



---

*Research article*

## Hygienic quality of dehydrated aromatic herbs marketed in Southern Portugal

Jessie Melo<sup>1,2</sup>, Chaiane Quevedo<sup>1</sup>, Ana Graça<sup>1</sup> and Célia Quintas<sup>1,2,\*</sup>

<sup>1</sup> Universidade do Algarve, Instituto Superior de Engenharia, Campus da Penha, 8005-139 Faro, Portugal

<sup>2</sup> Universidade do Algarve, Centre for Mediterranean Bioresources and Food (MeditBio), Campus de Gambelas, 8005-139 Faro, Portugal

\* **Correspondence:** Email: [cquintas@ualg.pt](mailto:cquintas@ualg.pt); Tel: +351289800100; +351289800124.

**Abstract:** Dehydrated aromatic herbs are highly valued ingredients, widely used at home level and by food processing industry, frequently added to a great number of recipes in the Mediterranean countries. Despite being considered low-moisture products and classified as GRAS, during pre and post-harvesting stages of production they are susceptible of microbial contamination. In Europe an increasing number of food recalls and disease outbreaks associated with dehydrated herbs have been reported in recent years. In this study the microbial quality of 99 samples of aromatic herbs (bay leaves, basil, coriander, oregano, parsley, Provence herbs, rosemary and thyme) collected from retail shops in the region of Algarve (Southern Portugal) was assessed. All the samples were tested by conventional methods and were assayed for the total count of aerobic mesophilic microorganisms, *Salmonella* spp., *Escherichia coli*, coagulase-positive staphylococci and filamentous fungi. Almost 50 % of the herbs did not exceed the aerobic mesophilic level of  $10^4$  CFU/g. The fungi count regarded as unacceptable ( $10^6$  CFU/g) was not found in any of the tested herbs, while 84 % of the samples ranged from  $\leq 10^2$  to  $10^4$  CFU/g. No sample was positive for the presence of *Salmonella* spp., *Escherichia coli* and staphylococci. The results are in compliance with the European Commission criteria although they point out to the permanent need of surveillance on the good standards of handling/cooking practices as well as the importance of avoiding contamination at production, retailing and distribution. The microbiological hazards associated with the pathogenic and toxigenic microbiota of dried herbs remain as a relevant public health issue, due to the fact that they are added to foods not submitted to any following lethal procedure. Control measures should be adopted in order to ensure that all phases of their supply chain respect the food safety standards.

---

**Keywords:** microbial quality; aromatic dried herbs; low moisture foods; foodborne pathogens; total count of microorganisms

---

## 1. Introduction

Production and trade of dried aromatic herbs have increased in the last quarter century and these compounds are currently present in almost all kinds of food preparation where they intensify flavor, aroma and color or provide a distinct sensorial characteristic [1]. Consumption of these commodities has also significantly increased in the last decade mainly due to a global trend for ethnic and regional cuisines where dry herbs enhance the particular characteristics of typical dishes, such as for example basil and oregano in Italian and Greek culinary [2]. In the Mediterranean countries they play an important role in the traditional diet and apart from being used as seasoning condiments they have been recognized as a valuable source of high value nutritional compounds such as antioxidants and essential oils [3].

Similar to other food products from agricultural origins dried aromatic herbs can be naturally exposed to microbial contamination during any stage of production since pre-harvesting to harvesting, processing, distribution, storage and use by consumers [4]. The dried herbs production chain is vulnerable to accidental or natural microbial contamination whose origin is hard to determine. Although present in food products only in minor quantities these ingredients can be carriers of pathogenic microorganisms and of some toxin and spore producers. Currently an increasing number of studies have demonstrated the ability of some foodborne pathogens such as *Salmonella* spp., *Escherichia coli* 0157:H7, *Cronobacter sakazaki*, *Staphylococcus aureus* to survive and grow in some aromatic dried herbs [5,6]. Recently new research showed the presence of *Bacillus cereus* in ten different aromatic dried herbs commercialized in Poland [7]. When these products are rehydrated and subjected to a favorable temperature range those microorganisms may find the ideal conditions to multiply and cause a foodborne infection or intoxication. In addition, pathogens which survive and grow in low moisture foods can acquire a virulence profile such as an increased thermal resistance, a higher tolerance to antimicrobials and a reduced infectious dose [5]. The safety risk is even higher considering those commodities are usually added directly to ready-to-eat foods/meals which do not require any heat treatment before consumption. Herbs and spices are the main source of spore-forming bacteria in products such as soups, cooked or stewed dishes and sauces [8]. These foodborne pathogens are also known for their capacity to survive and grow in desiccation conditions and persist in food processing environments. Foods from non-animal origin such as dried herbs along with spices, fruits, vegetables, seeds, nuts and cereals are recognized as critical hazards and have been associated with large outbreaks such as the 2011 VTEC (verotoxin-producing *Escherichia coli*) outbreak in Germany which cause 3793 cases, 2353 hospitalizations and 53 deaths [9]. According to the European Rapid Alert System for Foods and Feeds [10] the dried herbs and spices food category had one of the highest numbers of recalls and outbreak notifications in the recent years. Since 2013 more than 500 notifications have been associated with *Salmonella* spp. present in dried herbs and spices [11]. Therefore, most of the food recalls refer to processed foods which did not experience any lethal procedure such as thermal, chemical or radiation treatment.

Currently data on the microbiological quality of these products are scarce and not available. On the other hand, the drying process of herbs leads to a stable product which can attend the consumer's

demand during the whole year. Therefore, the consumer's assumption on the innate safety of low-moisture food products such as aromatic dried herbs may rise the opportunity for the occurrence of cross contamination. In fact, little is known about the potential impact of dried herbs on the survival, growth and tenacity of foodborne pathogenic microorganisms at this kind of food matrix. Currently legal regulations in the European Union still do not include microbiological criteria for these commodities but the European Commission (EC/2004) [12] refer that food products should be free of pathogenic bacteria which may represent a public health risk for the consumer. This recommendation states that *Salmonella*, should be absent in 25 g of a dried herb sample, *Escherichia coli* and *Bacillus cereus* should be present at less than  $10^2$  CFU/g and the total count of mesophilic bacteria should be acceptable between  $10^2$  and  $10^4$  CFU/ g of product. The contamination of dried herbs by yeasts and molds represents also a food safety hazard; some of those microorganisms may produce toxins originated from improper harvesting and storage. Some of these mycotoxins may cause adverse effects on human and animal health due to their neurotoxicity and the risk of being potentially carcinogenic and/or teratogenic [13].

Although dried herbs are present as minor ingredients in food/meal composition they can potentially contaminate a wide range of products. Therefore, the maintenance of a good microbial quality of dried herbs is crucial for avoiding the presence of these pathogens in food supply chain and reducing the public health concerns and hazards associated with their potential risks. Since food safety is one of the greatest concerns regarding these commodities the aim of this study was to evaluate the microbial quality at the point-of-sale of a group of eight types of dehydrated aromatic herbs commercialized at retail shops in the region of Algarve (Southern Portugal) and to assess their compliance to the current food safety standards.

## 2. Materials and methods

### 2.1. Sampling and sampling preparation

The commercialized dried aromatic herbs were purchased in individual packs of 8 g to 40 g from retail shops within the region of Algarve (Southern Portugal) and were subjected to examination before their use to date. They were transported to the laboratory in a cold box (below 4 °C) and analyzed immediately. In total the experimental material included 99 samples composed of eight different dried aromatic herbs (basil: 13; bay leaves: 12; coriander: 9; oregano: 16; parsley: 14; Provence herbs: 10; rosemary: 10; and thyme: 15). These samples were purchased from different companies and distinct brands and batches.

### 2.2. Microbiological analyses

The samples were determined for: Total count of aerobic mesophilic bacteria, total count of molds, *Escherichia coli*, *Salmonella* spp. and coagulase positive staphylococci. The assays were performed according to the ISO International Standards methods and the microbiological status of the dried aromatic herbs was assessed using the criteria outlined in Recommendation 2004/24/EC (European Commission (EC) 2004a and 2004b). The herb samples were submitted to decimal dilutions and aliquots were inoculated in the culture media proposed by the ISO International Standards. The aerobic mesophilic counts were performed according to the ISO 4833 (2003) and

aliquots were pour-plated in Plate Count Agar and incubated at 30 °C. The molds were enumerated following the ISO 21527-1(2008), spreading the aliquots on Dichloran Rose Bengal Chloramphenicol Agar following incubation at 25 °C. *Escherichia coli*  $\beta$ -glucuronidase positive was counted through the ISO 16649-2 (2001) using Tryptone Bile X-Glucuronide at 44 °C; the number of the coagulase positive staphylococci was determined according to the ISO 6888-1 (1999; Amd 1:2003) using Baird-Parker media followed by incubation at 37 °C and *Salmonella* sp. was detected according to the ISO 6579 (2002). For each sample, all the analyses were performed in duplicated.

### 3. Results and discussion

The hygienic quality of 99 samples of dehydrated aromatic herbs purchased in retail stores in Southern Portugal was assessed and the achieved results are summarized in Tables in 1 and 2. The mesophilic total counts ranged from under the detection limit to  $1.2 \times 10^7$  cfu/g and 47% of the samples did not exceed the  $10^4$  cfu/g level (Table 1).

**Table 1.** Total count of mesophilic microorganisms in the dried aromatic herbs samples (n—Number of samples of each species, <sup>a</sup>—Range in cfu per gram of dried herbs, <sup>b</sup>—Counts are given in cfu/g). The numbers in the table correspond to the samples in the indicated microbial range.

Samples	n	$\leq 10^{2a}$	$>10^2-10^4$	$>10^4-10^6$	$>10^6$	Range <sup>b</sup> (cfu/g)	Media <sup>b</sup> (cfu/g)
Basil	13	1	2	6	4	$<10-1.2 \times 10^7$	$1.6 \times 10^6$
Bay leaves	12	0	9	2	1	$1.9 \times 10^2-2.8 \times 10^6$	$2.5 \times 10^5$
Coriander	9	0	6	3	0	$1.3 \times 10^2-5.9 \times 10^5$	$7.4 \times 10^4$
Oregano	16	1	7	7	1	$4.5 \times 10-1.4 \times 10^6$	$2.3 \times 10^5$
Parsley	14	0	8	6	0	$8.5 \times 10^2-1.9 \times 10^5$	$3.2 \times 10^4$
Provence herbs	10	1	1	7	1	$<10-3.6 \times 10^6$	$4.2 \times 10^5$
Rosemary	10	2	4	3	1	$<10-2.3 \times 10^6$	$2.6 \times 10^5$
Thyme	15	0	5	7	3	$2.8 \times 10^2-3.1 \times 10^6$	$6.4 \times 10^5$

The lowest mesophilic microorganism counts were found on coriander and parsley ( $7.4 \times 10^4$ ,  $3.2 \times 10^4$  cfu/g) whereas the highest microbial counts ( $1.6 \times 10^6$ ,  $6.4 \times 10^5$  and  $4.2 \times 10^5$  cfu/g) were associated to basil, thyme and provence herbs respectively. Basil and thyme were the only herbs with higher counts than 6 Log cfu/g which represents only 11.1% of the samples. In case of bay leaves, oregano, province herbs and rosemary, just one sample of each exceeded  $10^6$  cfu/g, the limit value according to the International Commission on Microbiological Specifications for Foods above which the samples are classified as having unsatisfactory microbiological quality [14] (ICMSF, 1974). Other studies found the highest mesophilic microorganisms' counts in oregano, tarragon and basil [6] or in rosemary and bay leaf [15]. The diversity of results might be explained by the crop production, harvesting, storage conditions, and a complex distribution network of the herbs, which differ in each geographic region.

The mesophilic counts are considered a useful microbiological indicator of the initial quality of raw materials and whether good practices of manufacturing have been performed [16]. Therefore, the production and processing of herbs deal with several critical steps to ensure safe handling procedures and prevent microbial contamination. In fact, herbs are in general added to foods without further

cooking or processing and their microbial quality is of major importance to avoid spoilage and safety hazards.

In general, significant differences in results obtained by different sources may be addressed to diverse factors. Some aromatic dried herbs are rich sources of phenolic compounds and essential oils whose antimicrobial properties have been demonstrated by a large number of studies [17–19]. This might have an inhibitory effect on the survival and proliferation of the microbiota.

Concerning the filamentous fungi (Table 2) their counts were lower than the levels of mesophilic populations, ranging from under the detection limit to  $2.6 \times 10^5$  cfu/g. None of the samples showed higher counts than 6 Log cfu/g which is the limit above which some countries microbiological guidelines classify as unacceptable (Critères microbiologiques des d'énres alimentaires, Luxembourg, 2007). In 32.3% of the samples the level of filamentous fungi was inferior to 10 cfu/g (under the detection limit of the method). Parsley and coriander were the herbs with the lowest molds media counts ( $1.1 \times 10^2$  and  $6.9 \times 10^2$ , respectively) and bay leaves, thyme and rosemary, presented the highest counts ( $2.3 \times 10^4$ ,  $2.0 \times 10^4$  and  $1.6 \times 10^4$  cfu/g, respectively). The occurrence of molds in herbs, partially favoured by the climatic conditions (high temperature and humidity) of production areas can affect food quality and safety, as some species produce mycotoxins [13,16] and can potentially cause food poisoning. On the other hand molds may change the appearance, flavor and texture of the foods where they grow on affecting their organoleptic properties.

**Table 2.** Filamentous fungi in the dried aromatic herbs samples (n—Number of samples of each species, <sup>a</sup>—Range in cfu per gram of dried herbs, <sup>b</sup>—Counts are given in cfu/g). The numbers in the table correspond to the samples in the indicated microbial range.

Samples	n	$\leq 10^2$ <sup>a</sup>	$>10^2-10^4$	$>10^4-10^6$	$>10^6$	Range <sup>b</sup> (cfu/g)	Media <sup>b</sup> (cfu/g)
Basil	13	7	3	3	0	$<10-3.0 \times 10^4$	$5.3 \times 10^3$
Bay leaves	12	3	8	1	0	$<10-2.0 \times 10^5$	$2.3 \times 10^4$
Coriander	9	6	3	0	0	$<10-3.0 \times 10^3$	$6.9 \times 10^2$
Oregano	16	7	4	5	0	$<10-6.0 \times 10^4$	$1.2 \times 10^4$
Parsley	14	12	2	0	0	$<10-9.5 \times 10^2$	$1.1 \times 10^2$
Provence herbs	10	4	5	1	0	$<10-6.0 \times 10^4$	$6.5 \times 10^3$
Rosemary	10	4	4	2	0	$<10-1.3 \times 10^5$	$1.6 \times 10^4$
Thyme	15	8	5	2	0	$<10-2.6 \times 10^5$	$2.0 \times 10^4$

The analysis of the overall results obtained in the present study reveals an average good microbiological quality of the herbs and the majority of samples were of satisfactory or acceptable microbiological quality according to the Recommendation 2004/24/EC (European Commission (EC) 2004a).

No pathogens such as *Salmonella* spp., a food safety parameter at the marketplace and *Escherichia coli*, a food hygiene parameter [20,21] (EC, 2005; EC 2007), were detected in any of the herbs. According to increasing data based on [22,23] and on EU summary reports between 2004 and 2014 the presence of *Salmonella* spp. has been detected in several kinds of dried herbs. *Salmonella*-related outbreaks where herbs were the principal foodborne source have also been reported [24]. In fact 23% of herbs and spices RASFF reports, between 2004 and 2014, were associated to pathogenic microorganisms among which *Salmonella* spp. corresponded to 74%. Given the high ability of this

pathogen to survive in low–moisture environments for long periods and persist in food processing facilities *Salmonella* has had the highest number of RASFF notifications associated to the dried herbs category [8]. The contamination of dried herbs by *Escherichia coli*, referred by [23,24] and associated to a 12% frequency of RASSF reports in the same period, was not observed in any of the present samples indicating an acceptable hygiene quality of the analyzed herbs. On the other hand, pathogens such as *Staphylococcus aureus*, a toxigenic member of the coagulase positive staphylococci group and a microorganism associated to improper food handling, according to [15,25] has also been isolated from dried herbs. The absence of this microorganism at the present assay may reinforce the satisfactory microbiological quality of these products. However, the antimicrobial effect of dried herbs against pathogens such as rosemary, oregano and thyme on *Salmonella* spp., *Escherichia coli* and *Staphylococcus aureus* has been well documented [17,26–30] and further investigation may be needed in order to clarify the real meaning of negative results in pathogens screening tests.

#### 4. Conclusions

Dried aromatic herbs like any other low–moisture food, which once were considered microbiologically safe, have shown in recent years to be vehicles of foodborne pathogens implicated in disease outbreaks and recalls. The implementation of good hygienic practices along the whole supply chain, besides the application of validated lethal process before distribution at retail is fundamental to ensure their food safety standards.

The present study sheds some light on the importance of monitoring and keeping the surveillance of dried aromatic herbs supply chain but also raises several questions to address in future research such as the role of food matrix as the substrate for microbial growth as well as the inhibitory effects they might have on their transmission and virulence.

Further research is required to elucidate the interactions between natural antimicrobial compounds of dried herbs and the food matrix and also the possible extension of the inhibitory effects. This might be important to gain a real insight of microbial level of contaminations in dried herbs and preventing the under estimation of microbe viable counts. More data should be obtained in order to evaluate the microbiological quality of dried herbs used in dairy and meat industry as well as in ready-to-eat-foods at commercial and home level.

#### Acknowledgments

This study received Portuguese national funds from FCT-Foundation for Science and Technology through project UID/BIA/04325/2019 (Center for Mediterranean Bioresources and Food, MeditBio).

#### Conflict of interest

All authors declare no conflict of interest.

## References

1. Szűcs V, Szabó E, Lakner Z, et al. (2018) National seasoning practices and factors affecting the herb and spice consumption in Europe. *Food Control* 83: 147–156.
2. Kaefer CM, Milner JA (2008) The role of herbs and spices in cancer prevention. *J Nutr Biochem* 19: 347–361.
3. Opara EI, Chohan M (2014) Culinary herbs and spices: Their bioactive properties, the contribution of polyphenols and the challenges in deducing their true health benefits. *Int J Mol Sci* 15: 19183–19202.
4. Minarovicová J, Cabicarová T, Kačková E, et al. (2018) Culture-independent quantification of pathogenic bacteria in spices and herbs using real-time polymerase chain reaction. *Food Control* 83: 85–89.
5. Bechaut LR, Komitopoulou E, Beckers H, et al. (2013) Low-water activity foods: Increased concern as vehicles of foodborne pathogens. *J Food Protec* 76: 150–172.
6. Garbowska M, Berthold-Pluta A, Stasiak-Rózanska J (2015) Microbiological quality of selected spices and herbs including the presence of *Cronobacter* spp. *Food Microbiol* 49: 1–5.
7. Berthold-Pluta A, Pluta A, Garbowska M, et al. (2019) Prevalence and toxicity of *Bacillus cereus* in food products from Poland. *Foods* 8: 269, 1–12.
8. Van Doren JM, Neil K, Parish M, et al. (2013) Foodborne illness outbreaks from microbial contaminants in spices, 1973–2010. *Food Microbiol* 36: 456–464.
9. European Food Safety Authority & European Centre for Disease CPrevention and Control (2013) The European Union summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in 2011. *EFSA J* 11: 250. Available from: <http://dx.doi.org/10.2903/j.efsa.2013.3129>.
10. Rapid Alert System for Food and Feed (2018) The Rapid Alert System Report for food and Feed annual report. Luxembourg: European Commission. Available from: <http://dx.doi.org/10.2875/945461>.
11. Lins P (2018) Detection of *Salmonella* spp. in spices and herbs. *Food Control* 83: 61–68.
12. European Commission (EC) (2004) Commission Recommendation of 19 December 2003 concerning a coordinated programme for the official control of food stuffs for 2004 (2004/24/EC). *The Official Journal of European Union* 16: 29–37.
13. Kabac B, Dobson ADW (2017) Mycotoxins in spices and herbs—An update. *Crit Rev Food Sci Nutr*. 57: 18–34.
14. International Commission for Microbiological Specifications for Foods, IMCFS (1974) *Microrganisms in Foods 2: Sampling for Microbiological Analysis: Principles and Specific Applications*. Toronto, Canada: Blackwell Scientific Publications.
15. Sospedra I, Soriano JM, Mañes J (2010) Assessment of the microbiological safety of dried spices and herbs commercialized in Spain. *Plant Foods Hum Nutr* 65: 364–368.
16. Vitullo M, Ripabelli G, Fanelli M, et al. (2011) Microbiological and toxicological quality of dried herbs. *Lett Appl Microbiol* 52: 573–580.
17. Alves-Silva JM, Santos SMD, Pintado ME, et al. (2013) Chemical composition and in vitro antimicrobial, antifungal and antioxidant properties of essential oils obtained from some herbs widely used in Portugal. *Food Control* 32: 371–378.
18. El-Zaedi H, Martínez-Tomé J, Calín-Sanches Á, et al. (2016) Volatile composition of essential oils from different aromatic herbs grown in Mediterranean regions of Spain. *Foods* 5: 41.

19. Teixeira B, Marques A, Ramos C, et al. (2013) Chemical composition and bacterial and antioxidant properties of commercial essential oils. *Ind Crop Prod* 43: 587–595.
20. European commission (EC): Regulation 2073/2005. *EU Official Journal* 15/11/2005. L.338 1–26.
21. European commission (EC): Regulation 1441/2007. *EU Official Journal* 5/12/2007. L.332 1–12.
22. Zweifel C, Stephan R (2012) Spices and herbs as source of *Salmonella*-related foodborne diseases. *Food Res Int* 45: 76–99.
23. Sagoo SK, Little CL, Greenwood M, et al. (2009) Assessment of the microbiological safety of dried spices and herbs from production and retail premises in the United Kingdom. *Food Microbiol* 26: 39–43.
24. Banach JL, Stratakou I, van der Fels-Klerx HJ, et al. (2016). European alerting and monitoring data as inputs for the risk assessment of microbiological and chemical hazards in spices and herbs. *Food Control* 69: 237–249.
25. Thanh MD, Frenzel H, Fetsxh A, et al. (2018) Tenacity of *Bacillus cereus* and *Staphylococcus aureus* in dried spices and herbs. *Food Control* 83: 75–84.
26. Burts S (2004) Essential oils: Their antibacterial properties and potential applications in foods—A review. *Int J Food Microbiol* 94: 223–253.
27. Calo JR, Crandall PG, O'Bryan CA, et al. (2015) Essential oils as antimicrobials in food system—A review. *Food Control* 54: 111–119.
28. Garcia-Diez J, Alheiro J, Falco V, et al. (2016) Chemical characterization and antimicrobial properties of herbs and spices essential oils against pathogens and spoilage bacteria associated to dry-cured meat products. *J Essent Oil Res* 1: 1–9.
29. Korshidian N, Yousefi M, Khanniri E, et al. (2018) Potential application of essential oils as antimicrobial preservatives in cheese. *Innov Food Sci Emerg Technol* 45: 63–72.
30. Tajkarimi MM, Ibrahim SA, Cliver DO (2010) Antimicrobial herb and spice compounds in food. *Food Control* 21: 1199–1218.



AIMS Press

© 2020 the Author(s), licensee AIMS Press. This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>)