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Unintended Immunological Consequences of Biologic Therapy

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Abstract

Recent advances in the understanding of immune dysregulation in autoimmune diseases have enabled the development of new monoclonal antibody based drugs called biologics. Biologics have been used to target aberrant immune responses in many diseases, but patients with rheumatologic and other autoimmune diseases have benefited the most and improvements in outcomes have been significant. The use of biologics is not without hazard; however, as these agents block immune pathways adapted to protect the host. This has been borne out by increased rates of infections as well as induction of new autoimmune and hematologic adverse effects. As new drugs for the treatment of autoimmune conditions are entering the pipeline, it is incumbent on the practicing immunologist to understand the mechanism of these biologics and the implications of clinical use.

Keywords

Autoimmune disease; biologics; monoclonal antibody; primary immunodeficiency; secondary immunodeficiency; biologic safety

Introduction

Our understanding of immune function has made great progress over the last few decades. This progress has led to the development of monoclonal antibodies (mAbs, biologics) that have revolutionized the treatment of rheumatologic/autoimmune diseases [1]. Small molecules inhibitors such as the Janus kinase (JAK) inhibitors are also important drugs in treatment of autoimmune disease, but will not be addressed in this review. Biologics provide therapy that target specific aspects of the immune system in order to combat autoimmune

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Compliance with Ethics Guidelines

Conflict of Interest

Drs. Henrickson, Ruffner, and Kwan declare no conflicts of interest.

Human and Animal Rights and Informed Consent

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responses. Now there are effective therapies for chronic systemic inflammatory conditions that previously had high rates of morbidity and mortality. Although biologics are tremendously successful in treating chronic inflammatory diseases, there are significant risks as specific components of immunological function are suppressed by these drugs [2]. As some of these biologics may mimic known immunodeficiencies, this is an important topic for immunologists to understand. In this review, we present currently available therapies for autoimmune/inflammatory conditions (Table 1), the immune mechanisms that they target and known and possible adverse outcomes based on their targets.

Innate Immunity

Complement

The complement system targets pathogens using three key mechanisms: bacterial cytolysis, pathogen opsonization allowing targeting by phagocytes, and stimulating inflammation via release of anaphylatoxins. The importance of the complement system is emphasized by known deficiencies in complement components such as C5 which results in invasive *Neisseria* infections [3]. Recent data also indicates that complement is involved in other pathways and is key in immune downregulation and homeostasis. Indeed, derangements in complement activation (e.g. atypical hemolytic uremic syndrome (aHUS)) or regulation (e.g. paroxysmal nocturnal hemoglobinuria (PNH)) may lead to inflammatory disease [4].

Eculizumab is currently the only biologic targeting the complement pathway that is approved and is indicated for treatment of PNH [5, 6] and aHUS [5] Eculizumab is a humanized anti-C5 mAb that selectively blocks activation of C5, release of C5a, and initiation of the membrane attack complex [5, 6]. PNH is a rare disorder resulting from an acquired somatic mutation in phosphatidylinositol glycan class A (*PIGA*) gene in hematopoietic stem cells leading to loss of glycophosphatidylinositol (GPI) anchor synthesis and expression of GPI anchored proteins such as decay accelerating factor (DAF or CD55) and CD59, which are key in prevention of complement activation on host cells. PNH patients suffer from hemolytic anemia, thrombosis, and cytopenias at significant morbidity and mortality with limited options for treatment until the advent of eculizumab. Eculizumab was approved for PNH in 2 small randomized clinical trials that showed significant improvement in outcomes for hemolysis and thrombosis. However, a significant subset of patients were refractory to treatment likely due to C3b deposition on red cells causing clearance in the spleen. Based on this observation, drugs targeting C3 are currently in the pipeline for the treatment of PNH [5, 6].

As eculizumab blocks C5, there have been concerns about susceptibility to invasive meningococcal infection. Patients were given meningococcal vaccine during clinical trials, but 2 patients developed meningococcal serotype B infections that resolved with antibiotic therapy. As there have been cases of serotype B infections on eculizumab, it is now recommended that patients receive meningococcal vaccination and prophylactic antibiotics [3, 5]. With the introduction of Bexsero, a meningococcal serotype B vaccine, consideration for use of this vaccine with eculizumab treatment is likely. As the clinical trials and the post marketing data are sparse, the use of this drug should be done with the utmost of caution due to susceptibility to *N. meningitides.* Additionally, caution will be needed as therapeutics

targeting C3 are in development, and C3 deficiency engenders a broader range of susceptibility to infectious pyogenic bacteria such as pneumococcus.

Cytokines

TNF-α, IL-1, and IL-6 are innate pro-inflammatory cytokines that are induced by infection. TNF-α is the primary cytokine that induces the inflammatory cascade as it is able to induce both IL-1 and IL-6. IL-1 is also able to induce IL-6 directly. In concert, this cascade promotes leukocyte recruitment, the activation of T cells, and the elaboration of other proinflammatory molecules such as serum amyloid A and C-reactive protein [7]. These cytokines have crucial disease inducing roles in autoimmune diseases (e.g. rheumatoid arthritis (RA), inflammatory bowel disease (IBD)) and autoinflammatory diseases (e.g. cryopyrin associated periodic syndromes (CAPS)) and are thus important therapeutic targets [8].

TNF-a is induced by microbial products during the initiation of an infection and in turn initiates the inflammatory cascade. This makes TNF an important druggable target, but as it is at the apex of the pathway, blocking this cytokine also leaves patients susceptible to infection due to its broad effects in immunity. TNF inhibitors (TNFis) were the initial biologics that were approved for use in RA. Their immunological adverse effects are reviewed in detail in Her *et al* [1]. Suffice it to say that this class of drugs has a high rate of serious infections, including tuberculosis, that were not anticipated in the clinical trials and careful screening for Mtb worldwide and fungal infections (such as *Histoplasma*) is critical in endemic regions Autoimmune conditions and paradoxical inflammation have also been induced by TNFis, which necessitates discontinuation of treatment [1].

IL-1

IL-1 is a prototypic pro-inflammatory cytokine that shares many properties with TNF- α and consists of 2 forms, IL-1 α and IL-1 β , the main biologically active form being IL-1 β . Secretion of IL-1 β is tightly regulated and released upon inflammasome activation in monocyte/macrophages during infection and inflammation. IL-1 serves to enhance the effector functions of innate immune cells and also facilitates the action of B cells and CD4+ Th cells [9]. As IL-1 is a key inflammatory cytokine driving innate and adaptive responses, it is an important contributor to autoimmune (e.g. RA) [9] and autoinflammatory (e.g. CAPS) [8]conditions leading to chronic systemic inflammation and symptomatology. Due to the critical role of IL-1 in inflammatory diseases, the application of IL-1 blockade for therapeutic treatment has become an important aspect of biologic development for a multitude of conditions with anakinra, canakinumab, and rilonacept currently approved [9, 10, 8].

Although IL-1 is important in induction of inflammatory responses to pathogens, the use of IL-1 antagonists has been quite safe with mainly increases in common bacterial and viral infections. Unlike other classes of biologics, such as TNFis, there is no increase in opportunistic infections with the use of anakinra such as *M. tuberculosis* (Mtb) reactivation [8] although with longer acting canakinumab (anti-IL-1 β) there was a somewhat higher rate of infections (e.g. nasopharyngitis, urinary tract infection) [10][11]. Indeed, use of anakinra

in patients with inflammatory disease during active infections (e.g. hidradenitis suppurativa) or with treatment of autoimmune manifestations in immunodeficient patients (i.e. IBD in chronic granulomatous disease) controls inflammation without causing exacerbation or onset of infection. There are reported cases of neutropenia IL-1 blockade, but there have only been rare cases of neutrophil counts less than 500 cells/mL (all without significant infections) and neutropenia corrects with cessation of therapy [8].

IL-6

IL-6 is a pleiotropic cytokine induced by TNF-α and IL-1 and functions in T cell expansion and activation, B cell differentiation, and activation of acute phase responses. This cytokine affects immune function and metabolic pathways including lipid metabolism, vasculopathies, and endocrine disease. Although downstream from TNF-α and IL-1, IL-6 is critical to immunity as proven by individuals with Hyper IgE syndrome (HIES) who demonstrate decreased IL-6 and IL-17 activity allowing recurrent infections particularly with Gram-negative bacteria and fungi. Although uncommon, children who develop inhibitory IL-6 autoantibodies are susceptible to cutaneous staphylococcal infections [12]. Although IL-6 was thought to be a biomarker of inflammatory disease, it is clear that IL-6 is involved in disease induction in autoimmune disease (e.g. RA, IBD)[13]. Tocilizumab is an IL-6 receptor (IL-6R) blocking humanized mAb approved for treatment of RA [13] and polyarticular and systemic juvenile idiopathic arthritis (PJIA and SJIA)[13, 14]. Tocilizumab, as well as other IL-6R and IL-6 inhibiting biologics, are currently in clinical trials to treat other autoimmune/inflammatory conditions including systemic sclerosis and adult Still's disease [14].

Blocking IL-6 therapeutically is difficult as this cytokine is crucial in multiple immune pathways including Th17 induction and acute phase responses [14, 12]. As such, it would be predicted that biologics targeting this cytokine would increase infections and impact other aspects of immune function. In clinical trials, tocilizumab has had an increased rate of infections with respiratory tract infections being most common, but increased cutaneous infections were also noted [15, 16]. Mtb reactivation has also been reported although at a lower rate than with TNFis and screening for tuberculosis is recommended [1, 16]. Other adverse immune effects observed with tocilizumab include some cases of neutropenia that were not associated with serious infections [10, 17, 16] Finally, in trials for SJIA, there were reports of episodes of macrophage activation syndrome (MAS) upon withdrawal of drug, although SJIA patients have an overall increased rate of MAS [17]. There are a significant number of biologics targeting IL-6 currently in trials (Table 2) and based on the mechanism of action of this cytokine, other possible immunological derangements include a spectrum of infections similar to deficiencies that result in Th17 defects as each biologic will likely have differing off target effects.

Adaptive Immunity

IL-17

IL-17A is an IL-17 family member and has proinflammatory functions linked to secretion of TNF, IL-6, IL-1 and GM-CSF. IL-17A is fundamental to eradication of extracellular bacteria

and fungal infections, but is also associated with autoimmune diseases including RA, psoriasis and multiple sclerosis (MS)[18]. Therefore, while IL-17 blockade could improve immune dysregulation in selected autoimmune conditions, it potentially increases fungal and extracellular bacterial infection. In humans, IL-17 pathway deficiencies lead to chronic mucocutaneous candidiasis [19]. Additionally, Th17 cells are important in combating mycobacterial infections making Mtb reactivation a concern [20]. Secukinumab, an anti-IL-17A mAb, is approved for plaque psoriasis [21–24], ankylosing spondylitis (AS) [25] and psoriatic arthritis (PsA)[26, 27]. Studies and trials have demonstrated an overall increase in infections with secukinumab versus placebo, including the expected increase in candida infections (mostly mild to moderate oral or genital infections whose incidence was dose dependent)[23, 28, 27] [29] oral herpes infections [30], as well as tinea pedia and two cases of perianal dermatophytosis [28]. In addition, neutropenia, and Crohn's disease (CD) were noted to be increased versus placebo in some studies[21, 22, 24–26]. There was no Mtb reactivation or invasive fungal infections noted [30], but it is recommended to screen for TB prior to initiating therapy and to use caution if considering use in a patient with active CD

IL-12/IL-23

IL-23 is a member of the IL-12 family, which also includes IL-27 and IL-35. This complex family of heterodimeric cytokines share cytokine subunits, but have divergent, sometimes orthogonal, functions. For example, although IL-12 and IL-23 share IL-12p40, IL-12 induces Th1 differentiation while IL-23 plays a role in Th17 differentiation/maintenance [31]. As discussed previously, immunodeficiency patients with Th17 pathway mutations have susceptibility to mucocutaneous candidiasis, mycobacteria, salmonella and S. aureus induced cold abscesses, but these infections were not seen in ustekinumab trials [18]. Given that this cytokine subunit is shared with IL-12, it is important to consider that effect as well. Mendelian susceptibility to mycobacterial disease (MSMD) is a broad set of diseases (such as IL12B, IL12RB1 mutations), which impact production and response to IFN- γ and are characterized by susceptibility to infections including Mycobacteria (i.e. BCG vaccine strains, MTb, etc), as well as Candida and Salmonella, among others [32]. IL-12RB1 defects are the most common cause of MSMD, with multiple mutations and broad clinical phenotype with susceptibility to mycobacteria (non BCG), invasive salmonellosis, and Candida, in addition to less frequent infections with bacteria (Klebsiella, S. pneumoniae, Nocardia) and other fungi [32]. Patients with IL-12p40 deficiency lack IL-12 and IL-23, and are characterized by BCG-osis after vaccination, salmonellosis and infections similar to IL-12Rβ1 deficiency [32]. Ustekinumab is a human IgG1 anti-IL-12p40 mAb. It is approved for treatment of severe plaque psoriasis and PsA [33–35]. There was no mention of systemic candidal infections in trials [36, 37] although there were higher rates of infection more broadly, most commonly including upper respiratory tract infections/nasopharyngitis [38, 39], but also reported a case of asymptomatic Mtb reactivation [38, 40], herpes zoster [34, 37] and cellulitis [39, 34]. It is recommended to screen for TB prior to initiating therapy and not to treat patients with active TB and to treat latent TB in patients prior to initiating therapy. Malignancy was also reported with non-melanoma skin cancer being most common [37]. Finally, one case of reversible posterior leukencephalopathy syndrome was reported

IL-2Ra (CD25)

IL-2R α , one chain of the high affinity IL-2 receptor (IL-2R), is found on activated T cells and regulatory T (Treg) cells. Daclizumab, blocks the IL-2 binding site on IL-2R, has been approved for treatment of organ transplant rejection, and in a phase III trial of daclizumab showed superiority in relapsing remitting MS versus IFN β -1- α [41]. With regards to adverse events, when daclizumab was compared to IFN β -1-a for relapsing-remitting MS, there was a similar rate of herpes infection, upper respiratory tract infections and urinary tract infections and no episodes of PML [41]. If note, one study reported higher rates of aspergillus colonization in lung transplantation [42]. There were higher rates of elevated liver function tests and cutaneous reactions in the daclizumab group. In prior studies, higher infections were also noted [43, 44].

Basiliximab, a chimeric monoclonal antibody IL-2Ra antagonist, is approved for use in induction of tolerance and transplant rejection in kidney transplant and did not have a clear pattern of increased infection. In trials, there was concern with regards to possible increase in infections [45–49]. It carries a black box warning with regards to being administered by physicians familiar with immunosuppression and transplant.

T cell

CTLA-4

T cells require two signals for activation, TCR activation via cognate peptide-MHC complex and costimulation [50]. The prototypic costimulatory signal is transduced by CD28 on T cells binding to CD80/86 on antigen presenting cells. As T cell activation progresses, CTLA-4 is upregulated and transduces negative signals as it binds with higher affinity to CD80/86 blocking further interaction with CD28. CTLA-4 is also constitutively expressed on regulatory T cells and acts to preserve peripheral tolerance.

Abatacept, a fusion of the extracellular domain of CTLA-4 to an altered Fc segment of IgG1, was developed to block T cell costimulation by binding to CD80 and CD86, thus preventing T cell activation. It has been approved for RA and JIA [51–62]. By preventing proper activation of T cells, concern for opportunistic infection is appropriate. Prior to initiating abatacept, screening for latent TB is recommended and treatment for latent TB should be completed prior to abatacept initiation. In addition, screening for hepatitis infection is also recommendation given concern for reactivation. Common infections included upper respiratory tract infections, [54, 55, 58–60, 62, 63], nasopharyngitis [63, 52, 54, 58, 59, 62], bronchitis [55, 62], UTI [63, 52, 60, 62] and sinusitis [56]. A small increase in herpes simplex was noted versus placebo [53], as well as influenza and gastroenteritis [64]. Serious infections included pneumonia [63, 52, 55, 58, 59, 62], bronchitis [55], cellulitis [65, 60] and urinary tract infections [63, 62, 52], pyelonephritis and diverticulitis. Infections with *Candida* [53, 66], TB [53][67], noted to be in patients from endemic areas, herpes zoster [66], ABPA[53], histoplasmosis [66], malignancies, which were generally consistent with placebo other than increases in lymphoma and lung malignancy which were argued to be consistent with the RA population [53, 52, 51], and autoimmunity (generally

psoriasis, with a few episodes of development of ANA or anti-dsDNA antibodies[64]) were also reported [63, 66, 55] though some of the studies were not placebo controlled.Overall, meta-analyses of abatacept studies demonstrated no significant increase in serious [68] or overall infections [53, 51]. Of note, abatacept in combination with TNF antagonists was shown to increase infections without increasing efficacy and is not recommended. Interestingly, patients with LRBA deficiency (CTLA-4 deficiency based on defective CTLA-4 recycling) have shown improvement with abatacept administration, and has been shown recently to be effective in CTLA-4 haploinsufficient patients [69][70].

Belatacept, CTLA-4Ig with higher avidity, has been approved for kidney transplant rejection in EBV seropositive patients, given the black box concern for development of PTLD, with a higher risk in the EBV seronegative population, as well as in CMV seronegative patients or those who have received T cell therapies [71–73]. Adverse reactions included nasopharyngitis and serious infections including increased CMV [71]. There was increased malignancy (especially non-melanoma skin cancer) and fungal and viral infections at similar rates compared with use of cyclosporine alone [74, 73]. There was an increase in TB infections with belatacept versus cyclosporine, which was noted to generally be in endemic regions, and they recommend screening for and treating latent TB prior to initation[73]. Post-transplant lymphoproliferative disorder was also reported [72], more often in the CNS [73], and is a black box warning for this therapy [75]. Of note, PML has also been reported [73] as well as polyoma virus nephropathy (often with BK virus). They recommend prophylaxis for CMV and PCP for three months after transplant. Patients who are undergoing liver transplant are not recommended to receive belatacept given worse outcomes.

In the setting of cancer, much like chronic infection, adaptive immunity can become hampered in its ability to combat a chronic antigen. Specifically, T cells become exhausted [76], with inhibitory signals outweighing the effect of activating signals. In this setting, therapeutic activation is desired. When predicting possible adverse events, CTLA-4 knock out mice were noted to have rampant autoimmunity and eventual fatality [77]. In humans, CTLA-4 haploinsufficiency mutations yield a clinical phenotype consistent with common variable immune deficiency in addition to dysfunctional Tregs and autoimmunity [78]. Ipilimumab is a mAb that prevents CTLA-4 interaction with CD80/86 allowing reactivation of exhausted T cells by halting inhibitory signals [76]. Ipilimumab is approved for the treatment of metastatic melanoma and is in trials for non-small cell and small cell lung carcinoma, renal cell carcinoma, and prostate cancer [79-81]. Ipilimumab carries a black box warning for immune related adverse events (irAE), which include skin rash, inflammatory colitis, hypophysitis, elevated LFTs and other inflammatory conditions [82, 79, 83]. Interestingly, treatment guidelines for many irAEs have been developed with good acute improvement, though some serious irAEs require cessation of therapy and high dose corticosteroids, and other therapies depending on the severity and affected organ [84, 82].

Adhesion Molecules

Leukocyte trafficking from the vasculature into the peripheral tissues involves three critical steps: rolling, firm adhesion and transendothelial migration. Selectins mediate initial

tethering and subsequent rolling of leukocytes along the endothelium [85, 86]. Leukocyte rolling permits further cell-cell interactions between integrins and endothelial adressins required for firm adhesion [85, 86]. Integrins are composed of α and β subunits that initiate intracellular signaling activating leukocytes when bound to their cognate addressin[85, 86], ultimately allowing for transendothelial migration. Genetic defects in cell adhesion have been identified resulting in Leukocyte Adhesion Deficiency (LAD) syndromes [85]. Starting in childhood, LAD patients have significant defects in antibacterial and antifungal defense resulting in characteristic non-purulent infections (oomphalitis and periodontitis) as well as invasive infections of other sites [85].

Natalizumab is a recombinant humanized IgG4 mAb that targets a_4 integrin blocking the binding of both gut-specific $a_4\beta_7$ as well as $a_4\beta_1$ integrin (binds VCAM-1 on the blood brain barrier)[87]. Natalizumab is currently approved as monotherapy for relapsing forms of MS and as a second line agent for refractory IBD. A surprising outcome of natalizumab post-market surveillance was occurrence of progressive multifocal leukoencephalopathy (PML), a severe demyelinating disease associated with activation of latent JC virus infection in immunocompromised hosts [88, 89]. Efalizumab, a recombinant humanized neutralizing antibody to $a_L\beta_2$ (LFA-1) had been approved for the treatment of severe psoriasis, but was withdrawn from the market in 2009 due to occurrence of three fatal cases of PML[90]. It is thought that due to impaired trafficking of lymphocytes into the CNS, there is a loss of lymphocyte-mediated CNS immunosurveillance which leads to the development of PML. Although not as well-known, there have been reports of increased rates of varicella zoster and herpes simplex central nervous system opportunistic infections in patients on natalizumab [91].

Lymphocytes bearing $\alpha_4\beta_7$ integrin home to the gut where they bind mucosal adressin celladhesion molecule-1 (MadCAM-1) expressed specifically on gut-associated lymphoid tissues (GALT). Vedolizumab, a recombinant humanized IgG1 antibody, is selective for gut tissue and binds only the $\alpha_4\beta_7$ complex [86, 89]. Due to this selectivity, vedolizumab has a more limited infectious risk profile. No known cases of PML have been associated with vedolizumab. Not surprisingly, there were slight increases in gastroenteritis and enteric abscesses compared to placebo given that vedolizumab inhibits lymphocyte transmigration into the gut [89]. Although there are case reports raising concerns that decreased immunosurveillance of nasopharyngeal mucosa were causing systemic infections [89, 92], this was not statistically significant [89]. Vedolizumab is approved as second-line treatment of moderate to severe IBD TNFi non-responders.

B Cells

B cells have been implicated in numerous autoimmune conditions, particular those where prognosis or response to therapy can be associated with autoantibody status, e.g. systemic lupus erythematosus (SLE), or ANCA vasculitis. In addition to secreting harmful autoantibodies, B cells also release pro-inflammatory cytokines (lymphotoxin, TNFα and IL-6) and present antigen to autoreactive T cells [93–95]. Therefore, there has been considerable interest in development of B-cell targeted therapies for the treatment of autoimmune conditions.

CD20 is a glycosylated phospholipid expressed on circulating B cells, but not B-cell hematopoietic precursors, plasmablasts or plasma cells [96]. Rituximab is a chimeric anti-CD20 mAb which induces transient depletion of circulating peripheral B cells. Rituximab is approved for the treatment of refractory, indolent non-Hodgkins' lymphoma, with methotrexate for TNFi-refractory RA, and for granulomatous polyangiitis. Of note, rituximab monotherapy in systemic lupus erythematosus (SLE) was not successful as many patients experienced worsening lupus flares with B cell reconstitution, which may in part be secondary to B-cell activation factor (BAFF) driven repopulation of autoreactive B cells following rituximab [97].

In initial clinical trials, transient hypogammaglobulinemia was seen in 14% of patients and average B cell recovery was 6-9 months following rituximab treatment [98, 99]. However, there have since been numerous reports of delayed reconstitution of memory B cells with hypogammaglobulinemia and recurrent infections in a subset of patients treated with rituximab [100–103]. Of note, several newer anti-CD20 mAbs have been developed, including ocrelizumab, ofatumumab, and veltuzumab. Ofatumumab was approved in a multi-drug regimen for treatment of chronic lymphocytic leukemia. Multiple clinical trials are ongoing with these biologics for both oncologic and rheumatologic conditions.

Depletion of B cells with anti-CD20 antibodies may also result in disruption of combined immune function. Both rituximab and oftatumumab bear black-box warnings regarding risk of PML and hepatitis B reactivation, deriving from post-market rituximab use. PML has been reported following rituximab use in malignancy as well as SLE, RA, and autoimmune cytopenias [104, 105]. Hepatitis B reactivation has mainly been seen with combined chemotherapy regimens for hematologic malignancy with fatalities reported [105]. *Pneumocystis jiroveci* infections have also been seen with combination regimens for malignancy and with rituximab monotherapy for autoimmune conditions, illustrating the importance of B cells in induction of specific T-cell immunity[105]. This is analogous to X-linked agammaglobulinemia, or CD40-ligand deficiency, where defective B cell antigen presentation results in combined immunodeficiency. As only a small number of patients were affected by opportunistic infectious adverse events.

As noted previously, it is thought that in some SLE patients BAFF drives rapid reconstitution of autoreactive B cells following rituximab monotherapy [97]. BAFF is a TNF-superfamily member secreted by multiple cell types (T follicular helper cells, monocytes, and dendritic cells) that is critical for maintaining B cell homeostasis [106–108, 97]. Elevated plasma BAFF levels have been correlated with increased disease activity in SLE and return of anti-dsDNA antibodies following rituximab treatment [106, 97]. Belimumab, a human recombinant IgG targeting soluble BAFF only, is approved for the treatment of SLE. Data from trials demonstrated unchanged memory B cell populations, decreased naïve B cells, and decreased activated plasma cells. Retrospective analysis of a small cohort showed unchanged titers to tetanus, diphtheria, and pneumococcus, but perhaps decreased response to *de novo* antigen challenge (seasonal influenza vaccination)[109, 110]. Investigation into the immune consequences of BAFF inhibitor use in autoimmune disease is ongoing and include studies examining vaccine responses while on belimumab [111]. Long term follow

up data of SLE patients over four years of belimumab use (approximately half on concomitant steroid or mycophenolate) has been reassuring as rates of infection with belimumab were similar to those using systemic immunosuppressant alone (i.e. corticosteroids, antimalarial agents, azathioprine, mycophenolate mofetil, methotrexate, or leflunomide alone or in combination) [112]. Trials investigating combination therapy with rituximab followed by belimumab for prevention of B cell reconstitution in SLE and lupus nephritis are ongoing [97]. However, it remains unclear if combined anti-B cell therapy may create synergistic immunosuppressive effects such as prolonged hypogammaglobulinemia, increased incidence of PML, or other unexpected immunosuppression. Tabalumab and blisibimod (fusion peptide antagonist), two additional biologics that target soluble and membrane-bound BAFF, were examined in initial clinical trials for efficacy in SLE [108]. Despite initial positive data, further development of tabalumab was halted by its manufacturer due to perceived low therapeutic window compared to placebo. Blisibimod is currently in trials for SLE [113].

Atacicept is a human recombinant fusion protein containing the extracellular TACI receptor fused to IgG1 Fc and neutralizes BAFF and A PRoliferation Inducing Ligand (APRIL). APRIL is a TNF superfamily member of cytokines which acts in conjunction with BAFF by binding TACI, B-cell maturation antigen receptors on B cells to promote B cell development and proliferation [108]. Atacicept failed to meet 20% improvement criteria for RA, and in trials for MS, a paradoxical worsening of CNS lesions was seen [114]. During SLE trials, atacicept increased rates of opportunistic infection compared to placebo, including *Haemophilus, Legionella*, and *Klebsiella* [115]. Atacicept also significantly decreased IgG levels compared to placebo and this correlated with increased rates of invasive infections, some of which were fatal [115]. These factors led to discontinuation of trials for this biologic.

Conclusions

Biologics have vastly increased the number of effective treatments in autoimmune conditions. This has significantly improved treatment of patients with conditions that were previously difficult to treat or required corticosteroids which itself comes with numerous side effects. However, the success of these therapies must be tempered by the immunosuppressive effects of these agents. There can be predicted adverse outcomes as well as off target effects which have come to light with some biologics. As immunologists, we should be aware of these possible immunological safety issues with biologics as they continue to enter common use. This is particularly important as there is a dizzying array of biologics for not only autoimmune diseases (Table 2.), but also for oncologic and metabolic inflammatory diseases that are currently entering the therapeutic pipeline.

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References

Papers of particular interest, published recently, have been highlighted as:

• Of importance

• Of major importance

- ••1. Her M, Kavanaugh A. Alterations in immune function with biologic therapies for autoimmune disease. The Journal of allergy and clinical immunology. 2016; 137(1):19–27. A concise review of unintended consequences of biologic therapies, organized by the complication rather than the pathway altered by the therapy. Particularly highlighting TNFis and to a lesser extent rituximab. DOI: 10.1016/j.jaci.2015.10.023. [PubMed: 26768759]
- Timlin H, Bingham CO 3rd. Efficacy and safety implications of molecular constructs of biological agents for rheumatoid arthritis. Expert opinion on biological therapy. 2014; 14(7):893–904. DOI: 10.1517/14712598.2014.900536 [PubMed: 24720727]
- Schejbel L, Fadnes D, Permin H, Lappegard KT, Garred P, Mollnes TE. Primary complement C5 deficiencies - molecular characterization and clinical review of two families. Immunobiology. 2013; 218(10):1304–10. DOI: 10.1016/j.imbio.2013.04.021 [PubMed: 23743184]
- Ricklin D, Hajishengallis G, Yang K, Lambris JD. Complement: a key system for immune surveillance and homeostasis. Nature immunology. 2010; 11(9):785–97. DOI: 10.1038/ni.1923 [PubMed: 20720586]
- Wong EK, Kavanagh D. Anticomplement C5 therapy with eculizumab for the treatment of paroxysmal nocturnal hemoglobinuria and atypical hemolytic uremic syndrome. Translational research : the journal of laboratory and clinical medicine. 2015; 165(2):306–20. DOI: 10.1016/j.trsl. 2014.10.010 [PubMed: 25468487]
- Mastellos DC, Ricklin D, Yancopoulou D, Risitano A, Lambris JD. Complement in paroxysmal nocturnal hemoglobinuria: exploiting our current knowledge to improve the treatment landscape. Expert review of hematology. 2014; 7(5):583–98. DOI: 10.1586/17474086.2014.953926 [PubMed: 25213458]
- Brzustewicz E, Bryl E. The role of cytokines in the pathogenesis of rheumatoid arthritis--Practical and potential application of cytokines as biomarkers and targets of personalized therapy. Cytokine. 2015; 76(2):527–36. DOI: 10.1016/j.cyto.2015.08.260 [PubMed: 26321413]
- •8. Dinarello CA, Simon A, van der Meer JW. Treating inflammation by blocking interleukin-1 in a broad spectrum of diseases. Nature reviews Drug discovery. 2012; 11(8):633–52. Good review of the role of IL-1 in inflammation and broad range of conditions where IL-1 blockade is being considered for therapy. DOI: 10.1038/nrd3800 [PubMed: 22850787]
- Sims JE, Smith DE. The IL-1 family: regulators of immunity. Nature reviews Immunology. 2010; 10(2):89–102. DOI: 10.1038/nri2691
- Kone-Paut I, Galeotti C. Current treatment recommendations and considerations for cryopyrinassociated periodic syndrome. Expert review of clinical immunology. 2015; 11(10):1083–92. DOI: 10.1586/1744666X.2015.1077702 [PubMed: 26312542]
- Lachmann HJ, Kone-Paut I, Kuemmerle-Deschner JB, Leslie KS, Hachulla E, Quartier P, et al. Use of canakinumab in the cryopyrin-associated periodic syndrome. The New England journal of medicine. 2009; 360(23):2416–25. DOI: 10.1056/NEJMoa0810787 [PubMed: 19494217]
- •12. Hunter CA, Jones SA. IL-6 as a keystone cytokine in health and disease. Nature immunology. 2015; 16(5):448–57. Good review of IL-6 biology particularly cis and trans signaling and applicability of anti-IL-6 biologics to therapy. DOI: 10.1038/ni.3153 [PubMed: 25898198]
- Yao X, Huang J, Zhong H, Shen N, Faggioni R, Fung M, et al. Targeting interleukin-6 in inflammatory autoimmune diseases and cancers. Pharmacology & therapeutics. 2014; 141(2):125– 39. DOI: 10.1016/j.pharmthera.2013.09.004 [PubMed: 24076269]
- Kang S, Tanaka T, Kishimoto T. Therapeutic uses of anti-interleukin-6 receptor antibody. International immunology. 2015; 27(1):21–9. DOI: 10.1093/intimm/dxu081 [PubMed: 25142313]
- Rubbert-Roth A. Assessing the safety of biologic agents in patients with rheumatoid arthritis. Rheumatology (Oxford). 2012; 51(Suppl 5):v38–47. DOI: 10.1093/rheumatology/kes114 [PubMed: 22718926]
- 16. Campbell L, Chen C, Bhagat SS, Parker RA, Ostor AJ. Risk of adverse events including serious infections in rheumatoid arthritis patients treated with tocilizumab: a systematic literature review

and meta-analysis of randomized controlled trials. Rheumatology (Oxford). 2011; 50(3):552–62. DOI: 10.1093/rheumatology/keq343 [PubMed: 21078627]

- 17. Horneff G. Safety of biologic therapies for the treatment of juvenile idiopathic arthritis. Expert opinion on drug safety. 2015; 14(7):1111–26. DOI: 10.1517/14740338.2015.1042453 [PubMed: 26084637]
- Gaffen SL, Jain R, Garg AV, Cua DJ. The IL-23-IL-17 immune axis: from mechanisms to therapeutic testing. Nature reviews Immunology. 2014; 14(9):585–600. DOI: 10.1038/nri3707
- •19. Puel A, Cypowyj S, Marodi L, Abel L, Picard C, Casanova JL. Inborn errors of human IL-17 immunity underlie chronic mucocutaneous candidiasis. Current opinion in allergy and clinical immunology. 2012; 12(6):616–22. Examination of the effect of genetic mutations in the IL-17 pathway in causing chronic mucocutaneous candidiasis in patients. DOI: 10.1097/ACI. 0b013e328358cc0b [PubMed: 23026768]
- Gopal R, Monin L, Slight S, Uche U, Blanchard E, Fallert Junecko BA, et al. Unexpected role for IL-17 in protective immunity against hypervirulent Mycobacterium tuberculosis HN878 infection. PLoS pathogens. 2014; 10(5):e1004099.doi: 10.1371/journal.ppat.1004099 [PubMed: 24831696]
- 21. Paul C, Lacour JP, Tedremets L, Kreutzer K, Jazayeri S, Adams S, et al. Efficacy, safety and usability of secukinumab administration by autoinjector/pen in psoriasis: a randomized, controlled trial (JUNCTURE). Journal of the European Academy of Dermatology and Venereology : JEADV. 2015; 29(6):1082–90. DOI: 10.1111/jdv.12751 [PubMed: 25243910]
- 22. Ohtsuki M, Morita A, Abe M, Takahashi H, Seko N, Karpov A, et al. Secukinumab efficacy and safety in Japanese patients with moderate-to-severe plaque psoriasis: subanalysis from ERASURE, a randomized, placebo-controlled, phase 3 study. The Journal of dermatology. 2014; 41(12):1039–46. DOI: 10.1111/1346-8138.12668 [PubMed: 25354738]
- Langley RG, Elewski BE, Lebwohl M, Reich K, Griffiths CE, Papp K, et al. Secukinumab in plaque psoriasis--results of two phase 3 trials. The New England journal of medicine. 2014; 371(4):326–38. DOI: 10.1056/NEJMoa1314258 [PubMed: 25007392]
- 24. Blauvelt A, Prinz JC, Gottlieb AB, Kingo K, Sofen H, Ruer-Mulard M, et al. Secukinumab administration by pre-filled syringe: efficacy, safety and usability results from a randomized controlled trial in psoriasis (FEATURE). The British journal of dermatology. 2015; 172(2):484–93. DOI: 10.1111/bjd.13348 [PubMed: 25132411]
- Baeten D, Sieper J, Braun J, Baraliakos X, Dougados M, Emery P, et al. Secukinumab, an Interleukin-17A Inhibitor, in Ankylosing Spondylitis. The New England journal of medicine. 2015; 373(26):2534–48. DOI: 10.1056/NEJMoa1505066 [PubMed: 26699169]
- 26. McInnes IB, Mease PJ, Kirkham B, Kavanaugh A, Ritchlin CT, Rahman P, et al. Secukinumab, a human anti-interleukin-17A monoclonal antibody, in patients with psoriatic arthritis (FUTURE 2): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet. 2015; 386(9999):1137–46. DOI: 10.1016/S0140-6736(15)61134-5 [PubMed: 26135703]
- 27. Mease PJ, McInnes IB, Kirkham B, Kavanaugh A, Rahman P, van der Heijde D, et al. Secukinumab Inhibition of Interleukin-17A in Patients with Psoriatic Arthritis. The New England journal of medicine. 2015; 373(14):1329–39. DOI: 10.1056/NEJMoa1412679 [PubMed: 26422723]
- •28. Quach OL, Hsu S. Perianal Dermatophytosis During Secukinumab Therapy for Plaque Psoriasis. JAMA dermatology. 2015; :1–2. 2 cases of perianal dermatophytosis with secukinumab and summary of candida infections in the relevant trials. DOI: 10.1001/jamadermatol.2015.4992
- Hueber W, Sands BE, Lewitzky S, Vandemeulebroecke M, Reinisch W, Higgins PD, et al. Secukinumab, a human anti-IL-17A monoclonal antibody, for moderate to severe Crohn's disease: unexpected results of a randomised, double-blind placebo-controlled trial. Gut. 2012; 61(12): 1693–700. DOI: 10.1136/gutjnl-2011-301668 [PubMed: 22595313]
- Thaci D, Blauvelt A, Reich K, Tsai TF, Vanaclocha F, Kingo K, et al. Secukinumab is superior to ustekinumab in clearing skin of subjects with moderate to severe plaque psoriasis: CLEAR, a randomized controlled trial. Journal of the American Academy of Dermatology. 2015; 73(3):400– 9. DOI: 10.1016/j.jaad.2015.05.013 [PubMed: 26092291]
- Kleinewietfeld M, Manzel A, Titze J, Kvakan H, Yosef N, Linker RA, et al. Sodium chloride drives autoimmune disease by the induction of pathogenic TH17 cells. Nature. 2013; 496(7446):518–22. DOI: 10.1038/nature11868 [PubMed: 23467095]

- Bustamante J, Boisson-Dupuis S, Abel L, Casanova JL. Mendelian susceptibility to mycobacterial disease: genetic, immunological, and clinical features of inborn errors of IFN-gamma immunity. Seminars in immunology. 2014; 26(6):454–70. DOI: 10.1016/j.smim.2014.09.008 [PubMed: 25453225]
- 33. Papp KA, Langley RG, Lebwohl M, Krueger GG, Szapary P, Yeilding N, et al. Efficacy and safety of ustekinumab, a human interleukin-12/23 monoclonal antibody, in patients with psoriasis: 52week results from a randomised, double-blind, placebo-controlled trial (PHOENIX 2). Lancet. 2008; 371(9625):1675–84. DOI: 10.1016/S0140-6736(08)60726-6 [PubMed: 18486740]
- Leonardi CL, Kimball AB, Papp KA, Yeilding N, Guzzo C, Wang Y, et al. Efficacy and safety of ustekinumab, a human interleukin-12/23 monoclonal antibody, in patients with psoriasis: 76-week results from a randomised, double-blind, placebo-controlled trial (PHOENIX 1). Lancet. 2008; 371(9625):1665–74. DOI: 10.1016/S0140-6736(08)60725-4 [PubMed: 18486739]
- Griffiths CE, Strober BE, van de Kerkhof P, Ho V, Fidelus-Gort R, Yeilding N, et al. Comparison of ustekinumab and etanercept for moderate-to-severe psoriasis. The New England journal of medicine. 2010; 362(2):118–28. DOI: 10.1056/NEJMoa0810652 [PubMed: 20071701]
- 36. Kalb RE, Fiorentino DF, Lebwohl MG, Toole J, Poulin Y, Cohen AD, et al. Risk of Serious Infection With Biologic and Systemic Treatment of Psoriasis: Results From the Psoriasis Longitudinal Assessment and Registry (PSOLAR). JAMA dermatology. 2015; 151(9):961–9. DOI: 10.1001/jamadermatol.2015.0718 [PubMed: 25970800]
- Papp KA, Griffiths CE, Gordon K, Lebwohl M, Szapary PO, Wasfi Y, et al. Long-term safety of ustekinumab in patients with moderate-to-severe psoriasis: final results from 5 years of follow-up. The British journal of dermatology. 2013; 168(4):844–54. DOI: 10.1111/bjd.12214 [PubMed: 23301632]
- 38. Tsai TF, Ho JC, Song M, Szapary P, Guzzo C, Shen YK, et al. Efficacy and safety of ustekinumab for the treatment of moderate-to-severe psoriasis: a phase III, randomized, placebo-controlled trial in Taiwanese and Korean patients (PEARL). Journal of dermatological science. 2011; 63(3):154– 63. DOI: 10.1016/j.jdermsci.2011.05.005 [PubMed: 21741220]
- Igarashi A, Kato T, Kato M, Song M, Nakagawa H. Efficacy and safety of ustekinumab in Japanese patients with moderate-to-severe plaque-type psoriasis: long-term results from a phase 2/3 clinical trial. The Journal of dermatology. 2012; 39(3):242–52. DOI: 10.1111/j.1346-8138.2011.01347.x [PubMed: 21955098]
- 40. Tsai TF, Chiu HY, Song M, Chan D. A case of latent tuberculosis reactivation in a patient treated with ustekinumab without concomitant isoniazid chemoprophylaxis in the PEARL trial. The British journal of dermatology. 2013; 168(2):444–6. DOI: 10.1111/j.1365-2133.2012.11162.x [PubMed: 22816505]
- Kappos L, Wiendl H, Selmaj K, Arnold DL, Havrdova E, Boyko A, et al. Daclizumab HYP versus Interferon Beta-1a in Relapsing Multiple Sclerosis. The New England journal of medicine. 2015; 373(15):1418–28. DOI: 10.1056/NEJMoa1501481 [PubMed: 26444729]
- 42. Iversen M, Burton CM, Vand S, Skovfoged L, Carlsen J, Milman N, et al. Aspergillus infection in lung transplant patients: incidence and prognosis. European journal of clinical microbiology & infectious diseases : official publication of the European Society of Clinical Microbiology. 2007; 26(12):879–86. DOI: 10.1007/s10096-007-0376-3
- 43. Wynn D, Kaufman M, Montalban X, Vollmer T, Simon J, Elkins J, et al. Daclizumab in active relapsing multiple sclerosis (CHOICE study): a phase 2, randomised, double-blind, placebocontrolled, add-on trial with interferon beta. The Lancet Neurology. 2010; 9(4):381–90. DOI: 10.1016/S1474-4422(10)70033-8 [PubMed: 20163990]
- 44. Gold R, Giovannoni G, Selmaj K, Havrdova E, Montalban X, Radue EW, et al. Daclizumab highyield process in relapsing-remitting multiple sclerosis (SELECT): a randomised, double-blind, placebo-controlled trial. Lancet. 2013; 381(9884):2167–75. DOI: 10.1016/ S0140-6736(12)62190-4 [PubMed: 23562009]
- Nashan B, Moore R, Amlot P, Schmidt AG, Abeywickrama K, Soulillou JP. Randomised trial of basiliximab versus placebo for control of acute cellular rejection in renal allograft recipients. CHIB 201 International Study Group. Lancet. 1997; 350(9086):1193–8. [PubMed: 9652559]

- McKeage K, McCormack PL. Basiliximab: a review of its use as induction therapy in renal transplantation. BioDrugs : clinical immunotherapeutics, biopharmaceuticals and gene therapy. 2010; 24(1):55–76. DOI: 10.2165/11203990-00000000-00000
- Hardinger KL, Brennan DC, Klein CL. Selection of induction therapy in kidney transplantation. Transplant international : official journal of the European Society for Organ Transplantation. 2013; 26(7):662–72. DOI: 10.1111/tri.12043 [PubMed: 23279211]
- Perales MA, Ishill N, Lomazow WA, Weinstock DM, Papadopoulos EB, Dastigir H, et al. Longterm follow-up of patients treated with daclizumab for steroid-refractory acute graft-vs-host disease. Bone marrow transplantation. 2007; 40(5):481–6. DOI: 10.1038/sj.bmt.1705762 [PubMed: 17618322]
- Morris JA, Hanson JE, Steffen BJ, Chu AH, Chi-Burris KS, Gotz VP, et al. Daclizumab is associated with decreased rejection and improved patient survival in renal transplant recipients. Clinical transplantation. 2005; 19(3):340–5. DOI: 10.1111/j.1399-0012.2005.00344.x [PubMed: 15877795]
- Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and its ligands in tolerance and immunity. Annual review of immunology. 2008; 26:677–704. DOI: 10.1146/annurev.immunol. 26.021607.090331
- *51. Wells AF, Jodat N, Schiff M. A critical evaluation of the role of subcutaneous abatacept in the treatment of rheumatoid arthritis: patient considerations. Biologics: targets & therapy. 2014; 8:41–55. Summary of subcutaneous abatacept data, collating multiple clinical trials. DOI: 10.2147/BTT.S55783 [PubMed: 24600202]
- 52. Weinblatt ME, Schiff M, Valente R, van der Heijde D, Citera G, Zhao C, et al. Head-to-head comparison of subcutaneous abatacept versus adalimumab for rheumatoid arthritis: findings of a phase IIIb, multinational, prospective, randomized study. Arthritis and rheumatism. 2013; 65(1): 28–38. DOI: 10.1002/art.37711 [PubMed: 23169319]
- *53. Weinblatt ME, Moreland LW, Westhovens R, Cohen RB, Kelly SM, Khan N, et al. Safety of abatacept administered intravenously in treatment of rheumatoid arthritis: integrated analyses of up to 8 years of treatment from the abatacept clinical trial program. The Journal of rheumatology. 2013; 40(6):787–97. Summary of IV abatacept data, collating multiple clinical trials. DOI: 10.3899/jrheum.120906 [PubMed: 23588946]
- 54. Weinblatt M, Combe B, Covucci A, Aranda R, Becker JC, Keystone E. Safety of the selective costimulation modulator abatacept in rheumatoid arthritis patients receiving background biologic and nonbiologic disease-modifying antirheumatic drugs: A one-year randomized, placebo-controlled study. Arthritis and rheumatism. 2006; 54(9):2807–16. DOI: 10.1002/art.22070 [PubMed: 16947384]
- 55. Schiff M, Pritchard C, Huffstutter JE, Rodriguez-Valverde V, Durez P, Zhou X, et al. The 6-month safety and efficacy of abatacept in patients with rheumatoid arthritis who underwent a washout after anti-tumour necrosis factor therapy or were directly switched to abatacept: the ARRIVE trial. Annals of the rheumatic diseases. 2009; 68(11):1708–14. DOI: 10.1136/ard.2008.099218 [PubMed: 19074911]
- 56. Schiff M, Keiserman M, Codding C, Songcharoen S, Berman A, Nayiager S, et al. Efficacy and safety of abatacept or infliximab vs placebo in ATTEST: a phase III, multi-centre, randomised, double-blind, placebo-controlled study in patients with rheumatoid arthritis and an inadequate response to methotrexate. Annals of the rheumatic diseases. 2008; 67(8):1096–103. DOI: 10.1136/ ard.2007.080002 [PubMed: 18055472]
- 57. Nash P, Nayiager S, Genovese MC, Kivitz AJ, Oelke K, Ludivico C, et al. Immunogenicity, safety, and efficacy of abatacept administered subcutaneously with or without background methotrexate in patients with rheumatoid arthritis: results from a phase III, international, multicenter, parallel-arm, open-label study. Arthritis care & research. 2013; 65(5):718–28. DOI: 10.1002/acr.21876 [PubMed: 23097311]
- Kremer JM, Genant HK, Moreland LW, Russell AS, Emery P, Abud-Mendoza C, et al. Effects of abatacept in patients with methotrexate-resistant active rheumatoid arthritis: a randomized trial. Annals of internal medicine. 2006; 144(12):865–76. [PubMed: 16785475]
- 59. Keystone EC, Kremer JM, Russell A, Box J, Abud-Mendoza C, Elizondo MG, et al. Abatacept in subjects who switch from intravenous to subcutaneous therapy: results from the phase IIIb

ATTUNE study. Annals of the rheumatic diseases. 2012; 71(6):857–61. DOI: 10.1136/ annrheumdis-2011-200355 [PubMed: 22302417]

- 60. Kaine J, Gladstein G, Strusberg I, Robles M, Louw I, Gujrathi S, et al. Evaluation of abatacept administered subcutaneously in adults with active rheumatoid arthritis: impact of withdrawal and reintroduction on immunogenicity, efficacy and safety (phase Iiib ALLOW study). Annals of the rheumatic diseases. 2012; 71(1):38–44. DOI: 10.1136/annrheumdis-2011-200344 [PubMed: 21917824]
- 61. Genovese MC, Schiff M, Luggen M, Becker JC, Aranda R, Teng J, et al. Efficacy and safety of the selective co-stimulation modulator abatacept following 2 years of treatment in patients with rheumatoid arthritis and an inadequate response to anti-tumour necrosis factor therapy. Annals of the rheumatic diseases. 2008; 67(4):547–54. DOI: 10.1136/ard.2007.074773 [PubMed: 17921185]
- 62. Genovese MC, Covarrubias A, Leon G, Mysler E, Keiserman M, Valente R, et al. Subcutaneous abatacept versus intravenous abatacept: a phase IIIb noninferiority study in patients with an inadequate response to methotrexate. Arthritis and rheumatism. 2011; 63(10):2854–64. DOI: 10.1002/art.30463 [PubMed: 21618201]
- 63. Schiff M, Keiserman M, Codding C, Songcharoen S, Berman A, Nayiager S, et al. Clinical response and tolerability to abatacept in patients with rheumatoid arthritis previously treated with infliximab or abatacept: open-label extension of the ATTEST Study. Annals of the rheumatic diseases. 2011; 70(11):2003–7. DOI: 10.1136/annrheumdis-2011-200316 [PubMed: 21914628]
- 64. Ruperto N, Lovell DJ, Quartier P, Paz E, Rubio-Perez N, Silva CA, et al. Abatacept in children with juvenile idiopathic arthritis: a randomised, double-blind, placebo-controlled withdrawal trial. Lancet. 2008; 372(9636):383–91. DOI: 10.1016/S0140-6736(08)60998-8 [PubMed: 18632147]
- 65. Kremer JM, Westhovens R, Leon M, Di Giorgio E, Alten R, Steinfeld S, et al. Treatment of rheumatoid arthritis by selective inhibition of T-cell activation with fusion protein CTLA4Ig. The New England journal of medicine. 2003; 349(20):1907–15. DOI: 10.1056/NEJMoa035075 [PubMed: 14614165]
- 66. Schiff M, Weinblatt ME, Valente R, van der Heijde D, Citera G, Elegbe A, et al. Head-to-head comparison of subcutaneous abatacept versus adalimumab for rheumatoid arthritis: two-year efficacy and safety findings from AMPLE trial. Annals of the rheumatic diseases. 2014; 73(1):86–94. DOI: 10.1136/annrheumdis-2013-203843 [PubMed: 23962455]
- 67. Alten R, Kaine J, Keystone E, Nash P, Delaet I, Genovese MC. Long-term safety of subcutaneous abatacept in rheumatoid arthritis: integrated analysis of clinical trial data representing more than four years of treatment. Arthritis Rheumatol. 2014; 66(8):1987–97. DOI: 10.1002/art.38687 [PubMed: 24782324]
- 68. Salliot C, Dougados M, Gossec L. Risk of serious infections during rituximab, abatacept and anakinra treatments for rheumatoid arthritis: meta-analyses of randomised placebo-controlled trials. Annals of the rheumatic diseases. 2009; 68(1):25–32. DOI: 10.1136/ard.2007.083188 [PubMed: 18203761]
- 69. Lee S, Moon JS, Lee CR, Kim HE, Baek SM, Hwang S, et al. Abatacept alleviates severe autoimmune symptoms in a patient carrying a de novo variant in CTLA-4. The Journal of allergy and clinical immunology. 2016; 137(1):327–30. DOI: 10.1016/j.jaci.2015.08.036 [PubMed: 26478010]
- 70. Lo B, Zhang K, Lu W, Zheng L, Zhang Q, Kanellopoulou C, et al. AUTOIMMUNE DISEASE. Patients with LRBA deficiency show CTLA4 loss and immune dysregulation responsive to abatacept therapy. Science. 2015; 349(6246):436–40. DOI: 10.1126/science.aaa1663 [PubMed: 26206937]
- Vincenti F, Rostaing L, Grinyo J, Rice K, Steinberg S, Gaite L, et al. Belatacept and Long-Term Outcomes in Kidney Transplantation. The New England journal of medicine. 2016; 374(4):333– 43. DOI: 10.1056/NEJMoa1506027 [PubMed: 26816011]
- 72. Vincenti F, Charpentier B, Vanrenterghem Y, Rostaing L, Bresnahan B, Darji P, et al. A phase III study of belatacept-based immunosuppression regimens versus cyclosporine in renal transplant recipients (BENEFIT study). American journal of transplantation : official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2010; 10(3):535–46. DOI: 10.1111/j.1600-6143.2009.03005.x

- 73. Durrbach A, Pestana JM, Pearson T, Vincenti F, Garcia VD, Campistol J, et al. A phase III study of belatacept versus cyclosporine in kidney transplants from extended criteria donors (BENEFIT-EXT study). American journal of transplantation : official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2010; 10(3):547–57. DOI: 10.1111/j.1600-6143.2010.03016.x
- 74. Rostaing L, Vincenti F, Grinyo J, Rice KM, Bresnahan B, Steinberg S, et al. Long-term belatacept exposure maintains efficacy and safety at 5 years: results from the long-term extension of the BENEFIT study. American journal of transplantation : official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2013; 13(11):2875–83. DOI: 10.1111/ajt.12460
- 75. Archdeacon P, Dixon C, Belen O, Albrecht R, Meyer J. Summary of the US FDA approval of belatacept. American journal of transplantation : official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2012; 12(3):554–62. DOI: 10.1111/j.1600-6143.2011.03976.x
- Wherry EJ, Kurachi M. Molecular and cellular insights into T cell exhaustion. Nature reviews Immunology. 2015; 15(8):486–99. DOI: 10.1038/nri3862
- 77. Tivol EA, Borriello F, Schweitzer AN, Lynch WP, Bluestone JA, Sharpe AH. Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4. Immunity. 1995; 3(5):541–7. [PubMed: 7584144]
- Kuehn HS, Ouyang W, Lo B, Deenick EK, Niemela JE, Avery DT, et al. Immune dysregulation in human subjects with heterozygous germline mutations in CTLA4. Science. 2014; 345(6204): 1623–7. DOI: 10.1126/science.1255904 [PubMed: 25213377]
- 79. Robert C, Thomas L, Bondarenko I, O'Day S, Weber J, Garbe C, et al. Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. The New England journal of medicine. 2011; 364(26):2517–26. DOI: 10.1056/NEJMoa1104621 [PubMed: 21639810]
- Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, et al. Improved survival with ipilimumab in patients with metastatic melanoma. The New England journal of medicine. 2010; 363(8):711–23. DOI: 10.1056/NEJMoa1003466 [PubMed: 20525992]
- Buchbinder E, Hodi FS. Cytotoxic T lymphocyte antigen-4 and immune checkpoint blockade. The Journal of clinical investigation. 2015; 125(9):3377–83. DOI: 10.1172/JCI80012 [PubMed: 26325034]
- *82. Weber JS, Kahler KC, Hauschild A. Management of immune-related adverse events and kinetics of response with ipilimumab. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2012; 30(21):2691–7. Description of the management of the immune-related adverse events (irAE) with ipilimumab therapy. DOI: 10.1200/JCO. 2012.41.6750 [PubMed: 22614989]
- Lipson EJ, Drake CG. Ipilimumab: an anti-CTLA-4 antibody for metastatic melanoma. Clinical cancer research : an official journal of the American Association for Cancer Research. 2011; 17(22):6958–62. DOI: 10.1158/1078-0432.CCR-11-1595 [PubMed: 21900389]
- 84. Wolchok JD, Hodi FS, Weber JS, Allison JP, Urba WJ, Robert C, et al. Development of ipilimumab: a novel immunotherapeutic approach for the treatment of advanced melanoma. Annals of the New York Academy of Sciences. 2013; 1291:1–13. DOI: 10.1111/nyas.12180 [PubMed: 23772560]
- *85. van de Vijver E, van den Berg TK, Kuijpers TW. Leukocyte adhesion deficiencies. Hematology/ oncology clinics of North America. 2013; 27(1):101–16. viii. Useful review of leukocyte adhesion cascade. DOI: 10.1016/j.hoc.2012.10.001 [PubMed: 23351991]
- Ghosh N, Chaki R, Mandal SC. Inhibition of selective adhesion molecules in treatment of inflammatory bowel disease. International reviews of immunology. 2012; 31(5):410–27. DOI: 10.3109/08830185.2012.690794 [PubMed: 23083349]
- Engelhardt B. Molecular mechanisms involved in T cell migration across the blood-brain barrier. J Neural Transm (Vienna). 2006; 113(4):477–85. DOI: 10.1007/s00702-005-0409-y [PubMed: 16550326]
- *88. McGuigan C, Craner M, Guadagno J, Kapoor R, Mazibrada G, Molyneux P, et al. Stratification and monitoring of natalizumab-associated progressive multifocal leukoencephalopathy risk: recommendations from an expert group. Journal of neurology, neurosurgery, and psychiatry.

2016; 87(2):117–25. Clinical algorithm for stratifying risk of PML and defining monitoring paramaters for patients on natalizumab. DOI: 10.1136/jnnp-2015-311100

- Colombel JF, Sands BE, Rutgeerts P, Sandborn W, Danese S, D'Haens G, et al. The safety of vedolizumab for ulcerative colitis and Crohn's disease. Gut. 2016; doi: 10.1136/ gutjnl-2015-311079
- 90. Schwab N, Ulzheimer JC, Fox RJ, Schneider-Hohendorf T, Kieseier BC, Monoranu CM, et al. Fatal PML associated with efalizumab therapy: insights into integrin alphaLbeta2 in JC virus control. Neurology. 2012; 78(7):458–67. discussion 65. DOI: 10.1212/WNL.0b013e3182478d4b [PubMed: 22302546]
- 91. Fine AJ, Sorbello A, Kortepeter C, Scarazzini L. Central nervous system herpes simplex and varicella zoster virus infections in natalizumab-treated patients. Clinical infectious diseases : an official publication of the Infectious Diseases Society of America. 2013; 57(6):849–52. DOI: 10.1093/cid/cit376 [PubMed: 23728144]
- 92. Boland BS, Dulai PS, Chang M, Sandborn WJ, Levesque BG. Pseudomonas Meningitis During Vedolizumab Therapy for Crohn's Disease. The American journal of gastroenterology. 2015; 110(11):1631–2. DOI: 10.1038/ajg.2015.326 [PubMed: 26618431]
- 93. von Budingen HC, Palanichamy A, Lehmann-Horn K, Michel BA, Zamvil SS. Update on the autoimmune pathology of multiple sclerosis: B-cells as disease-drivers and therapeutic targets. European neurology. 2015; 73(3–4):238–46. DOI: 10.1159/000377675 [PubMed: 25824054]
- Jennette JC, Falk RJ. B cell-mediated pathogenesis of ANCA-mediated vasculitis. Seminars in immunopathology. 2014; 36(3):327–38. DOI: 10.1007/s00281-014-0431-y [PubMed: 24777746]
- 95. Chan VS, Tsang HH, Tam RC, Lu L, Lau CS. B-cell-targeted therapies in systemic lupus erythematosus. Cellular & molecular immunology. 2013; 10(2):133–42. DOI: 10.1038/cmi. 2012.64 [PubMed: 23455017]
- 96. Leandro MJ. B-cell subpopulations in humans and their differential susceptibility to depletion with anti-CD20 monoclonal antibodies. Arthritis research & therapy. 2013; 15(Suppl 1):S3.doi: 10.1186/ar3908
- Ehrenstein MR, Wing C. The BAFFling effects of rituximab in lupus: danger ahead? Nature reviews Rheumatology. 2016; doi: 10.1038/nrrheum.2016.18
- 98. McLaughlin P, Grillo-Lopez AJ, Link BK, Levy R, Czuczman MS, Williams ME, et al. Rituximab chimeric anti-CD20 monoclonal antibody therapy for relapsed indolent lymphoma: half of patients respond to a four-dose treatment program. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 1998; 16(8):2825–33. [PubMed: 9704735]
- 99. Maloney DG, Grillo-Lopez AJ, Bodkin DJ, White CA, Liles TM, Royston I, et al. IDEC-C2B8: results of a phase I multiple-dose trial in patients with relapsed non-Hodgkin's lymphoma. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 1997; 15(10): 3266–74. [PubMed: 9336364]
- 100. Makatsori M, Kiani-Alikhan S, Manson AL, Verma N, Leandro M, Gurugama NP, et al. Hypogammaglobulinaemia after rituximab treatment-incidence and outcomes. QJM : monthly journal of the Association of Physicians. 2014; 107(10):821–8. DOI: 10.1093/qjmed/hcu094 [PubMed: 24778295]
- **101. Kaplan B, Kopyltsova Y, Khokhar A, Lam F, Bonagura V. Rituximab and immune deficiency: case series and review of the literature. The journal of allergy and clinical immunology In practice. 2014; 2(5):594–600. Clear explanation of hypogammaglobulinemia secondary to rituximab and management. DOI: 10.1016/j.jaip.2014.06.003 [PubMed: 25213054]
- 102. Casulo C, Maragulia J, Zelenetz AD. Incidence of hypogammaglobulinemia in patients receiving rituximab and the use of intravenous immunoglobulin for recurrent infections. Clinical lymphoma, myeloma & leukemia. 2013; 13(2):106–11. DOI: 10.1016/j.clml.2012.11.011
- 103. Barmettler S, Price C. Continuing IgG replacement therapy for hypogammaglobulinemia after rituximab--for how long? The Journal of allergy and clinical immunology. 2015; 136(5):1407–9. DOI: 10.1016/j.jaci.2015.06.035 [PubMed: 26277594]
- 104. Vermeer NS, Straus SM, Mantel-Teeuwisse AK, Hidalgo-Simon A, Egberts AC, Leufkens HG, et al. Drug-induced progressive multifocal leukoencephalopathy: Lessons learned from contrasting

natalizumab and rituximab. Clinical pharmacology and therapeutics. 2015; 98(5):542–50. DOI: 10.1002/cpt.207 [PubMed: 26347128]

- Gea-Banacloche JC. Rituximab-associated infections. Seminars in hematology. 2010; 47(2):187– 98. DOI: 10.1053/j.seminhematol.2010.01.002 [PubMed: 20350666]
- *106. Vincent FB, Morand EF, Schneider P, Mackay F. The BAFF/APRIL system in SLE pathogenesis. Nature reviews Rheumatology. 2014; 10(6):365–73. Details role of role of BAFF/ April system in pathogenesis of lupus and outlines rationale for role in therapy. DOI: 10.1038/ nrrheum.2014.33 [PubMed: 24614588]
- 107. Vilas-Boas A, Morais SA, Isenberg DA. Belimumab in systemic lupus erythematosus. RMD open. 2015; 1(1):e000011.doi: 10.1136/rmdopen-2014-000011 [PubMed: 26509047]
- 108. Morais SA, Vilas-Boas A, Isenberg DA. B-cell survival factors in autoimmune rheumatic disorders. Therapeutic advances in musculoskeletal disease. 2015; 7(4):122–51. DOI: 10.1177/1759720X15586782 [PubMed: 26288664]
- 109. Stohl W, Hiepe F, Latinis KM, Thomas M, Scheinberg MA, Clarke A, et al. Belimumab reduces autoantibodies, normalizes low complement levels, and reduces select B cell populations in patients with systemic lupus erythematosus. Arthritis and rheumatism. 2012; 64(7):2328–37. DOI: 10.1002/art.34400 [PubMed: 22275291]
- 110. Chatham WW, Wallace DJ, Stohl W, Latinis KM, Manzi S, McCune WJ, et al. Effect of belimumab on vaccine antigen antibodies to influenza, pneumococcal, and tetanus vaccines in patients with systemic lupus erythematosus in the BLISS-76 trial. The Journal of rheumatology. 2012; 39(8):1632–40. DOI: 10.3899/jrheum.111587 [PubMed: 22707609]
- 111. ClinicalTrials.gov. A Study to Evaluate the Effect of Belimumab on Vaccine Responses in Subjects With Systemic Lupus Erythematosus (SLE).
- 112. Merrill JT, Ginzler EM, Wallace DJ, McKay JD, Lisse JR, Aranow C, et al. Long-term safety profile of belimumab plus standard therapy in patients with systemic lupus erythematosus. Arthritis and rheumatism. 2012; 64(10):3364–73. DOI: 10.1002/art.34564 [PubMed: 22674457]
- 113. ClinicalTrials.gov. CHABLIS-SC1: A Study of the Efficacy and Safety of Subcutaneous Blisibimod in Subjects With Systemic Lupus Erythematosus (CHABLIS-SC1).
- 114. Cogollo E, Silva MA, Isenberg D. Profile of atacicept and its potential in the treatment of systemic lupus erythematosus. Drug design, development and therapy. 2015; 9:1331–9. DOI: 10.2147/DDDT.S71276
- 115. Isenberg D, Gordon C, Licu D, Copt S, Rossi CP, Wofsy D. Efficacy and safety of atacicept for prevention of flares in patients with moderate-to-severe systemic lupus erythematosus (SLE): 52week data (APRIL-SLE randomised trial). Annals of the rheumatic diseases. 2015; 74(11):2006– 15. DOI: 10.1136/annrheumdis-2013-205067 [PubMed: 24951103]

Table 1

Currently Approved and Discontinued Biologics

Generic Name	Brand Name	Indications		
Innate Immunity				
Eculizumab	Soliris	Paroxysmal nocturnal hemoglobinuria		
Anakinra	Kineret	Rheumatoid arthritis (RA) Neonatal Onset Multisystem Inflammatory Disease		
Canakinumab	Ilaris	Familial cold autoinflammatory syndrome Muckle-Wells syndrome Systemic juvenile idiopathic arthritis (SJIA)		
Rilonacept	Arcalyst	Cryopyrin associated periodic syndromes		
Tocilizumab	Actemra	RA, SJIA, Polyarticular juvenile idiopathic arthritis		
Adaptive Immunity				
Secukinumab	Cosentyx	Plaque psoriasis, Psoriatic arthritis Ankylosing spondylitis		
Ustekinumab	Stelara	Plaque psoriasis, Psoriatic arthritis		
T cells				
Basiliximab	Simulect	Renal transplant rejection		
Abatacept	Orencia	RA, Juvenile idiopathic arthritis (JIA)		
Belatacept	Nulojix	Renal transplant rejection		
Ipilimumab	Yervoy	Melanoma, unresectable or metastatic; adjuvant		
Adhesion Molecules				
Natalizumab	Tysabri	Multiple sclerosis Ankylosing spondylitis Crohn's disease (CD)		
Vedolizumab	Entyvio	Inflammatory bowel disease (CD, ulcerative colitis)		
B cells				
Rituximab	Rituxan	RA Granulomatous polyangiitis, Microscopic polyangiitis		
Belimumab	Benlysta	Systemic lupus erythematosus		
Discontinued				
Daclizumab		Renal transplant rejection		
Efalizumab		Plaque psoriasis		
Atacicept		Systemic lupus erythematosus		

Table 2

Biologics in phase II/III trials

Generic Name	Target	Trial Indications
Gevokizumab	IL-1β	Diabetes, type II
Sifalimumab	IFN-a	Systemic lupus erythematosus
Rontalizumab	IFN-a	Systemic lupus erythematosus
Clazakizumab	IL-6	Rheumatoid arthritis
Sirutumab	IL-6	Rheumatoid arthritis Depression
Mavrilimumab	GM-CSFRa	Rheumatoid arthritis
Ixekizumab	IL-17	Plaque psoriasis
Brodalumab	IL-17R	Plaque psoriasis
Tildrakizumab	IL-23	Plaque psoriasis
Guselkumab	IL-23 p19	Plaque psoriasis
Oteliximab	CD3e	Diabetes, type I
Teplizumab	CD3	Diabetes, type I
Visilizumab	CD3	Inflammatory bowel disease
Blisibimod	BAFF	Systemic lupus erythematosus
Tabalumab	BAFF	Systemic lupus erythematosus