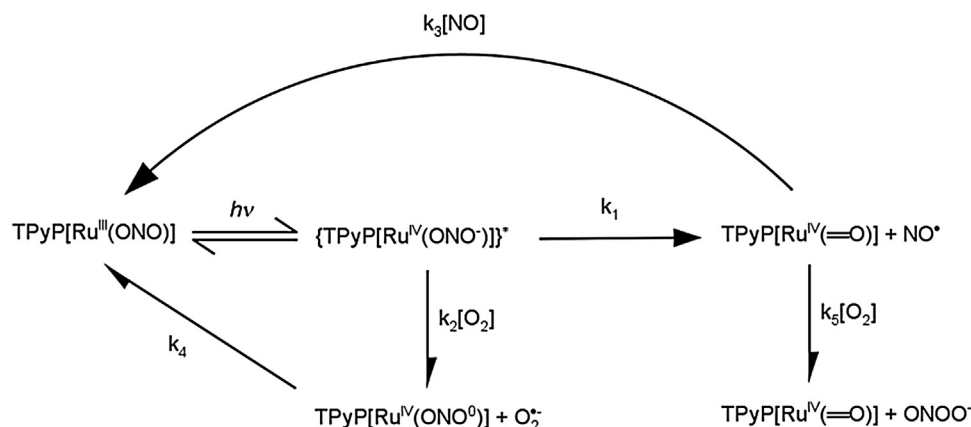
3.5. Determination of quantum yields for reactive nitrogen and oxygen species (RNOS)

RNOS quantum yield (Φ_{Δ}) values were calculated by Eq. (4) according to Fig. S16. The Φ_{Δ} values obtained for (4) and (5) were 0.07 and 0.78, respectively. The generation of the RNOS depends on the formation of the triplet state from the PS. Thus, the RNOS quantum yield is limited by a triplet quantum yield ($\Phi_{\Delta} \leq \Phi_T$). However, the Φ_{Δ} for compound (5) was 0.78 and Φ_T was 0.27, indicating that a second mechanism is involved in ROS formation. As the Φ_{Δ} for complex (5) (0.78) is consistent with the sum of quantum yields NO release (0.53) and triplet state (0.27), we believe that another mechanism involved is the photorelease of the NO radical.

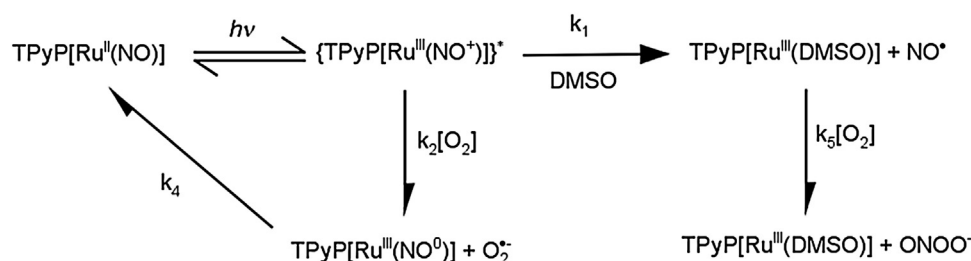
Therefore, both photodynamic processes participate in the formation of reactive oxygen species and reactive nitrogen species, contributing to the increase in photodynamic efficiency of the studied compounds. This result is in agreement with other photosensitizer–NO compounds [60]. Thus, Schemes 2 and 3 summarize the photochemical pathway for the photolysis of the investigated nitrosyl porphyrin ruthenium complexes.

4. Conclusions

In this paper, we report the synthesis and characterization of three new nitro/nitrosyl Ru-based complexes, which were used as precursors to obtain two new porphyrin ruthenium species containing nitric oxide (NO) or nitrogen dioxide (NO_2), able to generate NO. The NO photorelease, triplet state and reactive oxygen species formation by these ruthenium porphyrins were



Scheme 2. Photochemical pathway for the photolysis of {TPyP[Ru(NO₂)(5,5'-Mebipy)]₄}(PF₆)₄ (**4**) porphyrin. The direct photolysis of (**4**) produces NO• and involves a reducing process with superoxide (O₂^{•-}) anion formation and peroxyntirite (ONOO⁻). The direct photolysis can be reversible due to recombination reaction between NO• and TPyP[Ru^{IV}=O⁻].



Scheme 3. Photochemical pathway for the photolysis of {TPyP[Ru(NO)(5,5'-Mebipy)]₄}(PF₆)₁₂ (**5**) porphyrin. The direct photolysis of (**5**) is associated with nitrosyl ligand reduction (NO•→NO⁰) and {TPyP[Ru^{III}(solvent)(5,5'-Mebipy)]₄}(PF₆)₁₂ formation. The direct photolysis of (**5**) also involves a reducing process with superoxide (O₂^{•-}) anion formation and peroxyntirite (ONOO⁻).

evaluated and followed by photochemical and photophysical characterization. The results suggest that the {TPyP[Ru(NO₂)(5,5'-Mebipy)]₄}(PF₆)₄ (**complex 4**) and the {TPyP[Ru(NO)(5,5'-Mebipy)]₄}(PF₆)₁₂ (**complex 5**) porphyrins are able to produce reactive nitrogen species and reactive oxygen species on the same platform and the highest production was observed for complex (**5**). These results could improve the photodynamic action of these compounds, which could be potential drugs for cancer treatment and other diseases.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.jphotochem.2017.01.028>.

References

- [1] K. Plaetzer, B. Krammer, J. Berlanda, F. Berr, T. Kiesslich, Photophysics and photochemistry of photodynamic therapy: fundamental aspects, *Lasers Med. Sci.* 24 (2009) 259–268.
- [2] A.E. O'Connor, W.M. Gallagher, A.T. Byrne, Porphyrin and nonporphyrin photosensitizers in oncology: preclinical and clinical advances in photodynamic therapy, *Photochem. Photobiol.* 85 (2009) 1053–1074.
- [3] J.M. Dabrowski, L.G. Arnaut, Photodynamic therapy (PDT) of cancer: from local to systemic treatment, *Photochem. Photobiol.* 81 (2005) 1765–1780.
- [4] M. Kim, H.Y. Jung, H.J. Park, Topical PDT in the treatment of benign skin diseases: principles and new applications, *Int. J. Mol. Sci.* 16 (2015) 23259–23278.
- [5] D.A. Caminos, M.B. Spesia, E.N. Durantini, Photodynamic inactivation of *Escherichia coli* by novel meso-substituted porphyrins by 4-(3-N,N,N-trimethylammoniumpropoxy)phenyl and 4-(trifluoromethyl)phenyl groups, *Photochem. Photobiol. Sci.* 5 (2006) 56–65.
- [6] M.R. Detty, S.L. Gibson, S.J. Wagner, Current clinical and preclinical photosensitizers for use in photodynamic therapy, *J. Med. Chem.* 47 (2004) 3897–3915.
- [7] P. Braham, C. Herron, C. Street, R. Darveau, Antimicrobial photodynamic therapy may promote periodontal healing through multiple mechanisms, *J. Periodontol.* 80 (2009) 1790–1798.
- [8] L.M. Almeida, F.F. Zanoelo, K.P. Castro, I.E. Borissevitch, C.M.A. Soares, P.J. Gonçalves, Cell survival and altered gene expression following photodynamic inactivation of *paracoccidioides brasiliensis*, *Photochem. Photobiol.* 88 (2012) 992–1000.
- [9] G.B. Kharkwal, S.K. Sharma, Y.Y. Huang, T. Dai, M.R. Hamblin, Photodynamic therapy for infections: clinical applications, *Lasers Surg. Med.* 43 (2011) 755–767.
- [10] X. Zhou, D. Liu, T. Wang, X. Hu, J. Guo, K.C. Weerasinghe, L. Wang, W. Li, Synthesis and photophysical studies of triazine-linked porphyrin-perylene bisimide dyad with long-lived perylene triplet state, *J. Photochem. Photobiol. A* 274 (2014) 57–63.
- [11] L. Alonso, R.N. Sampaio, T.F.M. Souza, R.C. Silva, N.M. Barbosa Neto, A.O. Ribeiro, A. Alonso, P.J. Gonçalves, Photodynamic evaluation of tetracarboxyphthalocyanines in model systems, *J. Photochem. Photobiol. B* 161 (2016) 100–107.

- [12] M.P. Romero, N.R.S. Gobo, K.T. de Oliveira, Y. Iamamoto, O.A. Serra, S.R.W. Louro, Photophysical properties and photodynamic activity of a novel menthol-zinc phthalocyanine conjugate incorporated in micelles, *J. Photochem. Photobiol. A* 253 (2013) 22–29.
- [13] B. Pucelik, I. Gürol, V. Ahsen, F. Dumoulin, J.M. Dabrowski, Fluorination of phthalocyanine substituents: improved photoproperties and enhanced photodynamic efficacy after optimal micellar formulations, *Eur. J. Med. Chem.* 124 (2016) 284–298.
- [14] K. Ogawa, Y. Kobuke, Recent advances in two-photon photodynamic therapy, *Anticancer Agents Med. Chem.* 8 (2008) 269–279.
- [15] L. De Boni, D.S. Correa, D.L. Silva, P.J. Gonçalves, S.C. Zílio, G.G. Parra, I.E. Borissevitch, S. Canuto, C.R. Mendonça, Experimental and theoretical study of two-photon absorption in nitrofurans derivatives: promising compounds for photochemotherapy, *J. Chem. Phys.* 134 (2011) 014509.
- [16] G.C. Bolfarina, M.P. Siqueira-Moura, G.J.F. Demets, P.C. Morais, A.C. Tedesco, In vitro evaluation of combined hyperthermia and photodynamic effects using magnetoliposomes loaded with curcubit[7]uril zinc phthalocyanine complex on melanoma, *J. Photochem. Photobiol. B* 115 (2012) 1–4.
- [17] T. Gianferrara, I. Bratsos, E. Iengo, B. Milani, A. Ostric, C. Spagnul, E. Zangrando, Enzo Alessio, Synthetic strategies towards ruthenium porphyrin conjugates for anticancer activity, *Dalton Trans.* 48 (2009) 10742–10756.
- [18] T. Gianferrara, A. Bergamo, I. Bratsos, B. Milani, C. Spagnul, G. Sava, E. Alessio, Ruthenium-porphyrin conjugates with cytotoxic and phototoxic antitumor activity, *J. Med. Chem.* 53 (2010) 4678–4690.
- [19] S. Rani-Beeram, K. Meyer, A. McCrate, Y. Hong, M. Nielsen, S. Swavey, A fluorinated ruthenium porphyrin as a potential photodynamic therapy agent: synthesis, characterization, DNA binding, and melanoma cell studies, *Inorg. Chem.* 47 (2008) 11278–11283.
- [20] M. Pernot, T. Bastogne, N.P.E. Barry, B. Therrien, G. Koellensperger, S. Hann, V. Reshetov, M. Barberi-Heyob, Systems biology approach for in vivo photodynamic therapy optimization of ruthenium-porphyrin compounds, *J. Photochem. Photobiol. B* 117 (2012) 80–89.
- [21] A. Frei, R. Rubbiani, S. Tubafard, O. Blacque, P. Anstaett, A. Felgenträger, T. Maisch, L. Spiccia, G. Gasser, Synthesis, characterization, and biological evaluation of new Ru(II) polypyridyl photosensitizers for photodynamic therapy, *J. Med. Chem.* 57 (2014) 7280–7292.
- [22] R.N. Sampaio, W.R. Gomes, D.M.S. Araujo, A.E.H. Machado, R.A. Silva, A. Marletta, I.E. Borissevitch, A.S. Ito, L.R. Dinelli, A.A. Batista, S.C. Zílio, P.J. Gonçalves, N.M. Barbosa Neto, Investigation of ground- and excited-State photophysical properties of 5,10,15,20-Tetra(4-pyridyl)-21H,23H-porphyrin with ruthenium outlying complexes, *J. Phys. Chem. A* 116 (2012) 18–26.
- [23] L. Huang, Y. Xuan, Y. Koide, T. Zhiyentayev, M. Tanaka, M.R. Hamblin, Type I and Type II mechanisms of antimicrobial photodynamic therapy: an in vitro study on Gram-negative and Gram-positive bacteria, *Lasers Surg. Med.* 44 (2012) 490–499.
- [24] D.E.J.G.J. Dolmans, D. Fukumura, R.K. Jain, Photodynamic therapy for cancer, *Nat. Rev. Cancer* 3 (2003) 380–387.
- [25] M.G. Saaia, R.G. de Lima, A.C. Tedesco, R.S. da Silva, Nitric Oxide Production by visible light irradiation of aqueous solution of Nitrosyl Ruthenium Complexes, *Inorg. Chem.* 44 (2005) 9946–9951.
- [26] S.A. Cicillini, A.C.L. Prazias, A.C. Tedesco, O.A. Serra, R.S. da Silva, Nitric oxide and singlet oxygen photo-generation by light irradiation in the phototherapeutic window of a nitrosyl ruthenium conjugated with a phthalocyanine rare earth complex, *Polyhedron* 28 (2009) 2766–2770.
- [27] W. Xu, L.Z. Liu, M. Loizidou, M. Ahmed, I.G. Charles, The role of nitric oxide in cancer, *Cell Res.* 12 (2002) 311–320.
- [28] S. Mocellin, V. Bronte, D. Nitti, Nitric oxide, a double edged sword in cancer biology: searching for therapeutic opportunities, *Med. Res. Rev.* 27 (2007) 317–352.
- [29] J.C.G. Rotta, C.N. Lunardi, A.C. Tedesco, Nitric oxide release from the S-nitrosothiol zinc phthalocyanine complex by flash photolysis, *Braz. J. Med. Biol. Res.* 36 (2003) 587–594.
- [30] T.A. Heinrich, A.C. Tedesco, J.M. Fukuto, R.S. da Silva, Production of reactive oxygen and nitrogen species by light irradiation of a nitrosyl phthalocyanine ruthenium complex as a strategy for cancer treatment, *Dalton Trans.* 43 (2014) 4021–4025.
- [31] J.B. Godwin, T.J. Meyer, Preparation of ruthenium nitrosyl complexes containing 2,2'-bipyridine and 1,10-phenanthroline, *Inorg. Chem.* 10 (1971) 471–474.
- [32] J.B. Godwin, T.J. Meyer, Nitrosyl-nitrite interconversion in ruthenium complexes, *Inorg. Chem.* 10 (1971) 2150–2153.
- [33] Enraf-Nonius, Collect, Nonius BV, Delft, The Netherlands, 1997–2000.
- [34] Z. Otwinowski, W. Minor, Processing of X-ray diffraction data collected in oscillation mode, *Methods Enzymol.* 276 (1997) 307–326.
- [35] G.M. Sheldrick, ShelXS-97 Program for Crystal Structure Resolution, University of Göttingen, Göttingen, Germany, 1997.
- [36] R.H. Blessing, An empirical correction for absorption anisotropy, *Acta Crystallogr. A* 51 (1995) 33–38.
- [37] L.J. Farrugia, ORTEP-3 for windows – a version of ORTEP-III with a graphical user interface (GUI), *J. Appl. Crystallogr.* 30 (1997) 565–565.
- [38] P.J. Gonçalves, P.L. Franzen, D.S. Correa, L.M. Almeida, M. Takara, A.S. Ito, S.C. Zílio, I.E. Borissevitch, Effects of environment on the photophysical characteristics of mesotetrakis methylpyridiniumyl porphyrin (TMPyP), *Spectrochim. Acta A* 79 (2011) 1532–1539.
- [39] C.A. Strassert, G.M. Bilmes, J. Awruch, L.E. Dixelio, Comparative photophysical investigation of oxygen and sulfur as covalent linkers on octaalkylamino substituted zinc(II) phthalocyanines, *Photochem. Photobiol. Sci.* 7 (2008) 738–747.
- [40] C. Ian, G.L. Hug, Triplet-triplet absorption spectra of organic molecules in condensed phases, *J. Phys. Chem. Ref. Data* 15 (1986) 1–250.
- [41] N.A. Daghestanli, I.A. Degterev, G.B. Olivera, A.B. Seabra, M.G. de Oliveira, I.E. Borissevitch, Formation of cytotoxic intermediates in the course of photodecomposition of a nitroheterocyclic antiseptic quinifuryl, *J. Photochem. Photobiol. A* 184 (2006) 98–104.
- [42] T. Ohyashiki, M. Nunomura, T. Katoh, Detection of superoxide anion radical in phospholipid liposomal membrane by fluorescence quenching method using 1,3-diphenylisobenzofuran, *Biochim Biophys. Acta* 1421 (1999) 131–139.
- [43] M.G. Saaia, F.S. Oliveira, A.C. Tedesco, R.S. da Silva, Control of NO release by light irradiation from nitrosyl-ruthenium complexes containing polypyridyl ligands, *Inorg. Chim. Acta* 355 (2003) 191–196.
- [44] N. Nagao, D. Ooyama, K. Oomura, Y. Miura, S. Howell, M. Mukaida, Ligand nature of coordinated NO₂- in the oxidation reaction of Ru(II) complexes, [Ru(NO₂)(OH₂)(py)₄-2n(bpy)_n]+ (n=0, 1, 2), and their related complexes, *Inorg. Chim. Acta* 225 (1994) 111–121.
- [45] S.L. Queiroz, A.A. Batista, G. Oliva, M.T.P. Gambardella, R.H.A. Santos, K.S. MacFarlane, S.J. Retting, B.R. James, The reactivity of five-coordinate Ru(II) (1,4-bis(diphenylphosphino)butane) complexes with the N-donor ligands: ammonia, pyridine, 4-substituted pyridines, 2,2'-bipyridine, bis(o-pyridyl) amine, 1,10-phenanthroline, 4,7-diphenylphenanthroline and ethylenediamine, *Inorg. Chim. Acta* 267 (1998) 209–221.
- [46] A.Y. Ershov, P.V. Kucheryavii, A. B.Nikol'skii, Chemistry of ruthenium polypyridine complexes: IX. Nitro-Nitrosyl equilibrium in cis-[Ru(2, 2'-bpy) 2(L)NO₂]+ complexes, *Russ. J. Gen. Chem.* 74 (2004) 651–654.
- [47] M. Ghirrotti, C. Chiorboli, M.T. Indelli, F. Scandola, M. Casanova, E. Iengo, E. Alessio, Energy transfer pathways in pyridylporphyrin Re(I) adducts, *Inorg. Chim. Acta* 360 (2007) 1121–1130.
- [48] M.G. Saaia, R.G. de Lima, A.C. Tedesco, R.S. da Silva, Photoinduced NO release by visible light irradiation from pyrazi-bridged nitrosyl ruthenium complexes, *J. Am. Chem. Soc.* 125 (2003) 14718–14719.
- [49] T.A. Heinrich, A.C. Tedesco, J.M. Fukuto, R.S. da Silva, Photoinduced NO release by visible light irradiation from pyrazi-bridged nitrosyl ruthenium complexes, *Dalton Trans.* 43 (2014) 4021–4025.
- [50] R.V. Maximiano, E. Piovesan, S.C. Zílio, A.E.H. Machado, R. de Paula, J.A.S. Cavaleiro, I.E. Borissevitch, A.S. Ito, P.J. Gonçalves, N.M. Barbosa Neto, Excited-state absorption investigation of a cationic porphyrin derivative, *J. Photochem. Photobiol. A* 214 (2010) 115–120.
- [51] N.M. Barbosa Neto, D.S. Correa, L. De Boni, G.G. Parra, L. Misoguti, C.R. Mendonça, I.E. Borissevitch, S.C. Zílio, P.J. Gonçalves, Excited states absorption spectra of porphyrins—Solvent effects, *Chem. Phys. Lett.* 587 (2013) 118–123.
- [52] R.N. Sampaio, M.M. Silva, A.A. Batista, N.M. Barbosa Neto, Investigation of the photophysical and electrochemical properties of a free base tetrapyrrolyl porphyrin with meso carbon linked ruthenium(II) groups, *J. Photochem. Photobiol.* 315 (2016) 98–106.
- [53] I.M. Lorković, K.M. Miranda, B. Lee, S. Bernhard, J.R. Schoonover, P.C. Ford, Flash photolysis studies of the Ruthenium(II) porphyrins Ru(P)(NO)(ONO). multiple pathways involving reactions of intermediates with nitric oxide, *J. Am. Chem. Soc.* 120 (1998) 11674–11683.
- [54] K.S. Suslick, R.A. Watson, Photochemical reduction of nitrate and nitrite by manganese and iron porphyrins, *Inorg. Chem.* 30 (1991) 912–919.
- [55] M. Yamaji, Y. Hama, Y. Miyazaki, M. Hoshino, Photochemical formation of oxochromium(IV) tetraphenylporphyrin from nitrochromium(III) tetraphenylporphyrin in benzene, *Inorg. Chem.* 31 (1992) 932–934.
- [56] R.S. Corrêa, J.P. Barolli, M.I.F. Barbosa, J. Ellena, A.A. Batista, The effect of guest molecules on the conformation and molecular assembly of the fac-[RuCl₃(NO)(dppb)] complex, *J. Mol. Struct.* 1048 (2013) 11–17.
- [57] Z.N. da Rocha, R.G. de Lima, F.G. Doro, E. Tfouni, R.S. da Silva, Photochemical production of nitric oxide from a nitrosyl phthalocyanine ruthenium complex by irradiation with light in the phototherapeutic window, *Inorg. Chem. Commun.* 11 (2008) 737–740.
- [58] Z.A. Carneiro, J.C.B. De Moraes, F.P. Rodrigues, R.G. De Lima, C. Curti, Z.N. Da Rocha, M. Paulo, L.M. Bendhack, A.C. Tedesco, A.L.B. Formiga, R.S. da Silva, Photocytotoxic activity of a nitrosyl phthalocyanine ruthenium complex – A system capable of producing nitric oxide and singlet oxygen, *J. Inorg. Biochem.* 105 (2011) 1035–1043.
- [59] G.L. Squadrito, W.A. Pryor, Oxidative chemistry of nitric oxide: the roles of superoxide, peroxyxynitrite, and carbon dioxide, *Free Radic. Biol. Med.* 25 (1998) 392–403.
- [60] A. Fraix, A.R. Gonçalves, V. Cardile, A.C.E. Graziano, T.A. Theodossiou, K. Yannakopoulou, S. Sortino, A multifunctional bichromophoric nanoaggregate for fluorescence imaging and simultaneous photogeneration of RNOS and ROS, *Chem. Asian J.* 8 (2013) 2634–2641.