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From bench to bedside: in vitro and in vivo evaluation of a neonate-focused nebulized surfactant delivery strategy

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Abstract

Background: Non-invasive delivery of nebulized surfactant has been a neonatology long-pursued goal. Nevertheless, the clinical efficacy of nebulized surfactant remains inconclusive, in part, due to the great technical challenges of depositing nebulized drugs in the lungs of preterm infants. The aim of this study was to investigate the feasibility of delivering nebulized surfactant (*poractant alfa*) in vitro and in vivo with an adapted, neonate-tailored aerosol delivery strategy.

Methods: Particle size distribution of undiluted *poractant alfa* aerosols generated by a customized eFlow-Neos nebulizer system was determined by laser diffraction. The theoretical nebulized surfactant lung dose was estimated in vitro in a clinical setting replica including a neonatal continuous positive airway pressure (CPAP) circuit, a cast of the upper airways of a preterm neonate, and a breath simulator programmed with the tidal breathing pattern of an infant with mild respiratory distress syndrome (RDS). A dose-response study with nebulized surfactant covering the 100–600 mg/kg nominal dose-range was conducted in RDS-modelling, lung-lavaged spontaneously-breathing rabbits managed with nasal CPAP. The effects of nebulized *poractant alfa* on arterial gas exchange and lung mechanics were assessed. Exogenous alveolar disaturated-phosphatidylcholine (DSPC) in the lungs was measured as a proxy of surfactant deposition efficacy.

Results: Laser diffraction studies demonstrated suitable aerosol characteristics for inhalation (mass median diameter, MMD = 3 μm). The mean surfactant lung dose determined in vitro was 13.7% ± 4.0 of the 200 mg/kg nominal dose. Nebulized surfactant delivered to spontaneously-breathing rabbits during nasal CPAP significantly improved arterial oxygenation compared to animals receiving CPAP only. Particularly, the groups of animals treated with 200 mg/kg and 400 mg/kg of nebulized *poractant alfa* achieved an equivalent pulmonary response in terms of oxygenation and lung mechanics as the group of animals treated with instilled surfactant (200 mg/kg).

Conclusions: The customized eFlow-Neos vibrating-membrane nebulizer system efficiently generated respirable aerosols of undiluted *poractant alfa*. Nebulized surfactant delivered at doses of 200 mg/kg and 400 mg/kg elicited a pulmonary response equivalent to that observed after treatment with an intratracheal surfactant bolus of 200 mg/kg. This bench-characterized nebulized surfactant delivery strategy is now under evaluation in Phase II clinical trial (EUDRACT No.:2016–004547-36).

Keywords: Nebulized surfactant, *Poractant alfa*, eFlow-Neos, Respiratory distress syndrome, CPAP, Neonatal ventilation, Nebulizer

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Background

Current trends in the management of preterm infants with Respiratory Distress Syndrome (RDS) aim for gentler, non-invasive interventions as first-line treatment options. Over the last years, the classic ways of providing artificial respiratory support, oxygen therapy, and exogenous surfactant have been systematically challenged [1–5] in order to reduce the iatrogenic effects that contribute to the development of chronic lung disease [6, 7]. Particularly, clinical research has been directed to reduce the use of mechanical ventilation [8–10], a major risk factor associated with lung inflammation and the development of Broncho-Pulmonary Dysplasia (BPD). Therefore, non-invasive ventilation techniques are gaining momentum as the primary treatment of RDS [10–13].

Surfactant replacement therapy has also undergone a clear evolution towards less invasive administration protocols [14]. Aerosol delivery of exogenous surfactant in combination with nCPAP has been proposed as a feasible, truly non-invasive surfactant delivery method [15, 16]. The theoretical advantages of nebulization include minimal manipulation of the respiratory tract, improved pulmonary distribution [17], and the avoidance of the acute airway fluid load occurring immediately after surfactant instillation [16]. A gradual surfactant administration by nebulization may reduce some of the side effects associated with surfactant instillation, e.g. transient airway obstructions and reflux, hypercarbia, and hypoxia [16], and may therefore contribute to more stable systemic and cerebral hemodynamics [18–20]. Experimental studies with aerosolized surfactant, however, have shown controversial results, ranging from no effect at all of the aerosolized surfactant to an equivalent performance compared to intratracheal bolus instillation [17–30]. A few clinical studies have also attempted to deliver nebulized surfactant to preterm infants managed with CPAP [31–35]. These studies enrolled a limited number of patients, applied heterogeneous surfactant administration protocols, and used different nebulizers. So far, these clinical studies have demonstrated that nebulized surfactant is well tolerated and can be safely administered during non-invasive ventilation.

A great challenge in the field of aerosol delivery is to improve the extremely low lung deposition of inhaled drugs in preterm neonates, which has been reported to be lower than 1% of the nominal dose [36, 37]. Preterm neonates are forced nasal breathers, have a low functional residual capacity, high respiratory rate (RR), low tidal volume, and small airway caliber [38]. All these factors reduce the residence time of aerosol particles within the airways, which significantly reduces lung deposition. In addition, the bias flow of the ventilation support can dilute the concentration of surfactant droplets, leading to a relatively high surfactant loss through the

expiratory limb of the CPAP circuit [38]. To overcome these limitations, research efforts have been directed to characterize, even further, the particular challenges posed by preterm infants to aerosol delivery, as well as to develop novel aerosol-generating devices and infant-focused strategies aimed at improving lung deposition of nebulized surfactant [39–45].

In the present work, we investigated the feasibility of delivering nebulized surfactant *in vitro* and *in vivo* with a customized, neonate-focused aerosol delivery strategy. For that purpose, we first analyzed the aerosol characteristics of nebulized *poractant alfa* (Curosurf®, Chiesi Farmaceutici SpA, Parma, Italy) generated by a customized eFlow-Neos vibrating-membrane nebulizer system (PARI Pharma, Starnberg, Germany) under physiological relative humidity (RH) conditions. We further investigated surfactant deposition in a realistic *in vitro* CPAP circuit, which included a cast of the upper airways of a preterm infant and a breath simulator programmed with a neonatal breathing pattern. *In vitro* data were used to implement surfactant nebulization in a nCPAP-supported RDS animal model, in which a dose-response study was performed to assess the pulmonary efficacy of nebulized surfactant.

Materials and methods

Surfactant particle size characterization

The particle size distribution of surfactant aerosols (*poractant alfa*, 80 mg/mL) generated by the customized eFlow-Neos nebulizer system was determined *in vitro* by laser diffraction (Helos/BF, Sympatec GmbH, Clausthal-Zellerfeld, Germany). Briefly, this technique measures the angular variation in the intensity of scattered light when a laser is directed through a particle dispersion. The angular variation in the intensity of scattered light is inversely proportional to particle size, which allows determining the size distribution of the particles in the surfactant cloud. To perform laser diffraction experiments, the customized eFlow-Neos was filled with 2 mL of undiluted surfactant that was then continuously nebulized towards the detection area. Laser diffraction experiments were conducted at $30\% \pm 5$ or, alternatively, at $90\% \pm 5$ relative humidity (RH) conditions, at 37 °C. Each of these experiments was repeated five times using independent nebulizer units. The mass median diameter (MMD), the geometric standard deviation (GSD), and the fine particle fraction (FPF) were used to characterize surfactant aerosols.

Benchmark breath simulation experiments

A set-up composed of a CPAP system (Fabian HFO, Acutronic, Zug, Switzerland) with a humidifier (MR 730, Fisher & Paykel Healthcare), a customized eFlow-Neos nebulizer system, a cast of the upper airways (nose-

throat) of a premature infant (PrINT model) [41], infant nasal prongs (3520, Fisher & Paykel Healthcare), surfactant collection filters (PARI Filter PAD PZN: 00632160), and a breath simulator (Compas 2, PARI Pharma, Starnberg, Germany) [46, 47] was implemented in order to simulate a clinical setting of surfactant nebulization under non-invasive neonatal ventilation conditions. The PrINT model was developed by Minocchieri et al. by 3D reconstruction of a magnetic resonance of a premature infant born after 32 weeks of gestation (body weight of 1750 g) [41]. The infant nose-throat cast was 3D printed (Izu1 prototypen, Dornbirn, Austria) as a solid substance (material: DSM water clear ultra 10,122). The nose area was silicon-coated to achieve a tight connection between cast and prongs (Additional file 1: Figure S1).

The nebulizer was placed between the Y piece and the nasal prongs using a custom-made adaptor. Before surfactant nebulization, the set up was systematically checked for air leaks. The temperature of the system was 37 °C and the RH was set at 90% ± 5. Bias flow rate and CPAP level were set at 5 L/min and 5 cmH₂O, respectively. The breath simulator was programmed with the following neonatal configuration: RR of 70 bpm, a tidal volume (V_T) of 8.9 mL (5 mL/kg), and an inhalation/exhalation ratio of 40/60.

A volume of 4.37 mL of *poractant alfa* (80 mg/mL), the equivalent volume of a 200 mg/kg dose for a 1750 g infant, was loaded into the customized nebulizer reservoir and was continuously nebulized. Surfactant collection filters were placed in the expiratory limb of the CPAP system (exhalation filter) as well as at the distal airway of the PrINT cast (Filter for lung dose, Fig. 1a). Lung dose (LD) was defined as the amount of surfactant collected within the filter placed at the distal airway of the PrINT cast (In-Filter). A backup-trap was installed between the PrINT cast and the In-Filter in order to collect the surfactant liquid film formed from the already impacted aerosol particles. The backup-trap did not have a significant impact on the aerosol flow. The amounts of surfactant deposited in the CPAP circuit, exhalation filter, backup-trap, nasal prongs, and residual surfactant remaining in the nebulizer were also determined, after disassembling of the analytical setup. After nebulization of the full surfactant dose, the required nebulization time was noted. A rinsing solvent containing 50 mg of potassium nitrate and 85% v/v isopropanol was used to recover the surfactant from each compartment of the analytical set up. Samples for quantitative analysis were prepared by defined dilutions. The main active constituent of *poractant alfa*, i.e. Phosphatidylcholine (PC), was determined as lead compound in order to quantify the surfactant amount. A validated high-pressure liquid chromatography (HPLC) method using external standard calibration was used for quantifying PC distribution

in the set-up compartments. The method is sensitive enough to determine PC in the applied concentration range from 20 to 2100 µg/ml for *poractant alfa* if a dual wavelength detector (WATERS 2487, Waters Corporation, Milford, US) is used.

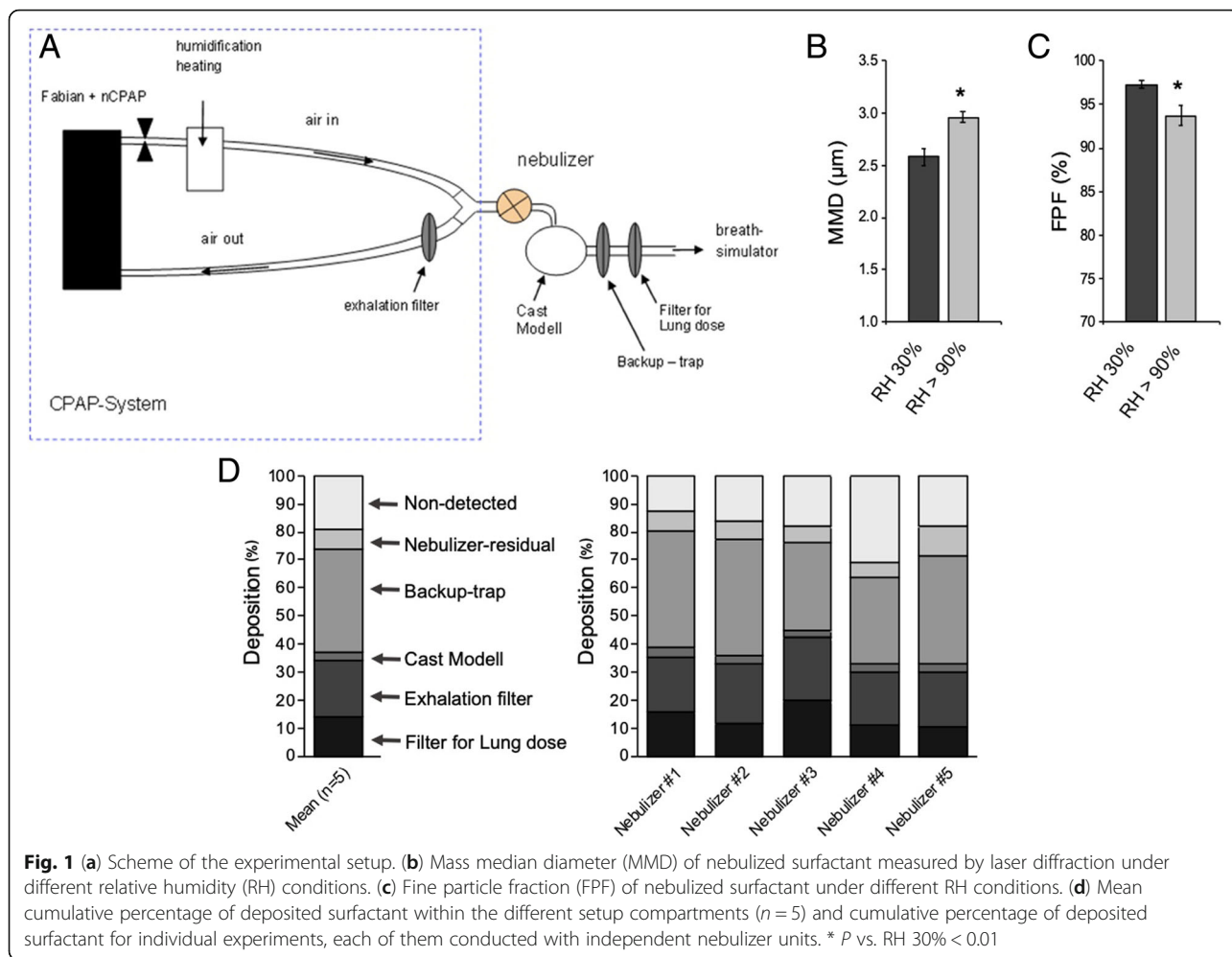
Animal preparation

The experiments were carried out in 6- to 7-week-old male rabbits (Charles River Laboratories, Calco, Italy). The experimental procedure was approved by the intramural Animal Welfare Body and the Italian Ministry of Health (Prot.n° 1300–2015-PR) and complied with the European and Italian regulations for animal care.

Rabbits (body weight of 1.5–2.5 kg) were initially sedated with intramuscular (i.m.) medetomidine (Domitor®, 2 mg/kg). The throat of the animals was first shaved and local anesthesia was applied in the anterior neck with lidocaine gel (Luan® 2.5%). Thirty minutes later, the animals received 50 mg/kg of ketamine (Imalgene 1000®, Merial-Boehringer Ingelheim, France) and 5 mg/kg of xylazine (Rompun®, Bayer, Germany) i.m. Rabbits, in the supine position, were intubated and stabilized on positive pressure ventilation (Fabian HFO, Acutronic, Zug, Switzerland) as previously described [48, 49]. Fraction of inspired oxygen (FiO_2) = 100%, Flow = 10 L/min, RR = 40 breaths/min, positive end-expiratory pressure (PEEP) = 3 cmH₂O, tidal volume (V_T) targeted to 7 ml/kg (with the peak inspiratory pressure, PIP, not exceeding 15 cmH₂O) and inspiratory time of 0.5 s. Airway flow, mean airway pressure (MAP) and V_T were monitored as long as the animals were intubated. Body temperature was constantly measured with a rectal probe and maintained at 37 degrees by placing a heating pad underneath the animal.

After endotracheal intubation, a catheter was inserted into the right jugular vein for continuous infusion of 1 mg/ml of ketamine and 0.1 mg/ml of xylazine in 0.9% saline solution (100 µl/min) to maintaining anesthesia at a steady level. Trometamol (tris-hydroxymethyl aminomethane, THAM, 1 M, Sigma-Aldrich, USA) was also infused during the surfactant depletion procedure for maintaining CO₂ level under control. A second catheter was inserted into the right carotid artery for blood sampling. After instrumentation, baseline blood gases were measured with an emogas analyzer (Radiometer Medical, Denmark).

Animals with an initial arterial oxygen partial pressure (PaO₂) value > 400 mmHg at PIP < 15 cmH₂O were included in the study. Repeated broncho-alveolar lavages (BALs) were performed by flushing the airways with 20 ml/kg of pre-warmed 0.9% NaCl solution, followed by a short recovery period in-between, until a PaO₂ value < 150 mmHg was reached. Then, if after 15 min of stabilization on mechanical ventilation



the respiratory failure was confirmed ($\text{PaO}_2 < 150$ mmHg, with PIP not exceeding 23 cmH₂O), the animal was included in the study.

Experimental groups

Forty-two animals were allocated to one of the six experimental groups. Animals in the **nCPAP group** ($n = 6$) were maintained in continuous nCPAP (Fabian HFO, Acutronic; 5 cmH₂O) for 180 min, using customized nasal prongs as an interface. Animals allocated in the **Inst-SURF group** ($n = 9$) received an intratracheal bolus of *poractant alfa* (200 mg/kg), using a modified InSurE approach, and were further managed with nCPAP (5 cmH₂O) for 180 min. After surfactant instillation, animals were managed with mechanical ventilation for 10 min ($V_T = 7$ mL/Kg, RR = 40/min and PEEP = 3 cmH₂O). Animals receiving nebulized surfactant were allocated in four different groups of escalating surfactant doses. The customized eFlow-Neos nebulizer was placed between the Y piece and the customized nasal prongs. Rabbits in the **Neb-**

SURF100 group ($n = 9$) received 100 mg/kg of nebulized *poractant alfa* while on nCPAP, whereas animals in the **Neb-SURF200** ($n = 9$), **Neb-SURF400 group** ($n = 9$), and **Neb-SURF600 group** ($n = 9$) received 200 mg/kg, 400 mg/kg, and 600 mg/kg of nebulized *poractant alfa*, respectively. All animals treated with nebulized surfactant were managed with nCPAP (5 cmH₂O) for 180 min, counting from the initiation of surfactant nebulization. For each animal experiment, a new customized eFlow-Neos nebulizer unit was used. The time required to nebulize the whole surfactant dose was annotated.

Gas exchange and respiratory indices

Arterial carbon dioxide partial pressure (PaCO_2), PaO_2 , and pH were measured (Radiometer Medical, Denmark) right after the induction of anesthesia (baseline) and after the induction of respiratory distress. Arterial blood gases were also measured right after placing the animals on nCPAP, 15 and 30 min after the start of nCPAP, and then every 30 min until the end of the experiment.

PaO₂ and PaCO₂ were also used to compute the oxygenation index (OI) and the ventilation efficacy index (VEI). OI and VEI were determined at those time intervals in which the animals were ventilated with a tracheal tube: at baseline, after inducing the respiratory distress and at the end of the 180 min follow-up period. Therefore, after the 180 min observational period, animals were shifted from nCPAP to invasive mechanical ventilation for a brief period of time, using exactly the same ventilation settings as at baseline (FiO₂ 100%, Flow = 10 L/min; RR = 40 bpm, PEEP = 3 cmH₂O, V_T targeted to 7 ml/kg, and inspiratory time of 0.5 s). The OI was calculated as follows:

$$\text{OI} = \text{FiO}_2 * \text{MAP} * 100 / \text{PaO}_2 \quad (1)$$

The VEI was calculated to evaluate the overall ventilation efficiency of mechanically ventilated animals independently from the ventilation settings [50]:

$$\text{VEI} = 3800 / [(\text{PIP} / \text{PEEP}) * \text{RR} * \text{PaCO}_2] \quad (2)$$

Ventilation indices were calculated at baseline (pre-BALs), after inducing respiratory distress (post-BALs), and at the end of the observational period.

Pulmonary mechanics

Dynamic compliance (C_{dyn}) was also determined in those time intervals in which the animals were ventilated with invasive mechanical ventilation: at baseline, resembling the “healthy” pulmonary status, right after inducing severe respiratory distress by repeated BALs, and at the end of the observational period, after re-intubation. C_{dyn} was calculated by dividing lung volume (ΔV, in mL) by the changes in pressure (ΔP) standardized by the animal’s weight.

$$C_{\text{dyn}} = \Delta V / (\Delta P * \text{Weight}) \quad (3)$$

Exogenous alveolar disaturated-phosphatidylcholine (DSPC) quantification

Intrapulmonary levels of exogenous surfactant were determined by administering *poractant alfa* labelled with U¹³C-PA-DPPC (*n* = 3 animals for each group). For tracer preparation, 0.2 mg (0.1 μl) of a suspension of U¹³C-PA-DPPC (Avanti Polar Lipids) in saline solution was carefully mixed with 200 mg (2.5 ml) of *poractant alfa*, which was then administered to distressed rabbits as described in the previous section. At the end of the experiment, BALs from all animal groups were collected and stored at -80 °C. Lipids were extracted according to the method described by Bligh and Dyer [51]. Disaturated-phosphatidylcholine (DSPC) was separated

by thin layer chromatography after treatment with osmium tetroxide [52]. Fatty acids of DSPC were derivatized as methyl ester by adding 2 mL 3 M HCl methanol and extracted with hexane. Quantitative analysis of DSPC was performed by a FID gas chromatograph (GC, Agilent 5890, Milan, Italy). The ¹³C enrichment of DSPC from the BALs was measured with gas chromatography-mass spectrometry (GC-MS, Agilent, 5973 Inert), as described elsewhere [53]. For data display, the intrapulmonary exogenous surfactant amount of the animals treated with nebulized surfactant was normalized taking the surfactant instillation group as reference.

Data analysis

In vitro and in vivo data are presented as mean ± SD. Raw data were analyzed and compared by repeated measures two-way analysis of variance (ANOVA) as a function of group and time, followed by Dunnett’s and Tukey’s *t* posthoc tests. Statistical analysis was performed using GraphPad software, version 6.0. A *P* < 0.05 was considered statistically significant.

Results

Particle size characterization of nebulized surfactant

Under low RH conditions (30% ± 5), the mean MMD of nebulized surfactant was 2.6 ± 0.1 μm. Increasing RH close to lung physiological conditions (90% ± 5) produced a slight, although significant, increase of the mean MMD to 3.0 ± 0.1 μm (Fig. 1b). Irrespective of RH conditions, all tested customized eFlow-Neos nebulizers showed a reproducible performance in terms of particle size with a consistent GSD of 1.5, indicative of a hetero-disperse aerosol yet with a narrow variability in terms of particle size distribution.

The slight increase of the MMD under high RH conditions was associated with a slight decrease of the FPF. This parameter represents the percentage of particles contained in the aerosol cloud with a diameter below 5 μm, also known as the respirable fraction. Mean FPF values determined under low and high humidity conditions were 97.2% ± 0.4 and 93.7% ± 1.1, respectively (Fig. 1c).

Benchmark breath simulation experiments

The mean nebulization time to deliver 4.37 mL of surfactant (80 mg/mL) was 18.9 min (range 13.3–23.3 min), which corresponds to an aerosol production rate of 18.5 mg phospholipid/min (range 15.0–26.3 mg/min). The determined mean LD was 13.7% ± 4.0 (range 10.2–19.8%, Fig. 1d and e), which would yield an estimated intrapulmonary surfactant dose of around 27 mg/kg in vivo if a 200 mg/kg dose was delivered. A fraction of generated aerosol was deposited on the backup-trap (36.7% ± 5.3), whereas 20.3% ± 1.5 was detected in the

exhalation filter of the CPAP circuit. Only $3.0\% \pm 0.6$ of the nebulized surfactant deposited within the PrINT cast and $7.2\% \pm 2.1$ was collected as a residual volume in the nebulizer chamber. Surfactant recovery was $81.0\% \pm 6.9$. The undetected surfactant fraction may have remained within the circuit as non-deposited particles that were further released to the environment during set up disassembling.

Gas exchange and respiratory indices

There were no significant differences between groups at baseline in terms of body weight, number of BALs needed to achieve the targeted respiratory failure, PaO_2 or C_{dyn} (Additional file 1: Table S1). All animals had similar mean PaO_2 at baseline (> 450 mmHg). However, following surfactant-depletion, a severe respiratory failure developed in all groups (mean $\text{PaO}_2 < 150$ mmHg). The PaO_2 in the nCPAP group remained extremely low, below 100 mmHg, despite a FiO_2 of 100%. PaO_2 values rapidly increased right after surfactant instillation, reaching mean PaO_2 values above 200 mmHg 15 min after treatment (Fig. 2a). PaO_2 values remained significantly higher throughout the whole experimental period in the Inst-SURF group compared to the nCPAP group ($P < 0.01$). In the groups of animals treated with nebulized surfactant, PaO_2 increased gradually over time achieving significantly higher values compared to untreated control animals ($P < 0.01$). However, whereas mean PaO_2 values of Neb-SURF200 and Neb-SURF400 groups were equivalent to the mean PaO_2 of the Inst-SURF group at 180 min, the mean PaO_2 of the Neb-SURF100 group was significantly lower ($P < 0.01$) compared to the latter group. The Neb-SURF600 group showed slightly lower mean PaO_2 values than Neb-SURF200 and Neb-SURF400 groups at 180 min.

Surfactant depletion by BALs was associated with severe hypercapnia and acidosis in all groups. Hypercapnia exacerbated even further in the nCPAP group, whereas surfactant-treated groups showed a downward trend of mean PaCO_2 values (Fig. 2b). However, a significant reduction of PaCO_2 in comparison to the nCPAP group was only observed for Inst-SURF, Neb-SURF200 and Neb-SURF400 groups ($P < 0.05$). Severe acidosis ($\text{pH} < 7.2$) persisted in the nCPAP group for the whole 180 min follow up period, with pH values that were significantly lower compared to any surfactant-treated group (Fig. 2c).

At baseline, OI was below 1.5 and the VEI above 0.15 in all study subjects. After BALs, mean OI increased to 10.84 ± 2.94 and the VEI dropped to 0.06 ± 0.01 (Fig. 3). At the end of the experimental period, OI was significantly lower in all surfactant-treated groups compared with the nCPAP group ($P < 0.01$). At 180 min, VEI remained low in the nCPAP group but increased in all surfactant-treated groups. However, a significant improvement of VEI

compared to the nCPAP group was only observed for Inst-SURF and Neb-SURF400 groups ($P < 0.05$).

Pulmonary mechanics

C_{dyn} significantly dropped in all groups after the induction of respiratory failure (Fig. 4). Mean C_{dyn} values measured after 180 min were relatively low in nCPAP (0.38 ± 0.06 mL/cmH₂O/kg) and Neb-SURF100 (0.46 ± 0.05 mL/cmH₂O/kg) groups. Conversely, mean C_{dyn} values significantly increased in the Inst-SURF, Neb-SURF200 and Neb-SURF400 groups after 180 min of non-invasive ventilation compared to the nCPAP group ($P < 0.05$).

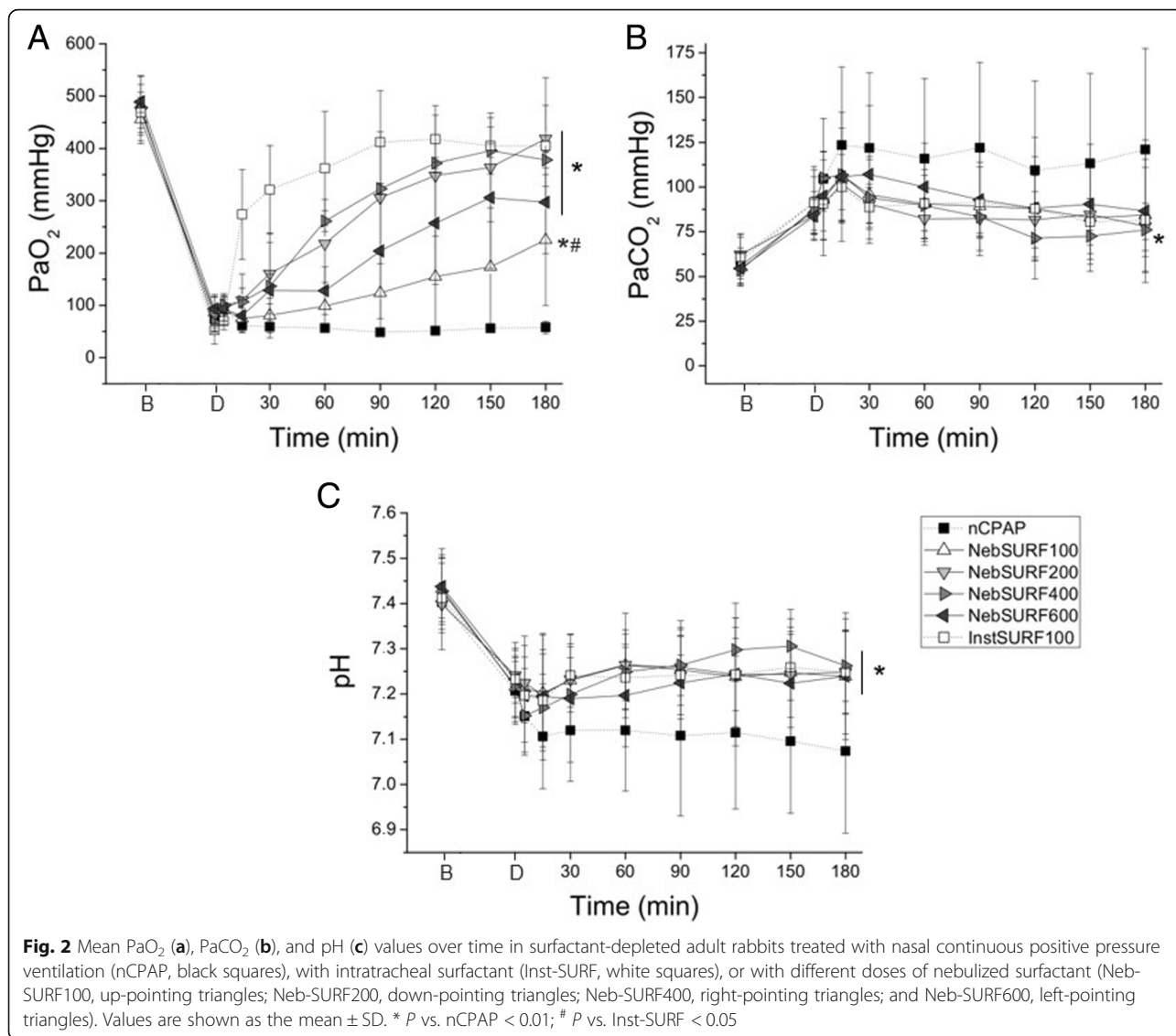
Alveolar DSPC quantification

The mean amount of exogenous DSPC in the Neb-SURF100 was 33-fold lower compared to the Inst-SURF group. Conversely, the Neb-SURF200 group registered an amount of exogenous DSPC 3.7-fold lower compared to the Inst-SURF group. Neb-SURF400 and Neb-SURF600 groups showed the highest mean intrapulmonary DSPC amounts just 1.8- and 1.2-fold lower compared to the animals which received intratracheal surfactant. These groups, however, showed a high inter-animal variability (Additional file 1: Figure S2).

Discussion

We investigated the feasibility of delivering undiluted nebulized surfactant with the customized eFlow Neos vibrating-membrane nebulizer system under realistic neonatal ventilation conditions. Laser diffraction studies demonstrated suitable aerosol characteristics for inhalation in terms of particle size distribution and fine particle fraction. The theoretical surfactant LD determined in vitro in a rational neonatal CPAP circuit, which included the simulated breathing pattern of a preterm infant with mild RDS, reached notable values for neonatal standards, as high as 19.8% of the nominal dose. Further, a dose-response study conducted in spontaneously-breathing lung-lavaged rabbits revealed a significant improvement of arterial oxygenation following treatment with nebulized surfactant. Particularly, the groups of animals treated with 200 mg/kg and 400 mg/kg of nebulized *poractant alfa* achieved an equivalent pulmonary response as the group of animals treated with instilled surfactant (200 mg/kg).

The concept of nebulized surfactant dates back to 1964 and was indeed one of the first attempts to treat preterm infants with RDS [54, 55]. Nevertheless, aerosol technology and lung deposition mechanisms were poorly understood at that time, which moved the field to the development of intratracheal instillation protocols [56, 57]. Since then, surfactant replacement therapy has undergone a constant evolution towards a



less invasive administration [4, 9, 58]. At present, the LISA method is experiencing worldwide clinical appraisal. This technique significantly reduces the need for mechanical ventilation and oxygen supplementation, and may as well reduce the incidence of BPD [59]. However, it still involves the acute fluid load of the infant’s airway, which may lead to transient airway obstruction, surfactant reflux, and hemodynamic imbalance [16].

In this study, we intended to develop a neonate-focused surfactant nebulization strategy, taking into consideration not only the particular characteristics of preterm infants but also the relatively high viscosity of surfactant. Finner et al. reported marked variability in surfactant dose output among single Aeroneb Pro (Aerogen) vibrating-membrane nebulizer units during a pilot clinical study evaluating the potential of aerosolized

surfactant for the prevention of RDS [34]. The authors attributed the variability in dose output to higher viscosity of surfactant compared to medications which are routinely delivered as aerosols. They discouraged the use of this device in combination with surfactant. Additional studies with nebulized surfactants have reported clogging of the pores of the vibrating membrane of the nebulizer, requiring surfactant dilution before nebulization [30, 60]. In the present study, the customized eFlow-Neos nebulizer system was able to produce respirable aerosols of *poractant alfa*, a highly concentrated surfactant, which is formulated at around 80 mg of phospholipids/mL. It is noteworthy that particle size distribution, as well as the output rate of the single customized eFlow-Neos nebulizing units, showed consistent results. We used new nebulizer units for each in vivo experiment, as well as 5 independent units for the in vitro

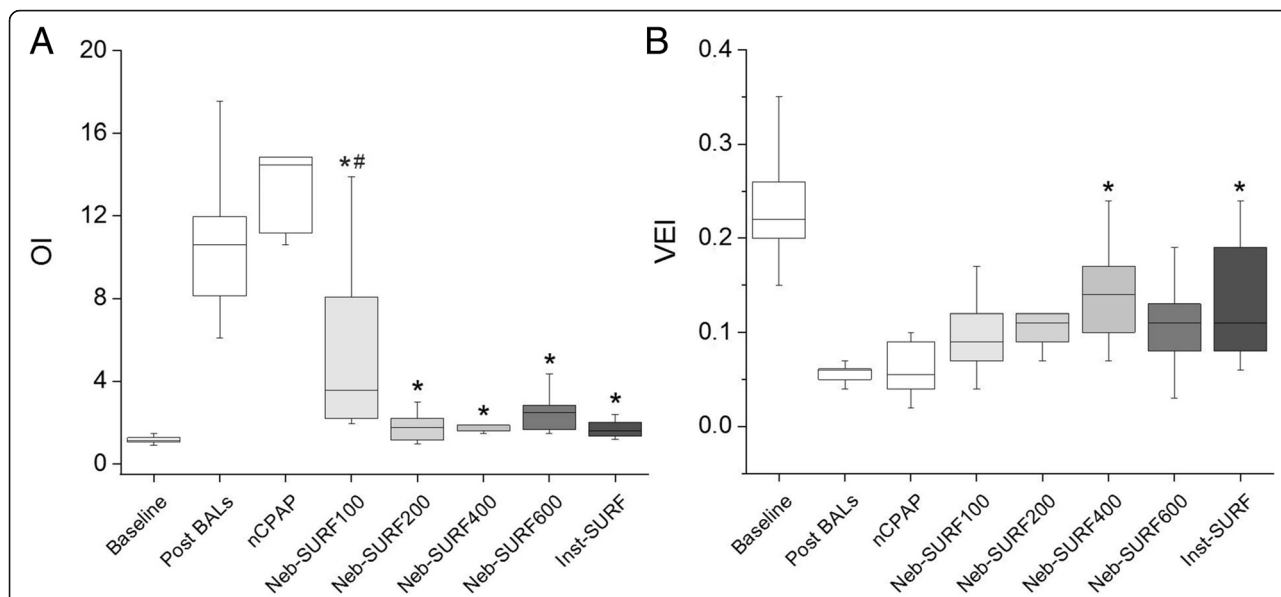


Fig. 3 Box-plots showing (a) the oxygenation index (OI) and (b) ventilation efficacy index (VEI) at baseline (from all animals), after inducing a respiratory distress (Post BALS, from all animals) and 180 min after treatment with just nasal continuous positive pressure ventilation (nCPAP), with different doses of nebulized surfactant (Neb-SURF100, Neb-SURF200, Neb-SURF400, Neb-SURF600) or with intratracheal surfactant (Inst-SURF). The boxes encompass the 25–75 percentiles. The horizontal line within the boxes represents the median. The whiskers indicate the maximum and minimum values observed for each group. * *P* vs. nCPAP *P* < 0.01; # *P* vs. Inst-SURF < 0.05

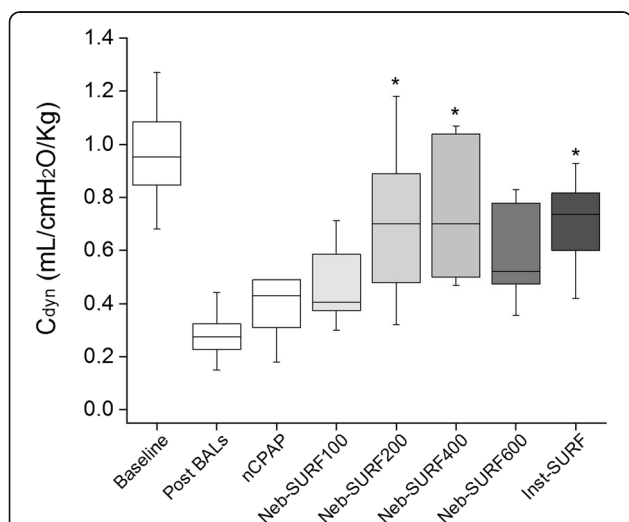


Fig. 4 Box-plots showing dynamic compliance (C_{dyn}) at baseline (from all animals), after inducing a respiratory distress (Post BALS, from all animals) and 180 min after treatment with just nasal continuous positive pressure ventilation (nCPAP), with different doses of nebulized surfactant (Neb-SURF100, Neb-SURF200, Neb-SURF400, and Neb-SURF600) or with intratracheal surfactant (Inst-SURF). The boxes encompass the 25–75 percentiles. The horizontal line within the boxes represents the median. The whiskers indicate the maximum and minimum values observed for each group. The whiskers indicate the maximum and minimum values observed for each group. * *P* vs. nCPAP < 0.05

study, overall analyzing the performance of 41 nebulizer units. All units showed a consistent output rate and performed well with undiluted *poractant alfa* (Additional file 1: Figure S3).

We further tested the customized eFlow-Neos nebulizer in a rational neonatal CPAP circuit. In order to maximize the surfactant LD, we placed the nebulizer between the Y piece and the nasal prongs. Surfactant aerosols were thus generated close to the nares of the PrINT model. This cast resembles the upper airways (nose-throat) of a premature infant born at a gestational age of 32 weeks and is a useful tool to estimate the LD of nebulized medications intended for delivery to preterm infants [41]. Minocchieri et al. investigated the theoretical LD of inhaled budesonide generated with another vibrating-membrane nebulizer prototype (e-Flow, PARI GmbH Starnberg, Germany). They found that the LD dropped from around 62 to 9% of the nominal dose if the bias flow-rate was increased from 1 L/min to 10 L/min [41]. In comparison to their set up, we used humidified air, a bias flow of 5 L/min, a CPAP support of 5 cmH₂O, and simulated the breathing pattern of a neonate with mild RDS. Under such a challenging configuration, the minimal and maximal surfactant LDs were 10.2 and 19.8% of the nominal dose (200 mg/kg), respectively, corresponding to surfactant doses of 20.4 mg/kg and 39.6 mg/kg. Since individual nebulizer units showed a consistent performance in terms of surfactant output and particle size, we assume that the variability

in LD observed for each single experiment may be rather related to slight changes in the positioning of the components of the circuit (e.g. small changes in the angle of the prongs) or to slight variations of the air-flow of the circuit from experiment to experiment. Nevertheless, considering that intrapulmonary doses as low as 2 mg/kg of nebulized surfactant have been associated with a significant improvement of lung mechanics [22], we performed a dose-response study using four escalating nominal doses of *poractant alfa* (dose-range 100–600 mg/kg).

The in vivo study was conducted in lung-lavaged, spontaneously-breathing rabbits managed with nCPAP [48]. A particular advantage of this model is that the animals do not spontaneously recover from the respiratory distress if they are just supported with non-invasive ventilation [48, 49]. Therefore, any improvement of oxygenation or lung mechanics observed over time can be attributed to lung deposition of active exogenous surfactant. A surfactant dose of 100 mg/kg produced already a significant improvement of arterial oxygenation. According to our in vitro study, a nominal dose of 100 mg/kg would achieve a LD of 13.7 mg/kg. Nevertheless, this dose did not suffice to revert the respiratory distress. On the other hand, nebulization of 200 and 400 mg/kg of surfactant was associated with a significant improvement of arterial oxygenation, respiratory indices, and pulmonary mechanics, achieving a pulmonary response at the same level of that observed for animals treated with an intratracheal bolus of 200 mg/kg of surfactant. It is noteworthy that such an improvement was achieved administering nebulized surfactant to spontaneously-breathing animals managed with nCPAP. These results are highly encouraging from a clinical point of view.

As a proxy of surfactant lung deposition, we determined the amount of exogenous DSPC in three animals of each surfactant-treated group. Although the limited sample size precluded a sound statistical test, a clear dose-dependent trend could be identified across the groups of animals treated with nebulized surfactant. Linner et al. determined by means of gamma scintigraphy that approximately 90% of the instilled surfactant reaches the lungs following intratracheal administration [60]. Therefore, we normalized the exogenous DSPC amount of the groups of animals treated with nebulized surfactant to the DSPC amount determined for the Inst-SURF group. The relative exogenous DSPC quantified in the BALs of Neb-SURF200 was almost 4.0-fold lower compared to the BALs of the Inst-SURF group, even though the elicited pulmonary response was equivalent in both groups. We speculate that a more uniform intrapulmonary spreading of surfactant after nebulization

might have accounted for the similar pulmonary outcome observed for both groups. Our data also suggest that a nominal surfactant dose of 400 mg/kg may be of further advantage because it provides a good short term pulmonary response but also contributes to increase the intrapulmonary surfactant pool. Surprisingly, a further increase of the nominal dose to 600 mg/kg did not produce additional benefits in terms of gas exchange or pulmonary mechanics. This was an unexpected result. In other animal models, high surfactant doses have been associated with increased inflammation and neutrophil migration [61] as well as with a reduction of the antibacterial defenses [62]. Nevertheless, we suspect that in this study the continuous nebulization of 600 mg/kg of surfactant may have produced surfactant accumulation in the airways, which would partly explain lower PaO₂ and slightly higher PaCO₂ values of this groups compared to Neb-SURF200 and Neb-SURF400 groups. Unfortunately, we could not determine the pulmonary distribution of nebulized surfactant in this work and we acknowledge this as a limitation of the study.

Conclusion

We have carried out a complete preclinical assessment of the customized eFlow-Neos nebulizer operated with *poractant alfa* for the treatment and prevention of RDS. The nebulizer efficiently generated respirable aerosols out of undiluted surfactant, yielding an estimated mean lung deposition of 13.7% in vitro. Benchmark experiments settled the basis to implement a dose-response study in spontaneously-breathing rabbits with severe respiratory distress. Nominal surfactant doses of 200 mg/kg and 400 mg/kg delivered with the customized eFlow neos nebulizer during nCPAP significantly improved oxygenation, respiratory indices, and lung compliance, eliciting a pulmonary response equal to the one observed for the group of animals treated with an intratracheal bolus of 200 mg/kg of surfactant. The lung improvement induced by surfactant nebulization was achieved with a lower intrapulmonary surfactant dose compared to the InSurE approach. The neonate-focused surfactant nebulization strategy described in this work is currently being evaluated in Phase II clinical trial (EUDRACT No.:2016–004547-36).

Additional file

Additional file 1: Table S1. Baseline and post lung injury (Post-BAL) characteristics of surfactant-depleted adult rabbits treated with nasal continuous positive pressure ventilation (nCPAP), with intratracheal surfactant (Inst-SURF), or with different doses of nebulized surfactant (Neb-SURF100, Neb-SURF200, Neb-SURF400, and Neb-SURF600). **Figure S1.** PrINT cast with silicon-coated nostrils. **Figure S2.** Nebulizer output assessment. (DOCX 239 kb)

Abbreviations

BAL: Broncho-Alveolar Lavage; BPD: Broncho-Pulmonary Dysplasia; CPAP: Continuous Positive Airway Pressure; DSPC: Disaturated-Phosphatidylcholine; FiO_2 : Fraction of Inspired Oxygen; FPF: Fine Particle Fraction; GSD: Geometric Standard Deviation; HPLC: High Pressure Liquid Chromatography; LD: Lung Dose; LISA: Less Invasive Surfactant Administration; MAP: Mean Airway Pressure; MMD: Mass Median Diameter; nCPAP: nasal Continuous Positive Airway Pressure; OI: Oxygenation Index; PaCO₂: Arterial carbon dioxide partial pressure; PaO₂: Arterial Oxygen Partial Pressure; PEEP: Positive End-expiratory Pressure; PIP: Peak Inspiratory Pressure; RDS: Respiratory Distress Syndrome; RH: Relative Humidity; RR: Respiratory Rate; VEI: Ventilation Efficacy Index; V_T: Tidal Volume

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Authors' contributions

CD, FR, XM, MS, AB, SB, ML, EP, MP, GP, GM, LL, SC, GV, MC, MS, BP, PC, FB, VC, FS conceived and planned the experiments. FR, CC, UH, MN, MS, BR, carried out the experiments. CD, FR, XM, MS, AB, EP, MP, SM, MS, GP, GM, SC, GV, MC, BP, PC, FB, VC, FS contributed to the interpretation of the results. XM, FS, CD, FB, FR took the lead in writing the manuscript. All authors provided critical feedback and helped shape the research, analysis and manuscript. All authors have revised and approved the final version of the manuscript.

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request. They are not immediately publically available because they have been obtained by a private-funded research activity.

Ethics approval and consent to participate

Experimental procedures were approved by the intramural Animal Welfare Body and the Italian Ministry of Health (Prot.n° 1300–2015-PR) and complied with the European and Italian regulations for animal care.

Consent for publication

Not applicable.

Competing interests

MS, AB, and UH are employees of Pari Pharma GmbH. ML, EP, MN, GV, FR, FB, FS, CC, SC, BP, SB, MP, are employees of Chiesi Farmaceutici S.p.A. XM, CD, and LL served as consultants for this study.

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References

- Gizzi C, Montecchia F, Panetta V, Castellano C, Mariani C, Campelli M, et al. Is synchronised NIPPV more effective than NIPPV and NCPAP in treating apnoea of prematurity (AOP)? A randomised cross-over trial. *Arch Dis Child - Fetal Neonatal Ed.* 2015;100:F17 LP–F23.
- Morley CJ, Davis PG, Doyle LW, Brion LP, Hascoet J-M, Carlin JB. Nasal CPAP or intubation at birth for very preterm infants. *N Engl J Med.* 2008;358:700–8.
- Vento M. Tailoring oxygen needs of extremely low birth weight infants in the delivery room. *Neonatology.* 2011;342–8.
- Göpel W, Kribs A, Ziegler A, Laux R, Hoehn T, Wieg C, et al. Avoidance of mechanical ventilation by surfactant treatment of spontaneously breathing preterm infants (AMV): an open-label, randomised, controlled trial. *Lancet.* 2011;378:1627–34.
- Rojas-Reyes MX, Morley CJ, Soll R. Prophylactic versus selective use of surfactant in preventing morbidity and mortality in preterm infants. *Cochrane Database Syst Rev.* 2012;3:CD000510.
- Jobe AH, Bancalari E. Bronchopulmonary dysplasia. *Am J Respir Crit Care Med.* 2001;163:1723–9.
- Vento M, Moro M, Escrig R, Arruza L, Villar G, Izquierdo I, et al. Preterm resuscitation with low oxygen causes less oxidative stress, inflammation, and chronic lung disease. *Pediatrics.* 2009;124:e439–49.
- Owen LS, Manley BJ. Nasal intermittent positive pressure ventilation in preterm infants: equipment, evidence, and synchronization. *Semin Fetal Neonatal Med.* 2017;21:146–53.
- Dargaville PA, Aiyappan A, Cornelius A, Williams C, De Paoli AG. Preliminary evaluation of a new technique of minimally invasive surfactant therapy. *Arch Dis Child - Fetal Neonatal Ed.* 2011;96:F243 LP–F248.
- Moretti C, Gizzi C, Montecchia F, Barbàra CS, Midulla F, Sanchez-Luna M, et al. Synchronized nasal intermittent positive pressure ventilation of the newborn: technical issues and clinical results. *Neonatology.* 2016;359–65.
- Dunn MS, Kaempf J, de Klerk A, de Klerk R, Reilly M, Howard D, et al. Randomized trial comparing 3 approaches to the initial respiratory Management of Preterm Neonates. *Pediatrics.* 2011;128:e1069 LP–e1076.
- Sweet DG, Carnielli V, Greisen G, Hallman M, Ozek E, Plavka R, et al. European consensus guidelines on the Management of Respiratory Distress Syndrome - 2016 update. *Neonatology.* 2017;111:107–25.
- Lemyre B, Davis PG, De Paoli AG, Kirpalani H. Nasal intermittent positive pressure ventilation (NIPPV) versus nasal continuous positive airway pressure (NCPAP) for preterm neonates after extubation. *Cochrane Database Syst Rev.* 2017.
- Niemark HJ, Hütten MC, Kramer BW. Surfactant for respiratory distress syndrome: new ideas on a familiar drug with innovative applications. *Neonatology.* 2017;111:408–14.
- Pillow J, Minocchieri S. Innovation in surfactant therapy II: surfactant administration by Aerosolization. *Neonatology.* 2012;101:337–44.
- Mazela J, Merritt TA, Finer NN. Aerosolized surfactants. *Curr Opin Pediatr.* 2007;19:155–62.
- Dijk PH, Heikamp A, Bambang Oetomo S. Surfactant nebulisation: lung function, surfactant distribution and pulmonary blood flow distribution in lung lavaged rabbits. *Intensive Care Med.* 1997;23:1070–6.
- Tashiro K, Yamada K, Li WZ, Matsumoto Y, Kobayashi T. Aerosolized and instilled surfactant therapies for acute lung injury caused by intratracheal endotoxin in rats. *Crit Care Med.* 1996;24:488–94.
- Dijk PH, Heikamp A, Oetomo SB. Surfactant nebulisation prevents the adverse effects of surfactant therapy on blood pressure and cerebral blood flow in rabbits with severe respiratory failure. *Intensive Care Med.* 1997;23:1077–81.
- Rey-Santano C, Mielgo VE, López-De-Heredia-Y-Goya J, Murgía X, Valls-I-Soler A. Cerebral effect of Intratracheal aerosolized surfactant versus bolus therapy in preterm lambs. *Crit Care Med.* 2016;44:e218–26.
- Henry MD, Rebello CM, Ikegami M, Jobe AH, Langenback EG, Davis JM. Ultrasonic nebulized in comparison with instilled surfactant treatment of preterm lambs. *Am J Respir Crit Care Med.* 1996;154:366–75.
- Lewis JF, Ikegami M, Jobe A H, Tabor B. Aerosolized surfactant treatment of preterm lambs. *J Appl Physiol.* 1991;70:869–76.
- Bahlmann H, Sun B, Nilsson G, Curstedt T, Robertson B. Aerosolized surfactant in lung-lavaged adult rats: factors influencing the therapeutic response. *Acta Anaesthesiol Scand.* 2000;44:612–22.

24. Dijk PH, Heikamp A, Oetomo SB. Surfactant nebulization versus instillation during high frequency ventilation in surfactant-deficient rabbits. *Pediatr Res*. 1998;44:699–704.
25. Ruppert C, Kuchenbuch T, Boensch M, Schmidt S, Mathes U, Hillebrand V, et al. Dry powder aerosolization of a recombinant surfactant protein-C-based surfactant for inhalative treatment of the acutely inflamed lung. *Crit Care Med*. 2010;38:1584–91.
26. Schermuly R, Schmehl T, Günther A, Grimminger F, Seeger W, Walmrath D. Ultrasonic nebulization for efficient delivery of surfactant in a model of acute lung injury: impact on gas exchange. *Am J Respir Crit Care Med*. 1997;156:445–53.
27. Rey-Santano C, Mielgo VE, Andres L, Ruiz-del-Yerro E, Valls-i-Soler A, Murgia X. Acute and sustained effects of aerosolized vs. bolus surfactant therapy in premature lambs with respiratory distress syndrome. *Pediatr Res Springer Nature*. 2013;73:639–46.
28. Rahmel DK, Pohlmann G, Iwatschenko P, Volland J, Liebisch S, Kock H, et al. The non-intubated, spontaneously breathing, continuous positive airway pressure (CPAP) ventilated pre-term lamb: a unique animal model. *Reprod Toxicol*. 2012;34:204–15.
29. Hütten MC, Kuypers E, Ophelders DR, Nikiforou M, Jellema RK, Niemarkt HJ, et al. Nebulization of Poractant alfa via a vibrating membrane nebulizer in spontaneously breathing preterm lambs with binasal continuous positive pressure ventilation. *Pediatr Res*. 2015;78:664–9.
30. Walther FJ, Hernández-Juviel JM, Waring AJ. Aerosol delivery of synthetic lung surfactant. Longo M, editor. *PeerJ*. 2014;2:e403.
31. Jorch G, Hartl H, Roth B, Kribs A, Gortner L, Schaible T, et al. To the editor: surfactant aerosol treatment of respiratory distress syndrome in spontaneously breathing premature infants. *Pediatr Pulmonol*. 1997;222–4.
32. Arroe M, Pedersen-Bjergaard L, Albertsen P, Bode S, Greison G, Jonsbo F. Inhalation of aerosolized surfactant exosurf to neonates treated with nasal continuous positive airway pressure. *Prenat Neonatal Med*. 1998;3:346–52.
33. Berggren E, Liljedahl M, Winblad B, Andreasson B, Curstedt T, Robertson B, et al. Pilot study of nebulized surfactant therapy for neonatal respiratory distress syndrome. *Acta Paediatr*. 2000;89:460–4.
34. Finer NN, Merritt TA, Bernstein G, Job L, Mazela J, Segal R. An open label, pilot study of Aerosurf® combined with nCPAP to prevent RDS in preterm neonates. *J Aerosol Med Pulm Drug Deliv*. 2010;23:303–9.
35. Minocchieri S, Berry CA, Pillow JJ. Nebulised surfactant to reduce severity of respiratory distress: a blinded, parallel, randomised controlled trial. *Arch Dis Child - Fetal Neonatal Ed*. 2018.
36. Fok TF, Monkman S, Dolovich M, Gray S, Coates G, Paes B, et al. Efficiency of aerosol medication delivery from a metered dose inhaler versus jet nebulizer in infants with bronchopulmonary dysplasia. *Pediatr Pulmonol*. 1996;21:301–9.
37. Köhler E, Jilg G, Avenarius S, Jorch G. Lung deposition after inhalation with various nebulisers in preterm infants. *Arch Dis Child Fetal Neonatal Ed*. 2008;93.
38. Mazela J, Polin RA. Aerosol delivery to ventilated newborn infants: historical challenges and new directions. *Eur J Pediatr*. 2011;433–44.
39. Goikoetxea E, Murgia X, Serna-Grande P, Valls-i-Soler A, Rey-Santano C, Rivas A, et al. In vitro surfactant and perfluorocarbon aerosol deposition in a neonatal physical model of the upper conducting airways. *PLoS One*. 2014;9:e106835.
40. Goikoetxea E, Rivas A, Murgia X, Antón R. Mathematical modeling and numerical simulation of surfactant delivery within a physical model of the neonatal trachea for different aerosol characteristics. *Aerosol Sci Technol*. 2017;51:168–77.
41. Minocchieri S, Burren JM, Bachmann MA, Stern G, Wildhaber J, Buob S, et al. Development of the premature infant nose throat-model (PiNT-model)-an upper airway replica of a premature neonate for the study of aerosol delivery. *Pediatr Res*. 2008;64:141–6.
42. Milesi I, Tingay DG, Zannin E, Bianco F, Tagliabue P, Mosca F, et al. Intratracheal atomized surfactant provides similar outcomes as bolus surfactant in preterm lambs with respiratory distress syndrome. *Pediatr Res*. 2016;80:92–100.
43. Murgia X, Gastiasoro E, Mielgo V, Alvarez-Diaz F, Lafuente H, Valls-i-Soler A, et al. Surfactant and perfluorocarbon Aerosolization by means of inhalation catheters for the treatment of respiratory distress syndrome: an In Vitro study. *J Aerosol Med Pulm Drug Deliv*. 2011;24:81–7.
44. Pohlmann G, Iwatschenko P, Koch W, Windt H, Rast M, de Abreu MG, et al. A novel continuous powder Aerosolizer (CPA) for Inhalative Administration of Highly Concentrated Recombinant Surfactant Protein-C (rSP-C) surfactant to preterm neonates. *J Aerosol Med Pulm Drug Deliv*. 2013;26:370–9.
45. Syedain ZH, Naqwi AA, Dolovich M, Somani A. In vitro evaluation of a device for intra-pulmonary aerosol generation and delivery. *Aerosol Sci Technol*. 2015;49:747–52.
46. Awad S, Williams DK, Berlinski A. Longitudinal evaluation of compressor/nebulizer performance. *Respir Care*. 2014;59:1053 LP–1061.
47. Berg EB, Picard RJ. In vitro delivery of budesonide from 30 jet nebulizer/compressor combinations using infant and child breathing patterns. *Respir Care*. 2009;54:1671–8.
48. Ricci F, Catozzi C, Murgia X, Rosa B, Amidani D, Lorenzini L, et al. Physiological, biochemical, and biophysical characterization of the lung-Lavaged spontaneously-breathing rabbit as a model for respiratory distress syndrome. *PLoS One*. 2017;12:e0169190.
49. Ricci F, Casiraghi C, Storti M, D'Alò F, Catozzi C, Ciccimarra R, et al. Surfactant replacement therapy in combination with different non-invasive ventilation techniques in spontaneously-breathing, surfactant-depleted adult rabbits. *PLoS One*. 2018;13:e0200542.
50. Cochrane CG, Revak SD, Merritt TA, Heldt GP, Hallman M, Cunningham MD, et al. The efficacy and safety of KL4-surfactant in preterm infants with respiratory distress syndrome. *Am J Respir Crit Care Med*. 1996;153:404–10.
51. Blich EG, Dier WJ. A rapid method of total lipid extraction and purification. *Can J Biochem Physiol*. 1959;37:911–7.
52. Verlato G, Cogo PE, Balzani M, Gucciardi A, Burattini I, De Benedictis F, et al. Surfactant status in preterm neonates recovering from respiratory distress syndrome. *Pediatrics*. 2008;122:102 LP–108.
53. Cogo PE, Facco M, Simonato M, Verlato G, Rondina C, Baritussio A, et al. Dosing of porcine surfactant: effect on kinetics and gas exchange in respiratory distress syndrome. *Pediatrics*. 2009;124:e950–7.
54. Robillard E, Alarie Y, Dagenais-Perusse P, Baril E, Guilbeault A. Microaerosol Administration of Synthetic β - γ -Dipalmitoyl-L- α -lecithin in the respiratory distress syndrome: a preliminary report. *Can Med Assoc J*. 1964;90:55–7.
55. Chu J, Clements JA, Cotton EK, Klaus MH, Sweet AY, Tooley WH. Neonatal Pulmonary Ischemia *Pediatrics*. 1967;40:709 LP–782.
56. Speer CP, Gefeller O, Groneck P, Laufkötter E, Roll C, Hanssler L, et al. Randomised clinical trial of two treatment regimens of natural surfactant preparations in neonatal respiratory distress syndrome. *Arch Dis Child Fetal Neonatal Ed*. 1995;72:F8–13.
57. Speer CP, Robertson B, Curstedt T, Halliday HL, Compagnone D, Gefeller O, et al. Randomized European multicenter trial of surfactant replacement therapy for severe neonatal respiratory distress syndrome: single versus multiple doses of Curosurf. *Pediatrics*. 1992;89:13–20.
58. Verder H, Robertson B, Greisen G, Ebbesen F, Albertsen P, Lundstrom K, et al. Surfactant therapy and nasal continuous positive airway pressure for newborns with respiratory distress syndrome. *N Engl J Med*. 1994;331:1051–5.
59. Aldana-Aguirre JC, Pinto M, Featherstone RM, Kumar M. Less invasive surfactant administration versus intubation for surfactant delivery in preterm infants with respiratory distress syndrome: a systematic review and meta-analysis. *Arch Dis Child - Fetal Neonatal Ed*. 2017;102:F17 LP–F23.
60. Linner R, Perez-de-Sa V, Cunha-Goncalves D. Lung deposition of nebulized surfactant in newborn piglets. *Neonatology*. 2015;107:277–82.
61. Sun Y, Wang YQ, Yang R, Zhu JJ, Le YY, Zhong JG, et al. Exogenous porcine surfactants increase the infiltration of leukocytes in the lung of rats. *Pulm Pharmacol Ther*. 2009;22:253–9.
62. Schrod L, Hornemann F, Von Stockhausen HB. Chemiluminescence activity of phagocytes from tracheal aspirates of premature infants after surfactant therapy. *Acta Paediatr Int J Paediatr*. 1996;85:719–23.

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