

Baseline AMH Level Associated With Ovulation Following Ovulation Induction in Women With Polycystic Ovary Syndrome

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Context: Anti-Müllerian hormone (AMH) reduces aromatase activity and sensitivity of follicles to FSH stimulation. Therefore, elevated serum AMH may indicate a higher threshold for response to ovulation induction in women with polycystic ovary syndrome (PCOS).

Objective: This study sought to determine the association between AMH levels and ovulatory response to treatment among the women enrolled into the Pregnancy in PCOS II (PPCOS II) trial.

Design and setting: This was a secondary analysis of data from a randomized clinical trial in academic health centers throughout the United States

Participants: A total of 748 women age 18–40 years, with PCOS and measured AMH levels at baseline, were included in this study.

Main Outcome Measures: Couples were followed for up to five treatment cycles to determine ovulation (midluteal serum progesterone > 5 ng/mL) and the dose required to achieve ovulation.

Results: A lower mean AMH and AMH per follicle was observed among women who ovulated compared with women who never achieved ovulation during the study (geometric mean AMH, 5.54 vs 7.35 ng/mL; $P = .0001$; geometric mean AMH per follicle, 0.14 vs 0.18; $P = .01$) after adjustment for age, body mass index, T, and insulin level. As AMH levels increased, the dose of ovulation induction medication needed to achieve ovulation also increased. No associations were observed between antral follicle count and ovulation.

Conclusions: These results suggest that high serum AMH is associated with a reduced response to ovulation induction among women with PCOS. Women with higher AMH levels may require higher doses of medication to achieve ovulation. (*J Clin Endocrinol Metab* 101: 3288–3296, 2016)

Anti-Müllerian hormone (AMH) is a dimeric glycoprotein member of the TGF- β family. In women, AMH is derived primarily from preantral and early antral follicles and has been shown in recent years to accurately reflect the follicular pool (1–3). Although the AMH level is used to assess the ovarian reserve in many scenarios, it has served most commonly to assess the likelihood of an adequate response during ovarian stimulation for assisted reproduction (4–8). In this context, AMH is predictive of the number of oocytes retrieved (9–15), and increasingly is being used to guide selection of the stimulation protocol (16–20).

During the last decade, serum AMH has also been evaluated in women with polycystic ovary syndrome (PCOS). Women with this disorder have an increased follicular pool and frequently present with oligo- or anovulation. Since 2003, the diagnostic criteria for PCOS has included an assessment of the follicular pool by specific ultrasound findings (21). There is support for the notion that AMH serves as a surrogate marker for the antral follicle count in the diagnosis of PCOS (22–28), and it has been posited that elevated AMH due to follicular excess, rather than facilitating ovulation, plays a critical role in the arrest of follicular growth that is characteristic of PCOS (29, 30).

An association between the high circulating AMH in women with PCOS and their chronic anovulation has long been noted, and seems to be due to several mechanisms. First, AMH directly inhibits aromatase activity in human and rodent granulosa cells (31–33). Further, Durlinger noted in an *in vitro* mouse model that direct culture with AMH inhibited the start of follicle growth from the pool of resting primordial follicles and attenuated FSH-stimulated preantral follicle growth (34). *In vitro* studies with human granulosa cells have shown an inhibition of aromatase activity and reduced follicular response to FSH in the presence of AMH (32, 35). Thus, excess AMH inhibits estradiol production and FSH action, stalling the FSH/E2-induced feed-forward loop that further increases the cellular FSH receptor and allows follicles to grow and mature. In this manner, it can be conceptualized that excess AMH is detrimental to the process of folliculogenesis.

Our objective was to evaluate the association between AMH level and ovulation in PCOS patients randomly assigned to treatment with either clomiphene citrate or letrozole. We hypothesized that a higher AMH level would be associated with lower ovulation rates in response to ovulation induction, and for those who did ovulate, that a higher AMH level would be associated with greater medication dose requirements to achieve ovulation.

Materials and Methods

Design and target population

This is a secondary analysis utilizing data from the PPCOS II trial, a multicenter, double-blind, randomized clinical trial (36, 37) comparing ovulation and live-birth rate in response to treatment with escalating doses of clomiphene citrate or letrozole for up to a total of five cycles. We have previously reported baseline AMH levels (38) and their change at end of study (36). The trial was registered on ClinicalTrials.gov as number NCT00719186. Each participating center's Institutional Review Board approved the study, and participants provided written informed consent. A Data Safety and Monitoring Board (DSMB) provided oversight.

The study design, methods, inclusion, and exclusion criteria have been described in detail elsewhere (36–38). In brief, 750 women with PCOS who were actively seeking pregnancy were enrolled. Females were between 18 and 40 years, with anovulation (defined as menstrual interval at least 45 days and/or ≤ 8 menses/y), combined with either hyperandrogenism (hirsutism or hyperandrogenemia) or polycystic appearing ovaries by ultrasound according to the Rotterdam criteria (39). Documentation of tubal patency was required, as well as a semen analysis with at least 14 million sperm per milliliter for the male partners. Exclusion criteria included contraindications to clomiphene or letrozole, poorly controlled diabetes, previously undiagnosed liver dysfunction, renal disease, significant anemia, history of thromboembolic events, or uncontrolled hypertension. Participants were randomly assigned to receive either an initial oral dose of 50 mg of clomiphene citrate or 2.5 mg of letrozole for 5 days per menstrual cycle and for up to five treatment cycles. The dose was increased in subsequent cycles in cases of nonresponse (midluteal progesterone < 3 ng/mL) or poor ovulatory response (progesterone levels indicative of ovulation but with values clustering just above the cutoff) (36–38), up to a maximum of 150 mg/d of clomiphene citrate or 7.5 mg/d of letrozole, both given for 5 days.

Data collection

Demographics and reproductive history were obtained using standardized forms, and waist circumference was measured to the nearest 0.1 cm. The degree of hirsutism was assessed at baseline using the modified Ferriman-Gallwey method (40). Fasting blood collected at baseline was used for all hormonal assays (38). Samples were batched and analyzed at the Ligand Assay and Analysis Core Laboratory at the University of Virginia. AMH was analyzed on samples stored at -80°C for approximately 13 months, using the Beckman-Coulter Gen 2 ELISA assay with no predilution. The sensitivity of the assay was to 0.25 ng/mL, with a range of 0.25–15 ng/mL. The interassay coefficient of variation was 7.0%, and the intraassay coefficient of variation was 3.0% (38). AMH levels were available for 748 of the 750 women enrolled onto the trial, and the data from the remaining 748 women were analyzed below. Insulin and LH were measured using the Immulite assay (Siemens Diagnostics), with all intra- and inter-assay coefficients of variation less than 6.2% (38). Fasting glucose levels were determined on a glucose analyzer using the glucose oxidase method, with all intra- and interassay coefficients of variation less than 1.5% (38). Insulin resistance (homeostasis model assessment for insulin resistance [HOMA-IR]) was calculated based on the homeostasis model (41). Total T was mea-

sured by RIA (Siemens Diagnostics), with intra- and interassay coefficients of variation less than 7.1% (38). This assay has been shown to have similar precision and quality control measures as commonly used liquid chromatography–tandem mass spectrometry assays of total T (42). Antral follicle count, which included both the left and right ovaries, was assessed during transvaginal ultrasound (using a 5–9-MHz vaginal probe, most centers used 5 MHz) at baseline, using standardized procedures as outlined previously (36). AMH per follicle was based on a concomitant AMH divided by the total antral follicle count.

The primary outcomes for this analysis were ovulation and the lowest dose required to achieve ovulation. Ovulation was defined as the first treatment cycle with a midluteal serum progesterone level greater than 5 ng/mL.

Statistical analysis

Descriptive statistics were used to compare characteristics between women by quartile of baseline AMH level, given that at present international standards for AMH thresholds do not exist (43). ANOVA, Kruskal-Wallis tests, χ^2 tests, or Fisher’s exact tests were used, as appropriate, to determine differences across quartiles. Geometric means and 95% confidence intervals (CIs) of AMH level, AMH level per follicle, and antral follicle count were compared by ovulatory status and by treatment group adjusted for age, body mass index (BMI), T, and insulin level. Geometric means were used given that AMH levels were not normally distributed. Further, geometric means and 95% CIs were compared by dose and treatment group adjusted for the same factors. The percentage of ovulatory and anovulatory cycles were compared by quartiles of AMH, AMH per follicle, and antral follicle count. Logistic regression models were used to

estimate odds ratios (ORs) and 95% CIs for the association between AMH, AMH per follicle, and antral follicle count, and achieved ovulation using both continuous and quartiles models. Unadjusted models were compared with models that 1) adjusted for age alone, 2) adjusted for age, BMI, T, and insulin level, 3) adjusted for age, BMI, T, insulin, and treatment group, and 4) adjusted for age, waist circumference, T, and insulin. Additional models were evaluated that assessed the association between AMH, AMH per follicle, and antral follicle count and ovulatory status per cycle using modified Poisson regression with robust errors to estimate the risk of an ovulatory cycle and 95% CIs accounting for multiple cycles per woman. The area under the receiver operating characteristics curve was also calculated. Analyses were conducted in SAS version 9.4 (SAS Institute).

Results

Demographic characteristics of participants overall, and by quartile of AMH level are shown in Table 1. Treatment arms were similar with regard to all characteristics assessed and are presented elsewhere (36). Women in the highest quartile of AMH level tended to be younger, have lower BMI, smaller waist circumference, lower insulin, glucose, HOMA-IR, as well as higher T and LH concentrations. No differences in hirsutism, smoking status, or treatment assignment were observed by AMH quartile.

Table 1. Demographic Characteristics of Women in PPCOS II by Quartile of Baseline AMH Level

Characteristics ^a	AMH (ng/mL) ^b					P-value ^c
	Overall	Quartile 1: <3.58	Quartile 2: 3.58–6.19	Quartile 3: 6.20–10.23	Quartile 4: >10.23	
n, (%)	748	187 (25)	187 (25)	187 (25)	187 (25)	
Age, y	28.9 ± 4.2	30.1 ± 4.6	28.2 ± 3.9	28.7 ± 4.1	28.5 ± 4.2	<.0001
BMI, kg/m ²	35.1 ± 9.2	39.7 ± 9.7	35.0 ± 8.6	34.5 ± 8.2	31.2 ± 8.3	<.0001
Waist circumference, cm	105.9 ± 20.5	114.7 ± 19.6	105.8 ± 19.0	105.3 ± 19.5	97.7 ± 20.3	<.0001
Hirsutism, mFG score						
Mean ± sd	17.0 ± 8.5	17.2 ± 8.5	17.5 ± 8.4	17.3 ± 8.8	15.9 ± 8.4	.27
mFG <8	97 (13)	17 (9)	22 (12)	28 (15)	30 (16)	.12
mFG 8–16	276 (37)	80 (43)	60 (32)	64 (34)	72 (39)	
mFG >16	375 (50)	90 (48)	105 (56)	95 (51)	85 (45)	
Insulin, μ U/mL	19.3 ± 27.0	24.8 ± 42.6	18.6 ± 17.2	18.9 ± 21.8	14.7 ± 17.4	<.0001
Glucose, mg/dL	86.0 ± 12.6	89.3 ± 15.2	86.0 ± 10.8	84.7 ± 12.0	84.2 ± 11.5	.0003
HOMA-IR	4.4 ± 9.1	6.1 ± 16.2	4.0 ± 3.6	4.2 ± 6.1	3.2 ± 4.2	<.0001
T, ng/dL	55.0 ± 28.8	40.3 ± 19.3	53.5 ± 31.0	57.6 ± 25.3	68.7 ± 30.7	<.0001
LH, mIU/mL	11.1 ± 8.5	8.6 ± 8.0	11.7 ± 8.8	11.3 ± 7.2	13.0 ± 9.4	<.0001
Current smoking ^d	111 (35)	25 (34)	31 (33)	28 (39)	27 (34)	.88
Treatment						
Clomiphene Citrate	375 (50)	91 (49)	90 (48)	99 (53)	95 (51)	.78
Letrozole	373 (50)	96 (51)	97 (52)	88 (47)	92 (49)	

Abbreviation: ; mFG, modified Ferriman-Gallway.

^a Values are Mean ± sd and n (%) as indicated.

^b AMH levels not available for two participants (one in each treatment group of the original trial).

^c Two-sided P-values for continuous variables calculated using ANOVA or the Kruskal-Wallis test, and for categorical variables using χ^2 tests or Fisher’s exact test where appropriate.

^d There were 430 values missing for current smoking status.

AMH levels were significantly lower among women who achieved ovulation over the trial (up to five treatment cycles), compared with women who did not achieve ovulation both overall, and within each treatment group ($P = .0001$) (Figure 1A). There were no significant differences between the treatment groups. AMH per follicle was also significantly lower among women who achieved ovulation compared with women who did not achieve ovulation overall, and among women treated with clomiphene, but not among those women treated with letrozole (Figure 1B). No differences in antral follicle count were observed between women who did and did not achieve ovulation either overall, or by treatment group (Figure 1C). A trend toward higher AMH levels at increased doses of ovulation induction medication was observed for both clomiphene citrate and letrozole (Table 2). Results were similar for AMH per follicle, although no differences were observed

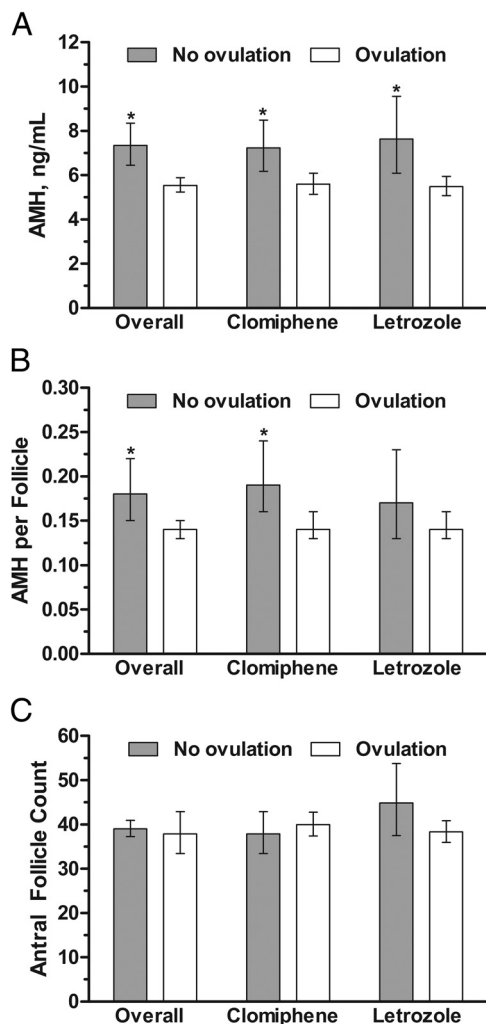


Figure 1. Comparison of geometric means (95% CIs) for (A) AMH concentrations, (B) AMH per follicle, and (C) antral follicle count by ovulatory status (comparing women who did not ovulate with women who achieved ovulation during the study) overall and by treatment group, adjusted for age, BMI, T, and insulin. *, $P < .05$ for comparison between ovulation and no-ovulation groups.

for antral follicle count. Importantly, we did not observe a level of AMH at or above which no one ovulated, and the area under the receiver operating characteristics curve was 0.59.

We also observed that women in the highest quartile of baseline AMH had a lower odds of ovulation compared with women in the lowest quartile (75% achieved ovulation in the fourth quartile compared with 87% in the first quartile; Table 3). This held after adjustment for age alone, as well as adjustment for age, BMI, T, and insulin, and additional adjustment for treatment group. Adjustment for waist circumference rather than BMI yielded similar results. Similar results were also observed for AMH per follicle, in that women in the second, third, and fourth quartiles compared with the first quartile had a lower odds of anovulation. Women in the third quartile of antral follicle count had a lower odds of ovulation compared with women in the lowest quartile. Similar results were observed when utilizing all cycles for analysis (Supplemental Table 1). Each one unit increase in AMH level and AMH per follicle was also associated with lower odds of ovulation after adjustment for age, BMI, T, and insulin levels, although the association was only marginally significant for AMH per follicle (AMH: OR, 0.94; 95% CI, 0.92–0.97; AMH per follicle: OR, 0.71; 95% CI, 0.49–1.03). No associations were observed with antral follicle count (OR, 1.00; 95% CI, 0.99–1.01)

Discussion

In a cohort of women with PCOS, we have observed that AMH levels were significantly lower among women who ovulated in response to medical treatment compared with women who never achieved an ovulatory cycle. Moreover, baseline AMH levels were lowest among women requiring the lowest dose to achieve ovulation, suggesting that high AMH is associated with a reduced response to ovulation induction, and women with higher AMH levels tended to need higher doses of ovulatory medication to achieve ovulation.

These results are consistent with prior studies of ovulation induction in smaller numbers of women with PCOS that observed an association of high AMH levels and lack of ovulation after treatment with laparoscopic ovarian diathermy and clomiphene (44, 45), and after ovulation induction with gonadotropins (4). We found that higher doses of ovulation induction agents were required to achieve ovulation for women with higher AMH levels, in agreement with the work of others (4, 44, 45). The prior studies associating high levels of AMH with a lack of response to ovarian stimulation suggested that anovulation

Table 2. AMH, AMH per Follicle, and Antral Follicle Count by Ovulation Induction Dose at First Ovulatory Response Overall and by Treatment Group Unadjusted and Adjusted For Age, BMI, T, and Insulin

Dose at First Ovulatory Response ¹	Unadjusted			Adjusted		
	Overall ²	Clomiphene	Letrozole	Overall	Clomiphene	Letrozole
AMH						
First dose	4.90 (4.38, 5.47) ^a	5.45 (4.66, 6.38) ^a	4.43 (3.79, 5.18) ^a	4.75 (4.31, 5.23) ^a	4.94 (4.29, 5.70) ^a	4.66 (4.08, 5.33) ^a
Second dose	6.07 (5.39, 6.83) ^{ab}	5.43 (4.53, 6.52) ^a	6.56 (5.60, 7.68) ^b	5.98 (5.40, 6.63) ^b	5.74 (4.89, 6.74) ^{ab}	6.05 (5.28, 6.93) ^b
Third dose	6.04 (5.34, 6.84) ^{ab}	6.17 (5.19, 7.34) ^a	5.92 (4.96, 7.07) ^{ab}	6.21 (5.58, 6.92) ^b	6.41 (5.50, 7.48) ^{ab}	5.97 (5.13, 6.95) ^{ab}
Never ovulated	6.87 (5.99, 7.88) ^b	6.81 (5.77, 8.04) ^a	6.97 (5.49, 8.85) ^b	7.09 (6.29, 7.99) ^b	7.01 (6.04, 8.13) ^b	7.33 (5.96, 9.01) ^b
<i>P</i> -trend	0.0002	0.04	0.002	<0.0001	0.0006	0.0004
AMH per follicle						
First dose	0.12 (0.10, 0.14) ^{ac}	0.13 (0.11, 0.17) ^a	0.11 (0.09, 0.13) ^a	0.12 (0.10, 0.13) ^a	0.12 (0.10, 0.14) ^a	0.11 (0.09, 0.14) ^a
Second dose	0.16 (0.14, 0.19) ^b	0.14 (0.11, 0.18) ^a	0.17 (0.14, 0.21) ^b	0.16 (0.14, 0.18) ^b	0.15 (0.12, 0.19) ^{ab}	0.16 (0.13, 0.19) ^b
Third dose	0.16 (0.14, 0.19) ^c	0.15 (0.12, 0.19) ^a	0.17 (0.14, 0.22) ^b	0.17 (0.15, 0.20) ^b	0.16 (0.13, 0.20) ^{ab}	0.18 (0.14, 0.21) ^b
Never ovulated	0.17 (0.14, 0.21) ^{bc}	0.18 (0.14, 0.23) ^a	0.15 (0.11, 0.21) ^{ab}	0.18 (0.15, 0.21) ^b	0.19 (0.15, 0.23) ^a	0.16 (0.12, 0.21) ^{ab}
<i>P</i> -trend	0.002	0.05	0.008	<0.0001	0.002	0.004
Antral Follicle Count						
First dose	41.3 (38.2, 44.6) ^a	40.9 (36.6, 45.7) ^a	41.6 (37.5, 46.3) ^{ab}	41.3 (38.3, 44.6) ^a	41.7 (37.4, 46.6) ^a	41.3 (37.1, 45.9) ^{ab}
Second dose	38.4 (35.4, 41.7) ^a	38.7 (34.1, 43.9) ^a	38.2 (34.3, 42.5) ^{ab}	38.4 (35.3, 41.6) ^a	37.9 (33.4, 42.9) ^a	38.5 (34.5, 42.9) ^{ab}
Third dose	36.9 (33.9, 40.2) ^a	40.0 (35.4, 45.1) ^a	34.1 (30.3, 38.5) ^a	36.8 (33.8, 40.1) ^a	39.8 (35.3, 44.9) ^a	34.2 (30.3, 38.7) ^a
Never ovulated	40.6 (36.9, 44.6) ^a	38.3 (34.1, 43.1) ^a	45.2 (38.4, 53.3) ^b	40.7 (36.9, 44.7) ^a	38.2 (34.0, 43.0) ^a	45.2 (38.3, 53.3) ^b
<i>P</i> -trend	0.45	0.52	0.65	0.45	0.39	0.73

^a Dose: For letrozole: first dose, 2.5 mg; second dose, 5.0 mg; third dose, 7.5 mg; and for clomiphene: first dose, 50 mg; second dose, 100 mg; third dose, 150 mg.

^b Values are geometric means and 95% CIs. Labeled means without a common letter differ. *P* < .05 (adjusted for multiple comparisons).

occurred at AMH levels that were lower than those observed in the present study. This discrepancy could be due to differences in the assays used given that the Beckman-Coulter assay tends to yield higher results (3, 46–52). Our findings differ from one prior study of 68 obese women with PCOS that observed higher AMH levels among women who responded to clomiphene citrate compared with those who did not respond (53). Importantly, the women in this prior study started the study receiving a relatively high dose of clomiphene (150 mg) compared with clinical practice protocols, and the protocol for the current study, which use an escalating dose schedule and start at 50 mg. This difference in protocol could partially explain the differences in findings between the studies. Notably, the women with PCOS in this prior study who did not respond to treatment had AMH levels that were even lower than the non-PCOS control group (53). Typically, AMH levels are higher among women with PCOS, which calls into question the comparability of the two study populations despite the fact that BMI was similar (the average BMI in the PCOS group in their study was 36.7 kg/m²). Although we found no associations with antral follicle count, it may be due to inaccuracy of counting in the high ranges given that the counts in this study were so high. Our results expand upon the findings in the previous studies by evaluating both clomiphene and letrozole, as well as accounting for potentially confounding factors, and using a much larger sample size (the largest previous study included only 68 women) from a well-designed randomized clinical trial.

These results are also in agreement with what is known from in vivo and in vitro studies regarding the important role of AMH in the regulation of ovarian function (54), and regulation of GnRH release (55). In particular, AMH has been shown to inhibit primordial follicle recruitment, as well as decrease the responsiveness of the follicle to FSH, which could have important downstream implications for ovulatory function (56–58). Elevated serum AMH reflects increased production of intraovarian AMH, which likely plays a role in the aberrations of follicle growth and ovulatory response seen among women with PCOS (59–61). In normal ovaries it has been shown that a reduction of AMH in follicles greater than 9 mm seems to be important in the selection of the dominant follicle (59–61). These findings may thus be particularly relevant for women with PCOS, given that women with PCOS are producing excess AMH in their granulosa cells, which could thus restrict the early stages of folliculogenesis and lead to anovulation (27, 32, 60–62). These AMH-induced processes make women with PCOS exceptionally challenging to treat because their large supply of preantral follicles, when recruited with exogenous FSH, can rapidly spiral into uncontrolled multiple follicle growth and risks for ovarian hyperstimulation (63). It should also be noted that other factors also influence follicle maturation, including obesity and insulin resistance (29, 64). Given that the women in this study had PCOS and were also obese, it may be that the effects were accentuated, although we adjusted for BMI as well as waist circumference to account for this factor.

Table 3. ORs and 95% CIs for the Association Between AMH levels, AMH per Follicle, Antral Follicle Count, and Ovulation

Model	Quartile 1	Quartile 2	Quartile 3	Quartile 4
Women, n	187	187	187	187
AMH, ng/mL	<3.58	3.58–6.19	6.20–10.23	>10.23
Anovulatory, n (%)	25 (13)	23 (12)	36 (19)	46 (25)
Ovulatory, n (%)	162 (87)	164 (88)	151 (81)	141 (75)
Unadjusted ^a	Reference	1.10 (0.60, 2.02)	0.65 (0.37, 1.13)	0.47 (0.28, 0.81)
Age adjusted	Reference	1.14 (0.62, 2.10)	0.66 (0.38, 1.16)	0.49 (0.28, 0.84)
Adjusted model 1	Reference	0.82 (0.43, 1.57)	0.45 (0.24, 0.83)	0.26 (0.14, 0.50)
Adjusted model 2	Reference	0.81 (0.42, 1.58)	0.45 (0.24, 0.84)	0.26 (0.13, 0.50)
Adjusted model 3	Reference	0.86 (0.45, 1.65)	0.49 (0.26, 0.89)	0.28 (0.15, 0.53)
AMH per follicle	<0.08	0.08–0.15	0.16–0.33	>0.33
Anovulatory, n (%)	21 (11)	34 (18)	32 (17)	39 (21)
Ovulatory, n (%)	163 (89)	150 (82)	153 (83)	145 (79)
Unadjusted	Reference	0.57 (0.32, 1.02)	0.62 (0.34, 1.12)	0.48 (0.27, 0.85)
Age adjusted	Reference	0.58 (0.32, 1.04)	0.63 (0.35, 1.13)	0.49 (0.27, 0.88)
Adjusted model 1	Reference	0.48 (0.26, 0.88)	0.49 (0.26, 0.93)	0.34 (0.18, 0.66)
Adjusted model 2	Reference	0.49 (0.26, 0.91)	0.48 (0.25, 0.92)	0.35 (0.18, 0.68)
Adjusted model 3	Reference	0.51 (0.28, 0.94)	0.48 (0.25, 0.92)	0.38 (0.20, 0.72)
Antral follicle Count	<27	27–40	41–58	>58
Anovulatory, n (%)	28 (15)	31 (16)	45 (25)	23 (13)
Ovulatory, n (%)	159 (85)	159 (84)	134 (75)	161 (87)
Unadjusted	Reference	0.90 (0.52, 1.58)	0.52 (0.31, 0.89)	1.23 (0.68, 2.23)
Age adjusted	Reference	0.89 (0.51, 1.55)	0.52 (0.30, 0.87)	1.21 (0.67, 2.20)
Adjusted model 1	Reference	0.84 (0.47, 1.49)	0.49 (0.28, 0.84)	1.21 (0.66, 2.24)
Adjusted model 2	Reference	0.84 (0.47, 1.51)	0.46 (0.27, 0.81)	1.23 (0.66, 2.27)
Adjusted model 3	Reference	0.74 (0.41, 1.33)	0.41 (0.23, 0.72)	1.15 (0.62, 2.14)

^a Logistic regression models to estimate ORs and 95% CIs. Adjusted model 1 is adjusted for age, BMI, T, and insulin. Adjusted model 2 is adjusted for the factors included in adjusted model 1, but is additionally adjusted for treatment. Adjusted model 3 is adjusted for age, waist circumference, T, and insulin.

Our study has many strengths, including measurement of AMH in a large cohort of over 700 women, making this the largest study of this kind to evaluate associations between AMH and response to ovulation induction in women with PCOS. Moreover, trial outcomes were well defined and well documented, and the study participants were monitored prospectively during frequent scheduled study visits to precisely document ovulation. One weakness is that given the variability in AMH assays and a lack of standardization, individual programs will need to assess their own cut points based on local assays. However, given that we did not identify a cut point above which no one ovulated, the observations that women with higher AMH levels may require higher doses of ovulation induction treatment still stand although this work does not reveal a predictive AMH threshold. It is also important to recognize that we evaluated AMH in relation to achieved ovulation during the study, which may have taken multiple cycles. As such, this may have potentially limited the power to detect associations even in this large cohort, although we observed similar associations when evaluating the association with ovulatory status during each cycle under study.

In summary, we observed that in women with PCOS, baseline serum AMH levels were higher among women

who did not respond to ovulation induction, although we did not find an absolute level of AMH above which women did not respond, and that AMH levels were significantly lower among women who ovulated. In addition, women with higher baseline AMH levels required higher doses of clomiphene or letrozole to achieve ovulation. These findings suggest that AMH may be a marker of ovarian resistance to ovulation induction.

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References

- Hansen KR, Hodnett GM, Knowlton N, Craig LB. Correlation of ovarian reserve tests with histologically determined primordial follicle number. *Fertil Steril*. 2011;95:170–175.
- Broekmans FJ, Visser JA, Laven JS, Broer SL, Themmen AP, Fauser BC. Anti-Müllerian hormone and ovarian dysfunction. *Trends Endocrinol Metab*. 2008;19:340–347.
- Broer SL, Broekmans FJ, Laven JS, Fauser BC. Anti-Müllerian hormone: Ovarian reserve testing and its potential clinical implications. *Hum Reprod Update*. 2014;20:688–701.
- Amer SA, Mahran A, Abdelmaged A, El-Adawy AR, Eissa MK, Shaw RW. The influence of circulating anti-Müllerian hormone on ovarian responsiveness to ovulation induction with gonadotrophins in women with polycystic ovarian syndrome: A pilot study. *Reprod Biol Endocrinol*. 2013;11:115.
- Broer SL, Dölleman M, Opmeer BC, Fauser BC, Mol BW, Broekmans FJ. AMH and AFC as predictors of excessive response in controlled ovarian hyperstimulation: A meta-analysis. *Hum Reprod Update*. 2011;17:46–54.
- Broer SL, Mol BW, Hendriks D, Broekmans FJ. The role of anti-müllerian hormone in prediction of outcome after IVF: Comparison with the antral follicle count. *Fertil Steril*. 2009;91:705–714.
- Jayaprakasan K, Campbell B, Hopkisson J, Johnson I, Raine-Fenning N. A prospective, comparative analysis of anti-Müllerian hormone, inhibin-B, and three-dimensional ultrasound determinants of ovarian reserve in the prediction of poor response to controlled ovarian stimulation. *Fertil Steril*. 2010;93:855–864.
- Jayaprakasan K, Deb S, Batcha M, et al. The cohort of antral follicles measuring 2–6 mm reflects the quantitative status of ovarian reserve as assessed by serum levels of anti-Müllerian hormone and response to controlled ovarian stimulation. *Fertil Steril*. 2010;94:1775–1781.
- Andersen AN, Witjes H, Gordon K, Mannaerts B. Predictive factors of ovarian response and clinical outcome after IVF/ICSI following a rFSH/GnRH antagonist protocol with or without oral contraceptive pre-treatment. *Hum Reprod*. 2011;26:3413–3423.
- Arce JC, LA Marca A, Mirner Klein B, Nyboe Andersen A, Fleming R. Antimüllerian hormone in gonadotropin releasing-hormone antagonist cycles: Prediction of ovarian response and cumulative treatment outcome in good-prognosis patients. *Fertil Steril*. 2013;99:1644–1653.
- Polyzos NP, Stoop D, Blockeel C, et al. Anti-Müllerian hormone for the assessment of ovarian response in GnRH-antagonist-treated oocyte donors. *Reprod Biomed Online*. 2012;24:532–539.
- Seifer DB, MacLaughlin DT, Christian BP, Feng B, Shelden RM. Early follicular serum mullerian-inhibiting substance levels are associated with ovarian response during assisted reproductive technology cycles. *Fertil Steril*. 2002;77:468–471.
- Majumder K, Gelbaya TA, Laing I, Nardo LG. The use of anti-Müllerian hormone and antral follicle count to predict the potential of oocytes and embryos. *Eur J Obstet Gynecol Reprod Biol*. 2010;150:166–170.
- Tsakos E, Tolikas A, Daniilidis A, Asimakopoulos B. Predictive value of anti-müllerian hormone, follicle-stimulating hormone and antral follicle count on the outcome of ovarian stimulation in women following GnRH-antagonist protocol for IVF/ET. *Arch Gynecol Obstet*. 2014;290:1249–1253.
- Lie Fong S, Baart EB, Martini E, et al. Anti-Müllerian hormone: A marker for oocyte quantity, oocyte quality and embryo quality? *Reprod Biomed Online*. 2008;16:664–670.
- Yates AP, Rustamov O, Roberts SA, et al. Anti-Müllerian hormone-tailored stimulation protocols improve outcomes whilst reducing adverse effects and costs of IVF. *Hum Reprod*. 2011;26:2353–2362.
- Broer SL, Mol B, Dölleman M, Fauser BC, Broekmans FJ. The role of anti-Müllerian hormone assessment in assisted reproductive technology outcome. *Curr Opin Obstet Gynecol*. 2010;22:193–201.
- La Marca A, D'Ippolito G. Ovarian response markers lead to appropriate and effective use of corifollitropin alpha in assisted reproduction. *Reprod Biomed Online*. 2014;28:183–190.
- Lan VT, Linh NK, Tuong HM, Wong PC, Howles CM. Anti-Müllerian hormone versus antral follicle count for defining the starting dose of FSH. *Reprod Biomed Online*. 2013;27:390–399.
- Fleming R, Broekmans F, Calhaz-Jorge C, et al. Can anti-Müllerian hormone concentrations be used to determine gonadotrophin dose

- and treatment protocol for ovarian stimulation? *Reprod Biomed Online*. 2013;26:431–439.
21. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome. *Fertil Steril*. 2004;81:19–25.
 22. Casadei L, Madrigale A, Puca F, et al. The role of serum anti-Müllerian hormone (AMH) in the hormonal diagnosis of polycystic ovary syndrome. *Gynecol Endocrinol*. 2013;29:545–550.
 23. Dewailly D, Gronier H, Poncelet E, et al. Diagnosis of polycystic ovary syndrome (PCOS): Revisiting the threshold values of follicle count on ultrasound and of the serum AMH level for the definition of polycystic ovaries. *Hum Reprod*. 2011;26:3123–3129.
 24. Pigny P, Jonard S, Robert Y, Dewailly D. Serum anti-Müllerian hormone as a surrogate for antral follicle count for definition of the polycystic ovary syndrome. *J Clin Endocrinol Metab*. 2006;91:941–945.
 25. Dewailly D, Lujan ME, Carmina E, et al. Definition and significance of polycystic ovarian morphology: A task force report from the Androgen Excess and Polycystic Ovary Syndrome Society. *Hum Reprod Update*. 2014;20:334–352.
 26. Eilertsen TB, Vanky E, Carlsen SM. Anti-Müllerian hormone in the diagnosis of polycystic ovary syndrome: Can morphologic description be replaced? *Hum Reprod*. 2012;27:2494–2502.
 27. Homburg R, Ray A, Bhide P, et al. The relationship of serum anti-Müllerian hormone with polycystic ovarian morphology and polycystic ovary syndrome: A prospective cohort study. *Hum Reprod*. 2013;28:1077–1083.
 28. Lauritsen MP, Bentzen JG, Pinborg A, et al. The prevalence of polycystic ovary syndrome in a normal population according to the Rotterdam criteria versus revised criteria including anti-Müllerian hormone. *Hum Reprod*. 2014;29:791–801.
 29. Jonard S, Dewailly D. The follicular excess in polycystic ovaries, due to intra-ovarian hyperandrogenism, may be the main culprit for the follicular arrest. *Hum Reprod Update*. 2004;10:107–117.
 30. Pigny P, Merlen E, Robert Y, et al. Elevated serum level of anti-Müllerian hormone in patients with polycystic ovary syndrome: Relationship to the ovarian follicle excess and to the follicular arrest. *J Clin Endocrinol Metab*. 2003;88:5957–5962.
 31. Grossman MP, Nakajima ST, Fallat ME, Siow Y. Müllerian-inhibiting substance inhibits cytochrome P450 aromatase activity in human granulosa lutein cell culture. *Fertil Steril*. 2008;89:1364–1370.
 32. Pellatt L, Rice S, Dilaver N, et al. Anti-Müllerian hormone reduces follicle sensitivity to follicle-stimulating hormone in human granulosa cells. *Fertil Steril*. 2011;96:1246–1251.e1
 33. Vigier B, Forest MG, Eychenne B, et al. Anti-Müllerian hormone produces endocrine sex reversal of fetal ovaries. *Proc Natl Acad Sci U S A*. 1989;86:3684–3688.
 34. Durlinger AL, Gruijters MJ, Kramer P, et al. Anti-Müllerian hormone attenuates the effects of FSH on follicle development in the mouse ovary. *Endocrinology*. 2001;142:4891–4899.
 35. Chang HM, Klausen C, Leung PC. Antimüllerian hormone inhibits follicle-stimulating hormone-induced adenyl cyclase activation, aromatase expression, and estradiol production in human granulosa-lutein cells. *Fertil Steril*. 2013;100:585–592.e1.
 36. Legro RS, Brzyski RG, Diamond MP, et al. Letrozole versus clomiphene for infertility in the polycystic ovary syndrome. *N Engl J Med*. 2014;371:119–129.
 37. Legro RS, Kunselman AR, Brzyski RG, et al. The Pregnancy in Polycystic Ovary Syndrome II (PPCOS II) trial: Rationale and design of a double-blind randomized trial of clomiphene citrate and letrozole for the treatment of infertility in women with polycystic ovary syndrome. *Contemp Clin Trials*. 2012;33:470–481.
 38. Legro RS, Brzyski RG, Diamond MP, et al. The Pregnancy in Polycystic Ovary Syndrome II study: Baseline characteristics and effects of obesity from a multicenter randomized clinical trial. *Fertil Steril*. 2014;101:258–269.e8
 39. Balen AH, Laven JS, Tan SL, Dewailly D. Ultrasound assessment of the polycystic ovary: International consensus definitions. *Hum Reprod Update*. 2003;9:505–514.
 40. Hatch R, Rosenfield RL, Kim MH, Tredway D. Hirsutism: Implications, etiology, and management. *Am J Obstet Gynecol*. 1981;140:815–830.
 41. Wallace TM, Levy JC, Matthews DR. Use and abuse of HOMA modeling. *Diabetes Care*. 2004;27:1487–1495.
 42. Legro RS, Schlaff WD, Diamond MP, et al. Total testosterone assays in women with polycystic ovary syndrome: Precision and correlation with hirsutism. *J Clin Endocrinol Metab*. 2010;95:5305–5313.
 43. Testing and interpreting measures of ovarian reserve. A committee opinion. *Fertil Steril*. 2015;103:e9–e17.
 44. Amer SA, Li TC, Ledger WL. The value of measuring anti-Müllerian hormone in women with anovulatory polycystic ovary syndrome undergoing laparoscopic ovarian diathermy. *Hum Reprod*. 2009;24:2760–2766.
 45. Mahran A, Abdelmegeed A, El-Adawy AR, Eissa MK, Shaw RW, Amer SA. The predictive value of circulating anti-Müllerian hormone in women with polycystic ovarian syndrome receiving clomiphene citrate: A prospective observational study. *J Clin Endocrinol Metab*. 2013;98:4170–4175.
 46. Clark CA, Laskin CA, Cadesky K. Anti-Müllerian hormone: Reality check. *Hum Reprod*. 2014;29:184–185.
 47. Craciunas L, Roberts SA, Yates AP, Smith A, Fitzgerald C, Pemberton PW. Modification of the Beckman-Coulter second-generation enzyme-linked immunosorbent assay protocol improves the reliability of serum antimüllerian hormone measurement. *Fertil Steril*. 2015;103:554–559 e551.
 48. Li HW, Ng EH, Wong BP, Anderson RA, Ho PC, Yeung WS. Correlation between three assay systems for anti-Müllerian hormone (AMH) determination. *J Assist Reprod Genet*. 2012;29:1443–1446.
 49. Nelson SM, Iliodromiti S, Fleming R, et al. Reference range for the antimüllerian hormone Generation II assay: A population study of 10,984 women, with comparison to the established Diagnostics Systems Laboratory nomogram. *Fertil Steril*. 2014;101:523–529.
 50. Rustamov O, Smith A, Roberts SA, et al. Anti-Müllerian hormone: Poor assay reproducibility in a large cohort of subjects suggests sample instability. *Hum Reprod*. 2012;27:3085–3091.
 51. Rustamov O, Smith A, Roberts SA, et al. The measurement of anti-Müllerian hormone: A critical appraisal. *J Clin Endocrinol Metab*. 2014;99:723–732.
 52. Wallace AM, Faye SA, Fleming R, Nelson SM. A multicentre evaluation of the new Beckman Coulter anti-Müllerian hormone immunoassay (AMH Gen II). *Ann Clin Biochem*. 2011;48:370–373.
 53. El-Halawaty S, Rizk A, Kamal M, et al. Clinical significance of serum concentration of anti-Müllerian hormone in obese women with polycystic ovary syndrome. *Reprod Biomed Online*. 2007;15:495–499.
 54. Durlinger AL, Visser JA, Themmen AP. Regulation of ovarian function: The role of anti-Müllerian hormone. *Reproduction*. 2002;124:601–609.
 55. Cimino I, Casoni F, Liu X, et al. Novel role for anti-Müllerian hormone in the regulation of GnRH neuron excitability and hormone secretion. *Nat Commun*. 2016;7:10055.
 56. Durlinger AL, Kramer P, Karels B, et al. Control of primordial follicle recruitment by anti-Müllerian hormone in the mouse ovary. *Endocrinology*. 1999;140:5789–5796.
 57. Durlinger AL, Gruijters MJ, Kramer P, et al. Anti-Müllerian hormone inhibits initiation of primordial follicle growth in the mouse ovary. *Endocrinology*. 2002;143:1076–1084.
 58. Weenen C, Laven JS, Von Bergh AR, et al. Anti-Müllerian hormone expression pattern in the human ovary: Potential implications for initial and cyclic follicle recruitment. *Mol Hum Reprod*. 2004;10:77–83.
 59. Pellatt L, Hanna L, Brincat M, et al. Granulosa cell production of

- anti-Müllerian hormone is increased in polycystic ovaries. *J Clin Endocrinol Metab.* 2007;92:240–245.
60. **Homburg R, Crawford G.** The role of AMH in anovulation associated with PCOS: A hypothesis. *Hum Reprod.* 2014;29:1117–1121.
61. **Pellatt L, Rice S, Mason HD.** Anti-Müllerian hormone and polycystic ovary syndrome: A mountain too high? *Reproduction.* 2010;139:825–833.
62. **Piltonen T, Morin-Papunen L, Koivunen R, Perheentupa A, Ruokonen A, Tapanainen JS.** Serum anti-Müllerian hormone levels remain high until late reproductive age and decrease during metformin therapy in women with polycystic ovary syndrome. *Hum Reprod.* 2005;20:1820–1826.
63. **Tal R, Seifer DB, Khanimov M, Malter HE, Grazi RV, Leader B.** Characterization of women with elevated antimüllerian hormone levels (AMH): Correlation of AMH with polycystic ovarian syndrome phenotypes and assisted reproductive technology outcomes. *Am J Obstet Gynecol.* 2014;211:59 e51–e58.
64. **Purcell SH, Moley KH.** The impact of obesity on egg quality. *J Assist Reprod Genet.* 2011;28:517–524.