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Smallest *Anopheles farauti* occur during the peak transmission season in the Solomon Islands

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Abstract

Background: Malaria transmission varies in intensity amongst Solomon Island villages where *Anopheles farauti* is the only vector. This variation in transmission intensity might be explained by density-dependent processes during *An. farauti* larval development, as density dependence can impact adult size with associated fitness costs and daily survivorship.

Methods: Adult anophelins were sampled from six villages in Western and Central Provinces, Solomon Islands between March 2014 and February 2017. The size of females was estimated by measuring wing lengths, and then analysed for associations with biting densities and rainfall.

Results: In the Solomon Islands, three anopheline species, *An. farauti*, *Anopheles hinesorum* and *Anopheles lungae*, differed in size. The primary malaria vector, *An. farauti*, varied significantly in size among villages. Greater rainfall was directly associated with higher densities of *An. farauti* biting rates, but inversely associated with body size with the smallest mean sized mosquitoes present during the peak transmission period. A measurable association between body size and survivorship was not found.

Conclusions: Density dependent effects are likely impacting the size of adult *An. farauti* emerging from a range of larval habitats. The data suggest that rainfall increases *An. farauti* numbers and that these more abundant mosquitoes are significantly smaller in size, but without any reduced survivorship being associated with smaller size. The higher malaria transmission rate in a high malaria focus village appears to be determined more by vector numbers than size or survivorship of the vectors.

Keywords: *Anopheles farauti*, *Anopheles hinesorum*, *Anopheles lungae*, Density-dependence, Wing length, Size variation, Solomon Islands

Background

Vector control with indoor residual spraying (IRS) and insecticide-treated nets (ITNs) is responsible for 80% of the reduction in *Plasmodium falciparum* cases in Africa between 2000 and 2015 [1]. The global malaria cases have since stabilized. Further reductions in malaria cases will require strengthened malaria control [2, 3]. ITNs are

most effective against vectors that blood feed indoors and late at night, while IRS is most effective when vectors rest indoors [3]. Despite increasing prevalence of insecticide resistance (physiological and behavioural), LLINs and IRS remain sufficiently effective to provide significant malaria control [4–6]. At the present time, the only WHO-recommended strategy that targets malaria vectors outside of houses is larval source management (LSM); but LSM is only recommended in areas with seasonal transmission or where the larval habitats are few in number, fixed in location and easily accessible (including urban areas) [7].

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Table 1 Timeline of anopheline surveys in Western and Central Provinces, Solomon Islands

Village	Sample period							
	2015		2016				2017	
	Aug	Nov	Jan	Jun	Aug	Nov	Feb	Jul
Jack Harbour	×				×	×	×	
Kinamara	×	×		×				
New Mala	×			×	×			×
Saeragi	×				×	×		×
Tuguivili		×		×	×	×	×	×
Haleta			×		×			×

Study period

Anophelines were sampled on multiple occasions in each village between July 2015 and July 2017 (Table 1). Adult anopheline densities up to August 2016 were previously reported from these study villages [4, 29]; subsequent collections to July 2017 are updated here.

Sampling and processing of adult anophelines

Anophelines were caught using human landing catches (HLC) from 18.00 to 00.00 at 10 outdoor sites that were used during each of the four nights per village during each survey, as previously described [4, 29]. Captured anophelines were identified morphologically [31] before storage in 100% ethanol by collection hour and sample site.

Mosquito size was estimated by measuring wing lengths [16]. Individual specimens were dried on a triple vented petri dish for 5 min. Wings were then mounted on double sided sticky tape on a microscope slide. Using a Nikon SMZ-745T microscope with a scaled eye piece, wings were measured under 6.7× from the alular notch to apical margin (excluding the fringe) along the R1 vein.

Individual mosquitoes were identified to species by PCR using the internal transcribed spacer region II of ribosomal DNA (ITS2) [32]. For villages where only *An. farauti* sensu stricto was captured by HLC, a subset of samples was analysed to confirm species identifications. For villages with more than one anopheline species, all samples for which wing lengths were measured were identified by PCR. The rainfall data was sourced from the Munda Airstrip in Western Province (Bureau of Meteorology, Solomon Islands, unpublished data).

Statistical analysis

Data on mosquito surveys, wing lengths and molecular analyses are available from the James Cook University Tropical Data Hub [33]. Differences in the species composition between the study villages were compared using a Chi-squared contingency table. Generalized linear models (GLM) with a gaussian distribution were

used for the following analyses: (a) differences in wing lengths between mosquito species, and (b) differences in the wing length of *An. farauti* between villages. A GLM with a negative binomial distribution was used to analyse differences in adult biting density between villages. The GLMs and sequential post hoc analyses, Tukey–Kramer HSD, were conducted in SAS JMP V14.0.0.

The relationship between wing lengths and concurrent adult biting densities were directly compared with a Spearman's rank correlation in villages where the wing lengths of > 200 wings were measured (e.g., Jack Harbour village, Tuguivili and Haleta). Both factors were $\log(x + 1)$ transformed prior to analyses. A generalized estimating equation (GEE) compared the relationship between: (a) rainfall and density, and (b) density and wing lengths. Rainfall was summed for the 14-day window prior to the date of mosquito collection. The GEE was conducted using SPSS V24, had a normal distribution and incorporated study period as a random factor.

The mean wing lengths of female anopheline mosquitoes captured at each sampling station were projected geographically in QGIS (v3.4). The spatial analysis was only conducted in Jack Harbour where high densities of *An. farauti* were captured. Clusters of sampling sites where larger mosquitoes were captured were detected using SaTScan (v9.6) using a normal model.

Results

Species distributions

A total of 10,973 anophelines were collected during 1005 man-nights of HLC collections. Members of both the *An. farauti* sensu lato (s.l.) ($n = 8529$) and *An. lungae* (s.l.) ($n = 48$) complexes were captured in Western Province, while only members of the *An. farauti* complex ($n = 2396$) were captured in Haleta, Central Province. PCR analysis of all members of the *An. farauti* complex confirmed that 93% of specimens were *An. farauti* ($n = 937/1005$) and 7% were *An. hinesorum* ($n = 68/1005$). Of the *An. lungae* complex specimens, 98% were confirmed by PCR

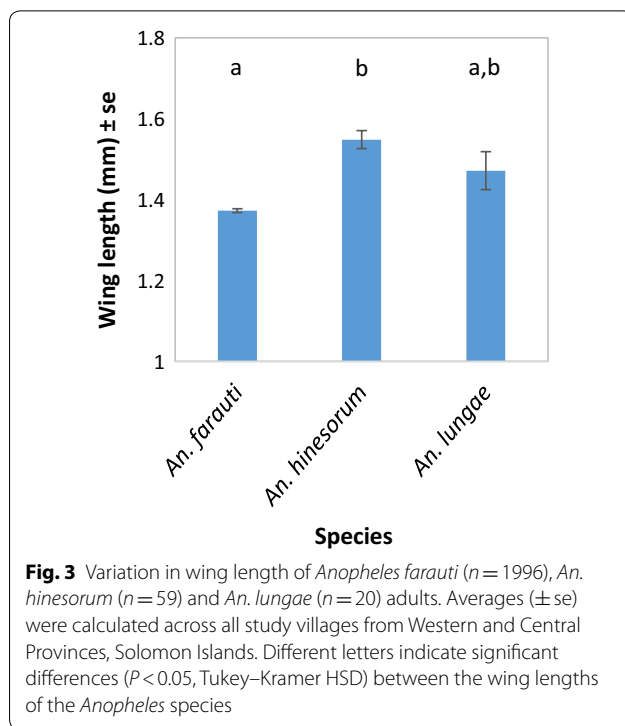
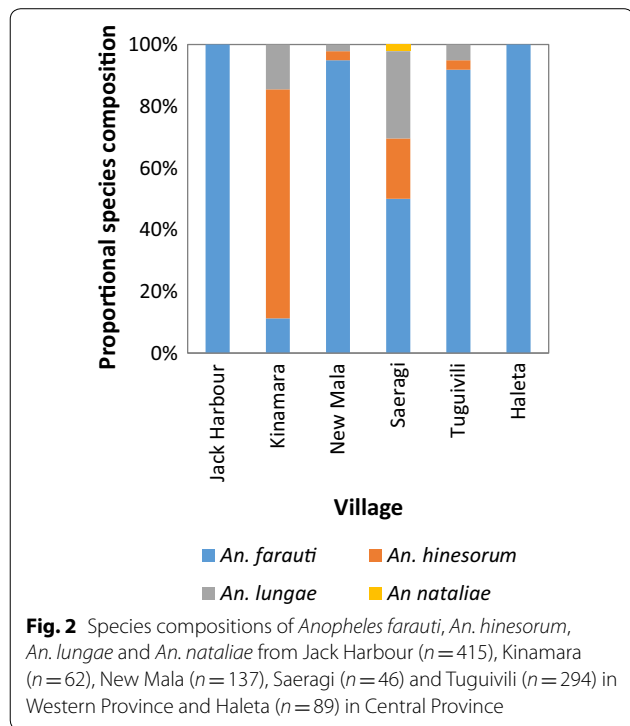
as being *An. lungae* ($n=40/41$) and 2% were *Anopheles nataliae* ($n=1/41$). Species compositions varied significantly by village ($\chi^2=5.53$, $DF=5$, $P<0.001$) (Fig. 2). *Anopheles farauti* was the dominant species in Jack Harbour, Haleta, Tuguivili and New Mala, with 100% of anophelines captured in Jack Harbour and Haleta being *An. farauti*. The dominant species in Kinamara was *An. hinesorum*. In Saeragi there was a mixture of species comprising 50% *An. farauti*, 20% *An. hinesorum*, 28% *An. lungae* and 2% *An. nataliae* (Fig. 2).

Anopheline species size

The wings of 2074 female anophelines were measured. Wing length varied significantly by anopheline species ($\beta=0.193$, $SE=0.0174$, $P<0.000$; Fig. 3) with *An. hinesorum* being significantly larger than *An. farauti* (post hoc: $P<0.0001$). Mean *An. farauti* size (as determined by wing length) varied by village ($\beta=0.053$, $SE=0.014$, $P<0.001$; Fig. 4), with the smallest *An. farauti* found in Jack Harbour, while larger *An. farauti* were found in New Mala, Haleta and Kinamara (post hoc: $P<0.05$).

Associations of *Anopheles farauti* size with population density, rainfall and distribution

The density of *An. farauti* varied significantly by village ($\beta=1.404$, $SE=0.5956$, $P<0.0001$; Fig. 5). Highest adult biting densities were found in Jack Harbour, Haleta and Tuguivili. There was a negative exponential correlation between wing length and adult biting density by



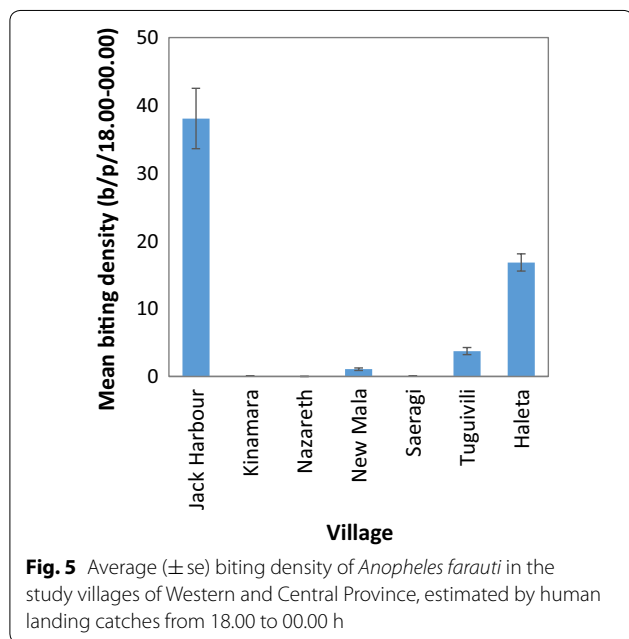
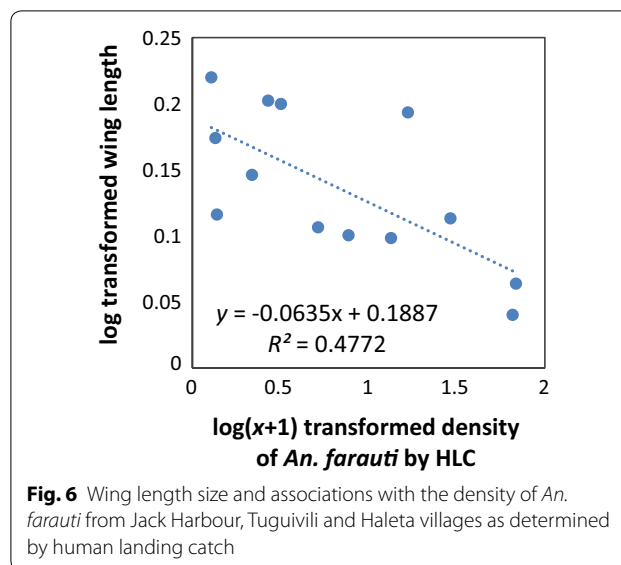
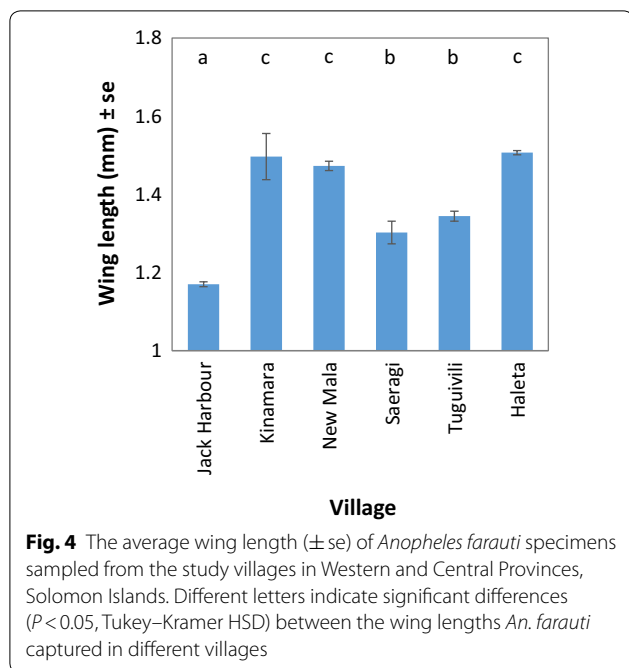
villages (Fig. 6). After log + 1 transformation, the relationship was linear and the variables were significantly correlated ($r=-0.709$, $P=0.0088$).

The density of *An. farauti* varied significantly in Jack Harbour by sampling period ($\beta=0.387$, $SE=0.039$, $P<0.0001$; Fig. 7), and was thereby incorporated into the sequential GLMMs as a random factor. The density of host seeking *An. farauti* was positively associated with rainfall in the 14-day window prior to mosquito collections ($\beta=0.0243$, $SE=0.0036$, $P<0.0001$, Fig. 7), with larger *An. farauti* populations being negatively associated with the size of the individual *An. farauti* ($\beta=-0.0002$, $SE=0.0046$, $P=0.007$, Fig. 7).

A spatial analysis of the distribution of the wing size of *An. farauti* in Jack Harbour was unable to identify any biologically meaningful patterns, indicating that there is one interspersed population within the isolated village. Although the spatial analysis did identify significant clusters with larger mosquitoes (see Additional file 1), each cluster contained only one or two sampling locations equally dispersed across the village consistent with *An. farauti* being a single population.

Discussion

Malaria transmission efficacy is a function of multiple vector parameters including size of the biting anopheline populations, survivorship, human blood feeding

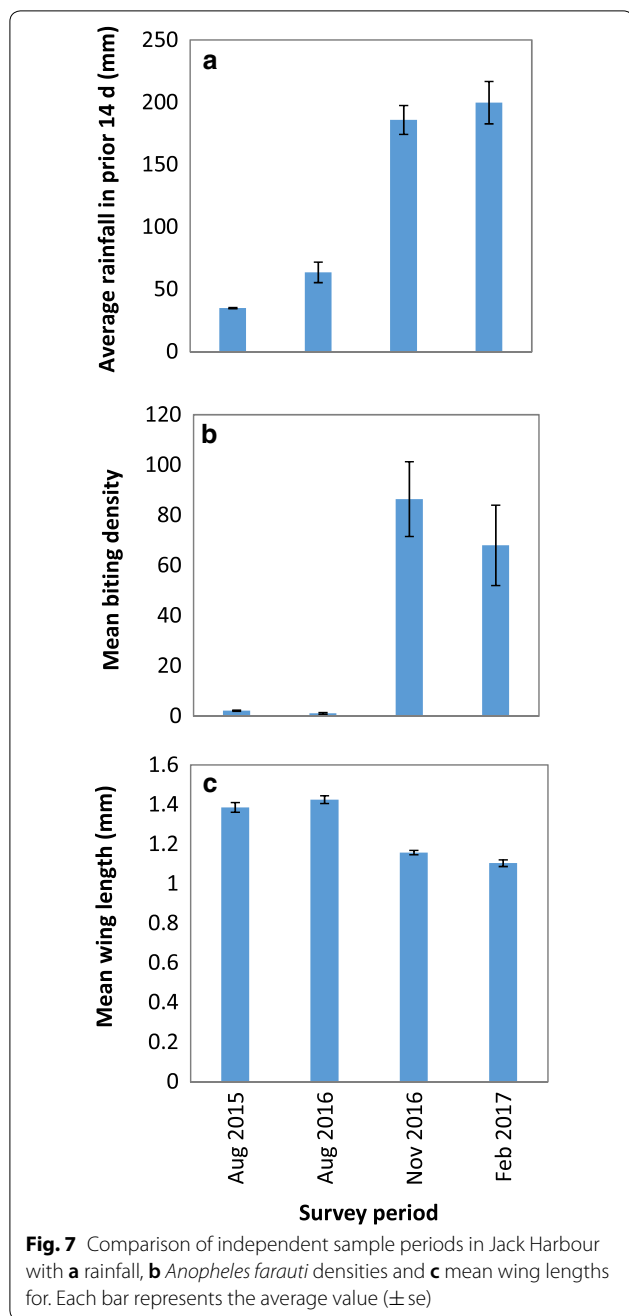


factors, such as temperature, water and resource availability (exogenous processes) [10–12].

Considering that the size of adult mosquitoes is governed by competition during the immature aquatic stages, understanding the population dynamics of mosquitoes in larval habitats is important. Many anophelines larvae are found in large habitats (swamps, lagoons, ponds) [27], for which the relationship between larval density and adult fitness has only been recently demonstrated [13, 38]. In the Solomon Islands, *An. farauti* are often associated with large fresh or brackish water swamps (e.g., Jack Harbour and Haleta villages). These habitats are believed to produce most of the adult *An. farauti* despite the low densities of larvae found (unpublished data).

Here, the finding that the mean *An. farauti* body size was negatively associated with higher densities suggests a density dependent feedback occurring during the larval stages. The smallest *An. farauti* were found in Jack Harbour, which is dominated by a large swamp as the primary larval habitat. This habitat consistently had low densities of larvae (unpublished data). It is hypothesised that density dependent effects are outcomes affected by a combination of both the density of the larval population and the ability of the environment to support the population. Our observations suggest that this swamp habitat may possibly be quite nutrient poor and thus density dependent effects may be exerted as small *An. farauti* adults from low larval densities. Following rainfall, *An. farauti* populations increased in number and these more abundant mosquitoes were smaller still in size; this observation is also consistent with interacting environmental and density dependent influences on *An. farauti* populations (the relationship between rainfall

frequency and susceptibility to infection [34, 35], with the population dynamics of most taxa being influenced by both environmental and density dependent processes [36, 37]. The size of the biting populations and survivorship are two dominant transmission parameters with the potential to be impacted by density dependent feedback. For mosquito species, the influence of density dependent feedback on population growth is often overlooked, as population growth is heavily influenced by environmental



and anopheline population size is well documented for a range of anopheline species [39–41]).

A previous study in these same villages established an association between the human biting rate of *An. farauti* and a malaria transmission focus in Jack Harbour [29]. Significant differences in mean *An. farauti* sizes among the villages within and outside this high malaria transmission focus were observed with the smallest *An. farauti* found in the malaria focus. However, identical estimates of *An. farauti* survivorship (by parous rates) were found

in the focus village (Jack Harbour) and a village outside the focus (New Mala) [29], suggesting that any impact on survivorship associated with adult size was not of a magnitude that could be measured. Furthermore, the peak transmission period in the high malaria focus village corresponded with the period of highest abundance but smallest sized *An. farauti* [13]. This suggests that density dependence effects were insufficient to limit malaria transmission in the Solomon Islands, and that the density of *An. farauti* adult population is the strongest determinant of malaria transmission rates. This is not consistent with the dogma of smaller mosquitoes being less fit and therefore not as likely to survive long enough to transmit malaria [10, 15, 28]. Similarly, previous research with *Anopheles gambiae* in Tanzania, has observed that the success of host seeking females is not linked to population densities [11]. The observation in this study (that a population of smaller mosquitoes are not always less fit) is based on data from only two villages (in which a sufficient sample of mosquitoes could be collected to estimate parity) and would need confirming by additional observations. Any fitness loss associated with smaller mosquitoes appears to be outweighed by greater impact on transmission resulting from the greater numbers of mosquitoes present during the high transmission season.

Conclusions

The findings here support a number of hypotheses. Firstly, *An. farauti* populations are directly associated with rainfall and inversely associated with the body size of individual mosquitoes. Secondly, density dependence impacts are likely occurring in a variety of habitats including large habitats. These impacts are expressed as variations in adult *An. farauti* size. The smallest *An. farauti* occurred during the peak transmission season suggesting that small mosquitoes are capable of adequately surviving long enough to transmit malaria. These results minimise concerns about whether density dependence might produce fitter vectors and suggests that decisions on whether to integrate larval control with ITN or IRS strategies for malaria vector control should be based predominantly on the capacity of national vector borne diseases control programs to effectively treat larval habitats.

Additional file

Additional file 1. Spatial clusters of locations where larger *An. farauti* were captured within Jack Harbour as detected with SatScan.

Abbreviations

API: annual parasite incidence; HLC: human landing catch; IRS: indoor residual spraying; LLIN: long lasting insecticidal net; LSM: larval source management; SIMHMS: Solomon Islands Ministry of Health and Medical Services; WHO: World Health Organization.

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Authors' contributions

Study design, manuscript preparation and data analysis: KMcl, TLR, TRB. Data collection: KMcl, AA, HB, OJ. Laboratory processing: KMcl, RDC, NB. All authors read and approved the final manuscript.

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Availability of data and materials

The dataset supporting the conclusions of this article are available in the JCU Tropical Data Hub repository at: <https://doi.org/10.25903/5caedbc8a62cb>.

Ethics approval and consent to participate

Ethical approvals were obtained from the National Health Research & Ethics Committee, Solomon Islands (2011-05-02, HRE002/16), the James Cook University Human Research Ethics Committee, Australia (H4914 and H6488). Meetings were held with community leaders, study participants and village residents, where the aims, the possible risks and potential benefits of the study were explained in Solomon Islands Pidgin. Mosquito collectors were then recruited from village residents and enrolled in the study after the risks were explained and an informed consent agreement signed.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Bhatt S, Weiss DJ, Cameron E, Bisanzio D, Mappin B, Dalrymple U, et al. The effect of malaria control on *Plasmodium falciparum* in Africa between 2000 and 2015. *Nature*. 2015;526:207–11.
- Alonso P, Noor AM. The global fight against malaria is at crossroads. *Lancet*. 2017;390(10112):2532–4.
- World Health Organization. Global technical strategy for malaria 2016–2030. Geneva: World Health Organization; 2015.
- Russell TL, Beebe NW, Bugoro H, Apairamo A, Chow WK, Cooper RD, et al. Frequent blood feeding enables insecticide-treated nets to reduce transmission by mosquitoes that bite predominately outdoors. *Malar J*. 2016;15:156.
- Killeen GF, Govella NJ, Lwetoijera DW, Okumu FO. Most outdoor malaria transmission by behaviourally-resistant *Anopheles arabiensis* is mediated by mosquitoes that have previously been inside houses. *Malar J*. 2016;15:225.
- Huijben S, Paaijmans KP. Putting evolution in elimination: winning our ongoing battle with evolving malaria mosquitoes and parasites. *Evol Appl*. 2018;11(4):415–30.
- Tusting LS, Thwing J, Sinclair D, Fillinger U, Gimnig J, Bonner KE, et al. Mosquito larval source management for controlling malaria. *Cochrane Database Syst Rev*. 2013;8:CD008923.
- Ferguson HM, Dornhaus A, Beeche A, Borgemeister C, Gottlieb M, Mulla MS, et al. Ecology: a prerequisite for malaria elimination and eradication. *PLoS Med*. 2010;7:e1000303.
- Russell TL, Beebe NW, Cooper RD, Lobo NF, Burkot TR. Successful malaria elimination strategies require interventions that target changing vector behaviours. *Malar J*. 2013;12:56.
- Lyimo E, Takken W, Koella JC. Effect of rearing temperature and larval density on larval survival, age at pupation and adult body size of *Anopheles gambiae*. *Entomol Exp Appl*. 1992;63:265–71.
- Charlwood JD, Smith T, Kihonda J, Heiz B, Billingsley PF, Takken W. Density independent feeding success of malaria vectors (Diptera: Culicidae) in Tanzania. *Bull Entomol Res*. 1995;85:29–35.
- Churcher T, Dawes E, Sinden R, Christophides G, Koella J, Basanez M-G. Population biology of malaria within the mosquito: density-dependent processes and potential implications for transmission-blocking interventions. *Malar J*. 2010;9:311.
- Russell TL, Lwetoijera DW, Knols BGJ, Takken W, Killeen GF, Ferguson HM. Linking individual phenotype to density-dependent population growth: the influence of body size on the population dynamics of malaria vectors. *Proc R Soc B*. 2011;278:3142–51.
- White M, Griffin J, Churcher T, Ferguson N, Basanez M-G, Ghani A. Modelling the impact of vector control interventions on *Anopheles gambiae* population dynamics. *Parasit Vectors*. 2011;4:153.
- Moller-Jacobs L, Murdock C, Thomas M. Capacity of mosquitoes to transmit malaria depends on larval environment. *Parasit Vectors*. 2014;7:593.
- Lyimo EO, Takken W. Effects of adult body size on fecundity and the pre-gravid rate of *Anopheles gambiae* females in Tanzania. *Med Vet Entomol*. 1993;7:328–32.
- Briegleb H. Fecundity, metabolism, and body size in *Anopheles* (Diptera: Culicidae), vectors of malaria. *J Med Entomol*. 1990;27:839–50.
- Armbruster P, Hutchinson RA. Pupal mass and wing length as indicators of fecundity in *Aedes albopictus* and *Aedes geniculatus* (Diptera: Culicidae). *J Med Entomol*. 2002;39:699–704.
- Gimnig JE, Ombok M, Otieno S, Kaufman MG, Vulule JM, Walker ED. Density-dependent development of *Anopheles gambiae* (Diptera: Culicidae) larvae in artificial habitats. *J Med Entomol*. 2002;39:162–72.
- Blackmore MS, Lord CC. The relationship between size and fecundity in *Aedes albopictus*. *J Vector Ecol*. 2000;25(2):212–7.
- Ameneshewa B, Service MW. The relationship between female body size and survival rate of the malaria vector *Anopheles arabiensis* in Ethiopia. *Med Vet Entomol*. 1996;10:170–2.
- Saul A. Estimation of survival rates and population size from mark-recapture experiments of bait-caught haematophagous insects. *Bull Entomol Res*. 1987;77:589–602.
- Landry S, DeFoliart G, Hogg D. Adult body size and survivorship in a field population of *Aedes triseriatus*. *J Am Mosq Control Assoc*. 1988;4:121–8.
- Lyimo EO, Koella JC. Relationship between body size of adult *Anopheles gambiae* s.l. and infection with the malaria parasite *Plasmodium falciparum*. *Parasitology*. 1992;104:233–7.
- WHO. World malaria report. Geneva: World Health Organization; 2018.
- Cooper RD, Frances SP. Malaria vectors on Buka and Bougainville islands, Papua New Guinea. *J Am Mosq Control Assoc*. 2002;18:100–6.

27. Russell TL, Burkot TR, Bugoro H, Apairamo A, Beebe NW, Chow WK, et al. Larval habitats of the *Anopheles farauti* and *Anopheles lungae* complexes in the Solomon Islands. *Malar J*. 2016;15:164.
28. Dietz K. Density-dependence in parasite transmission dynamics. *Parasitol Today*. 1988;4:91–7.
29. Burkot TR, Bugoro H, Apairamo A, Cooper RD, Echeverry DF, Odabasi D, et al. Spatial–temporal heterogeneity in malaria receptivity is best estimated by vector biting rates in areas nearing elimination. *Parasit Vectors*. 2018;11:606.
30. Waltmann A, Darcy AW, Harris I, Koepfli C, Lodo J, Vahi V, et al. High rates of asymptomatic, sub-microscopic *Plasmodium vivax* infection and disappearing *Plasmodium falciparum* malaria in an area of low transmission in Solomon Islands. *PLoS Negl Trop Dis*. 2015;9:e0003758.
31. Belkin JN. The mosquitoes of the South Pacific (Diptera, Culicidae). Berkeley: University of California Press; 1962.
32. Beebe NW, Saul A. Discrimination of all members of the *Anopheles punctulatus* complex by polymerase chain reaction-restriction fragment length polymorphism analysis. *Am J Trop Med Hyg*. 1995;53:478–81.
33. McLaughlin K, Russell TL, Apairamo A, Bugoro H, Oscar J, Cooper RD, et al. Dataset describing the longitudinal density and wing length of anophelines in Solomon Islands. James Cook University Tropical Data Hub. 2019. <https://doi.org/10.25903/5caedbc8a62cb>.
34. Brady OJ, Godfray HCJ, Tatem AJ, Gething PW, Cohen JM, McKenzie FE, et al. Vectorial capacity and vector control: reconsidering sensitivity to parameters for malaria elimination. *Trans R Soc Trop Med Hyg*. 2016;110:107–17.
35. Cohuet A, Harris C, Robert V, Fontenille D. Evolutionary forces on Anophelines: what makes a malaria vector? *Trends Parasitol*. 2010;26:130–6.
36. Sibly RM, Barker D, Denham MC, Hone J, Page M. On the regulation of populations of mammals, birds, fish and insects. *Science*. 2005;309:607–10. <https://doi.org/10.1126/science.1110760>.
37. Brook BW, Bradshaw CJA. Strength of evidence for density dependence in abundance time series of 1198 species. *Ecology*. 2006;87:1445–51.
38. Yang G-J, Brook BW, Whelan PI, Cleland S, Bradshaw CJA. Endogenous and exogenous factors controlling temporal abundance patterns of tropical mosquitoes. *Ecol Appl*. 2008;18:2028–40.
39. Bugoro H, Hii J, Russell T, Cooper R, Chan B, Iro'ofa C, et al. Influence of environmental factors on the abundance of *Anopheles farauti* larvae in large brackish water streams in Northern Guadalcanal, Solomon Islands. *Malar J*. 2011;10:262.
40. Smith J, Tahani L, Bobogare A, Bugoro H, Otto F, Fafale G, et al. Malaria early warning tool: linking inter-annual climate and malaria variability in northern Guadalcanal, Solomon Islands. *Malar J*. 2017;16:472.
41. Abiodun GJ, Maharaj R, Witbooi P, Okosun KO. Modelling the influence of temperature and rainfall on the population dynamics of *Anopheles arabiensis*. *Malar J*. 2016;15:364.

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