

Acta Scientiae Veterinariae, 2017. 45: 1458.

RESEARCH ARTICLE Pub. 1458

ISSN 1679-9216

# A New Approach to Blood Parameters in Dogs with Hemorrhagic Enteritis

Handan Hilal Arslan<sup>1</sup>, Murat Guzel<sup>1</sup>, Yucel Meral<sup>1</sup>, Duygu Dalgin<sup>1</sup>, Guvenc Gokalp<sup>2</sup> & Umit Ozcan<sup>1</sup>

#### ABSTRACT

**Background:** Some blood parameters have diagnostic and prognostic importance for the infections in human medicine. However, there is insufficient research regarding the importance of blood parameters and their correlations in veterinary medicine. Increased blood cell distribution width (RDW) and platelet activity can link with the important inflammatory markers. The main objective of the present study was the evaluation of the relationship among some important blood parameters namely RDW, platelet count (PLT), platelet distribution width (PDW), mean platelet volume (MPV), plateletcrit (PCT), their potential usage in the diagnosis and determination of the clinical severity in dogs with hemorrhagic enteritis. Materials, Methods & Results: In this study, the case records of 29 dogs with hemorrhagic enteritis were evaluated and the records of 10 healthy dogs were used as controls. The animals of the study group were presented at the Ondokuz Mayis University, Veterinary Internal Medicine Clinic. The complete blood count (CBC), which includes the total WBC, RBC, hematocrit (HCT), hemoglobin concentration (Hgb), MCV, mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), RDW, PLT, MPV, PCT, and PDW, was determined. Significant positive correlations between RDW and RBC, HCT, MCHC, PLT and PDW, and a negative correlation with MCV, were determined. PDW was positively correlated with the lymphocyte count, MCHC and RDW, and negatively correlated with PCT. PLT was negatively correlated with MCV and MPV and positively correlated with RBC and RDW. In addition, MPV was positively correlated with MCV and MCH, and negatively correlated with PLT. Furthermore, there were significant differences between the granulocyte, WBC, HCT, RDW and PDW values (P < 0.001) and monocyte count, Hgb and MCV (P < 0.05), of the study and control groups.

*Discussion:* Acute hemorrhagic enteritis has various causes in dogs such as idiopathic hemorrhagic gastroenteritis and a number of viral, bacterial and parasitic agents. Hematological and biochemical parameters are not specific to enteric diseases, but these paremeters can provide clinically helpful information for differential diagnosis, response to treatment, and prognosis. In this frame, the evaluation of MCV and RDW in combination, and the determination of the mean red cell size and the extent of heterogeneity of the red cell population, can be especially useful to the diagnosis of different red blood cell disorders. In the present study, differences in RDW and MCV values were statistically significant between the study and control groups (P < 0.05). Increased RDW and decreased MCV can be good indicators of hemorragic diseases and in the present study, in addition to these findings, decreased Hgb and Hct confirmed anemia in dogs with hemorrhagic enteritis. The other key findings of this study were statistically significant relationships between RDW, PLT and PDW, which could be important indicators of inflammation in dogs with hemorrhagic enteritis. These parameters should be evaluated carefully in clinical cases of hemorrhagic enteritis. However, due to nature of retrospective studies, there were some limitations (the lack of another control group of dogs suffering from other hemorrhagic diseases) lack of serial measurements of the blood parameters and further studies should be carried out on dogs with hemorrhagic enteritis for a more detailed evaluation and confirmation of the findings of this study.

**Keywords:** Blood parameters, hemorrhagic enteritis, mean platelet volume (MPV), platelets, platelet distribution width (PDW), red blood cell distribution width (RDW).

Received: 3 March 2017

Accepted: 8 June 2017

Published: 13 July 2017

<sup>&</sup>lt;sup>1</sup>Department of Internal Medicine, Faculty of Veterinary Medicine, Ondokuz Mayis University, Samsun, Turkey. <sup>2</sup>Bogazlayan Vocational School, Bozok University, Yozgat, Turkey. CORRESPONDENCE: H.H. Arslan [hharslan@omu.edu.tr - Fax: +90 (362) 4576922]. Department of Internal Medicine, Faculty of Veterinary Medicine, Ondokuz Mayis University. 55139 Kurupelit, Samsun, Turkey.

### INTRODUCTION

Hematological and biochemical parameters are not specific enough to reveal the cause of enteric diseases, but they can provide clinically helpful information for differential diagnosis, response to treatment, and prognosis [2,3,5]. To evaluate these parameters, the levels of red blood cells (RBCs), white blood cells (WBCs) and platelets (PLT) are measured electronically [13].

Red blood cell distribution width (RDW) is a very useful parameter for the evaluation of the anisocytosis, the degree of heterogeneity of erythrocyte volume, and is used in laboratory haematology for the diagnosis of different anemias [15]. Additionally, it has been reported that RDW can be used as a marker of inflammation and there was a relationship between increased RDW and bloodstream infection [6].

Platelets appear to be important in a variety of pathological conditions in dogs [8,12]. Plateletcrit (PCT), mean platelet volume (MPV) and platelet distribution width (PDW) are important parameters in the examination of platelet activation [20]. MPV and PDW are the mean platelet size and the variation in platelet size, respectively. The PCT, which is derived from the platelet count and the MPV, indicates the percentage of platelets in a decilitre of blood, in a similar manner to hematocrit determination for erythrocytes [17].

Against that background, the aim of this study was the evaluation of the relationship among some important blood parameters, namely RDW, PLT, PDW, MPV PCT, and their potential usage in the diagnosis and determination of the clinical severity in dogs with hemorrhagic enteritis.

# MATERIALS AND METHODS

#### Collection of blood samples

The blood test results of 29 dogs with hemorrhagic enteritis (2-12 months old), which were presented at the Ondokuz Mayis University, Veterinary Internal Medicine Clinic, were used as the study group. The blood test results of 10 clinically healthy dogs were used as the control group. Blood samples were collected into tubes containing EDTA.

# Hematological analysis

The samples were analyzed with an automated blood analyzer (Abacus Junior Vet - Mindray Bc-5000)<sup>1</sup>. The complete blood count (CBC), which includes the total WBC, lymphocytes, monocytes, granulocytes, RBC, hematocrit (HCT), hemoglobin concentration (Hgb), MCV, mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), RDW, PLT, MPV, PCT, and PDW, was determined.

# Data Analysis

Pearson's correlation test was used for the determination of the degree of correlation among the investigated blood parameters. In addition, after the determination of normality of the data, for statistical comparison of the groups, the Independent Samples *t*-Test was employed for parametric values and the Mann-Whitney U test was used for non-parametric values. Differences were considered significant when *P* values were less than 0.05.

#### RESULTS

In this study, significant positive correlations between RDW and RBC, HCT, MCHC, PLT and PDW, and a negative correlation with MCV, were determined. PDW was positively correlated with the lymphocyte count, MCHC and RDW, and negatively correlated with PCT. PLT was negatively correlated with MCV and MPV and positively correlated with RBC and RDW. In addition, MPV was positively correlated with MCV and MCH, and negatively correlated with PLT (Table 1).

Furthermore, there were significant differences between the granulocyte, WBC, HCT, RDW and PDW values (P < 0.001) and monocyte count, Hgb and MCV (P < 0.05), of the study and control groups (Table 2). However, there was no significant difference between the RBC, MCH and MCHC values of the groups (P > 0.05).

#### DISCUSSION

RDW is mainly used for the detection and classification of anemias. In addition, the evaluation of MCV and RDW in combination, and the determination of the mean red cell size and the extent of heterogeneity of the red cell population, can be especially useful to the diagnosis of different red blood cell disorders. When MCV is insufficient for a diagnosis, RDW can be an additional finding. In addition, this information has a potential use in the screening and monitoring of marrow function in iron deficiency [11].

Parameters	Correlation analysis Pearson Correlation	ABC	LYM 410*	MONO 430*	GRA 921**	- RBC	HGB	HCT - 076	MCV - 064	MCH 052	MCHC 072	- 065	PLT - 254	PCT - 747	VAM 790	
WBC	Sig. (2-tailed)	-	.024	.020	000.	.688	-192	.006	.742	760. 790	.710	.736	.192	-272	.73	. ++
	N	29	29	29	29	29	29	29	29	29	29	29	28	28	28	
	Pearson Correlation	.419*	1	.091	.036	.163	.173	.197	.093	.212	.275	.207	.007	202	.130	_
ГΥМ	Sig. (2-tailed) N	.024 29	70	.639 29	.855 90	995. 29	.369 29	.305 29	.632 29	.270 29	.149 20	.282 29	179. 80	.304 28	509 80	
	Pearson Correlation	.430*	.091		.379*	020	.047	003	084	.021	.119	078	012	024	000-	
MONO	Sig. (2-tailed)	.020	.639		.043	.917	809.	.988	.663	.916	.537	.688	.952	.902	976.	
	Ň	29	29	29	29	29	29	29	29	29	29	29	28	28	28	
	Pearson Correlation	.921**	.036	.379*		149	221	161	108	034	035	145	294	199	.020	
GRA	Sig. (2-tailed)	000.	.855	.043	Q	.441	.248	.404 8	.578	.860	.855	.452	.129	.311	.918	
		67	67	67	67	67	67	67	67	67	67	67	87	87	87	
C	Pearson Correlation	8/0'-	.103 002	020	149	-	<b>.043</b>	<b>645</b>	14/	240	111.	80C.	404.	100.		
KBC	Sig. (2-tailed)	000. 00	995. DC	716. 00	144. 0 c	00	000.	000.	,44. 0 c	961. 00	80C. 0C	c00.	CIU. 86	166.	28U. 8C	_
	Degreon Correlation	- 137	173	047	- 771	£43**	- 1	740**	343	471**	570**	730	218	015	00.	
aun	Circ (7 toiled)	701-	360	(±0;	177		-		040	010		().4. C1C	996	010. 110		
2	DIG. (2-laileu) N	29	29 29	-005 29	29 29	29	29	29	29	29	-001 29	212	28	28	28.28	
	Pearson Correlation	076	.197	003	161	.943**	.749**	-	.150	062	.265	.454*	.301	023	173	0
HCT	Sig. (2-tailed)	969.	.305	.988	.404	000.	000.		.436	.750	.165	.013	.120	908.	.379	
	N	29	29	29	29	29	29	29	29	29	29	29	28	28	28	
	Pearson Correlation	064	.093	084	108	147	.343	.150	-	.561**	.239	439*	477*	.073	.421	
MCV	Sig. (2-tailed)	.742	.632	.663	.578	.447	.068	.436		.002	.211	.017	.010	.713	.026	
	Z	29	29	29	29	29	29	29	29	29	29	29	28	28	28	
	Pearson Correlation	.052	.212	.021	034	246	.471 <sup>**</sup>	062	.561**	1	.787**	134	288	081	.380	
MCH	Sig. (2-tailed)	067.	.270	.916	.860	99I.	.010	057.	.002	00	000.	.490	.137	.683	.046	
		67	67	710	67	67	**922	575	67	** <b>EGE</b>	67	\$7 \$7	070	070	07	
C	Pearson Correlation	710.	C/7.	911. Ecz	050	111.	6/c.	207.	657.	18/	-	.411 700	052	249	107.	
MCHC	Sig. (2-tailed) N	01/.	.149 20	16C. 00	CC8. DC	80C. 0C	100.	C01.	117. 112.	000.	00	170. 00	1/8. 86	202. 80	081. 28	
	Pearson Correlation	065	207	078	- 145	.508**	239	454*	439*	- 134	.411*	-	392*	- 224	- 011	
RDWc	Sio (2-tailed)	736	282	688	452	2002	212	013	017	490	027	1	039	251	957	
)	N N	29	29	29	29	29	29	29	29	29	29	29	28	28	28	
	Pearson Correlation	254	.007	012	294	.454*	.218	.301	477*	288	032	.392*		.141	589*	*
PLT	Sig. (2-tailed)	.192	.971	.952	.129	.015	.266	.120	.010	.137	.871	.039		.474	.001	
	Ν	28	28	28	28	28	28	28	28	28	28	28	28	28	28	
	Pearson Correlation	242	202	024	199	.001	.015	023	.073	081	249	224	.141	1	.073	
PCT	Sig. (2-tailed)	.214	.304	.902 30	.311	.997 20	.941 30	.908 0	.713	.683	.202	.251	.474		.712	
	2	78	87	28	87	787	87	78	87	87	87	78	87	87	. 78	
ţ	Pearson Correlation	.007	.130	900'-	070	-327	1.60	173	.421	.180	197	110		.073 	-	
MPV	Sig. (2-tailed)	./34	60C.	.976 97	918. 918	48U.	770	615. 00	97 <u>0</u> .	.040	180	1 CY.	100.	71/.	ĉ	
	Z (	782		-78	87	87	28	87	87	28	87	\$7 87	78		780	
	Pearson Correlation	.203	.494	245	.042	.303	.259	.290	176	.169	.393 <sup>°</sup>	.580	024	414	080.	
PDW	Sig. (2-tailed)	.301 30	.008 008	.208	.834 3	.117	.183	.134	.370	.390 32	.039 3	.001	.905 20	.028	.687	
	Z	28	28	28	28	28	28	28	X	28	× X	× ×	× X	SC SC	c X	

Parameter	Control $(n = 10)$	Study group $(n = 29)$
WBC (10 <sup>9</sup> /L)	$12.00 \pm 0.73$	5.89 ± 0.78**
Lymphocytes (10 <sup>9</sup> /L)	$1.98 \pm 0.22$	$1.98 \pm 0.30$
Monocytes (10 <sup>9</sup> /L)	$0.65 \pm 0.12$	$0.28 \pm .0.49*$
Granulocytes (10 <sup>9</sup> /L)	$9.36 \pm 0.65$	$3.61 \pm 0.69 * *$
RBC (10 <sup>12</sup> /L)	$6.61 \pm 0.18$	$5.88 \pm 0.30$
HGB (g/dL)	$15.91 \pm 0.52$	$13.21 \pm 0.81*$
HCT (%)	$45.45 \pm 1.41$	$36.97 \pm 2.09*$
MCV (fL)	$68.76 \pm 0.88$	$63.76 \pm 0.97*$
MCH (pg)	$24.08 \pm 0.36$	$23.33 \pm 1.12$
MCHC (g/L)	$34.73 \pm 0.31$	$35.40 \pm 1.84$
RDWc (%)	$14.40 \pm 0.48$	$17.69 \pm 0.85 **$
PLT (10 <sup>9</sup> /L)	$352.20 \pm 28.20$	$449.11 \pm 55.32$
PCT (mL/L)	$0.91 \pm 0.40$	$0.56 \pm 0.12$
MPV (fL)	$9.47 \pm 0.30$	$9.68 \pm 0.34$
PDW (%)	$15.48 \pm 0.09$	$33.43 \pm 1.93 **$

Table 2. Blood parameters of the control group and the study group of dogs with hemorrhagic enteritis (Mean ± S.E.M.).

\*Significantly different from the control group (\*P < 0.05; \*\*P < 0.001).

In the present study, differences in RDW and MCV values were statistically significant between the study and control groups (P < 0.05). Increased RDW and decreased MCV can be good indicators of hemorragic diseases [10,14] and in the present study, in addition to these findings, decreased Hgb and Hct confirmed anemia in dogs with hemorrhagic enteritis. In addition, a previous study reported that RDW can be used to evaluate the prognosis of septic shock and severe sepsis in human patients [7]. Unterer et al. [19] reported that there is an increased probability of bacterial translocation due to damage to the intestinal epithelial barrier and increased risk of sepsis in the case of acute intestinal diseases. In the present study, the mean RDW value of the study group was significantly higher than in the control group (P < 0.01). This finding may show that increased RDW level can also be important in veterinary medicine and indicate the severity of sepsis and therefore be prognostic for hemorrhagic enteritis in dogs.

Furthermore, increased RDW is linked with the important inflammatory markers, interleukin-6 and tumor necrosis factor. The cytokines may have a role in the suppression of the maturation and reduction of the half-life of RBCs [6,10,14]. The RDW status represents a surrogate marker for illness severity and the early stage of acute illness and thereby correlates with the clinical prognosis [22]. RDW also has prognostic value in patients with acute myocardial infarction, cardiac arrest, congestive heart failure, critical illness, pneumonia and pulmonary embolism. The mechanism changing RDW range is not clearcut in these diseases, but it has been associated with the inflammatory process [6]. In the present study, increased RDW could also be reflecting severe clinical inflammation in dogs with hemorrhagic enteritis.

Platelets are intimately involved in homeostasis, inflammation, immunity, tissue regeneration and other physiological and pathological processes [18]. Some researchers have reported that, as for RDW, platelets are very important in the pathogenesis of local and systemic inflammation related disorders. Platelet activation has been reported in particular diseases, such as acute systemic inflammatory response syndrome (SIRS), disseminated intravascular coagulation (DIC), gastrointestinal disorders, inflammatory bowel disease and septicemia, in animals and human beings [16,18].

The three platelet parameters, namely PCT, MPV and PDW, are important indicators of platelet activation [20]. Activated platelets have some chemotactic substances such as the platelet-derived growth factor and lipopolysaccharides. These substances facilitate the binding of leukocytes to the endothelium and their extra-vasation, and they may either stimulate or inhibit the inflammatory responses of leukocytes. The platelets themselves also contain a group of pro-inflammatory compounds and are therefore accepted to be mediator and effectors cells in inflammation [4,8,21].

In the present study, although the mean platelet count of the study group was higher than that of the control group, the difference was not statistically significant. Several studies have presented data which infer a correlation between higher MPV values and active inflammatory disease [9]. Researchers have re-

ported a nonlinear, inverse relationship between MPV and platelet count in dogs and humans. However, in dogs, there is contradictory information as to whether there is a statistically significant relationship between MPV and PLT [1,23]. In this study, a negative correlation between MPV and PLT was determined in dogs with hemorrhagic enteritis.

Increased numbers of larger platelets increase platelet heterogeneity (PDW). The PDW is also elevated as a bone marrow response to thrombocytopenia. In human medicine, divergences from the expected relationship between MPV and PLT have been used to differentiate the causes of thrombocytopenia in terms of their importance for determination of the current situation and prognosis. PDW may be a more sensitive indicator of increased proportions of macroplatelets than MPV because the latter may be decreased by the presence of smaller platelets and cellular debris [1].

In the present study, mean MPV values were almost the same in the control group as in the study group, whereas the mean PDW values of the study group were two times higher than the control group (P < 0.001). Furthermore, PDW was positively correlated with lymphocyte numbers. Therefore, abnormal RDW and PDW counts may be indicative of a severe response to inflammation in dogs with hemorrhagic enteritis and be useful new parameters for the diagnosis of hemorrhagic enteritis.

#### CONCLUSIONS

In conclusion, in the present study there was a significant relationship between RDW, PLT and PDW, which could be an important indicator of inflammation in dogs with hemorrhagic enteritis. Therefore, these parameters should be evaluated carefully in clinical cases of hemorrhagic enteritis. However, it should be noted that the data was obtained from a retrospective study, which implies limitations such as the lack of another control group of dogs suffering from other hemorrhagic diseases and lack of serial measurements of the blood parameters. For a more detailed evaluation and confirmation of the findings of this study, further studies should be performed on dogs with hemorrhagic enteritis.

#### MANUFACTURER

<sup>1</sup>Shenzhen Mindray Bio-Medical Electronics Co. Nanshan, Shenzhen, China.

*Acknowledgements*. The authors thank Gregory T. Sullivan of the University of Queensland for editing the English in an earlier version of this manuscript.

**Declaration of interest.** The authors report no conflicts of interest. The authors alone are responsible for the content and writing of paper.

#### REFERENCES

- 1 Bommer N.X., Shaw D.J., Milne E. & Ridyard A.E. 2008. Platelet distribution width and mean platelet volume in the interpretation of thrombocytopenia in dogs. *Journal of Small Animal Practice*. 49: 518-24.
- 2 Castro T.X., Cubel Garcia Rde C., Gonçalves L.P., Costa E.M., Marcello G.C., Labarthe N.V. & Mendes-de-Almeida F. 2013. Clinical, hematological, and biochemical findings in puppies with coronavirus and parvovirus enteritis. *The Canadian Veterinary Journal*. 54(9): 885-888.
- **3 Decaro N. & Buonavoglia C. 2012.** Canine parvovirus A review of epidemiological and diagnostic aspects, with emphasis on type 2c.*Veterinary Microbiology*. 155: 1-12.
- 4 Deuel T.F., Senior R.M., Huang J.S. & Griffin G.L. 1982. Chemotaxis of monocytes and neutrophils to plateletderived growth factor. *Journal of Clinical Investigation*. 69(4): 1046-1049.
- **5 Groschwitz K.R. & Hogan S.P. 2009.** Intestinal barrier function: molecular regulation and disease pathogenesis. *Journal of Allergy and Clinical Immunology*. 124(1): 3-20.
- 6 Jo Y.H., Kim K., Lee J.H. Kang C., Kim T., Park H.M., Kang K.W., Kim J. & Rhee J.E. 2013. Red cell distribution width is a prognostic factor in severe sepsis and septic shock. *The American Journal of Emergency Medicine*. 31: 545-548.
- 7 Kisaoglu A., Bayramoglu A., Ozogul B., Atac K., Emet M. & Atamanalp S.S. 2014. Sensitivity and specificity of red cell distribution width in diagnosing acute mesenteric ischemia in patients with abdominal pain. *World Journal of Surgery*. 38: 2770-2776.
- 8 Klinger M.H. 1997. Platelets and inflammation. Anatomy and Embryology. 196: 1-11.
- 9 Leader A., Pereg D. & Lishner M. 2012. Are platelet volume indices of clinical use? A multidisciplinary review. *Annals of Medicine*. 44: 805-816.

- 10 Lippi G., Targher G., Montagnana M., Salvagno G.L., Zoppini G. & Guidi G.C. 2009. Relation between red blood cell distribution width and inflammatory biomarkers in a large cohort of unselected outpatients. *Archives of Pathology* & *Laboratory Medicine*. 133: 628-632.
- 11 Monzon C.M., Beaver B.D. & Dillon TD. 1987. Evaluation of erythrocyte disorders with mean corpuscular volume (MCV) and red cell distribution width (RDW). *Clinical Pediatrics*. 26(12): 632-638.
- 12 Moritz A., Walcheck B.K. & Weiss D.J. 2005. Evaluation of flow cytometric and automated methods for detection of activated platelets in dogs with inflammatory disease. *American Journal of Veterinary Research*. 66: 325-329.
- 13 Neiger R., Hadley J. & Pfeiffer D.U. 2002. Differentiation of dogs with regenerative and non-regenerative anaemia on the basis of their red cell distribution width and mean corpuscular volume. *Veterinary Record.* 150: 431-434.
- 14 Prittie J. 2004. Canine parvoviral enteritis: a review of diagnosis, management, and prevention. *Journal of Veterinary Emergency and Critical Care.* 14: 167-176.
- 15 Salvagno G.L., Sanchis-Gomar F., Picanza A. & Lippi G. 2015. Red blood cell distribution width: A simple parameter with multiple clinical applications. *Critical Reviews in Clinical Laboratory Sciences*. 52: 86-105.
- 16 Segura D., Monreal L., Armengou L., Tarancón I., Brugués R. & Escolar G. 2007. Mean platelet component as an indicator of platelet activation in foals and adult horses. *Journal of Veterinary Internal Medicine*. 21: 1076-1082.
- 17 Smith J.R., Smith K.F. & Brainard B.M. 2014. Platelet parameters from an automated haematology analyzer in dogs with inflammatory clinical diseases. *The Veterinary Journal*. 201: 406-411.
- 18 Tanju C., Ekrem G., Berksoy E.A. & Nur A. 2014. Mean platelet volume as a negative marker of inflammation in children with rotavirus gastroenteritis. *Iran Journal of Pediatrics*. 24: 617-622.
- 19 Unterer S., Strohmeyer K., Kruse B.D., Sauter-Louis C. & Hartmann K. 2011. Treatment of aseptic dogs with hemorrhagic gastroenteritis with amoxicillin/clavulanic acid: a prospective blinded study. *Journal of Veterinary Internal Medicine*. 25: 973-979.
- **20** Wiwanitkit V. 2004. Plateletcrit, mean platelet volume, platelet distribution width: its expected values and correlation with parallel red blood cell parameters. *Clinical and Applied Thrombosis/Hemostasis*. 10: 175-178.
- 21 Wright G.G., Read P.W. & Mandell G.L. 1988. Lipopolysaccharide releases a priming substance from platelets that augments the oxidative response of polymorphonuclear neutrophils to chemotactic peptide. *The Journal of Infectious Diseases*. 157(4): 690-696.
- 22 Yang X. & Du B. 2014. Red cell distribution width: the crystal ball in the hands of intensivists? *Journal of Thoracic Diseases*. 6: 64-65.
- 23 Yilmaz Z., Eralp O. & Ilcol Y.O. 2008. Evaluation of platelet count and its association with plateletcrit, mean platelet volume, and platelet size distribution width in a canine model of endotoxemia. *Veterinary Clinical Pathology*. 37: 159-163.

