

ResearchOnline@JCU

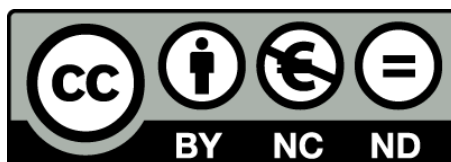
This is the **Accepted Version** of a paper published in the
Journal Estuarine, Coastal and Shelf Science:

Waltham, Nathan J., and Sheaves, Marcus (2017) *Acute thermal tolerance of tropical estuarine fish occupying a man-made tidal lake, and increased exposure risk with climate change*. Estuarine, Coastal and Shelf Science, 196. pp. 173-181.

<https://doi.org/10.1016/j.ecss.2017.06.032>

© 2017. This manuscript version is made available under
the CC-BY-NC-ND 4.0 license

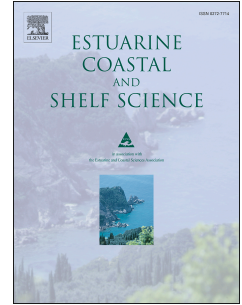
<http://creativecommons.org/licenses/by-nc-nd/4.0/>



Accepted Manuscript

Acute thermal tolerance of tropical estuarine fish occupying a man-made tidal lake, and increased exposure risk with climate change

Nathan J. Waltham, Marcus Sheaves



PII: S0272-7714(16)30585-6

DOI: [10.1016/j.ecss.2017.06.032](https://doi.org/10.1016/j.ecss.2017.06.032)

Reference: YECSS 5522

To appear in: *Estuarine, Coastal and Shelf Science*

Received Date: 7 November 2016

Revised Date: 8 May 2017

Accepted Date: 25 June 2017

Please cite this article as: Waltham, N.J., Sheaves, M., Acute thermal tolerance of tropical estuarine fish occupying a man-made tidal lake, and increased exposure risk with climate change, *Estuarine, Coastal and Shelf Science* (2017), doi: 10.1016/j.ecss.2017.06.032.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1
2 Acute thermal tolerance of tropical estuarine fish occupying a man-
3 made tidal lake, and increased exposure risk with climate change

4
5
6 Nathan J. Waltham^{*1,2}, Marcus Sheaves^{1,2}

7
8 ¹Centre for Tropical Water & Aquatic Ecosystem Research (TropWATER), James Cook
9 University, Queensland, 4811, Australia.

10 ²College of Science & Engineering, James Cook University, Townsville, Queensland, 4811,
11 Australia.

12
13 *Author Tel + 61 7 4781 4191; fax + 61 7 4781 5589

14 E-mail address: nathan.waltham@jcu.edu.au

15
16 Keywords: Acute Effect Temperature (AET), fish, thermal refugia, tropical estuary, climate
17 change

18 Running title: Acute thermal tolerance and climate risks for tropical estuary fish

19

20 **Highlights**

- 21 • Constructed tidal lakes provide new and additional habitat for fish
- 22 • Summer lake surface water temperature frequency exceeded acute thresholds for fish
- 23 • Summer lake bottom water provide important thermal refugia for fish
- 24 • Future climate change will double surface acute thermal exposure
- 25 • Current bottom water refugia will almost certainly be reduced under future climate

26

ACCEPTED MANUSCRIPT

27 **Abstract**

28 Understanding acute hyperthermic exposure risk to animals, including fish in tropical estuaries,
29 is increasingly necessary under future climate change. To examine this hypothesis, fish (upper
30 water column species - glassfish, *Ambassis vachellii*; river mullet, *Chelon subviridis*; diamond
31 scale mullet, *Ellochelon vaigiensis*; and ponyfish, *Leiognathus equulus*; and lower water bottom
32 dwelling species – whiting *Sillago analis*) were caught in an artificial tidal lake in tropical north
33 Queensland (Australia), and transported to a laboratory tank to acclimate (3wks). After
34 acclimation, fish (between 10 to 17 individuals each time) were transferred to a temperature
35 ramping experimental tank, where a thermoline increased (2.5°C/hr; which is the average
36 summer water temperature increasing rate measured in the urban lakes) tank water temperature
37 to establish threshold points where each fish species lost equilibrium (defined here as Acute
38 Effect Temperature; AET). The coolest AET among all species was 33.1°C (*S. analis*), while
39 the highest was 39.9°C (*A. vachellii*). High frequency loggers were deployed (November and
40 March representing Austral summer) in the same urban lake where fish were sourced, to
41 measure continuous (20min) surface (0.15m) and bottom (0.1m) temperature to derive thermal
42 frequency curves to examine how often lake temperatures exceed AET thresholds. For most
43 fish species examined, water temperature that could be lethal were exceeded at the surface, but
44 rarely, if ever, at the bottom waters suggesting deep, cooler, water provides thermal refugia for
45 fish. An energy-balance model was used to estimate daily mean lake water temperature with
46 good accuracy ($\pm 1^\circ\text{C}$; $R^2 = 0.91$, modelled vs lake measured temperature). The model was used
47 to predict climate change effects on lake water temperature, and the exceedance of thermal
48 threshold change. A 2.3°C climate warming (based on 2100 local climate prediction) raised lake
49 water temperature by 1.3°C. However, small as this increase might seem, it led to a doubling of
50 time that water temperatures were in excess of AET thresholds at the surface, but also the
51 bottom waters that presently provide thermal refugia for fish.

52

53 **1. Introduction**

54 Despite being incredibly productive habitats for fish (Blaber et al., 2010; Manson et al., 2005;
55 Nagelkerken et al., 2015), across much of the world tropical estuaries continue to be modified
56 for human gain (Rozas, 1992; Wen et al., 2010). An example of this modification occurs where
57 property developers excavate large tracts of natural wetlands (e.g., mangroves, saltmarsh), or dig
58 out terrestrial habitat to create artificial, urban water development, designed to increase extent of
59 usable waterfront land (Lindall et al., 1973; Waltham and Connolly, 2013). Residential urban
60 waterways have been built on most continents, and collectively contribute to over 4,000 km
61 linear of engineered habitat for fish (Waltham and Connolly, 2011). In utilising these built
62 waterways fish (Claassens, 2016; Waltham and Connolly, 2006) are susceptible to
63 contamination and poor water quality (Maxted et al., 1997), and hydraulic connectivity with
64 downstream estuaries may be altered (Zigic et al., 2002). Furthermore, their position in low
65 lying areas of coastal floodplains raise concerns about vulnerability to sea level rise, shoreline
66 erosion (Harvey and Stocker, 2015), and that climate change might reduce the utility of these
67 man-made habitats for fish (Waltham and Connolly, 2011).

68
69 Animals spend a significant proportion of time (and energy) avoiding or escaping stimuli
70 (predation, chemical contamination, noise) that could cause physical harm that reduces fitness or
71 causes death (Connell, 1993). One causal stimulus contributing to animal avoidance is exposure
72 to high temperature (Brett, 1956). Determining effects of temperature on animal behaviour and
73 movement has received increasing attention prompted by climate change concerns and how
74 future, warmer temperature may cause range shifts in distribution of native (James et al., 2017;
75 Stewart et al., 2013; Welbergen et al., 2008), or invasive species (Carveth et al., 2006), or in
76 some cases extinction of vulnerable species (Thomas et al., 2004). For many aquatic species,
77 including fish, temperature directly controls metabolic rate, and can influence growth, resource
78 allocation for reproduction and ultimately, population size (Armstrong et al., 2013; Jobling,

79 1995). Evidence shows that growth rate and development in fish tend to increase with
80 temperature up to an optimum, provided sufficient food is available (Eaton and Scheller, 1996).
81 However, the long term (chronic) effects of exposure to elevated water temperature can include
82 reduced year class strength (Brown et al., 2016; Nunn et al., 2003), cessation of growth, and
83 increased susceptibility to environmental stresses such as low concentrations of dissolved
84 oxygen (Pearson et al., 2003). Exposure to extreme temperature causes acute hyperthermic (or
85 hypothermic) response, requiring animals to thermoregulate or they will die (Coulter et al.,
86 2016; McCauley and Casselman, 1981). Determining the temperature threshold (defined here as
87 Acute Effect Temperature, AET) provides insight into thermal exposure risk, necessary for
88 species protection and conservation.

89
90 This paper reports fine time-interval resolution (20 min) continuous water temperature
91 measurements made in a residential man-made tidal lake in Townsville, northern Queensland,
92 Australia. We used these data to quantify how water temperature changes as the austral summer
93 evolves, and how water temperature varies between the surface and bottom layer in tidal built
94 lakes. We then determine the AET for five common estuarine fish that occupy the lakes using
95 laboratory manipulative experiments, to assess how often lake water temperature approach and
96 exceed these thresholds. Advancements in water thermal energy modelling provides the
97 opportunity to predict temperature exposure risk to aquatic animals using readily available daily
98 weather data (McJannet et al., 2014; McJannet et al., 2012; Wallace et al., 2015). We then use
99 an energy balance model to simulate how climate change might influence the thermal exposure
100 risk for fish occupying engineered tidal lakes.

101

102 **2. Methods**

103 *2.1 Study area*

104 Ross Creek is a small (8 km linear) transitional (Elliott and Whitfield, 2011) estuary in tropical
105 north Queensland (-19.270688° S, 146.788279° E) that flows into Cleveland Bay, and the Great
106 Barrier Reef lagoon, Figure 1a (Sheaves and Johnston, 2010). Located adjacent to Ross Creek
107 is a large constructed residential tidal lake estate, built in the early 1990s as a way to increase
108 residential real estate with waterfrontage (Waltham and Sheaves, 2015), and to treat water
109 quality (sediment and nutrient load reductions) discharged from the surrounding urban and
110 industrial estates before reaching the main estuary and Great Barrier Reef lagoon. The lake
111 system is approximately 7.5 ha, average water depth is between 1.9 to 2.5 m (150ML). The lake
112 has two sections that are connected via a narrow concrete open channel (approximately 150 m
113 long, 10 m width and 1 m depth) which allows water exchange and fish passage between the
114 lakes. A long concrete channel extends from the lake, joining with Ross Creek approximately
115 3.5 km upstream from the mouth of the creek. A series of four engineered hydraulic arms
116 separate the concrete channel from Ross Creek estuary, and are synchronised to open based on
117 the tidal height of the downstream Ross Creek (though can be manually opened during extreme
118 flood events) (Causeway Floodgate Procedures, Townsville City Council, unpublished manual).
119 The hydraulic control structure permits tidal exchange with Ross Creek, in such a way that it
120 reduces the tidal prism, which is necessary to circumvent situations where increased tidal prism
121 compromises engineering rock walls or bridge foundations, and contributes to erosion along the
122 lake edges (Zigic et al., 2002). Fish visit the lakes and can return to the estuary during times
123 when the hydraulic gates are open. The lakes holds a subset of fish species found in the adjacent
124 estuaries (Sheaves et al., 2012) including a number of diadromous species common throughout
125 the region (Sheaves and Johnston, 2010; Sheaves et al., 2010; Waltham and Davis, 2016).
126 During summer months the lakes become hypoxic, a consequence of high ambient air and water
127 temperature (which reduces the solubility of oxygen in water available for fish), in addition to
128 high densities of oxygen consuming phytoplankton and sediment benthic algae; a trait that

129 contributes to poor water quality and fish kills in coastal waters of Queensland (Dunn et al.,
130 2012).

131

132 2.2 Estuary fish acute temperature effects experiments

133 In this study, a subset of local estuarine fish species were examined, including glass perch
134 (*Ambassis vachellii*), river mullet (*Chelon subviridis*), diamond scale mullet (*Ellochelon*
135 *vaigiensis*), and pony fish (*Leiognathus equulus*) – representing upper water column
136 assemblage; and the whiting (*Sillago analis*) – representing benthic dwelling assemblage. Fish
137 were collected in the lake using a seine net (10mm mesh, 1.8m drop), and transported to the
138 laboratory for acclimation (from the collection site to the laboratory was 30 min, using three 90
139 L containers each with battery aerators). The laboratory had a single 800 L saltwater tank
140 (salinity 33), set up on a re-circulatory system with water exchange set approximately 10 L/min
141 (MARFU, James Cook University).

142

143 In the laboratory, fish were acclimated to a constant temperature (28°C; $\pm 2^\circ\text{C}$) for three weeks
144 prior to the Acute Effect Temperature (AET) exposure experiment. This acclimation
145 temperature represents approximately the summer average daily water column temperature in
146 the lakes (based on historical water quality monitoring undertaken by Townsville City Council
147 since 1994 - unpublished data). Fish were fed aquaculture pellets (Ridley AgriProducts Pty Ltd)
148 every 2-3 days; all fish were feeding during the acclimation period suggesting that they were not
149 stressed prior to the temperature exposure experiment.

150

151 In the AET experiment an experimental glass aquarium tank (0.7 x 0.4 x 0.6 m; ~150 L) was
152 designed specifically for the experiment. Two circulatory pumps were placed in the tank to
153 ensure the tank was well mixed. Water in the experiment tank was continuously replaced at a
154 rate of 2 L/min with water on the acclimation tank system. Photoperiod in the aquarium

155 laboratory was maintained at 12:12h dark:light cycle. The experimental tank was cleaned after
156 each experiment, resulting in an approximate 80% water exchange.

157

158 Between 10 and 16 individual fish were transplanted from the acclimation tank to the
159 experimental tank 2-3 days prior to the AET experiment tank so that fish would acclimate to the
160 new tank setting. During the experimental tank acclimation period, conditions (i.e., water
161 temperature (28°C) and photoperiod) remained the same as the acclimation tank.

162

163 At the start of each AET experiment, the water circulation pipe was closed so the tank was a
164 single experimental unit. A programmable thermo-controller (Thermoline, Eurotherm 3216
165 Control) was used to increase the water temperature at a linear rate of approximately 2-3°C per
166 hour with the experiment commencing at the acclimation temperature (this rate is similar to
167 diurnal water temperature changes experienced in the lake, see below). The time elapsed and
168 water temperature on the thermocline readout display were recorded when fish (one at a time)
169 lost equilibrium or displayed erratic behaviour (Burrows and Butler, 2012). Fish were then
170 immediately placed into a separate recovery container filled with room temperature (28°C)
171 water for up to 30 mins before being relocated to a separate holding tank (to avoid repeated use
172 of fish) that was also on the main water circulatory system (fish total length was measured
173 before release; there were no linear relationship between fish size and AET, for each fish species
174 examined here). The experimental tank was drained, left to cool for 24 hrs, refilled with
175 seawater from the main acclimation tank, ready for the next experiment. Fish AET statistics
176 were determined and are presented in Table 1.

177

178 2.3 *Lake water temperature logging*

179 To profile water temperature characteristics in the urban tidal lakes, Hobo temperature loggers
180 (Onset Corporation Bourne, Massachusetts) were deployed at two depths at approximately the

181 deepest point in both lakes: 1) surface; 0.2 m below water surface; and 2) bottom; 0.1 m above
182 the lake bottom. The surface logger was attached to the underside of a 0.15 m diameter buoy to
183 shield it from the sun at all times as direct exposure could produce erroneous results. Loggers
184 were set to record data every 20 min from 31 October 2015 to 30 March 2016 (this logging
185 period represents the summer months for the region, and thereby the maximum likely
186 temperature that fish would be exposed too). This logging frequency was necessary to derive
187 water temperature frequency distributions for the purposes of assessing exposure risks (Wallace
188 et al., 2017; Wallace et al., 2015). The same logger configuration was deployed in Lake 2
189 (Keyatta Lake) (Fig 1.), unfortunately these temperature loggers, after 21 December 2015, failed
190 and no further data were retrieved. This limited our ability to generate exposure risk plots and
191 to model the temperature. Data for Lake 2 are presented in the Supplementary Notes (see Fig.
192 S1), however, is not included further in this study.

193

194 2.4 *Estimating lake water temperatures*

195 Water temperature was estimated using the energy balance model (McJannet et al., 2014;
196 Wallace et al., 2015). The model was originally developed for estimating daily evaporation
197 from open water bodies of various sizes (ranging from waterbodies ~ 60 m wide, ~ 600 km² in
198 area), but it can also calculate the daily mean water body temperature in order to specify the
199 changes in heat storage (to a well-mixed water column). The main input of energy to the model
200 is solar radiation and the main loss occurs via heat conduction to the atmosphere and
201 evaporation. It is also possible for energy to enter/leave water if there is flow, however, this
202 effect can be ignored here given the tidal exchange is small, when considering the total lake
203 volume (approximately < 0.01% of total lake volume is exchanged each day).

204

205 The lake water temperature model requires daily weather data, which were obtained from the
206 Australian Government SILO database (<http://www.nrw.qld.gov.au/silo/>). The SILO database

207 consists of interpolated meteorological variables on a 0.05° (5 km) grid for the whole of
208 Australia (Jeffrey et al., 2001). The variables available from SILO used in the temperature
209 model are air temperature, vapour pressure, solar radiation and rainfall, and the way these
210 variables are used to calculate all of the terms in the model are described by (McJannet et al.,
211 2008). The model also requires daily mean wind speed (to calculate the evaporation rate) and as
212 this is not available in the SILO database, a fixed wind speed of 1.3 m s^{-1} was applied in the
213 model; the consequence of this assumption is discussed later in the paper. Evaporation rate is
214 also dependent on the water body size, in terms of both surface area and depth. Water area
215 affects the 'wind function' used in calculating evaporation (McJannet et al., 2012). Water depth
216 primarily affects heat storage and the model is run from the beginning of the year so that its
217 depth predictions match waterhole depth measurements made during the model period
218 November 2015 to March 2016 (Supplementary Fig S3). Sensitivity analysis shows that
219 altering the water surface area or depth by a factor of 2 only changes modelled water
220 temperature by 0.8% and 0.2% respectively.

221

222 **3. Results and Discussion**

223 *3.1 Lake water temperature*

224 During the logging period, weather conditions were generally fine during November to February
225 period, as is typical for this time of year in the region. In this period the water column
226 consistently exhibited pronounced diel temperature periodicity and occasional diurnal
227 stratification (Fig 2). Typically one or two hours after sunrise each day the near-surface water
228 temperature began to rise at almost a linear rate for a period of 8 hrs reaching daily maxima as
229 high as 40.4°C (mean 33) during the early evening hours (14:00 to 17:00). The mean increase
230 in water temperature at the near-surface during the day (06:00 to 14:00) was 2.8°C h^{-1} (max 6.5
231 $^\circ\text{C h}^{-1}$). For the remaining 16 hrs of the day the near-surface water temperatures gradually

232 declined reaching a minimum 24.0 °C (mean 30.8 °C; max 38.2 °C), shortly before sunrise
233 (04:00 to 08:00).

234

235 In order to properly quantify the temperature regime in constructed urban tidal lakes, would
236 require logging water temperature over a number of spatiotemporal scales, in order to
237 incorporate the full range of engineering designs of these urban lakes (e.g., where flow is
238 controlled using tidal gates, tidal pipes, rock bund walls, as each have varying differences in the
239 hydrodynamic exchange of tidal water with the downstream primary estuary (Waltham and
240 Connolly, 2007)), and to also examine among year differences in thermal regimes. Data here
241 were collected during a single summer period, and therefore provide an indicative guide of the
242 annual minimum and maximum conditions expected at this time of the year. The methodology
243 applied here of determining acute thermal thresholds (laboratory experiments), combining with
244 high frequency continuous water temperature data, in addition to water balance models to
245 examine future exposure risks under climate change (Wallace et al., 2015), are the key focus of
246 this paper; these methods are transferable to elsewhere, to examine thermal exposure risks to
247 tropical estuarine fish species.

248

249 Water temperatures near the bottom increased at a more gradual rate each day, with an
250 increasing trend sustained for a longer period (15 hrs), therefore daily maximum was reached
251 after sunset (mean 29.3 °C; max 36.5 °C). The mean hourly (during the day) increase in the
252 near-lake bottom was 0.3 °C h⁻¹ (max 1.8 °C h⁻¹). The fact that bottom water temperature
253 continued to increase (lag) after sunset is a consequence of either continuing thermal heat
254 exchange after sunset (conduction), or the effect of partial mixing with the warmer surface
255 water.

256

257 An important fact in the data here is that lake water temperature changed across the logging
258 period (Fig 2a), where the water column was vertically well mixed (surface and bottom waters
259 remained similar), but at other times the column was diurnally stratified, where surface and
260 bottom waters were separated by several degrees Celsius (Fig. 2b). Changes in mixing often
261 coincided with rainfall events, the most notable occurring on 10 January 2016 (91.8 mm over 72
262 hours, Townsville airport, station number 32040), where the initial influx of cool rainwater
263 decreased surface water temperature by 6 °C in just 3 hours, compared to only 3 °C in this time
264 period in the bottom water temperature. It can be seen that rainfall causes dramatic changes in
265 the thermal regime in the lake, where diurnally stratified profile then became vertically well
266 mixed, which occurred on 28 December 2015 after 91.8 mm of rainfall fell in 3 days (Fig 2b).
267 The period where the near-surface waters (and to a lesser extent bottom waters) were coolest
268 occurred between 20 February and 18 March 2016, where a series of rainfall events (totaling
269 561mm over several weeks) occurred in the region. In this time series, after each rainfall event
270 surface water temperature progressively increased again until the next rainfall event. In fact,
271 between 11 and 18 March 2016 the bottom water temperature was higher than at the surface,
272 which indicates the ability for deeper waters to store thermal energy for relatively long periods.
273

274 3.2 Fish temperature threshold experiments

275 The AET of five estuarine fish species ranged between 33.1 °C (minimum) and 39.9 °C
276 (maximum), Table 1. Of these fish species, the range between lowest AET and the highest for
277 an individual species was 6.7 °C, which occurred for the whiting, *Sillago analis*, while the
278 smallest difference (0.9 °C) was found for diamond scale mullet, *Ellochelon viagensis*.
279 Interestingly, in a study of thermal tolerance of marine fish in Indonesia, Eme and Bennett
280 (2009) revealed a similar thermal tolerance for *E. viagensis* of 38°C when acclimated at about
281 25°C, compared to (AET₅₀) 39.5°C when acclimated at 28°C in the present study.

282

283 The narrow range in AET has been found in other studies using estuarine fish species (Cheng et
284 al., 2013; Heath et al., 1993; Rajaguru and Ramachandran, 2001), but also tropical freshwater
285 fish species (Burrows and Butler, 2012). Table 1 provides an overview of the percentage of
286 time that each fish species exceeded the thresholds using the surface and bottom logging
287 temperature data. For example, the surface dwelling glass perch, *Ambassis vachellii*, exceeded
288 the minimum AET (35.1°C) 26% of the logging time at the surface, and less than 3% of the
289 logging time at the bottom waters. In contrast the benthic dwelling whiting, *Sillago analis*,
290 minimum AET threshold (33.1°C) was exceeded 58% of the logging time at the surface, and
291 28% of the logging time at the bottom waters. When exposed to water temperature above these
292 thermal thresholds, we assume fish would search for cooler thermal refugia. Both lakes have
293 limited edge vegetation for shading (Fig. 1B); to this end the most likely response for fish would
294 be to descend the water column to the bottom, cooler lake waters (Figure 3b), where neither of
295 the above thresholds are exceeded during the summer months. The need to migrate through the
296 water column has been also suggested for freshwater fish occupying ephemeral waterholes
297 within the tropical seasonal rivers of northern Australia (Wallace et al., 2017). While providing
298 thermal refugia, the cooler bottom waters have critically low, hypoxic, dissolved oxygen
299 concentrations (unpublished data Townsville City Council). By continually adjusting position
300 to regulate against high water temperature and dissolved oxygen, fish would use important
301 energy reserves leaving them more susceptible to critical water quality conditions,
302 compromising reproductive fitness and predation susceptibility (Eaton et al., 1995). The energy
303 balance model used here shows that thermoregulation will increase under future climate change,
304 for some fish this increase is considerable. For example, the ponyfish, *Leiognathus equulus*,
305 AET is exceeded between 14% of the logging time (based on maximum AET), but could be as
306 high as 47% (using the minimum AET) at the surface, and up to 14% of the logging time in the
307 bottom waters (based on minimum AET) under future climate conditions (Table 1).

308

309 The acclimation tank temperature used in our experiment (28 °C) is commonly reported in the
310 literature (Burrows and Butler, 2012). Clearly these data show that aquatic organisms inhabiting
311 transitional coastal waters are exposed to constantly fluctuating water temperature, which raises
312 questions regarding the validity of acclimating at a constant temperature (Rajaguru and
313 Ramachandran, 2001); it would seem advisable to simulate the natural diel temperature
314 periodicity of the animals' environment during acclimation (Coulter et al., 2016). We advocate
315 here that fluctuating acclimation temperatures are probably most appropriate in laboratory
316 experiments, however, based on our field measured temperature data, determining what diel
317 range to simulate would be difficult.

318

319 3.3 *Modelling lake water temperature*

320 The water temperature model used here predicted measured temperature to within 1°C during
321 periods when the lake was well mixed (Supplementary Fig S2 and S3). Underestimation of
322 water temperature in the modelling has been shown in other studies McJannet et al., (2008) and
323 Wallace et al., (2017), where those studies attributed the underestimation in wind speed (which
324 may be different to the 1.3m^{-s} applied here in the model; increasing the wind speed to 2m^{-s} (as
325 applied in freshwater waterholes in northern Australia; Wallace et al., 2015; Wallace et al.,
326 2017) contributed to further model underestimation ($R^2 = 0.81$). For this reason the absolute
327 accuracy of the modelling when using readily available government weather climate data is
328 probably between 1 and 2°C, however, precision could be improved with installation of weather
329 stations, recording continuous weather conditions, that are located immediately adjacent to
330 water body of interest.

331

332 Concern about the potential impacts of climate change on coastal transitional waters is
333 widespread. For example, authors of studies in the US and Europe have found that climate-
334 warming increases may reduce habitat availability, while for others it will increase leading to

335 shifts in species distributions (Buisson et al., 2008; Eaton and Scheller, 1996; Robins et al.,
336 2016; Sinokrot et al., 1995). Future changes in climate in northern Queensland could affect the
337 thermal environment of constructed urban tidal lakes. For the proposed increase in air
338 temperature of 2.7°C by 2100 (Hennessy et al., 2008), the model predicts water temperature will
339 increase accordingly by 1.3 °C. The modelling suggests that the period of exposure to acute
340 thermal conditions increases, particularly at the near-surface water layer. It seems that deeper
341 lake areas might provide important thermal refugia, where water temperatures under future
342 climate conditions remain below the thermal threshold for the fish species examined here. On
343 this basis it seems probable that fish occupying the deep waters are shielded from future climate,
344 however, fish species associated with near-surface waters may need to migrate down the water
345 column, more often, to find thermal relief. In the future, vertical migration in the water column
346 may increase expose to critically low dissolved oxygen (Marshall and Elliott, 1998; O'connell et
347 al., 2000). Fish in estuaries may be also subjective to salinity which can vary seasonally, tidally
348 and following rainfall (Araujo et al., 2000; Marshall and Elliott, 1998; Whitfield et al., 1981).
349 The interaction between salinity and water temperature has been previously shown to influence
350 thresholds in estuarine fish (including for analogous Ambassidae species) (Blaber, 1973; Martin,
351 1988), and should be investigated in future research.

352

353 **4. Conclusions**

354 Once in a constructed artificial urban lake, at least some estuarine fish species are faced with
355 acute thermal exposure stress during summer months, and in response, would need to actively
356 search for thermal refugia, including potentially accessing the cooler, lake, bottom waters.

357 Using a water energy balance model, it seems that fish occupying the near-surface waters will
358 spend more hours of the day searching for thermal refugia under future climate change, in some
359 cases up to twice the amount of time each day that they currently invest. Whether fish can
360 successfully achieve this will be influenced by other factors, such as available oxygen, salinity

361 or prey abundance, but would indeed still require fish to be continually moving in the water
362 column. We believe that the methodology presented here is transferable to other transitional
363 water locations.

364

365 **Acknowledgements**

366 Funding for this project was provided by TropWATER (Centre for Tropical Water and Aquatic
367 Ecosystem Research), and College of Science and Engineering, James Cook University, internal
368 grant awarded to NW. We thank Prof J Wallace (TropWATER) for assistance with climate
369 change modelling, M Kuehlcke for laboratory assistance, and staff from MARFU (Marine
370 Aquaculture Research Facility Unit), James Cook University. This study was completed under
371 ethics approval (A2150) and general fisheries permit (151660).

372

373 **References**

- 374 Araujo, F.G., Williams, W.P., Bailey, R.G., 2000. Fish assemblages as indicators of water quality
375 in the Middle Thames Estuary, England (1980-1989). *Estuaries* 23, 305-317. [https://doi:
376 10.2307/1353323](https://doi.org/10.2307/1353323)
- 377 Armstrong, J.B., Schindler, D.E., Ruff, C.P., Brooks, G.T., Bentley, K.E., Torgersen, C.E., 2013.
378 Diel horizontal migration in streams: juvenile fish exploit spatial heterogeneity in thermal
379 and trophic resources. *Ecology* 94, 2066-2075. [https://doi: 10.1890/12-1200.1](https://doi.org/10.1890/12-1200.1)
- 380 Blaber, S.J., 1973. Temperature and salinity tolerance of juvenile *Rhabdosargus holubi*
381 [Steindachner (Teleostei: Sparidae)]. *J. Fish Biol.* 5, 593-598. [https://doi: 10.1111/j.1095-
382 8649.1973.tb04492.x](https://doi.org/10.1111/j.1095-8649.1973.tb04492.x)
- 383 Blaber, S.J.M., Griffiths, S.P., Pillans, R., 2010. Changes in the fish fauna of a tropical Australian
384 estuary since 1990 with reference to prawn predators and environmental change.
385 *Estuarine, Coastal and Shelf Science* 86, 692-696.
386 <https://doi.org/10.1016/j.ecss.2009.12.012>

- 387 Brett, J., 1956. Some principles in the thermal requirements of fishes. *The Quarterly Review of*
388 *Biology* 31, 75-87.
- 389 Brown, C.A., Sharp, D., Mochon Collura, T.C., 2016. Effect of climate change on water
390 temperature and attainment of water temperature criteria in the Yaquina Estuary, Oregon
391 (USA). *Estuarine, Coastal and Shelf Science* 169, 136-146.
392 <https://doi.org/10.1016/j.ecss.2015.11.006>
- 393 Buisson, L., Thuiller, W., Lek, S., Lim, P., Grenouillet, G., 2008. Climate change hastens the
394 turnover of stream fish assemblages. *Global Change Biology* 14, 2232-2248.
395 [https://doi: 10.1111/j.1365-2486.2008.01657.x](https://doi:10.1111/j.1365-2486.2008.01657.x)
- 396 Burrows, D., Butler, B., 2012. Primary studies of temperature regimes and temperature
397 tolerance of aquatic fauna in freshwater habitats of northern Australia. Australian Centre
398 of Tropical Freshwater Research (Report 12/08), James Cook University, Australia.
399 [https://research.jcu.edu.au/tropwater/resources/12%2008%20NAWFA%20Stream%20Te](https://research.jcu.edu.au/tropwater/resources/12%2008%20NAWFA%20Stream%20Temperature%20Project.pdf)
400 [mperature%20Project.pdf](https://research.jcu.edu.au/tropwater/resources/12%2008%20NAWFA%20Stream%20Temperature%20Project.pdf) (accessed March 2017)
- 401 Carveth, C.J., Widmer, A.M., Bonar, S.A., 2006. Comparison of upper thermal tolerances of
402 native and nonnative fish species in Arizona. *Transactions of the American Fisheries*
403 *Society* 135, 1433-1440. <https://doi.org/10.1577/T05-025.1>
- 404 Cheng, S.Y., Chen, C.S., Chen, J.C., 2013. Salinity and temperature tolerance of brown-marbled
405 grouper *Epinephelus fuscoguttatus*. *Fish Physiology and Biochemistry* 39, 277-286.
406 [https://doi: 10.1007/s10695-012-9698-x](https://doi:10.1007/s10695-012-9698-x)
- 407 Claassens, L., 2016. An artificial water body provides habitat for an endangered estuarine
408 seahorse species. *Estuarine, Coastal and Shelf Science* 180, 1-10.
409 <https://doi.org/10.1016/j.ecss.2016.06.011>
- 410 Connell, D., 1993. *Water Pollution : Causes and effects in Australia and New Zealand*. University
411 of Queensland Press, Brisbane.

- 412 Coulter, D.P., Sepúlveda, M.S., Troy, C.D., Höök, T.O., 2016. Species-specific effects of
413 subdaily temperature fluctuations on consumption, growth and stress responses in two
414 physiologically similar fish species. *Ecology of Freshwater Fish* 25, 465-475. [https://doi:
415 10.1111/eff.12227](https://doi.org/10.1111/eff.12227)
- 416 Dunn, R.J.K., Welsh, D.T., Jordan, M.A., Waltham, N.J., Lemckert, C.J., Teasdale, P.R., 2012.
417 Benthic metabolism and nitrogen dynamics in a sub-tropical coastal lagoon:
418 Microphytobenthos stimulate nitrification and nitrate reduction through photosynthetic
419 oxygen evolution. *Estuarine, Coastal and Shelf Science* 113, 272-282.
420 <https://doi.org/10.1016/j.ecss.2012.08.016>
- 421 Eaton, J.G., McCormick, J.H., Stefan, H.G., Hondzo, M., 1995. Extreme value analysis of fish
422 temperature field database. *Ecological Engineering* 4, 289-305.
423 [https://doi.org/10.1016/0925-8574\(95\)92708-R](https://doi.org/10.1016/0925-8574(95)92708-R)
- 424 Eaton, J.G., Scheller, R.M., 1996. Effects of climate warming on fish thermal habitat in streams
425 of the United States. *Limnology and Oceanography* 41, 1109-1115.
426 [https://doi: 10.4319/lo.1996.41.5.1109](https://doi.org/10.4319/lo.1996.41.5.1109)
- 427 Elliott, M., Whitfield, A.K., 2011. Challenging paradigms in estuarine ecology and management.
428 *Estuarine, Coastal and Shelf Science* 94, 306-314.
429 <https://doi.org/10.1016/j.ecss.2011.06.016>
- 430 Eme, J., Bennett, W.A., 2009. Critical thermal tolerance polygons of tropical marine fishes from
431 Sulawesi, Indonesia. *J. Therm. Biol.* 34, 220-225.
432 <https://doi.org/10.1016/j.jtherbio.2009.02.005>
- 433 Harvey, N., Stocker, L., 2015. Coastal residential waterways, science and policy-making: The
434 Australian experience. *Estuarine, Coastal and Shelf Science* 155, A1-A13.
435 <https://doi.org/10.1016/j.ecss.2014.12.019>

- 436 Heath, A.G., Turner, B.J., Davis, W.P., 1993. Temperature preferences and tolerances of three
437 fish species inhabiting hyperthermal ponds on mangrove islands. *Hydrobiologia* 259, 47-
438 55. [https://doi: 10.1007/BF00005964](https://doi.org/10.1007/BF00005964)
- 439 Hennessy, K., Webb, L., Ricketts, J., Macadam, I., 2008. *Climate Change Projections for the*
440 *Townsville Region*. CSIRO, Canberra, Australia.
- 441 James, C.S., Reside, A.E., VanDerWal, J., Pearson, R.G., Burrows, D., Capon, S.J., Harwood,
442 T.D., Hodgson, L., Waltham, N.J., 2017. Sink or swim? Potential for high faunal turnover
443 in Australian rivers under climate change. *J. Biogeogra.* 44, 489-501.
444 [https://doi: 10.1111/jbi.12926](https://doi.org/10.1111/jbi.12926)
- 445 Jeffrey, S.J., Carter, J.O., Moodie, K.B., Beswick, A.R., 2001. Using spatial interpolation to
446 construct a comprehensive archive of Australian climate data. *Environmental Modelling*
447 *and Software with Environment Data News* 16, 309-330. [https://doi.org/10.1016/S1364-](https://doi.org/10.1016/S1364-8152(01)00008-1)
448 [8152\(01\)00008-1](https://doi.org/10.1016/S1364-8152(01)00008-1)
- 449 Jobling, M., 1995. *Environmental Biology of Fishes*. Chapman and Hall, London, UK.
- 450 Lindall, W.N., Hall, R.J., Saloman, C.H., 1973. Fishes, macroinvertebrates and hydrological
451 conditions of upland canals in Tampa Bay, Florida. *Fishery Bulletin* 71, 155-163.
- 452 Manson, F.J., Loneragan, N.R., Harch, B.D., Skilleter, G.A., Williams, L., 2005. A broad-scale
453 analysis of links between coastal fisheries production and mangrove extent: A case study
454 for northeastern Australia. *Fisheries Research* 74, 69-85.
455 <https://doi.org/10.1016/j.fishres.2005.04.001>
- 456 Marshall, S., Elliott, M., 1998. Environmental influences on the fish assemblage of the Humber
457 Estuary, U. K. *Estuarine, Coastal and Shelf Science* 46, 175-184.
458 <https://doi.org/10.1006/ecss.1997.0268>
- 459 Martin, T., 1988. Interaction of salinity and temperature as a mechanism for spatial separation of
460 three co-existing species of Ambassidae (Cuvier)(Teleostei) in estuaries on the south-east
461 coast of Africa. *J. Fish Biol.* 33, 9-15. [https://doi: 10.1111/j.1095-8649.1988.tb05553.x](https://doi.org/10.1111/j.1095-8649.1988.tb05553.x)

- 462 Maxted, J.R., Eskin, R.A., Weisberg, S.B., Chaillou, J.C., Kutz, F.W., 1997. The ecological
463 condition of dead-end canals of the Delaware and Maryland coastal bays. *Estuaries* 20,
464 319-327. [https://doi: 10.2307/1352347](https://doi.org/10.2307/1352347)
- 465 McCauley, R., Casselman, J., 1981. The final preferendum as an index of the temperature
466 optimum for growth in Fish. *Proceedings of a World Symposium on Aquaculture, Heated*
467 *Effluents and Recirculating Systems* 2, 82-93.
- 468 McJannet, D., Marvanek, S., Kinsey-Henderson, A., Petheram, C., Wallace, J., 2014. Persistence
469 of in-stream waterholes in ephemeral rivers of tropical northern Australia and potential
470 impacts of climate change. *Mar. Freshwa. Res.* 65, 1131-1144.
471 <https://doi.org/10.1071/MF14035>
- 472 McJannet, D.L., Webster, I.T., Cook, F.J., 2012. An area-dependent wind function for estimating
473 open water evaporation using land-based meteorological data. *Environmental Modelling*
474 *and Software.* 31, 76-83. <https://doi.org/10.1016/j.envsoft.2011.11.017>
- 475 McJannet, D.L., Webster, I.T., Stenson, M.P., Sherman, B.S., 2008. Estimating open water
476 evaporation for the Murray Darling Basin. A report to the Australian Government from the
477 CSIRO Murray-Darling Basin Sustainable Yields Project, CSIRO, Australia.
- 478 Nagelkerken, I., Sheaves, M., Baker, R., Connolly, R.M., 2015. The seascape nursery: a novel
479 spatial approach to identify and manage nurseries for coastal marine fauna. *Fish and*
480 *Fisheries* 16, 362-371. [https://doi: 10.1111/faf.12057](https://doi.org/10.1111/faf.12057)
- 481 Nunn, A., Cowx, I., Frear, P., Harvey, J., 2003. Is water temperature an adequate predictor of
482 recruitment success in cyprinid fish populations in lowland rivers? *Freshwater Biology* 48,
483 579-588. [https://doi: 10.1046/j.1365-2427.2003.01033.x](https://doi.org/10.1046/j.1365-2427.2003.01033.x)
- 484 O'connell, M., Baldwin, D.S., Robertson, A., Rees, G., 2000. Release and bioavailability of
485 dissolved organic matter from floodplain litter: influence of origin and oxygen levels.
486 *Freshwater Biology* 45, 333-342. [https://doi: 10.1111/j.1365-2427.2000.00627.x](https://doi.org/10.1111/j.1365-2427.2000.00627.x)

- 487 Pearson, R.G., Crossland, M., Butler, B., Mansaring, S., 2003. Effects of cane-field drainage on
488 the ecology of tropical waterways. . Report on Sugar Research Development Corporation,
489 James Cook University, Townsville Australian Centre for Tropical Freshwater Research
490 (JCU016 & JCU024).
- 491 Rajaguru, S., Ramachandran, S., 2001. Temperature tolerance of some estuarine fishes. *Journal of*
492 *Thermal Biology* 26, 41-45. [https://doi.org/10.1016/S0306-4565\(00\)00024-3](https://doi.org/10.1016/S0306-4565(00)00024-3)
- 493 Robins, P.E., Skov, M.W., Lewis, M.J., Giménez, L., Davies, A.G., Malham, S.K., Neill, S.P.,
494 McDonald, J.E., Whitton, T.A., Jackson, S.E., Jago, C.F., 2016. Impact of climate change
495 on UK estuaries: A review of past trends and potential projections. *Estuarine, Coastal and*
496 *Shelf Science* 169, 119-135. <https://doi.org/10.1016/j.ecss.2015.12.016>
- 497 Rozas, L.P., 1992. Comparison of nekton habitats associated with pipeline canals and natural
498 channels in Louisiana salt marshes. *Wetlands* 12, 136-146. [https://doi:](https://doi.org/10.1007/BF03160594)
499 [10.1007/BF03160594](https://doi.org/10.1007/BF03160594)
- 500 Sheaves, M., Johnston, R., 2010. Implications of spatial variability of fish assemblages for
501 monitoring of Australia's tropical estuaries. *Aquatic Conservation: Marine and Freshwater*
502 *Ecosystems* 20, 348-356. [https://doi: 10.1002/aqc.1093](https://doi.org/10.1002/aqc.1093)
- 503 Sheaves, M., Johnston, R., Connolly, R.M., 2010. Temporal dynamics of fish assemblages of
504 natural and artificial tropical estuaries. *Marine Ecology Progress Series* 410, 143-157.
505 <https://doi.org/10.3354/meps08655>
- 506 Sheaves, M., Johnston, R., Connolly, R.M., 2012. Fish assemblages as indicators of estuary
507 ecosystem health. *Wetlands Ecology and Management* 20, 477-490. [https://doi:](https://doi.org/10.1007/s11273-012-9270-6)
508 [10.1007/s11273-012-9270-6](https://doi.org/10.1007/s11273-012-9270-6)
- 509 Sinokrot, B., Stefan, H., McCormick, J., Eaton, J., 1995. Modeling of climate change effects on
510 stream temperatures and fish habitats below dams and near groundwater inputs. *Climate*
511 *Change* 30, 181-200. [https://doi: 10.1007/BF01091841](https://doi.org/10.1007/BF01091841)

- 512 Stewart, B.A., Close, P.G., Cook, P.A., Davies, P.M., 2013. Upper thermal tolerances of key
513 taxonomic groups of stream invertebrates. *Hydrobiologia* 718, 131-140. [https://doi:](https://doi:10.1007/s10750-013-1611-9)
514 [10.1007/s10750-013-1611-9](https://doi:10.1007/s10750-013-1611-9)
- 515 Thomas, C.D., Cameron, A., Green, R.E., Bakkenes, M., Beaumont, L.J., Collingham, Y.C.,
516 Erasmus, B.F., De Siqueira, M.F., Grainger, A., Hannah, L., 2004. Extinction risk from
517 climate change. *Nature* 427, 145-148. <https://doi:10.1038/nature02121>
- 518 Wallace, J., Waltham, N., Burrows, D., 2017. A comparison of temperature regimes in dry season
519 waterholes in the Flinders and Gilbert catchments in northern Australia. *Mar.Freshwa.*
520 *Res.* 68, 650-667. <https://doi.org/10.1071/MF15468>
- 521 Wallace, J., Waltham, N.J., Burrows, D.W., McJannet, D., 2015. The temperature regimes of dry-
522 season waterholes in tropical northern Australia: potential effects on fish refugia.
523 *Freshwater Science* 34, 663-678. [https://doi: 10.1086/681278](https://doi:10.1086/681278).
- 524 Waltham, N., Davis, A., 2016. Fish survey Lilliesmere Creek: Pre-fish barrier construction.
525 TropWATER Report 16/73, James Cook University, Australia.
526 <https://research.jcu.edu.au/tropwater/publications/1573KalamiaCreekfishsurvey.pdf>
527 (accessed March 2017).
- 528 Waltham, N.J., Connolly, R.M., 2006. Trophic strategies of garfish, *Arrhamphus sclerolepis*, in
529 natural coastal wetlands and artificial urban waterways. *Mar. Bio.* 148, 1135-1141.
530 [https://doi: 10.1007/s00227-005-0154-7](https://doi:10.1007/s00227-005-0154-7)
- 531 Waltham, N.J., Connolly, R.M., 2007. Artificial waterway design affects fish assemblages in
532 urban estuaries. *J. Fish Biol.* 71, 1613-1629. [https:// doi: 10.1111/j.1095-](https://doi:10.1111/j.1095-8649.2007.01629.x)
533 [8649.2007.01629.x](https://doi:10.1111/j.1095-8649.2007.01629.x)
- 534 Waltham, N.J., Connolly, R.M., 2011. Global extent and distribution of artificial, residential
535 waterways in estuaries. *Estuarine, Coastal and Shelf Science* 94, 192-197.
536 <https://doi.org/10.1016/j.ecss.2011.06.003>

- 537 Waltham, N.J., Connolly, R.M., 2013. Artificial tidal lakes: Built for humans, home for fish.
538 Ecological Engineering 60, 414-420. [https://doi: 10.1086/681278](https://doi.org/10.1086/681278)
- 539 Waltham, N.J., Sheaves, M., 2015. Expanding coastal urban and industrial seascape in the Great
540 Barrier Reef World Heritage Area: Critical need for coordinated planning and policy.
541 Marine Policy 57, 78-84. [https://doi: 10.1086/681278](https://doi.org/10.1086/681278)
- 542 Welbergen, J.A., Klose, S.M., Markus, N., Eby, P., 2008. Climate change and the effects of
543 temperature extremes on Australian flying-foxes. Proceedings of the Royal Society of
544 London B: Biological Sciences 275, 419-425. [https://doi: 10.1098/rspb.2007.1385](https://doi.org/10.1098/rspb.2007.1385)
- 545 Wen, C.K., Pratchett, M.S., Shao, K.T., Kan, K.P., Chan, B.K., 2010. Effects of habitat
546 modification on coastal fish assemblages. J. Fish Biol. 77, 1674-1687. [https://doi:10.1111/j.1095-8649.2010.02809.x](https://doi.org/10.1111/j.1095-8649.2010.02809.x)
- 547
- 548 Whitfield, A.K., Blaber, S.J.M., Cyrus, D.P., 1981. Salinity ranges of some southern African fish
549 species occurring in estuaries. South African Journal of Zoology 16, 151-155.
- 550 Zigic, S., King, B.A., Lemckert, C., 2002. Mixing between two canals connected by an automated
551 bi-directional gated structure, Gold Coast, Australia. Estuarine, Coastal and Shelf Science
552 55, 59-66. <https://doi.org/10.1006/ecss.2001.0886>
- 553
- 554

Table 1. Temperature tolerance experiments for each species examined. Summary statistics provided demonstrating the range in AET. Current and future climate threshold exceedance (%) for both surface and bottom logger data.

Family/species	Number	Size range (TL, mm)	Statistic	28(°C) acclimation experiment	Current climate threshold exceedance (%)		Future climate threshold exceedance (%)	
					Surface	Bottom	Surface	Bottom
Ambassidae								
<i>Ambassis vachellii</i>	61	27-47	Lowest observed AET	35.10	26	47	3	15
			AET ₁₀	37.30				
			AET ₅₀	38.70	5	14	0	0
			AET ₉₀	39.60				
			Highest observed AET	39.90	1	3	0	0
Leiognathidae								
<i>Leiognathus equulus</i>	29	21-47	Lowest observed AET	34.90	26	47	3	15
			AET ₁₀	36.30				
			AET ₅₀	37.05	11	21	0	1
			AET ₉₀	37.39				
			Highest observed AET	37.90	5	14	0	0
Mugilidae								
<i>Chelon subviridis</i>	8	48-189	Lowest observed AET	37.30	11	21	0	1
			AET ₁₀	37.58				
			AET ₅₀	39.00	2	6	0	0
			AET ₉₀	39.50				
			Highest observed AET	39.50	1	3	0	0
<i>Ellochelon viagiensis</i>	21	50-82	Lowest observed AET	38.90	2	6	0	0
			AET ₁₀	38.90				
			AET ₅₀	39.50	1	3	0	0
			AET ₉₀	39.50				
			Highest observed AET	39.80	<1	3	0	0
Silliginidae								
<i>Sillago analis</i>	12	65-145	Lowest observed AET	33.10	6	16	28	54
			AET ₁₀	37.20				
			AET ₅₀	38.50	5	6	0	0
			AET ₉₀	39.80				
			Highest observed AET	39.80	<1	3	0	0

1 **List of figures**

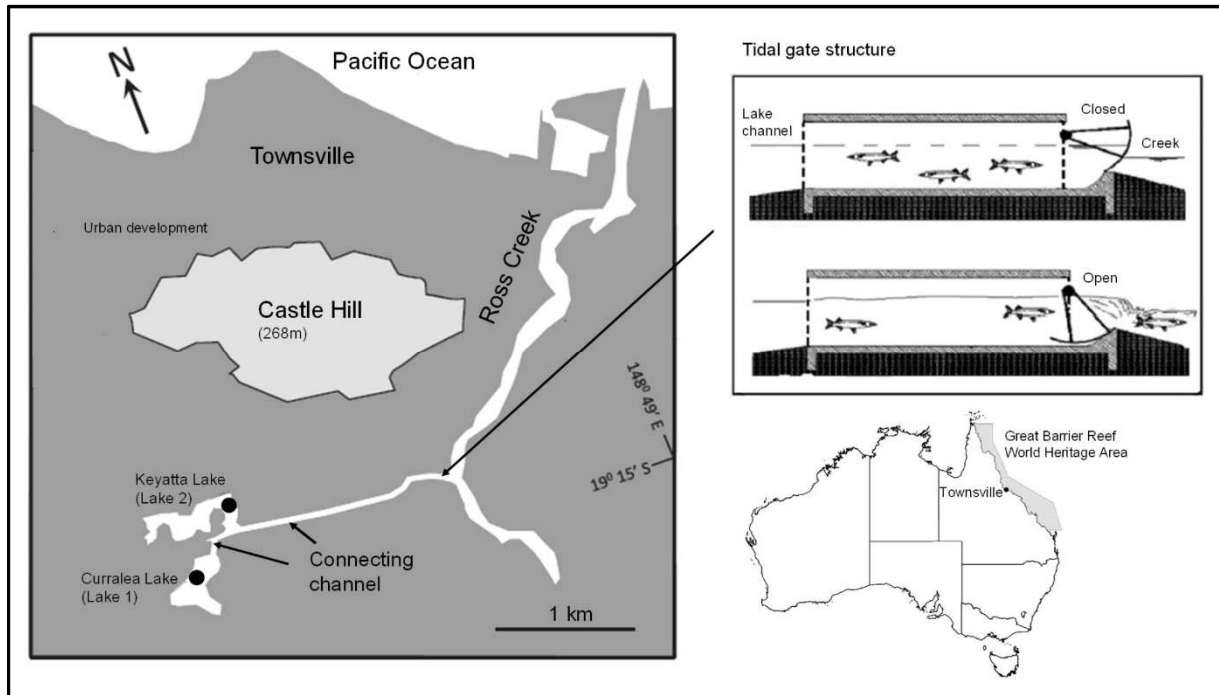
2 Figure 1. A) Curralea Lake (Lake 1) and Keyatta Lake (Lake 2), and connecting channels
3 adjacent to Ross Creek, Townsville, Australia. Dark grey fill indicates urban, industrial or
4 commercial areas. B) Photo illustrating limited riparian shading provided around these lakes.

5
6 Figure 2. High frequency of water temperature recorded at the surface (~0.2m; black) and
7 bottom (~2.4m; grey) in Lake 1. A) full data set; and B) subset of the logging data from (A),
8 along with rainfall data for this logging period.

9
10 Figure 3. The percentage of time (based on 20 min data between 31 October 2015 and 31
11 March 2016) water temperature in Lake 1. In both graphs, black curve line is measured water
12 temperature for the current survey period, broken black curve line is the future modelled
13 climate change data for the same time period. (a) lake near surface; and b) lake bottom
14 waters. Threshold lines present minimum *Ambassis vachellii* AET (black line), and
15 maximum AET (grey line measured temperature (not shown given <1%; Table 1)).

16
17

18 A)



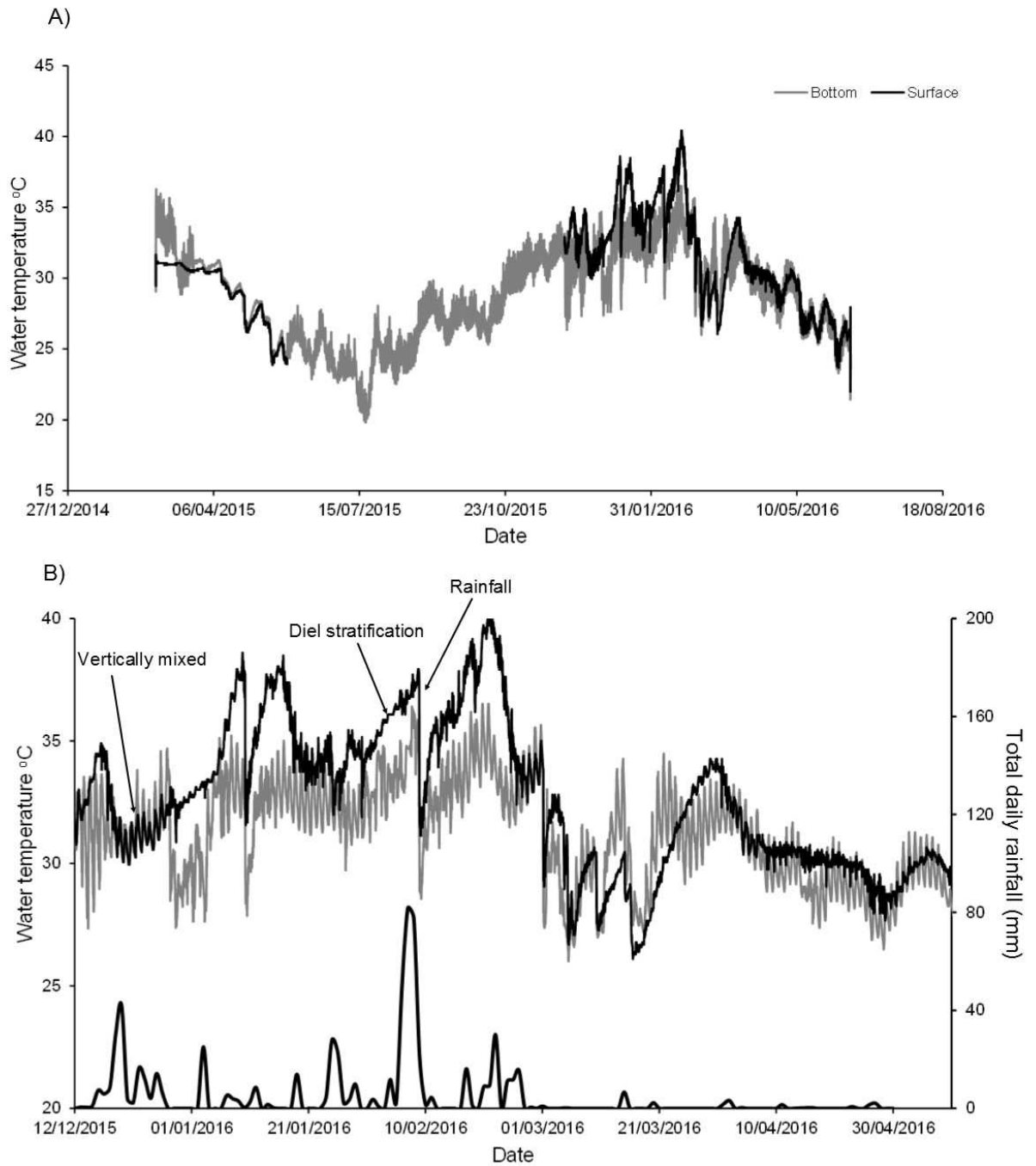
19

20 B)



21

22 Fig 1.

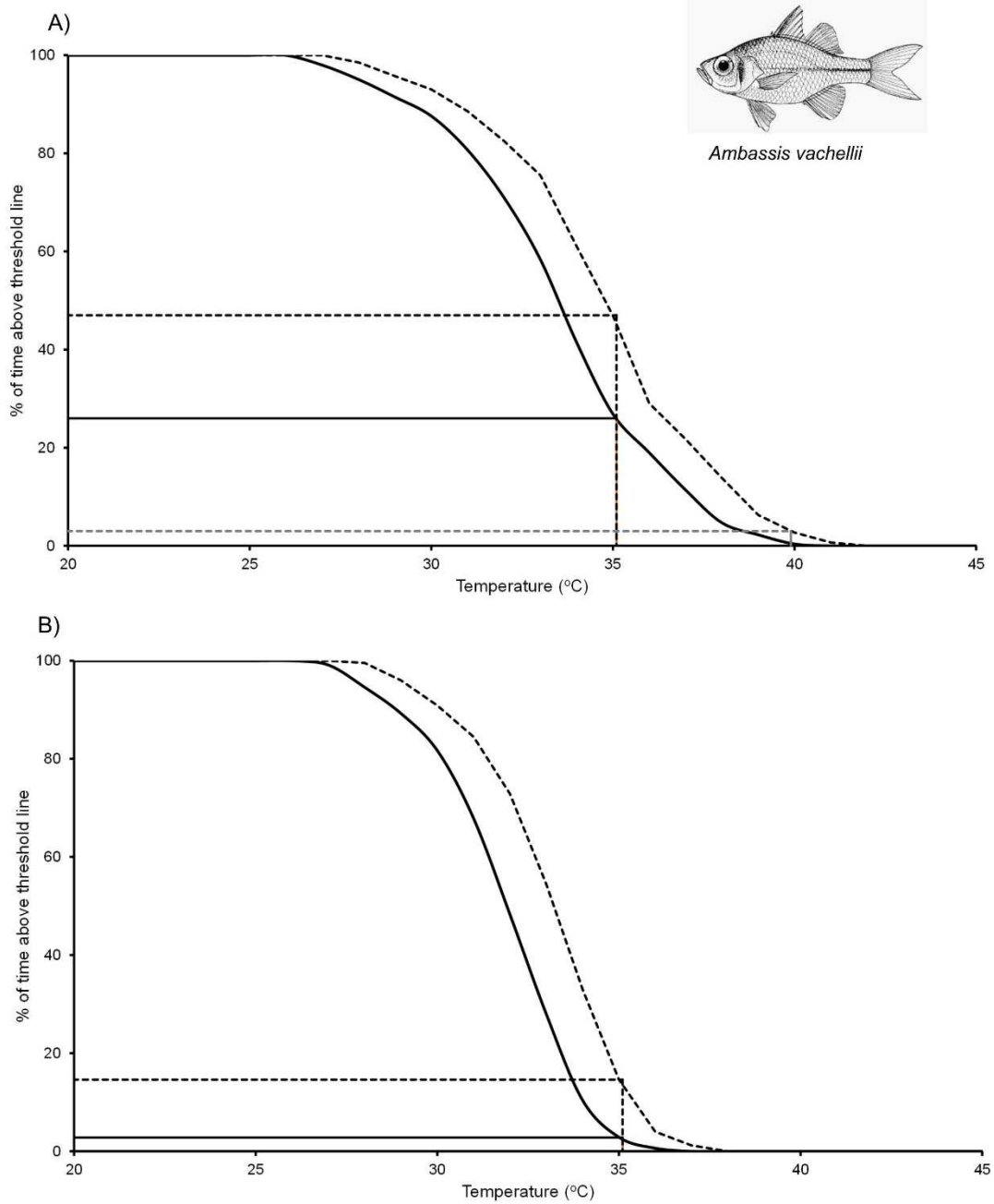


23

24

25 Fig 2.

26



27

28 Fig 3.

29