

Dynamic analysis of MCF-10A and MCF-7: A simulation approach

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Abstract. Cancer is one of the leading causes of death in the world. Breast cancer is the most common form of cancer among women and is responsible for 15 % of all cancer related deaths. Though there are significant advancements in cancer treatment strategies, clinical tumour treatment methods currently employed are often accompanied by severe side effects as they induce damage to the normal cells along with the cancer cells. The alterations in the biophysical and biomechanical properties of a cell as it undergoes transformation from a normal to cancerous cell results in changes in its dynamic characteristics. These changes can be utilised to induce selective cytotoxicity of tumour cells. In the present study, two simulation models (homogenous and non-homogenous) of normal (MCF-10A) and cancerous (MCF-7) breast cells are developed. A finite element approach using Ansys is adopted to investigate the variation in dynamic characteristics of the cells using the two modelling approaches. Results indicate that the natural frequencies of cells modelled as a homogenous system is greater than that of cells whose sub-cellular material properties are considered for analysis. A comparison of the first four natural frequencies using the two modelling approaches for both MCF-10A and MCF-7 cells are illustrated and the corresponding mode shapes reported. A plot highlighting the variation in the natural frequencies of MCF-10A and MCF-7 using the two modelling approaches is presented.

Keywords: MCF-10A, MCF-7, cancer, simulation, dynamics, natural frequency, mode shapes.

1. Introduction

The human body contains trillions of cells which grow, divide and die in a conventional manner. Replication of DNA during the interphase stage forms a crucial step in cell division. An error during the replication process induces changes in the genome of the cell, giving rise to mutations. Some mutations result in uncontrolled, uncoordinated and undesirable cell division leading to cancer. The World Health Organisation (WHO) estimates that breast cancer is the most common form of cancer affecting 2.1 million women each year and is responsible for 15 % of all cancer related deaths. The treatment methods currently employed to treat cancer include chemotherapy, radiation therapy, surgery, targeted therapy and immunotherapy. Radiotherapy and chemotherapy are known to induce damage to healthy cells surrounding the tumour, giving rise to numerous side effects which negatively impacts the quality of life of a patient. The transformation of a normal cell to cancerous cell is accompanied by various alterations in its biophysical and biomechanical properties. Mechano-oncology deals with utilisation of the differences in these properties at cellular and subcellular levels to specifically target and annihilate cancer cells. Dynamic analysis of cancer cells by experimental, mathematical and simulation approaches gives an insight into understanding the differences in mechanical properties and helps in development of treatment strategies that specifically target tumor cells. Experimental analysis in vivo is extremely expensive, requires considerable amount of skill and time, thereby posing a great challenge to researchers. Experimental constraints have led to researchers adopting a numerical and analytical approach to study cellular systems. On the other hand, simulation approach allows

effective and detailed analysis of the biomechanical behaviour of cellular systems. Simulation techniques potentially reduce the cost and time in the development and analysis of cellular models due to ease of use and accessibility, helping a larger scientific community to illustrate the effects of different inputs on a biological system.

The mechanical properties of cancer cells are investigated using various techniques. Lekka et al., [1, 2] used scanning force microscopy and atomic force microscopy to determine the elastic properties of human epithelial cells and melanoma cells respectively. Suresh [3] utilised micropipette aspiration to study the properties of various normal and cancer cells. Literature mentioned above reveals that there is significant reduction in the stiffness of cancer cells as compared to their normal counterparts. This change in stiffness results in a corresponding variation of its dynamic characteristics such as its natural frequency. Precise knowledge of tumour specific frequencies would provide a firm platform for the development of new treatment strategies that would either target cancer cells or aid the existing techniques to treat cancer. Numerous attempts are made to specifically target cancer cells due to the change in their natural frequency as compared to that of normal cells. Jagannathan et al., [4] modelled and analysed breast cancer cells, MCF-7 (Michigan Cancer Foundation-7) and prostate cancer cells, LNCap (Lymph Node Carcinoma of the Prostate) to determine their natural frequencies. Geltmeier et al., [5] determined the natural frequencies of normal breast cells, MCF-10A (Michigan Cancer Foundation-10A) and cancerous (MCF-7) breast cells and utilized low frequency ultrasound to specifically target and destroy breast tumour cells (MCF-7). Roberts [6] made use of histotripsy as an ablative therapy to annihilate preclinical models of normal prostate cells (benign prostatic hyperplasia or BPH). Zimmerman et al., [7] utilised radiofrequency electromagnetic fields that are amplitude-modulated at tumour specific frequencies, to disrupt the mitotic spindle and inhibit cell division. Vadalà et al., [8] presented a review on employing pulsed electromagnetic field (PEMF) and possibility of it being combined with other available therapies to treat various types of cancers. Fraldi et al., [9] developed a frequency-based hypothesis for mechanically targeting and selectively attacking cancer cells. A mechanical model of the cell comprising of various sub-cellular elements is utilised in evaluating the frequency response of the cell. Schuster et al., [10] carried out an investigation to study the effect of low, medium and high energy density ultrasound at three individual frequencies on inhibiting the growth of cancer cells.

The literature review indicates that there is very little information provided on the modelling techniques used to generate the cellular models and the boundary conditions applied for analysis. Obscurity exists in the variation of natural frequencies of cells obtained by considering or neglecting the mechanical properties of subcellular elements. In the present study, an effort is made to develop two models, namely a non-homogenous model and a homogeneous model to investigate the dynamic characteristics of normal and cancerous breast cells. Material properties of subcellular elements are taken into account in the case of non-homogenous model whereas the cell is assumed to have uniform material properties throughout for the homogenous model. A comparison of the natural frequencies for the two models of MCF-10A and MCF-7 is reported.

2. Modelling and simulation

The shape of cells in general is hard to define. Extensive variability in cell size and shape is noticed in tumour cells. Based on reported literature on the study of breast cells, the shape of the cell is assumed to be spherical [4, 5] with appropriate modifications in geometry and boundary conditions. Two types of breast cells, namely MCF-10A and MCF-7 are considered for the current study. Homogenous (model I) and non-homogenous (model II) simulation models of both MCF-10A and MCF-7 are developed using SolidWorks. The interaction of cells with the physiological environment associated with cell culture is taken into consideration while modelling the cell and incorporating the necessary boundary conditions.

A cell consists of numerous sub-cellular elements out of which the cell membrane, cytoplasm and nucleus largely contribute to its functioning. The cell membrane is the outermost layer of the

cell, made up of a phospholipid bilayer implanted with proteins. The cytoplasm is a viscous material that is present between the membrane and nucleus. The nucleus, which controls the major functioning of the cell, is considered as a rigid body, suspended in the cytoplasm.

For model I, the cell is assumed to be composed of a homogenous material and the material properties of individual sub-cellular elements namely the cytoplasm, cell membrane and nucleus are neglected as shown in Fig. 1.

Model II is developed considering the properties of sub-cellular elements as shown in Fig. 2. The properties of the cytoskeleton are assumed to be incorporated within the material properties of the cytoplasm. The geometrical parameters and material properties considered for the model I is listed in Table 1.

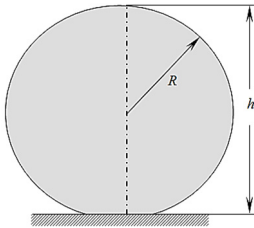


Fig. 1. Model I

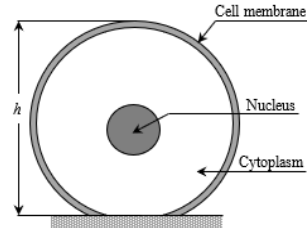


Fig. 2. Model II

Table 1. Geometrical parameters and material properties of model I [4, 5]

Cell type	Volume (μm^3)	Density (g/cm^3)	Young's modulus (kPa)	Poisson's ratio
MCF-10A	678	1.020	0.730	0.49
MCF-7	3,375	0.994	0.425	0.49

It is observed that there is a considerable difference in the mechanical properties between healthy and cancer breast cells. This difference forms a key factor that can be used to distinguish a cancer cell from a normal cell. The geometrical parameters and mechanical properties considered for the cellular constituents of model II is tabulated in Table 2.

Table 2. Geometrical parameters and material properties of model II [5]

Cellular elements	Parameter	MCF-7	MCF-10A
Cell	Volume (μm^3)	3,375	678
Cell membrane	Thickness (nm)	200	200
	Density (g/cm^3)	1.5	1.5
	Young's Modulus (kPa)	1.125	1.1
	Poisson's ratio	0.49	0.49
Cytoplasm	Density (g/cm^3)	1.05	1.05
	Young's modulus (kPa)	0.15	0.25
	Poisson's ratio	0.49	0.49
Nucleus	Volume (μm^3)	1122	360
	Density (g/cm^3)	1.3	1.3
	Young's modulus (kPa)	0.15	0.25
	Poisson's ratio	0.49	0.49

Based on the known volume of the cell, the radius for MCF-10A and MCF-7 is calculated. Boundary conditions pertaining to the in-vitro cell culture environment in a petri dish are considered for analysis. The radius of the contact area between the cell membrane and the petri dish is calculated using the formulation from [11] as:

$$b = \left(\frac{JR_c^3}{\eta} \right)^{1/4} t^{1/4}, \quad (1)$$

where b is the radius of contact area at the base, R_c is the initial radius of the cell, $J = 100 \mu\text{J}/\text{m}^2$, corresponds to adhesion energy per unit area, $\eta = 300 \text{ Pa}\cdot\text{s}$, is the viscosity and $t = 10 \text{ s}$, is the characteristic deforming time.

Due to the interaction of the cell with the surface of the petri dish, the height of the cell reduces while the volume of the cell remains constant. The height of the cell is calculated using the simple relation:

$$h = R + \sqrt{R^2 - b^2}, \tag{2}$$

where h is the obtained height of the cell, R is the radius of the cell after spreading and b is the radius of the base of the cell in contact with the petri dish.

The developed models are imported into Ansys Workbench for analysis. An adaptive size function is used to mesh the model. A fixed boundary condition is incorporated between the base of the cell and the surface of the petri dish. The inter-cellular adhesion is determined from [12] and is given a fixed boundary condition. Modal analysis is carried out for models I and II of MCF-10A and MCF-7 and the corresponding natural frequencies are extracted. The natural frequencies corresponding to the first four modes for both model I and II of MCF-10A are tabulated in Table 3.

Table 3. Natural frequencies for model I and II of MCF-10A

Mode	Natural frequency (Hz)		% error
	Model I	Model II	
1	22,942	14,240	37.93
2	22,998	14,284	37.89
3	30,563	21,047	31.13
4	36,471	21,797	40.23

Table 4. Natural frequencies for model I and II of MCF-7

Mode	Natural frequency (Hz)		% error
	Model I	Model II	
1	9,611	5,768	39.98
2	9,657	5,799	39.95
3	12,854	8,718	32.17
4	16,469	9,406	42.88

The natural frequencies corresponding to the first four modes for models I and II of MCF-7 are presented in Table 4 along with the percentage error in the natural frequencies.

As observed in Tables 3 and 4, an approximate 36 % error in natural frequencies is observed between model I and II due to the difference in the modelling technique adopted for dynamic analysis. A comparison between the natural frequencies obtained for model I of MCF-10A and MCF-7 is displayed in Table 5.

Table 5. Comparison of natural frequencies for model I of MCF-10A and MCF-7

Mode	Natural frequency (Hz)		% reduction in natural frequency
	MCF-10A	MCF-7	
1	22,942	9,611	58.11
2	22,998	9,657	58.01
3	30,563	12,854	57.94
4	36,471	16,469	54.84

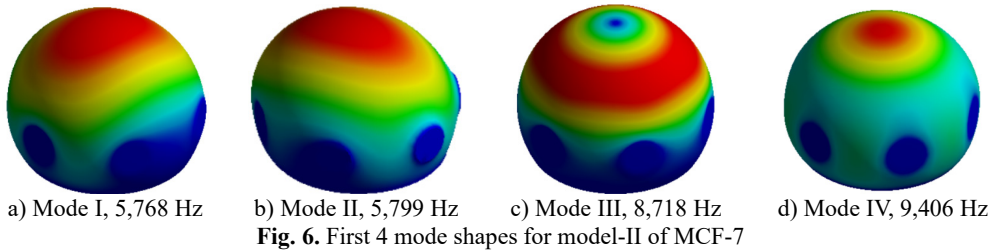
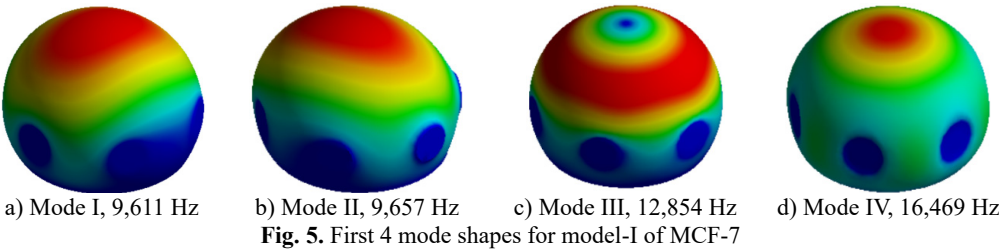
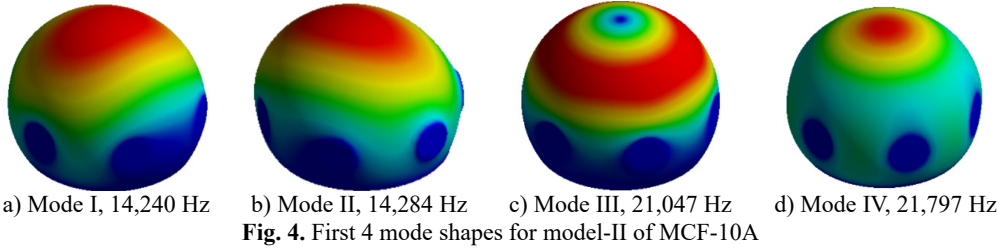
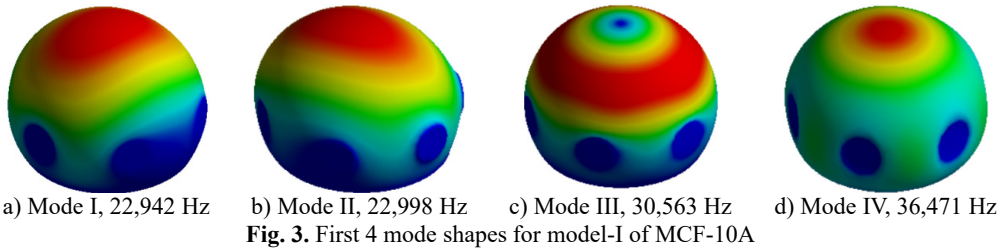
Table 6. Comparison of natural frequencies for model II of MCF-10A and MCF-7

Mode	Natural frequency (Hz)		% reduction in natural frequency
	MCF-10A	MCF-7	
1	14,240	5,767.6	59.49
2	14,284	5,798.8	59.40
3	21,047	8,718.1	58.58
4	21,797	9,405.9	56.84

The natural frequencies corresponding to the first four modes for model II of MCF-10A and MCF-7 are compared and presented in Table 6.

From Tables 5 and 6, it is observed that there is approximately a 60 % reduction in the natural frequencies as the cell undergoes transformation from a healthy to a cancer cell.

Figs. 3 and 5 indicate that the first and second natural frequencies for model I of both the cells correspond to a lateral mode of vibration along the z and x direction of the system respectively. A twisting mode shape about the vertical axis y -axis is observed for the third natural frequency of the system. Finally, the fourth mode shape shows an axial mode of vibration along the y -direction.



Figs. 4 and 6 represent the fundamental mode shapes for model II of MCF-10A and MCF-7. A lateral mode of vibration in the z and x direction of the system is observed for the first and second natural frequencies respectively. Similar to model I, the third mode shape depicts a twisting mode of vibration about the y axis of the system. The 4th mode is observed to be an axial mode of vibration along the y -axis.

Figs. 7 and 8 graphically show the error in natural frequencies for first four modes of Model I and II of MCF-10A and MCF-7 respectively.

Fig. 9 graphically represents the reduction in natural frequencies for the first four modes of Model I of MCF-10A and MCF-7. Fig. 10 graphically demonstrates the reduction in natural frequencies for the first four modes of Model II of MCF-10A and MCF-7.

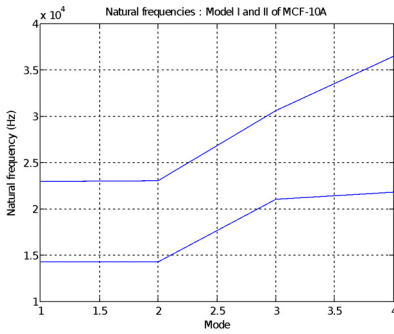


Fig. 7. Natural frequencies of Model I and II of MCF-10A

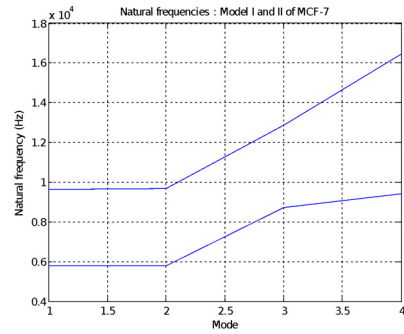


Fig. 8. Natural frequencies of Model I and II of MCF-7

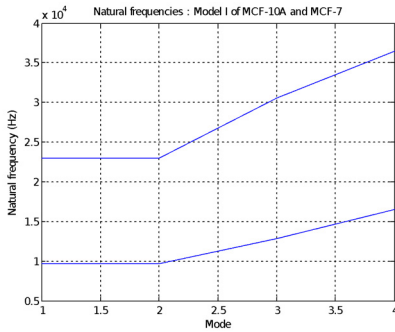


Fig. 9. Natural frequencies of Model I of MCF-10A and MCF-7

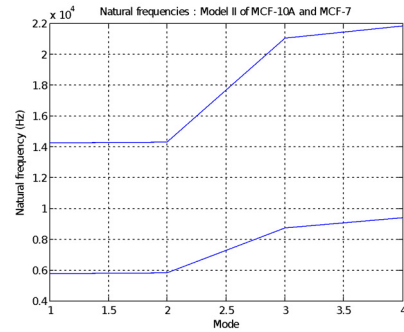


Fig. 10. Natural frequencies of Model II of MCF-10A and MCF-7

It is noticed that the natural frequencies of MCF-10A is about 2.4 times that of MCF-7. The reduction in natural frequencies observed in the case of cancer cells is attributed to the decrease in its stiffness as the cell turns cancerous.

3. Conclusions

The current study establishes a basic model of an adherent breast cell for the purpose of modelling and simulation. Homogeneous and non-homogeneous models are developed to evaluate the dynamic characteristics of normal and cancerous breast cells. Results indicate that the natural frequencies of cells that are modelled as a homogeneous system is approximately 36 % more than that of the cells whose sub-cellular material properties are considered for analysis. The natural frequencies of normal breast cells (MCF-10A) is found to be about 2.4 times that of breast tumour cells (MCF-7). This difference in natural frequencies due to increased cellular mass and decreased stiffness of cancer cells can be exploited for the development of new strategies that specifically identify and target cancer cells.

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