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## **Pulmonary xenon-129 MRI: New opportunities to unravel enigmas in respiratory medicine**

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## Early View

ERJ Methods

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## **Pulmonary $^{129}\text{Xe}$ MRI: New Opportunities to Unravel Enigmas in Respiratory Medicine**

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**Key Words:** xenon-129, magnetic resonance imaging, pulmonary disease, pulmonary functional imaging

### **ERJ Methods**

### **Take Home Message:**

$^{129}\text{Xe}$  MRI provides rapid, sensitive, non-invasive, high spatial resolution and simultaneous measurements of pulmonary ventilation, tissue microstructure and gas exchange, and is poised for routine clinical assessments in patients with chronic lung disease.

## **Introduction**

Chest x-ray computed tomography (CT) is the imaging modality of choice for the non-invasive, quantitative evaluation of chronic lung diseases because thoracic CT protocols are nearly ubiquitously available and provide rapid, high-resolution images of airway and parenchymal structure and anatomy. Pulmonary magnetic resonance imaging (MRI) has not been used clinically, mainly because of complexity (pulmonary MRI signal decays rapidly), cost and because conventional MRI is dependent on proton-density (hydrogen atoms in tissue) which is exceptionally low (around  $0.1 \text{ g/cm}^3$ ) in the healthy lung because it is mainly air and not tissue or water-filled [1, 2]. Using conventional MRI methods, the lungs mainly appear as dark, signal-deficient voids and the pulmonary MRI signal is further degraded because of the millions of lung air-tissue interfaces that cause local magnetic field distortions, and respiratory and cardiac motion. While anatomical proton MRI of the lung is now developing rapidly to overcome these technical limitations, it still does not provide information beyond that of a low-dose CT so there is a drive towards functional MRI. One of these approaches involves the use of inhaled gas contrast agents, primarily hyperpolarised (or magnetised) helium-3 ( $^3\text{He}$ ) and xenon-129 ( $^{129}\text{Xe}$ ), both of which provide a way to rapidly ( $<10 \text{ s}$ ) and directly visualise inhaled gas distribution with high spatial resolution ( $\sim 3 \text{ mm}$  x, y and z planes). Pulmonary functional MRI is also possible using inhaled fluorinated gas [3], oxygen-enhanced techniques [4] and free-breathing proton methods [5, 6], however hyperpolarised inhaled gas MRI has been the most widely used and described. Because inhaled contrast gases have different resonant frequencies than hydrogen protons as used for conventional MRI, these methods inherently have no background signal and excellent contrast.

While inhalation of hyperpolarised gases to measure pulmonary function was originally discovered using  $^{129}\text{Xe}$  [7], the field was dominated by  $^3\text{He}$  MRI until recently because of superior  $^3\text{He}$  MRI image quality with low volume (<500 mL) inhaled doses. However, the limited worldwide supply of  $^3\text{He}$  [8] has driven the field back to  $^{129}\text{Xe}$  gas because it is naturally abundant and turnkey polarisation technologies have significantly advanced [9-11].  $^{129}\text{Xe}$  also has modest solubility in biologic tissues and resonates at different frequencies when dissolved in different tissues, so it provides novel alveolar tissue and capillary blood information. Here, we provide an overview of the key concepts and methods for  $^{129}\text{Xe}$  MRI, discuss the current state-of-the-art and how  $^{129}\text{Xe}$  MRI may be applied in the future.

### **How does $^{129}\text{Xe}$ MRI work?**

$^{129}\text{Xe}$  MRI is dependent on equipment that polarises  $^{129}\text{Xe}$  atoms which effectively increases the nuclear polarisation by ~100,000 times. Commercial (Polarean Inc., Durham, NC; Xemed LLC, Durham, NH) and custom-built [10, 11] polariser systems operate via spin-exchange optical pumping [12] whereby circularly polarised light bombards a glass cell housing rubidium and  $^{129}\text{Xe}$ . The circularly polarised light is absorbed by rubidium and subsequent collisions between polarised rubidium and  $^{129}\text{Xe}$  transfer angular momentum to  $^{129}\text{Xe}$  and increase its nuclear-spin polarisation. Typically, isotopically enriched  $^{129}\text{Xe}$  (~86% by volume) is used to further increase the fraction of  $^{129}\text{Xe}$  atoms that are polarised in a given volume.  $^{129}\text{Xe}$  flows through the glass cell at a constant rate and the hyperpolarised  $^{129}\text{Xe}$  is frozen as it leaves the cell and remains frozen until the desired amount is accumulated, upon which it is thawed and dispensed into a bag for patient inhalation. The  $^{129}\text{Xe}$  flow rate through the cell can be adjusted to accumulate the desired  $^{129}\text{Xe}$  dose within a desired time frame.  $^{129}\text{Xe}$  doses vary from 250 mL up to 1.0 L [13],

and doses are typically diluted up to 1.0 L using medical-grade nitrogen (N<sub>2</sub>) or helium-4. <sup>129</sup>Xe hyperpolariser equipment is typically located in a small room adjacent to the MRI suite, within close proximity to deliver the polarised gas to the patient as quickly as possible.

<sup>129</sup>Xe MRI is feasible on both 1.5 T and 3.0 T field strength scanners. To acquire the signal however, a radiofrequency coil is fitted around the patient chest and this is specifically tuned to the resonance frequency of <sup>129</sup>Xe. There are a number of different <sup>129</sup>Xe coils, however as shown in Figure 1, the most common are quadrature rigid birdcage or flexible vests (Clinical MR Solutions, Brookfield, WI; RAPID Biomedical, Rimpar, Germany). Rigid birdcage coils provide more homogeneous MRI signal throughout the lungs (ie, no artifactual differences in MRI signal that are unrelated to anatomy) and increased signal-to-noise but they have patient size limitations; flexible vest coils can accommodate a larger range of patient sizes but may require additional image corrections after MRI acquisition [14] to account for signal differences caused by the way the patient and coil interact. Additional receive array-coils may be placed on the patients' chest inside the rigid or flexible design coils and this improves signal-to-noise ratios and accelerates acquisition times which enables very short breath-hold scans (~5 seconds). Figure 1A shows <sup>129</sup>Xe MRI equipment including hyperpolariser, gas delivery bag and radiofrequency coils, and patient set-up in MRI scanner.

<sup>129</sup>Xe MRI is primarily performed under breath-hold conditions, though dynamic multi-breath protocols have also been applied [15]. While laying supine, patients are coached to inhale the gas mixture from passive end expiration or functional residual capacity and hold their breath while the image is acquired, which may take from 5-16 seconds. Pulmonary <sup>129</sup>Xe MRI

acquisitions may include static ventilation, diffusion-weighted or dissolved-phase. Static ventilation images are the most commonly reported  $^{129}\text{Xe}$  MRI acquisition and this provides regional maps of pulmonary gas distribution. A conventional proton ( $^1\text{H}$ ) image is also typically acquired in the same scanning session and at the same lung volume so that the anatomical and functional images may be co-registered to distinguish the edges of the thoracic cavity during image analysis and segmentation. Figure 1B shows a  $^{129}\text{Xe}$  gas static ventilation coronal image (cyan) and corresponding anatomical proton image (greyscale), and the two co-registered demonstrating a regional ventilation map. A corresponding CT slice is also shown in Figure 1 as well as the 3D segmented airway tree (yellow) which may be co-registered to the ventilation map to demonstrate structure-function relationships.

Diffusion-weighted  $^{129}\text{Xe}$  MRI methods may also be employed and these estimate the self-diffusion of  $^{129}\text{Xe}$  gas within the terminal airways and their restriction within the airspaces on a voxel-wise basis [16], which provides an excellent surrogate measurement of alveolar and terminal bronchiole enlargement.

$^{129}\text{Xe}$  has a large, loosely-bound electron cloud, making it sensitive to its surroundings and soluble in biological tissue; once dissolved in biological tissues,  $^{129}\text{Xe}$  exhibits a different resonance frequency. Thus,  $^{129}\text{Xe}$  MRI can also provide *in vivo* measurements of pulmonary gas exchange [17, 18]. The so-called dissolved-state refers to  $^{129}\text{Xe}$  dissolved in the alveolar membrane and in red blood cells.  $^{129}\text{Xe}$  that has diffused into biological tissues experiences a different chemical environment and this is reflected via a shift in the  $^{129}\text{Xe}$  resonance frequency (chemical shift) from gaseous  $^{129}\text{Xe}$ , as shown in Figure 2B. In the same manner, there is also a difference in the resonance frequency between  $^{129}\text{Xe}$  in the tissue and  $^{129}\text{Xe}$  in the blood, which

enables simultaneous, independent imaging of these three states or compartments: 1) gas, 2) tissue barrier plus plasma, and 3) red blood cell. The gas state reflects the inhaled gas and has the largest measurable signal.  $^{129}\text{Xe}$  dissolved in the tissue barrier and blood plasma have indistinguishable chemical shifts and combine for the second largest measurable signal with a ~197 ppm shift from gaseous  $^{129}\text{Xe}$  [19].  $^{129}\text{Xe}$  dissolved in red blood cells exhibits an additional ~20 ppm shift beyond the tissue-plasma signal and makes up the smallest, measurable  $^{129}\text{Xe}$  signal [19]; moreover, the red blood cell signal is oxygen-dependent and may undergo another measurable ~5 ppm shift based on blood oxygenation [20]. Tuning the scanner allows acquisition of images of all three compartments, and acquisition protocols have been developed to simultaneously acquire quantitative images from all three compartments within a single breath-hold [18].

### **Current state-of-the-art**

$^{129}\text{Xe}$  MRI methods have an excellent safety profile in patients with respiratory disease [21, 22], including asthma [23-25], COPD [16, 26-29], cystic fibrosis [30-32], pulmonary vascular disease [33], idiopathic pulmonary fibrosis [34, 35], lung cancer [36] and lymphangiomyomatosis [37].

In healthy volunteers,  $^{129}\text{Xe}$  gas distribution is typically highly homogeneous and gas signal fills the entire lung whereas in patients with lung disease such as in Figure 1, focal MRI signal voids or ventilation defects and patchy gas distribution are often observed. Ventilation defects are commonly quantified as the ventilation defect percent (VDP) [38] – the volume of ventilation defects normalised to the thoracic cavity volume. In a similar manner, using diffusion-weighted MRI, the apparent diffusion coefficient (ADC), a measure of airspace enlargement, may be



estimated.  $^{129}\text{Xe}$  ADC is low in healthy participants and increases with increasing airspace size and the extent of emphysema [16, 26]. Diffusion-weighted MRI may also be used to generate morphological airspace measurements [39, 40], for example mean linear intercept analogous to histology. Normalised biomarkers of the ratio of  $^{129}\text{Xe}$  dissolved in the tissue barrier and red blood cells [18], have been shown to be particularly relevant in pulmonary fibrosis. A unique feature of  $^{129}\text{Xe}$  MRI is that it provides a way to regionally quantify gas exchange [17, 41] in regions of the lung that are ventilated and models have been developed to extract subcomponents of gas exchange and structural measurements [42-44].

At the current time,  $^{129}\text{Xe}$  MRI is approved for clinical use in the United Kingdom and clinical approval is pending in the United States.

### **How is $^{129}\text{Xe}$ MRI likely to be used in the future?**

The infrastructure needed to enable  $^{129}\text{Xe}$  MRI is available in ~12 respiratory imaging sites worldwide using a wide variety of different scanners, polarisers and coils. Whilst technical developments will enable faster  $^{129}\text{Xe}$  polarisation times and larger volumes of polarised gas, multi-centre clinical trials are now poised to demonstrate the inter- and intra-site reproducibility of  $^{129}\text{Xe}$  MRI biomarkers so that clinical trials of new treatments and interventions using MRI may be powered. A comprehensive  $^{129}\text{Xe}$  examination (including localiser scan, anatomical scan, ventilation, diffusion-weighted, perfusion and gas exchange scans) in a patient may be easily performed within 15 minutes, often with patients inside the MRI bore for approximately 5 minutes and this is certainly compatible with current clinical imaging workflows.

Highly sensitive  $^{129}\text{Xe}$  MRI VDP promises therapy studies utilising smaller sample sizes to evaluate response to therapy. The broad array of  $^{129}\text{Xe}$  MRI biomarkers may provide endpoints

for clinical trials of novel treatments for asthma, COPD, cystic fibrosis and especially pulmonary fibrosis. The spatially resolved functional information provided by  $^{129}\text{Xe}$  MRI may perhaps guide placement of endobronchial valves, which are currently guided using structural information from CT. Moreover,  $^{129}\text{Xe}$  MRI may provide a way to generate quantitative pathological evidence for treatment responses, for example following bronchial thermoplasty [45, 46] or novel biologic treatments for asthma [47], where patients experience improved quality of life and reduced exacerbation frequency often in the absence of  $\text{FEV}_1$  improvements. Another valuable application of  $^{129}\text{Xe}$  MRI may be in the prediction of pulmonary exacerbations, which has been demonstrated using  $^3\text{He}$  MRI [48, 49]. Because  $^{129}\text{Xe}$  MRI appears to be more sensitive to ventilation abnormalities [23, 26], it is expected to also predict or explain pulmonary exacerbations of COPD, cystic fibrosis and asthma.

It remains difficult to predict response to treatment in many respiratory diseases, which is becoming increasingly important as novel, expensive therapies continue to be developed. Imaging phenotypes are widely recognised in COPD and pulmonary fibrosis and the combination of available  $^{129}\text{Xe}$  MRI biomarkers may provide novel phenotypes to support regulatory and treatment decisions. Moreover,  $^{129}\text{Xe}$  MRI provides a regional map of lung structure-function that can be likened to a fingerprint or “lung-print”, to support individualised therapy decisions or n=1 studies in individual patients.

$^{129}\text{Xe}$  MRI is particularly invaluable for longitudinal monitoring of disease progression, disease phenotypes and/or treatment response (in and outside of clinical trials) because it poses no radiation burden on patients. The lack of ionising radiation is particularly needed for examinations in vulnerable populations such as children with chronic lung disease.

In summary,  $^{129}\text{Xe}$  MRI provides rapid, sensitive, non-invasive and simultaneous measurements of pulmonary ventilation, lung tissue microstructure as well as diffusion within the alveolus and into the alveolar tissue and red blood cells, providing new opportunities to more deeply investigate lung diseases and unravel enigmas in respiratory medicine.

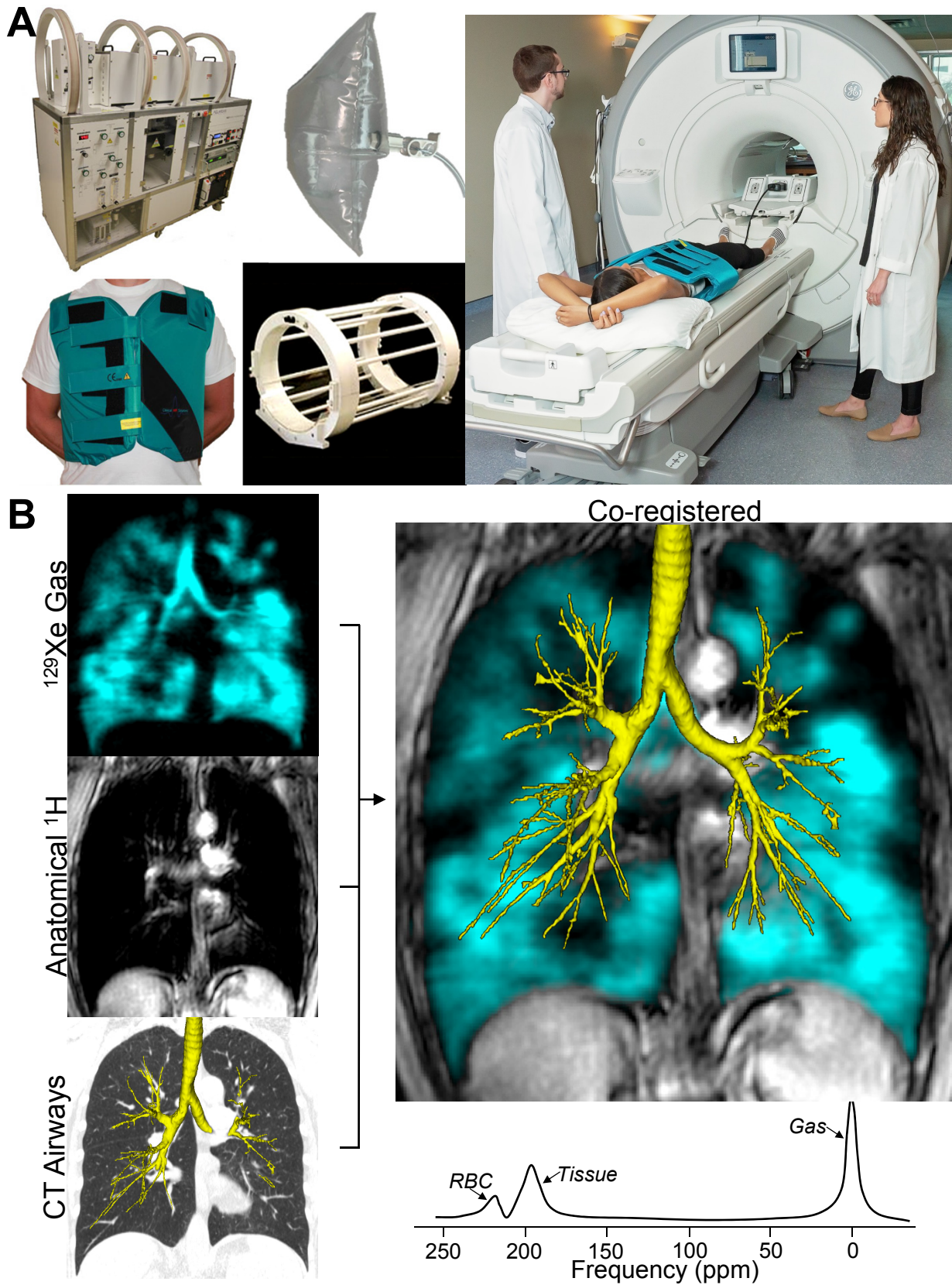
## REFERENCES

1. Bergin CJ, Glover GM, Pauly J. Magnetic resonance imaging of lung parenchyma. *J Thorac Imaging* 1993; 8(1): 12-17.
2. Johnson KM, Fain SB, Schiebler ML, Nagle S. Optimized 3D ultrashort echo time pulmonary MRI. *Magn Reson Med* 2013; 70(5): 1241-1250.
3. Couch MJ, Ball IK, Li T, Fox MS, Ouriadov AV, Biman B, Albert MS. Inert fluorinated gas MRI: a new pulmonary imaging modality. *NMR Biomed* 2014; 27(12): 1525-1534.
4. Edelman RR, Hatabu H, Tadamura E, Li W, Prasad PV. Noninvasive assessment of regional ventilation in the human lung using oxygen-enhanced magnetic resonance imaging. *Nat Med* 1996; 2(11): 1236-1239.
5. Bergin CJ, Pauly JM, Macovski A. Lung parenchyma: projection reconstruction MR imaging. *Radiology* 1991; 179(3): 777-781.
6. Bauman G, Puderbach M, Deimling M, Jellus V, Chefd'hotel C, Dinkel J, Hintze C, Kauczor HU, Schad LR. Non-contrast-enhanced perfusion and ventilation assessment of the human lung by means of fourier decomposition in proton MRI. *Magn Reson Med* 2009; 62(3): 656-664.
7. Albert MS, Cates GD, Driehuys B, Happer W, Saam B, Springer CS, Jr., Wishnia A. Biological magnetic resonance imaging using laser-polarized  $^{129}\text{Xe}$ . *Nature* 1994; 370(6486): 199-201.
8. Shea DA, Morgan D. The helium-3 shortage: Supply, demand, and options for congress. In; 2010: Congressional Research Service, Library of Congress; 2010.
9. Hersman FW, Ruset IC, Ketel S, Muradian I, Covrig SD, Distelbrink J, Porter W, Watt D, Ketel J, Brackett J, Hope A, Patz S. Large production system for hyperpolarized  $^{129}\text{Xe}$  for human lung imaging studies. *Acad Radiol* 2008; 15(6): 683-692.
10. Nikolaou P, Coffey AM, Walkup LL, Gust BM, Whiting N, Newton H, Barcus S, Muradyan I, Dabaghyan M, Moroz GD, Rosen MS, Patz S, Barlow MJ, Chekmenev EY, Goodson BM. Near-unity nuclear polarization with an open-source  $^{129}\text{Xe}$  hyperpolarizer for NMR and MRI. *Proc Natl Acad Sci U S A* 2013; 110(35): 14150-14155.
11. Norquay G, Parnell SR, Xu X, Parra-Robles J, Wild JM. Optimized production of hyperpolarized  $^{129}\text{Xe}$  at 2 bars for in vivo lung magnetic resonance imaging. *J Appl Phys* 2013; 113(4): 044908.
12. Walker TG, Happer W. Spin-exchange optical pumping of noble-gas nuclei. *Rev Mod Phys* 1997; 69(2): 629.
13. He M, Robertson SH, Kaushik SS, Freeman MS, Virgincar RS, Davies J, Stiles J, Foster WM, McAdams HP, Driehuys B. Dose and pulse sequence considerations for hyperpolarized ( $^{129}\text{Xe}$ ) ventilation MRI. *Magn Reson Imaging* 2015; 33(7): 877-885.
14. He M, Driehuys B, Que LG, Huang YT. Using Hyperpolarized  $^{129}\text{Xe}$  MRI to Quantify the Pulmonary Ventilation Distribution. *Acad Radiol* 2016; 23(12): 1521-1531.
15. Horn FC, Rao M, Stewart NJ, Wild JM. Multiple breath washout of hyperpolarized ( $^{129}\text{Xe}$ ) and ( $^3\text{He}$ ) in human lungs with three-dimensional balanced steady-state free-precession imaging. *Magn Reson Med* 2017; 77(6): 2288-2295.
16. Kaushik SS, Cleveland ZI, Cofer GP, Metz G, Beaver D, Nouis J, Kraft M, Auffermann W, Wolber J, McAdams HP, Driehuys B. Diffusion-weighted hyperpolarized  $^{129}\text{Xe}$  MRI in healthy volunteers and subjects with chronic obstructive pulmonary disease. *Magn Reson Med* 2011; 65(4): 1154-1165.

17. Qing K, Ruppert K, Jiang Y, Mata JF, Miller GW, Shim YM, Wang C, Ruset IC, Hersman FW, Altes TA, Mugler JP, 3rd. Regional mapping of gas uptake by blood and tissue in the human lung using hyperpolarized xenon-129 MRI. *J Magn Reson Imaging* 2014; 39(2): 346-359.
18. Kaushik SS, Robertson SH, Freeman MS, He M, Kelly KT, Roos JE, Rackley CR, Foster WM, McAdams HP, Driehuys B. Single-breath clinical imaging of hyperpolarized (129)Xe in the airspaces, barrier, and red blood cells using an interleaved 3D radial 1-point Dixon acquisition. *Magn Reson Med* 2016; 75(4): 1434-1443.
19. Mugler JP, 3rd, Driehuys B, Brookeman JR, Cates GD, Berr SS, Bryant RG, Daniel TM, de Lange EE, Downs JH, 3rd, Erickson CJ, Happer W, Hinton DP, Kassel NF, Maier T, Phillips CD, Saam BT, Sauer KL, Wagshul ME. MR imaging and spectroscopy using hyperpolarized 129Xe gas: preliminary human results. *Magn Reson Med* 1997; 37(6): 809-815.
20. Norquay G, Leung G, Stewart NJ, Wolber J, Wild JM. (129) Xe chemical shift in human blood and pulmonary blood oxygenation measurement in humans using hyperpolarized (129) Xe NMR. *Magn Reson Med* 2017; 77(4): 1399-1408.
21. Shukla Y, Wheatley A, Kirby M, Svenningsen S, Farag A, Santyr GE, Paterson NA, McCormack DG, Parraga G. Hyperpolarized 129Xe magnetic resonance imaging: tolerability in healthy volunteers and subjects with pulmonary disease. *Acad Radiol* 2012; 19(8): 941-951.
22. Driehuys B, Martinez-Jimenez S, Cleveland ZI, Metz GM, Beaver DM, Nouls JC, Kaushik SS, Firszt R, Willis C, Kelly KT, Wolber J, Kraft M, McAdams HP. Chronic obstructive pulmonary disease: safety and tolerability of hyperpolarized 129Xe MR imaging in healthy volunteers and patients. *Radiology* 2012; 262(1): 279-289.
23. Svenningsen S, Kirby M, Starr D, Leary D, Wheatley A, Maksym GN, McCormack DG, Parraga G. Hyperpolarized (3) He and (129) Xe MRI: differences in asthma before bronchodilation. *J Magn Reson Imaging* 2013; 38(6): 1521-1530.
24. Qing K, Mugler JP, 3rd, Altes TA, Jiang Y, Mata JF, Miller GW, Ruset IC, Hersman FW, Ruppert K. Assessment of lung function in asthma and COPD using hyperpolarized 129Xe chemical shift saturation recovery spectroscopy and dissolved-phase MRI. *NMR Biomed* 2014; 27(12): 1490-1501.
25. Ebner L, He M, Virgincar RS, Heacock T, Kaushik SS, Freeman MS, McAdams HP, Kraft M, Driehuys B. Hyperpolarized 129Xenon Magnetic Resonance Imaging to Quantify Regional Ventilation Differences in Mild to Moderate Asthma: A Prospective Comparison Between Semiautomated Ventilation Defect Percentage Calculation and Pulmonary Function Tests. *Invest Radiol* 2017; 52(2): 120-127.
26. Kirby M, Svenningsen S, Owrangi A, Wheatley A, Farag A, Ouriadov A, Santyr GE, Etemad-Rezai R, Coxson HO, McCormack DG, Parraga G. Hyperpolarized 3He and 129Xe MR imaging in healthy volunteers and patients with chronic obstructive pulmonary disease. *Radiology* 2012; 265(2): 600-610.
27. Virgincar RS, Cleveland ZI, Kaushik SS, Freeman MS, Nouls J, Cofer GP, Martinez-Jimenez S, He M, Kraft M, Wolber J, McAdams HP, Driehuys B. Quantitative analysis of hyperpolarized 129Xe ventilation imaging in healthy volunteers and subjects with chronic obstructive pulmonary disease. *NMR Biomed* 2013; 26(4): 424-435.
28. Matin TN, Rahman N, Nickol AH, Chen M, Xu X, Stewart NJ, Doel T, Grau V, Wild JM, Gleeson FV. Chronic Obstructive Pulmonary Disease: Lobar Analysis with Hyperpolarized (129)Xe MR Imaging. *Radiology* 2017; 282(3): 857-868.

29. Ruppert K, Qing K, Patrie JT, Altes TA, Mugler JP, 3rd. Using Hyperpolarized Xenon-129 MRI to Quantify Early-Stage Lung Disease in Smokers. *Acad Radiol* 2019; 26(3): 355-366.
30. Kanhere N, Couch MJ, Kowalik K, Zanette B, Rayment JH, Manson D, Subbarao P, Ratjen F, Santyr G. Correlation of Lung Clearance Index with Hyperpolarized (129)Xe Magnetic Resonance Imaging in Pediatric Subjects with Cystic Fibrosis. *Am J Respir Crit Care Med* 2017; 196(8): 1073-1075.
31. Couch MJ, Thomen R, Kanhere N, Hu R, Ratjen F, Woods J, Santyr G. A two-center analysis of hyperpolarized (129)Xe lung MRI in stable pediatric cystic fibrosis: Potential as a biomarker for multi-site trials. *J Cyst Fibros* 2019; 18(5): 728-733.
32. Rayment JH, Couch MJ, McDonald N, Kanhere N, Manson D, Santyr G, Ratjen F. Hyperpolarised (129)Xe magnetic resonance imaging to monitor treatment response in children with cystic fibrosis. *Eur Respir J* 2019; 53(5).
33. Dahhan T, Kaushik SS, He M, Mammarrappallil JG, Tapson VF, McAdams HP, Sporn TA, Driehuys B, Rajagopal S. Abnormalities in hyperpolarized (129)Xe magnetic resonance imaging and spectroscopy in two patients with pulmonary vascular disease. *Pulm Circ* 2016; 6(1): 126-131.
34. Wang JM, Robertson SH, Wang Z, He M, Virgincar RS, Schrank GM, Smigla RM, O'Riordan TG, Sundry J, Ebner L, Rackley CR, McAdams P, Driehuys B. Using hyperpolarized (129)Xe MRI to quantify regional gas transfer in idiopathic pulmonary fibrosis. *Thorax* 2018; 73(1): 21-28.
35. Weatherley ND, Stewart NJ, Chan HF, Austin M, Smith LJ, Collier G, Rao M, Marshall H, Norquay G, Renshaw SA, Bianchi SM, Wild JM. Hyperpolarised xenon magnetic resonance spectroscopy for the longitudinal assessment of changes in gas diffusion in IPF. *Thorax* 2019; 74(5): 500-502.
36. Tahir BA, Hughes PJC, Robinson SD, Marshall H, Stewart NJ, Norquay G, Biancardi A, Chan HF, Collier GJ, Hart KA, Swinscoe JA, Hatton MQ, Wild JM, Ireland RH. Spatial Comparison of CT-Based Surrogates of Lung Ventilation With Hyperpolarized Helium-3 and Xenon-129 Gas MRI in Patients Undergoing Radiation Therapy. *Int J Radiat Oncol Biol Phys* 2018; 102(4): 1276-1286.
37. Walkup LL, Roach DJ, Hall CS, Gupta N, Thomen RP, Cleveland ZI, McCormack FX, Woods JC. Cyst Ventilation Heterogeneity and Alveolar Airspace Dilation as Early Disease Markers in Lymphangiomyomatosis. *Ann Am Thorac Soc* 2019; 16(8): 1008-1016.
38. Kirby M, Heydarian M, Svenningsen S, Wheatley A, McCormack DG, Etemad-Rezai R, Parraga G. Hyperpolarized 3He magnetic resonance functional imaging semiautomated segmentation. *Acad Radiol* 2012; 19(2): 141-152.
39. Sukstanskii AL, Yablonskiy DA. Lung morphometry with hyperpolarized 129Xe: theoretical background. *Magn Reson Med* 2012; 67(3): 856-866.
40. Thomen RP, Quirk JD, Roach D, Egan-Rojas T, Ruppert K, Yusen RD, Altes TA, Yablonskiy DA, Woods JC. Direct comparison of (129) Xe diffusion measurements with quantitative histology in human lungs. *Magn Reson Med* 2017; 77(1): 265-272.
41. Kaushik SS, Freeman MS, Cleveland ZI, Davies J, Stiles J, Virgincar RS, Robertson SH, He M, Kelly KT, Foster WM, McAdams HP, Driehuys B. Probing the regional distribution of pulmonary gas exchange through single-breath gas- and dissolved-phase 129Xe MR imaging. *J Appl Physiol (1985)* 2013; 115(6): 850-860.
42. Chang YV. MOXE: a model of gas exchange for hyperpolarized 129Xe magnetic resonance of the lung. *Magn Reson Med* 2013; 69(3): 884-890.

43. Stewart NJ, Leung G, Norquay G, Marshall H, Parra-Robles J, Murphy PS, Schulte RF, Elliot C, Condliffe R, Griffiths PD, Kiely DG, Whyte MK, Wolber J, Wild JM. Experimental validation of the hyperpolarized (129) Xe chemical shift saturation recovery technique in healthy volunteers and subjects with interstitial lung disease. *Magn Reson Med* 2015; 74(1): 196-207.
44. Stewart NJ, Chan HF, Hughes PJC, Horn FC, Norquay G, Rao M, Yates DP, Ireland RH, Hatton MQ, Tahir BA, Ford P, Swift AJ, Lawson R, Marshall H, Collier GJ, Wild JM. Comparison of (3) He and (129) Xe MRI for evaluation of lung microstructure and ventilation at 1.5T. *J Magn Reson Imaging* 2018.
45. Castro M, Rubin AS, Laviolette M, Fiterman J, De Andrade Lima M, Shah PL, Fiss E, Olivenstein R, Thomson NC, Niven RM, Pavord ID, Simoff M, Duhamel DR, McEvoy C, Barbers R, Ten Hacken NH, Wechsler ME, Holmes M, Phillips MJ, Erzurum S, Lunn W, Israel E, Jarjour N, Kraft M, Shargill NS, Quiring J, Berry SM, Cox G. Effectiveness and safety of bronchial thermoplasty in the treatment of severe asthma: a multicenter, randomized, double-blind, sham-controlled clinical trial. *Am J Respir Crit Care Med* 2010; 181(2): 116-124.
46. Chupp G, Laviolette M, Cohn L, McEvoy C, Bansal S, Shifren A, Khatri S, Grubb GM, McMullen E, Strauven R, Kline JN. Long-term outcomes of bronchial thermoplasty in subjects with severe asthma: a comparison of 3-year follow-up results from two prospective multicentre studies. *Eur Respir J* 2017; 50(2).
47. McGregor MC, Krings JG, Nair P, Castro M. Role of Biologics in Asthma. *Am J Respir Crit Care Med* 2019; 199(4): 433-445.
48. Kirby M, Pike D, Coxson HO, McCormack DG, Parraga G. Hyperpolarized (3)He ventilation defects used to predict pulmonary exacerbations in mild to moderate chronic obstructive pulmonary disease. *Radiology* 2014; 273(3): 887-896.
49. Mummy DG, Kruger SJ, Zha W, Sorkness RL, Jarjour NN, Schiebler ML, Denlinger LC, Evans MD, Fain SB. Ventilation defect percent in helium-3 magnetic resonance imaging as a biomarker of severe outcomes in asthma. *J Allergy Clin Immunol* 2018; 141(3): 1140-1141 e1144.



**Figure 1.  $^{129}\text{Xe}$  MRI infrastructure requirements and image acquisition**

(A)  $^{129}\text{Xe}$  hyperpolariser, gas delivery bag, radiofrequency coils (flexible vest and rigid birdcage), and patient set-up for MRI. (B)  $^{129}\text{Xe}$  gas, anatomical proton ( $^1\text{H}$ ), and CT airways (yellow) images co-registered (cyan), with  $^{129}\text{Xe}$  gas, tissue plus plasma and red blood cell (RBC) spectrum.