

Vasectomy in Spotted Paca (*Cuniculus paca*)

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ABSTRACT

Background: *Cuniculus paca* is the second largest neotropical rodent. It is not endangered, but your habitat has been destroyed and the specie has been hunted, because of its prized meat. In this context captive breeding is an alternative to reduce the hunt. Then, adult male vasectomy is an interesting alternative for *Cuniculus paca* since the animal does not lose libido and maintain cyclicity of females into the enclosure. This technique is a method of sterilization which the vas deferens is surgically clamped, cut, or otherwise sealed and thus prevents the release of sperm when a male ejaculates. The aim of this study was to describe the vasectomy technique on a male spotted paca kept in captive.

Case: A captive adult male of *Cuniculus paca*, lived in Brazilian wild fauna breeding for scientific research. It was maintained on precinct with no other animal, ate fruits, vegetables, tubers and rodent chow and water offered *ad libitum*. It was submitted to bilateral vasectomy to maintain reproductive behavior on bevy, but not impregnate females. The anaesthesia was performed using ketamine hydrochloride (25 mg/kg IM) and midazolam (0.5 mg/kg IM) as premedication, and isoflurane in open system by facemask diluted in 100% O₂ for induction and maintenance. Immediately after induction, was performed epidural anesthesia using 4 mg/kg of lidocaine hydrochloride without vasoconstrictor associated in the same syringe with 0.2 mg/kg of methadone hydrochloride to promote analgesia. As the testes in this species are inside the abdomen, the surgical approach was made by paraprepuical skin incision and ventral midline abdominal incision. After access the cavity, the testes were located and the vaginal tunics were incised to access the vas deferens. After exposed, both were doubly ligated, sectioned and removed a segment of approximately 1 cm of each duct. Finally, the occlusion of subcutaneous and muscle layers were made using 2-0 absorbable and skin with 2-0 non-absorbable sutures. Postoperatively, benzathine penicillin (30,000 IU/kg IM once), tramadol (4 mg/kg IM once) and meloxicam (0.3 mg/kg SC SID for 3 days) were administered. Ten days post-surgery, the animal was fully recovered and after twenty days, it was transferred to enclosure of females. None were fertilized after vasectomy, and there was no change in reproductive behavior among individuals.

Discussion: Zoos and breeding have a problem when animal population increases too much, thus vasectomy is an important alternative to avoid this, because it does not lose sexual behavior, but cannot impregnate. The spotted paca presented intracavitary testicles without elevation Skin evidence to scrotum formation, however may also present inguinal regions testes beside penis or scrotum, most evident in reproductive season, similarly occurs in rats and agoutis. In the present case was performed bilaterally vasectomy to have satisfactory results for spermatogenesis reduction, confirming was indicated by some studies in rats and rabbits. Some studies in monkeys and rodents showed that after some time the spermatic cells decrease. In the present study sexual behavior and quality of patient ejaculate, were not completely monitor, to not interfere on management, but there was no female fertilized in a period of 1 year. This procedure showed to be feasible and an easy implementation for maintenance of the estrous cycle of females.

Keywords: Cuniculidae, reproduction, rodent, surgery, vas deferens.

INTRODUCTION

Spotted paca is not endangered, according to the International Union for Conservation of Nature [16], although there no specimen in certain parts of its distribution due to habitat destruction [15] and poaching, once its meat is highly prized [13,16]. Thus, the use of captive breeding in this species is interesting since it could promote a reduction in hunting and trafficking pressure and also provide an alternative source of protein [10]. Positive factor to this is its good adaptability to captivity, which contributes directly to the research related to breeding, conservation purposes and improvement in zootecnical production [5].

In captivity, contraception techniques are often important for maintenance of males and females housed in the same enclosure. Vasectomy is a highly effective contraceptive method which can play an important role in reducing the growth of a population [18].

Moreover, it shows an excellent alternative in the breeding of wild species in captivity, keeping reproductive behavior of males and females. This surgical procedure leads to interruption of the passage of sperm through the vas deferens, by ligation, excision or section of part of these [6,8,18].

Considering the few data in the literature, the aim of this study was to describe vasectomy technique in a captive spotted paca.

CASE

A male spotted paca, adult, kept in breeding specimens of Brazilian fauna for scientific purposes, recorded by the Brazilian Institute of Environment and Renewable Natural Resources (IBAMA, 482,508 record), underwent bilateral vasectomy in order not to change the its libido, keep the cyclicity of females and do not fertilize them. It was kept in an isolated cage, receiving daily handling similar to other animals, feeding on fruits, vegetables and various tubers, rodent chow, and water offered *ad libitum*.

For the surgical procedure, it was submitted to anesthesia by using ketamine (ketaminaTM)¹ [25 mg/kg IM] and midazolam (DormonidTM)² [0.5 mg/kg IM] as premedication and isoflurane (IsoforineTM)² via face mask in an open system diluted in 100% O₂ for induction and maintenance.

Immediately after induction, epidural anesthesia was performed using 4 mg/kg of lidocaine hydrochloride (LidovetTM)³ without vasoconstrictor as-

sociated in the same syringe with 0.2 mg/kg methadone hydrochloride (MytedonTM)² to promote analgesia.

Animal was positioned in dorsal recumbency on a surgical table. As the testes in this species are located inside the abdominal cavity, the access was performed by incision in paraprepucial ventral skin of the abdomen. After access to the cavity, the testes were located and vaginal tunics were sectioned for location of the vas deferens (Figure 1). After exposure, both were doubly linked, sectioned and removed a segment of about 1 cm in length of each conduit (Figure 2).

Finally, the occlusion of muscle and subcutaneous layers were made using 2-0 absorbable and skin was closed with 2-0 non-absorbable sutures. Postoperatively, benzathine penicillin (Pronto PenTM)³ [30,000 IU/kg IM once], tramadol (TramadolTM)² [4 mg/kg IM once] and meloxicam (MaxicamTM)⁴ [0.3 mg/kg SC SID for 3 days] were administered.

Ten days post-surgery, the animal was fully recovered and after twenty days, it was transferred to enclosure of females. None were fertilized after vasectomy, and there was no change in reproductive behavior among individuals.

DISCUSSION

The no controlled population increase of captive animals is a problem faced by zoos and farms [19]. Thus, vasectomy is an interesting alternative in order to not lose the male libido. As long as androgens continue to be produced by the Leydig cells, keeps cyclicity in females, continuing the reproductive behavior in the group [6].



Figure 1. Vasectomy in *Cuniculus paca*. Ventral right paraprepucial access for vas deferens localization.

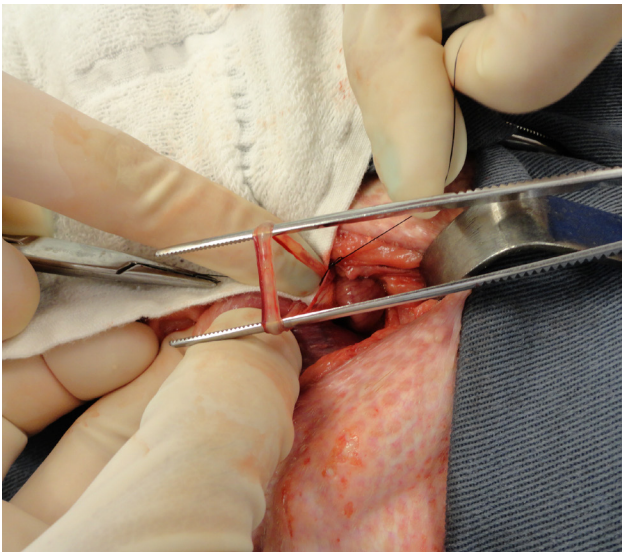


Figure 2. Vasectomy in *Cuniculus paca*. Exposure and link of the vas deferens.

The male presented intracavitary testicles without evidence of skin rise to the formation of the scrotum, but may also have the testicles in the inguinal regions beside to the penis or scrotum, most evident in the breeding season, just as occurs in rats and agoutis. The vas deferens was surrounded by the tunica vaginalis, cremaster muscle and was shown as a relatively straight tube having uniform diameter across the thread [2,3,12].

In this case, it was performed bilateral vasectomy to get satisfactory results for reduction of spermatogenesis, as suggested for some studies in rabbits and rats [8,11,14], however, other authors had reduced motility and sperm count in semen from both testes when done unilateral vasectomy on African giant rat (*Gambianus cricetomys*) [4] and mice [17].

In a study using capuchin monkeys (*Cebus apella*), after 105 postoperative days 100% of the animals showed a negative ejaculate for sperm cells [19]. It was observed testicular fibrosis in mice 12 to 16 months after vasectomy, resulting in infertility [17]. In

another study, it was observed that surgery induced deterioration in the seminiferous tubules associated with increased DNA fragmentation in testicular cells [18]. In the present study we could not completely monitor the behavior and the quality of patient ejaculate, to not interfere on the handling, but there was no female fertilized in a period of 1 year. However, some studies in rodents showed that decreasing in testosterone levels occurs eight weeks after vasectomy [4].

The decrease in the number and sperm motility occurs because developing anti-sperm autoantibody [17,20]. It happens gradually, and we should avoid placing the male with breeding females soon after surgery [19]. There are also sperm granuloma formation at the vasectomy site and orchitis that contribute to infertility, as well as oxidative stress that leads to apoptosis, but no one knows for sure the pathogenesis of post-vasectomy infertility [1,9].

Sperm granuloma formation is directly linked to the effective reduction of spermatogenesis. In rats, spermatogenic damage depends mainly on the post-operative development of the sperm granuloma, thus when granuloma formation is small or when there was no formation, sperm production should be significantly low [11].

Overall, we can conclude this procedure proved to be feasible and suggests an easy alternative for the management of males and females kept in the same enclosure, without gonadectomy.

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