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## **How can an understanding of plant-pollinator interactions contribute to global food security?**

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## **Abstract**

Pollination of crops by animals is an essential part of global food production, but evidence suggests that wild pollinator populations may be declining while a number of problems are besetting managed honey bee colonies. Animal-pollinated crops grown today, bred in an environment where pollination was less likely to limit fruit set, are often suboptimal in attracting and sustaining their pollinator populations. Research into plant-pollinator interactions is often conducted in a curiosity-driven, ecological framework, but may inform breeding and biotechnological approaches to enhance pollinator attraction and crop yield. In this article we review key topics in current plant-pollinator research that have potential roles in future crop breeding for enhanced global food security.

## 1. Introduction

Pollination of crops by insects and vertebrates is an essential part of global food production. Recent reports have estimated that 35% of global crop yield depends on the activity of animal pollinators [1]. Yet, at the same time, evidence has started to accumulate that wild pollinator populations may be declining, while a number of different biotic and abiotic problems are besetting managed honey bee colonies [2]. In light of these two statements it is clear that pollination biology has a large, and increasingly urgent, role to play in protecting and maintaining global food supply. Crop breeding programmes select for agronomic traits such as yield or disease resistance [3]. Therefore, it is highly likely that insect-pollinated crops grown today, bred in an environment where pollinators were more abundant and pollination was less likely to limit fruit set, are suboptimal in terms of their floral traits with respect to attracting and sustaining their pollinator populations. Our aim in this article is to review a range of aspects of plant-pollinator research that have potential utility for crop breeders, and to highlight areas of the field where increased research effort might yield important agronomic improvements.

## 2. Pollinators and crop production

While wind-pollinated cereals still account for the majority of calories produced worldwide for human consumption, crop dependence on animal pollinators is increasing, and at a greater rate than honey bee colony production [4]. Around 75 % of the 115 highest producing crops worldwide benefit from greater yields when pollinated by animals, mainly bees [1] (Figure 1), with brazil nut, watermelon, cantaloupe, *Cucurbita* spp., cocoa, and vanilla the most pollinator-dependent of these [5]. Pollinators can also improve the quality of crops [6] and the nutritional value of pollinator dependent crops is perhaps even more important than their economic value. Pollinator-dependent crop production of vital micronutrients accounts for a large proportion of total production in many areas; for example the reliance on pollinators for vitamin A is close to 50% in Thailand [7].

Research over the past few decades has documented declines in both wild pollinator populations and the abundance and health of managed honey bee hives. These two issues are largely separate, though have become conflated in the popular media [8]. “Colony collapse disorder”, parasites such as varroa, and, more recently, neonicotinoid pesticides have all been implicated in the reduction of the number of honey bee hives at a regional level, particularly in Europe and North America [9-11]. In contrast, the loss of diversity and abundance of wild pollinators such as social bumblebees, solitary bees, hoverflies, wasps and butterflies has mainly been attributed to changes in land use and reduction in the area

of habitat available for foraging and life cycle completion, [12-14] a process, which in the UK at least, has deep historical roots [15\*\*].

The relative importance of honey bees and wild pollinators for agriculture depends upon both the crop and local management practices; in the UK and parts of the USA it is thought that wild pollinators are responsible for the large majority of crop pollination [16,17]. Wild bees have also been shown to improve fruit set in a wide variety of crop systems, independent of the abundance of honey bees [18\*\*]. This implies that future habitat conservation efforts, particularly at a landscape scale, could provide both immediate and long-term benefits to agriculture in the form of locally enhanced crop production and insurance against fluctuations in wild pollinator abundance. Examples of such efforts are enhanced agri-environmental schemes on farms and appropriate restoration of post-industrial areas, including landfill sites [19,20]. There is some evidence that conservation programmes in northern Europe may have slowed the rate of decline of some pollinators, but more research is required to substantiate this trend [15\*\*,21\*\*].

### **3. Plant-pollinator interaction research with implications for food security**

#### **a. Crops as a resource for pollinators**

Insect-pollinated crops can provide a substantial nutritional resource to pollinator communities, which may help maintain their populations over generations. Mass flowering crops such as oilseed rape/canola (*Brassica napus*) are of particular interest in this respect as they provide a bonanza of food for pollinators [22]. However, because this reward is transient, the benefits to pollinators have been found to be species specific and dependent on timing [23-26\*]. Phenological matching of crop flowering and pollinator activity could result in a two-fold benefit, improving the reproductive potential of pollinators and promoting more efficient pollination of crops both in current and subsequent years.

In the example of oilseed rape, key lifecycle stages of the red mason bee *Osmia bicornis* (a solitary bee) occur concurrently with flowering, leading to increased reproductive success for the bees [25,26\*]. Contrastingly, for bumblebees (*Bombus* spp.), while the temporary increase in resources that early flowering oilseed rape provides may help colony founding [22,24,27], a lack of food later in the season can lead to this not translating into an increase in the number of queens and males [24]. This can be resolved by complementing early flowering crops with late flowering crops in adjacent fields, or strips of semi-natural flower-rich vegetation, ensuring a stable foraging supply over a longer period [28,29]. An alternative approach would be to coordinate crop flowering with key life cycle stages for pollinators, by breeding for different flowering times as appropriate. In crops such as oilseed rape and the

field bean (*Vicia faba*) QTLs for both flowering time and duration have been identified and the molecular basis of these traits is well defined [30-32].

### **b. Exploiting multiple pollinators**

The ability of flowering plants to utilize multiple pollinators can be beneficial in ensuring fruit set. The majority of flowering plants engage in flexible relationships with their pollinators that are variable within populations, and over both time and space [33]. This variability can be at the level of species within broad taxonomic/functional groups (e.g. use of different large bee species) or use of pollinators from very different groups (e.g. bees and birds, or butterflies and bees). Much of this flexibility depends upon the local ecological context in which a plant finds itself [33-35].

This also applies to most crop plants, where the effective pollinators will be determined by the plant's floral biology and the locality, season and year in which it is planted. For example, in manipulative caged plant experiments it was shown that various bumblebees are the main pollinators of field beans, with honey bees (*Apis mellifera*) and red mason bees playing a secondary role; in contrast the same groups plus hoverflies (*Episyrphus balteatus*) are equally good at pollinating oilseed rape [36]. Under field conditions, the importance of these different groups will depend upon their visitation rate to flowers, which is largely determined by their abundance, which will fluctuate over time and space. These findings probably apply to the majority of crops grown worldwide as most are relatively generalised in their interactions with local and managed pollinators.

Looking at which floral traits change in response to different groups of pollinators over the course of evolution may provide insights into which traits would be useful to target in breeding programmes to improve pollinator visitation. If a floral trait changes repeatedly in a phylogenetic context it is available for selection to target. However, if a floral trait is fixed, either no genetic variability is present, or else the trait is constrained by its developmental or functional basis. For example, flower colour is a trait with astonishing lability over evolutionary time and the potential to discriminate between pollinator types. Multiple evolutionary shifts from hummingbird to moth pollination in *Aquilegia* have been associated with independent losses of anthocyanin production [37]. The molecular basis of many such shifts is attributable to changes in the transcription factors that regulate the pigment synthetic pathway (reviewed by [38]), and similar changes can cause variations in the pattern of floral pigment, generating spots, veins and other nectar guides that are differentially used by different pollinators [39,40\*,41]. However changing a single trait such as flower colour alone is unlikely to affect the type and diversity of pollinators to a crop

because it must be backed up by a change in the quality or quantity of the reward being offered, and its accessibility to particular pollinators.

### **c. Attracting more of whatever pollinators there are**

From the pollinator's perspective, it may be a floral trait's utility as an indicator of reward rather than its inherent attractiveness that is most important (e.g. [42,43\*\*]). The importance of honest signals for foraging decisions of bees was recently demonstrated by Knauer and Schiestl [43\*\*] who showed that bumblebees specifically selected for a volatile signal which correlated with floral reward in *Brassica rapa*. Floral scent may be particularly useful for pollinators as an honest trait [44], although the most honest traits are those physically linked to the reward itself [45].

In order to successfully utilise floral traits for crop improvement, it is important to understand how they attract pollinators. Recent experiments with volatile cues have highlighted the complexity of their function. Experimentally augmenting floral scent increased pollinator attraction for *Hesperis matronalis* [46], but only with specific components of its floral bouquet. In contrast, a similar experiment with *Curcubita pepo* had no effect on pollinators but increased visits from florivores [47]. Similar dynamics can occur with visual cues [48,49] for example the showy bracts of *Dalechampia scandens* are attractive to both pollinators and seed predators [48]. Balancing these potential costs of trade-offs from traits with multiple effects will be necessary for selecting appropriate crop breeding strategies [49].

### **d. Manipulating floral reward**

Since pollinators often visit flowers to harvest nectar and pollen, breeding programmes that improve reward production, nutritive value or availability perhaps have the most potential to enhance pollination and fruit set of crops reliant on both wild and managed pollinator populations. The mean reward value of nectar encountered by a pollinator has been shown to be positively correlated with the proportion of flowers visited on a plant, the time spent on a flower, flower constancy, and the number of unrewarding flowers that will be visited before moving to a new patch [50-52]. These factors should enhance overall pollination rates of a crop but may also increase the proportion of self pollen received [52]. For crop systems such as the field bean, which benefit from both self and non-self pollen [53], this will not necessarily have a negative impact, but for others the optimum size of the reward to deliver maximum yields may need to be more carefully assessed.

Nectar is rarely just a simple sugar solution. Beside sugars, nectar can contain amino acids, nectarin proteins, volatile compounds, minerals, and secondary metabolites such as

phenolics and alkaloids [54,55\*\*,56 and references within]. The additives to the nectar listed above may have nutritive value, but some also have the potential to modify pollinator behaviour, or defend against microbes and nectar thieves. For example, the amino acids proline and phenylalanine act as phagostimulants, causing insects to visit more flowers than they would usually. Other compounds such as non-protein amino acids (reviewed by [57]) and caffeine [55\*\*] also have the capability to increase the number of visits a flower receives.

Recent studies have identified transcription factors from the MYB family as regulators of nectar production in *Arabidopsis* (*AtMYB57*) and *Nicotiana* (*NtMYB305*) [58,59]. MYB305 has been shown to regulate nectarin production through *NECTARIN* genes [60] and starch accumulation during nectary development and thus the amount of nectar secreted [59]. Nectar secretion is also under the control of plant growth regulators jasmonic acid and auxin [58,61,62]. Downstream, the sugar transporter *AtSWEET9* is involved in the export of sugars into the nectary [63\*\*]. This export is coupled with the hydrolysis of sucrose into glucose and fructose by *CELL WALL INVERTASE 4*, providing a sink for sucrose export and allowing sugars to accumulate [64]. One area worthy of more attention is how the ratio of sucrose:hexose in nectar is controlled and what consequences this has for the attractiveness of a crop to different pollinators.

For pollinators that obtain their nutrition solely from flowers, pollen is a vital source of nitrogen from which to rear offspring. As well as proteins and amino acids, pollen contains lipids, vitamins, and sometimes starch [65,66]. As the chemical composition of pollen varies between plant species [66], its nutritional value to pollinators will also differ. Experienced pollen foragers are able to determine the nutritional quality of pollen and preferentially choose those containing a greater content of essential amino acids leucine, isoleucine and valine [66,67]. Therefore, a potential route to increase pollinator visitation to crops could be to manipulate pollen quantity by altering the expression of pollen-specific nutrient transporters, such as amino acid transporters (eg. *AtLHT* genes, *NsAAP1*, *LeProT1*; reviewed by [68]) which have been identified in *Arabidopsis*, tobacco and tomato.

Ultimately, when looking to increase crop yields through the enhancement of floral yields, there is still a lot of research on the genetic basis of the quantity and quality of floral reward produced to be done, which will allow a more targeted approach to crop breeding. Nectar is a much less costly reward for plants to produce in terms of nitrogen (and also in terms of gametes not available for reproduction) and may therefore hold most promise for improving pollinator visitation. However, in the context of trying to support pollinator populations by



providing resources for reproduction it will also be important to consider whether crops provide a suitable pollen source to allow reproduction to occur.

#### **e. Optimising efficiency of reward acquisition**

The energetic reward provided by a plant can also be increased by reducing the costs incurred by a pollinator in obtaining it. The symmetry of a flower influences how easily a pollinator can acquire the reward by specifying the presence and position of landing platforms and occluded palates that require force to open. In *Antirrhinum majus* a transcriptional network comprising TCP and MYB family transcription factors has been shown to specify zygomorphy (bilateral symmetry) [69-72]. The mutants generated during these studies have provided insight into how pollinators forage to optimise their energy return. In wild type flowers bumblebees alight on the landing platform and generate sufficient force to open the palate and access the nectar, whereas other insects cannot; for example honey bees, which can only exert around a tenth of the force of bumblebees [73]. Mutants with all their petals ventralised are closed and inaccessible to pollinators (BJG, unpublished data). In contrast, mutants with all their petals dorsalised are open and readily accessible to a range of pollinators, including honey bees, but experience reduced visits by long-tongued bumblebees, likely as a result of nectar depletion by other foragers [74]. These extreme variants hint at the possibility of optimising floral symmetry and pollinator visitation using less extreme natural variation. In alfalfa it has been shown that the force required to open a flower is heritable and an easier to open flower is correlated with a greater rate of flower opening under field conditions [75].

At a finer scale, the shape of epidermal cells on the flower surface can influence the energetic efficiency of foraging. Whitney et al. [76] showed that *Bombus terrestris* foragers prefer flowers with conical epidermal cells to those with flat epidermal cells, but only when flowers were oriented vertically. Similarly, Alcorn et al. [77] found that flowers with conical epidermal cells were preferred when flowers were moving, but not when they were stationary. Conical epidermal cells enhance foraging efficiency by providing a surface into which the tarsal claws of bees can grip. Variation has been found within and between species in the extent and distribution of conical epidermal cells ([78]; EJB and BJG, unpublished), suggesting that this trait has potential as a breeding target for optimised pollinator attraction.

## **4. Conclusions**

Although the various traits discussed above, and summarized in Figure 2, have potential as targets of crop breeding, there are surprisingly few published examples of research

specifically analyzing or targeting yield through animal pollination. Perhaps the most extensive studies have been by Suso and colleagues, who have investigated how a number of floral traits affect outcrossing of the field bean (*Vicia faba*) [eg. 79,80\*]. Traits such as standard petal dimensions, number of inflorescences, ovary length, and pollen content are all associated with changes in outcrossing level and by extension with animal pollination.

Much more work is needed to define how pollination biology can contribute to crop production, and it will also be important to consider the interaction between plant and pollinator in the wider context of the many other factors influencing pollinator numbers. However, as substantial variation appears to be present in a wide range of crops and floral traits [54,80\*,81] - from the level of floral display right down to the size of reproductive structures – we are confident that there is great potential for additional research in this area to inform the breeding of higher yielding, more insect-friendly crops.

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## Figure Legends

**Figure 1. Crops and their pollinators.** The pollinator shown is not necessarily the principal pollinator. **A** Sunflower with *Aglais io*. **B** Radish with beetle and fly pollinators. **C** Blueberry with *Bombus ternarius*. **D** Tomato with *Bombus terrestris*. **E** Apple with *Apis mellifera*. **F** Field bean with *Bombus hortorum*. **G** Coffee with *Apis mellifera*. **For image credits please refer to the Acknowledgements.**

**Figure 2. Floral traits affecting pollinator visitation.** The floral traits highlighted at the level of the crop, plant and flower may make useful targets for crop breeding programmes to enhance pollination rates.

## REFERENCES

1. Klein AM, Vaissiere BE, Cane JH, Steffan-Dewenter I, Cunningham SA, Kremen C, Tscharntke T: **Importance of pollinators in changing landscapes for world crops.** *Proc Biol Sci* 2007, **274**:303-313.
2. Vanbergen AJ, Baude M, Biesmeijer JC, Britton NF, Brown MJF, Brown M, Bryden J, Budge GE, Bull JC, Carvell C, Challinor AJ, Connolly CN, Evans DJ, Feil EJ, Garratt MP, Greco MK, Heard MS, Jansen VAA, Keeling MJ, Kunin WE, Marris GC, Memmott J, Murray JT, Nicolson SW, Osborne JL, Paxton RJ, Pirk CWW, Polce C, Potts SG, Priest NK, Raine NE, Roberts S, Ryabov EV, Shafir S, Shirley MDF, Simpson SJ, Stevenson PC, Stone GN, Termansen M, Wright GA **Threats to an ecosystem service: pressures on pollinators.** *Front Ecol Environ* 2013, **11**:251-259
3. Kobayashi K, Tsukamoto S, Tanaka A, Niikura S, Ohsawa R: **Selective flower visitation behavior by pollinators in a radish F1 seed production field.** *Breed Sci* 2010, **60**:203-211.
4. Aizen MA, Harder LD: **The global stock of domesticated honey bees is growing slower than agricultural demand for pollination.** *Curr Biol* 2009, **19**:915-918.
5. Garibaldi LA, Aizen MA, Klein AM, Cunningham SA, Harder LD: **Global growth and stability of agricultural yield decrease with pollinator dependence.** *Proc Natl Acad Sci U S A* 2011, **108**:5909-5914.
6. Klatt BK, Holzschuh A, Westphal C, Clough Y, Smit I, Pawelzik E, Tscharntke T: **Bee pollination improves crop quality, shelf life and commercial value.** *Proc Biol Sci* 2014, **281**:20132440.
7. Chaplin-Kramer R, Dombeck E, Gerber J, Knuth KA, Mueller ND, Mueller M, Ziv G, Klein AM: **Global malnutrition overlaps with pollinator-dependent micronutrient production.** *Proc Biol Sci* 2014, **281**:20141799.
8. Ollerton J, Price V, Armbruster WS, Memmott J, Watts S, Waser NM, Totland Ø, Goulson D, Alarcon R, Stout JC, et al.: **Overplaying the role of honey bees as pollinators: a comment on Aebi and Neumann (2011).** *Trends Ecol Evol* 2012, **27**:141-142.
9. Potts S, Roberts S, Dean R, Marris G, Brown M, Jones R, Neumann P, Settele J: **Declines of managed honey bees and beekeepers in Europe.** *J Apic Res* 2010, **49**:15.
10. vanEngelsdorp D, Hayes J, Underwood RM, Caron D, Pettis J: **A survey of managed honey bee colony losses in the USA, fall 2009 to winter 2010.** *J Apic Res* 2011, **50**:1-10.
11. Godfray HCJ, Blacquière T, Field LM, Hails RS, Petrokofsky G, Potts SG, Raine NE, Vanbergen AJ, McLean AR: **A restatement of the natural science evidence base concerning neonicotinoid insecticides and insect pollinators.** *Proc R Soc B* 2014, **281**:20140558.

12. Biesmeijer JC, Roberts SP, Reemer M, Ohlemuller R, Edwards M, Peeters T, Schaffers AP, Potts SG, Kleukers R, Thomas CD, et al.: **Parallel declines in pollinators and insect-pollinated plants in Britain and the Netherlands.** *Science* 2006, **313**:351-354.

13. Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE: **Global pollinator declines: trends, impacts and drivers.** *Trends Ecol Evol* 2010, **25**:345-353.

14. Goulson D, Lye GC, Darvill B: **Decline and conservation of bumble bees.** *Ann Rev Entomol* 2008, **53**:191-208.

\*\*15. Ollerton J, Erenler H, Edwards M, Crockett R: **Extinctions of aculeate pollinators in Britain and the role of large-scale agricultural changes.** *Science* 2014, **346**:1360-1362.

The first analysis of rates of extinction of pollinators for any country. The authors showed that the peak period of extinction began much earlier than previously believed, in the decade following the First World War, and continued until the late 1950s.

16. Breeze TD, Bailey AP, Balcombe KG, Potts SG: **Pollination services in the UK: How important are honeybees?** *Agric Ecosyst Environ* 2011, **142**:137-143.

17. Winfree R, Williams NM, Gaines H, Ascher JS, Kremen C: **Wild bee pollinators provide the majority of crop visitation across land-use gradients in New Jersey and Pennsylvania, USA.** *J Appl Ecol* 2008, **45**:793-802.

\*\*18. Garibaldi LA, Steffan-Dewenter I, Winfree R, Aizen MA, Bommarco R, Cunningham SA, Kremen C, Carvalheiro LG, Harder LD, Afik O, Bartomeus I, Benjamin F, Boreux V, Cariveau D, Chacoff NP, Dudenhöffer JH, Freitas BM, Ghazoul J, Greenleaf S, Hipólito J, Holzschuh A, Howlett B, Isaacs R, Javorek SK, Kennedy CM, Krewenka KM, Krishnan S, Mandelik Y, Mayfield MM, Motzke I, Munyuli T, Nault BA, Otieno M, Petersen J, Pisanty G, Potts SG, Rader R, Ricketts TH, Rundlöf M, Seymour CL, Schüepp C, Szentgyörgyi H, Taki H, Tscharrntke T, Vergara CH, Viana BF, Wanger TC, Westphal C, Williams N, Klein AM: **Wild Pollinators Enhance Fruit Set of Crops Regardless of Honey Bee Abundance** *Science* 2013, **339**:1608-1611.

These authors collected data from across the globe for 41 pollinator dependent crop systems. They showed that whilst honeybees have a greater positive affect on pollen deposition, wild bees increase fruit set independent of the number of honeybee visits, suggesting honeybees are not a substitute for wild pollinators.

19. Dicks LV, Showler DA, Sutherland WJ, Dicks LV, Showler DA, Sutherland WJ: *Bee conservation. Evidence for the effects of interventions:* Pelagic Publishing; 2010.

20. Tarrant S, Ollerton J, Rahman ML, Tarrant J, McCollin D: **Grassland Restoration on Landfill Sites in the East Midlands, United Kingdom: An Evaluation of Floral Resources and Pollinating Insects.** *Restor Ecol* 2013, **21**:560-568.

\*\*21. Carvalheiro LG, Kunin WE, Keil P, Aguirre-Gutiérrez J, Ellis WN, Fox R, Groom Q, Hennekens S, Van Landuyt W, Maes D, et al.: **Species richness declines and biotic homogenisation have slowed down for NW-European pollinators and plants.** *Ecol Lett* 2013, **16**:870-878.

The authors used long-term data sets to show that there may be grounds for optimism that conservation efforts in recent decades may have succeeded in reducing the rate of decline of pollinating insects and plants.

22. Westphal C, Steffan-Dewenter I, Tschardt T: **Mass flowering crops enhance pollinator densities at a landscape scale.** *Ecol Lett* 2003, **6**:961-965.
23. Diekötter T, Kadoya T, Peter F, Wolters V, Jauker F: **Oilseed rape crops distort plant-pollinator interactions.** *J Appl Ecol* 2010, **47**:209-214.
24. Westphal C, Steffan-Dewenter I, Tschardt T: **Mass flowering oilseed rape improves early colony growth but not sexual reproduction of bumblebees.** *J Appl Ecol* 2009, **46**:187-193.
25. Jauker F, Peter F, Wolters V, Diekötter T: **Early reproductive benefits of mass-flowering crops to the solitary bee *Osmia rufa* outbalance post-flowering disadvantages.** *Basic Appl Ecol* 2012, **13**:268-276.
- \*26. Holzschuh A, Dormann CF, Tschardt T, Steffan-Dewenter I: **Mass-flowering crops enhance wild bee abundance.** *Oecologia* 2013, **172**:477-484.

The effect of oilseed rape (*Brassica napus*) fields and semi-natural grassland on reproduction of the solitary bee *Osmia bicornis* was assessed using trap nests. Although nest colonisation and number of brood cells was reliant on the presence of grasslands, both these measures significantly increased if oilseed rape fields were also present.

27. Herrmann F, Westphal C, Moritz RF, Steffan-Dewenter I: **Genetic diversity and mass resources promote colony size and forager densities of a social bee (*Bombus pascuorum*) in agricultural landscapes.** *Mol Ecol* 2007, **16**:1167-1178.
28. Riedinger V, Renner M, Rundlöf M, Steffan-Dewenter I, Holzschuh A: **Early mass-flowering crops mitigate pollinator dilution in late-flowering crops.** *Landsc Ecol* 2014, **29**:425-435.
29. Rundlöf M, Persson AS, Smith HG, Bommarco R: **Late-season mass-flowering red clover increases bumble bee queen and male densities.** *Biol Conserv* 2014, **172**:138-145.
30. Cruz-Izquierdo S, Avila CM, Satovic Z, Palomino C, Gutierrez N, Ellwood SR, Phan HT, Cubero JI, Torres AM: **Comparative genomics to bridge *Vicia faba* with model and closely-related legume species: stability of QTLs for flowering and yield-related traits.** *Theor Appl Genet* 2012, **125**:1767-1782.
31. Chen G, Geng J, Rahman M, Liu X, Tu J, Fu T, Li G, McVetty PBE, Tahir M: **Identification of QTL for oil content, seed yield, and flowering time in oilseed rape (*Brassica napus*).** *Euphytica* 2010, **175**:161-174.
32. Wang N, Qian W, Suppanz I, Wei L, Mao B, Long Y, Meng J, Muller AE, Jung C: **Flowering time variation in oilseed rape (*Brassica napus* L.) is associated with allelic variation in the *FRIGIDA* homologue *BnaA.FRI.a*.** *J Exp Bot* 2011, **62**:5641-5658.
33. Waser NM, Chittka L, Price MV, Williams NM, Ollerton J: **Generalization in pollination systems, and why it matters.** *Ecology* 1996, **77**:1043-1060.
34. Fenster CB, Armbruster WS, Wilson P, Dudash MR, Thomson JD: **Pollination syndromes and floral specialization.** *Ann Rev Ecol Evol Syst* 2004, **35**:375-403.
35. Ollerton J, Killick A, Lamborn E, Watts S, Whiston M: **Multiple meanings and modes: on the many ways to be a generalist flower.** *Taxon* 2007, **56**:717-728.

36. Garratt MPD, Coston DJ, Truslove CL, Lappage MG, Polce C, Dean R, Biesmeijer JC, Potts SG: **The identity of crop pollinators helps target conservation for improved ecosystem services.** *Biol Conserv* 2014, **169**:128-135.
37. Hodges SA, Derieg NJ: **Adaptive radiations: From field to genomic studies.** *Proc Natl Acad Sci U S A* 2009, **106**:9947-9954.
38. Wessinger CA, Rausher MD: **Lessons from flower colour evolution on targets of selection.** *J Exp Bot* 2012, **63**:5741-5749.
39. Shang YJ, Venail J, Mackay S, Bailey PC, Schwinn KE, Jameson PE, Martin CR, Davies KM: **The molecular basis for venation patterning of pigmentation and its effect on pollinator attraction in flowers of *Antirrhinum*.** *New Phytol* 2011, **189**:602-615.
- \*40. Whitney HM, Milne G, Rands SA, Vignolini S, Martin C, Glover BJ: **The influence of pigmentation patterning on bumblebee foraging from flowers of *Antirrhinum majus*.** *Naturwissenschaften* 2013, **100**:249-256.

This paper demonstrates that venation patterning is not innately attractive to naïve bees, suggesting that previous studies demonstrating a field preference of bees for venation patterning indicates that bees can learn to use coloured veins as nectar guides.

41. Martins TR, Berg JJ, Blinka S, Rausher MD, Baum DA: **Precise spatio-temporal regulation of the anthocyanin biosynthetic pathway leads to petal spot formation in *Clarkia gracilis* (Onagraceae).** *New Phytol* 2013, **197**:958-969.
42. Katzenberger TD, Lunau K, Junker RR: **Salience of multimodal flower cues manipulates initial responses and facilitates learning performance of bumblebees.** *Behav Ecol and Sociobiol* 2013, **67**:1587-1599.
- \*\*43. Knauer AC, Schiestl FP: **Bees use honest floral signals as indicators of reward when visiting flowers.** *Ecol Lett* 2015, **18**:135-143.

This study demonstrates the potential importance of honest signals to pollinators and considers various mechanisms under which honest signalling could evolve. Bumblebees specifically selected for an honest volatile signal from *Brassica rapa*. When the identity of honest and dishonest volatile signals was artificially manipulated, the bees still selected the honest signal, independently of the signal's identity.

44. Wright GA, Schiestl FP: **The evolution of floral scent: the influence of olfactory learning by insect pollinators on the honest signalling of floral rewards.** *Funct Ecol* 2009, **23**:841-851.
45. Bolstad GH, Armbruster WS, Pelabon C, Perez-Barrales R, Hansen TF: **Direct selection at the blossom level on floral reward by pollinators in a natural population of *Dalechampia schottii*: full-disclosure honesty?** *New Phytol* 2010, **188**:370-384.
46. Majetic CJ, Raguso RA, Ashman T-L: **The sweet smell of success: floral scent affects pollinator attraction and seed fitness in *Hesperis matronalis*.** *Funct Ecol* 2009, **23**:480-487.
47. Theis N, Adler LS: **Advertising to the enemy: enhanced floral fragrance increases beetle attraction and reduces plant reproduction.** *Ecology* 2012, **93**:430-435.

48. Pérez-Barrales R, Bolstad GH, Pélabon C, Hansen TF, Armbruster WS: **Pollinators and seed predators generate conflicting selection on *Dalechampia* blossoms.** *Oikos* 2013, **122**:1411-1428.
49. Theis N, Barber NA, Gillespie SD, Hazzard RV, Adler LS: **Attracting mutualists and antagonists: Plant trait variation explains the distribution of specialist floral herbivores and pollinators on crops and wild gourds.** *Am J Bot* 2014, **101**:1314-1322.
50. Dreisig H: **How long to stay on a plant: the response of bumblebees to encountered nectar levels.** *Arthropod-Plant Interact* 2012, **6**:315-325
51. Grüter C, Moore H, Firmin N., Helanterä H, Ratnieks FL: **Flower constancy in honey bee workers (*Apis mellifera*) depends on ecologically realistic rewards.** *J Exp Biol* 2011, **214**:1397 -1402
52. Hodges SA: **The Influence of Nectar Production on Hawkmoth Behavior, Self Pollination, and Seed Production in *Mirabilis multiflora* (Nyctaginaceae)** *Am J Bot* 1995, **82**:197-204
53. Kambal AE, Bond DA, **Toynbee-Clarke G, 1976 A study on the pollination mechanism in field beans ( *Vicia faba* L.)** *J Agri Sci* **88**:519-26
54. Soto VC, Maldonado IB, Gil RA, Peralta IE, Silva MF, Galmarini CR: **Nectar and Flower Traits of Different Onion Male Sterile Lines Related to Pollination Efficiency and Seed Yield of F1 Hybrids.** *J Econ Entomol* 2013, **106**:1386-1394.
- \*\*55. Wright GA, Baker DD, Palmer MJ, Stabler D, Mustard JA, Power EF, Borland AM, Stevenson PC: **Caffeine in floral nectar enhances a pollinator's memory of reward.** *Science* 2013, **339**:1202-1204.
- These authors show that coffee and citrus species produce varying concentrations of caffeine in their nectar. They demonstrate using laboratory experiments with *Apis mellifera* that caffeine, at nectar relevant concentrations, increases how long bees remember a scent associated with a sucrose reward. Thus, a bee is more likely to remember a flower producing caffeine in its nectar than one that is not.
56. Nepi M, Soligo C, Nocentini D, Abate M, Guarnieri M, Cai G, Bini L, Puglia M, Bianchi L, Pacini E: **Amino acids and protein profile in floral nectar: Much more than a simple reward.** *Flora* 2012, **207**:475-481.
57. Nepi M, Bartomeus I: **Beyond nectar sweetness: the hidden ecological role of non-protein amino acids in nectar.** *J Ecol* 2014, **102**:108-115.
58. Bender RL, Fekete ML, Klinkenberg PM, Hampton M, Bauer B, Malecha M, Lindgren K, J AM, Perera MA, Nikolau BJ, et al.: ***PIN6* is required for nectary auxin response and short stamen development.** *Plant J* 2013, **74**:893-904.
59. Liu G, Thornburg RW: **Knockdown of MYB305 disrupts nectary starch metabolism and floral nectar production.** *Plant J* 2012, **70**:377-388.
60. Liu G, Ren G, Guirgis A, Thornburg RW: **The MYB305 transcription factor regulates expression of nectarin genes in the ornamental tobacco floral nectary.** *Plant Cell* 2009, **21**:2672-2687.
61. Wang W, Liu G, Niu H, Timko MP, Zhang H: **The F-box protein CO11 functions upstream of MYB305 to regulate primary carbohydrate metabolism in tobacco (*Nicotiana tabacum* L. cv. TN90).** *J Exp Bot* 2014, **65**:2147-2160.

62. Radhika V, Kost C, Boland W, Heil M: **The role of jasmonates in floral nectar secretion.** *PLoS One* 2010, **5**:e9265.

\*\*63. Lin IW, Sosso D, Chen LQ, Gase K, Kim SG, Kessler D, Klinkenberg PM, Gorder MK, Hou BH, Qu XQ, et al.: **Nectar secretion requires sucrose phosphate synthases and the sugar transporter SWEET9.** *Nature* 2014, **508**:546-549.

The authors describe a model for floral nectar secretion, showing that SWEET9 is involved in sugar secretion, and this mechanism is likely to be conserved across the eudicots. They also show that sugar biosynthesis genes *SPS1F* and *SPS2F* are up-regulated in *Arabidopsis* nectaries. These genes may provide potential targets for increasing nectar production in crops.

64. Ruhlmann JM, Kram BW, Carter CJ: **CELL WALL INVERTASE 4 is required for nectar production in *Arabidopsis*.** *J Exp Bot* 2010, **61**:395-404.

65. Roulston TH, Cane JH: **Pollen nutritional content and digestibility for animals** *Plant Syst Evol* 2000, **222**:187-209

66. Somme L, Vanderplanck M, Michez D, Lombaerde I, Moerman R, Wathelet B, Wattiez R, Lognay G, Jacquemart AL: **Pollen and nectar quality drive the major and minor floral choices of bumble bees** *Apidologie* 2015, **46**: 92-106.

67. Cook SM, Awmack CS, Murray DA, Williams IH: **Are honey bees' foraging preferences affected by pollen amino acid composition?** *Ecol. Entomol.* 2003, **28**: 622-627.

68. Tegeder M, Rentsch D: **Uptake and Partitioning of Amino Acids and Peptides** *Molecular Plant* 2010, **3**:997-1011.

69. Luo D, Carpenter R, Copsey L, Vincent C, Clark J, Coen E: **Control of organ asymmetry in flowers of *Antirrhinum*.** *Cell* 1999, **99**:367-376.

70. Corley SB, Carpenter R, Copsey L, Coen E: **Floral asymmetry involves an interplay between TCP and MYB transcription factors in *Antirrhinum*.** *Proc Natl Acad Sci U S A* 2005, **102**:5068-5073.

71. Galego L, Almeida J: **Role of *DIVARICATA* in the control of dorsoventral asymmetry in *Antirrhinum* flowers.** *Gene Dev* 2002, **16**:880-891.

72. Raimundo J, Sobral R, Bailey P, Azevedo H, Galego L, Almeida J, Coen E, Costa MMR: **A subcellular tug of war involving three MYB- like proteins underlies a molecular antagonism in *Antirrhinum* flower asymmetry.** *Plant J* 2013, **75**:527-538.

73. Cordoba SA, Cocucci AA: **Flower power: its association with bee power and floral functional morphology in papilionate legumes.** *Ann Bot* 2011, **108**:919-931.

74. Comba L, Corbet SA, Barron A, Bird A, Collinge S, Miyazaki N, Powell M: **Garden flowers: Insect visits and the floral reward of horticulturally-modified variants.** *Ann Bot* 1999, **83**:73-86.

75. Knapp EE, Teuber LR: **Selection progress for ease of floret tripping in alfalfa.** *Crop Sci* 1994, **34**:323-326.

76. Whitney HM, Chittka L, Bruce TJ, Glover BJ: **Conical epidermal cells allow bees to grip flowers and increase foraging efficiency.** *Curr Biol* 2009, **19**:948-953.



77. Alcorn K, Whitney H, Glover B: **Flower movement increases pollinator preference for flowers with better grip.** *Funct Ecol* 2012, **26**:941-947.

78. Ojeda I, Santos-Guerra A, Caujapé-Castells J, Jaén-Molina R, Marrero Á, Cronk QC: **Comparative micromorphology of petals in macaronesian *Lotus* (Leguminosae) reveals a loss of papillose conical cells during the evolution of bird pollination.** *Int J Plant Sci* 2012, **173**:365-374.

79. Suso MJ, Nadal S, Roman B, Gilsanz S: ***Vicia faba* germplasm multiplication – floral traits associated with pollen-mediated gene flow under diverse between-plot isolation strategies.** *Ann Appl Biol* 2008, **152**:201-208.

\*80. Suso MJ, del Rio R: **A crop-pollinator inter-play approach to assessing seed production patterns in faba bean under two pollination environments.** *Euphytica* 2015, **201**:231-251.

These authors grew six genotypes of field bean under open-pollinated and pollinator-exclusion for three consecutive years. Floral traits such as standard petal and ovary dimensions showed signs of selection in the different environments, however, genotype appeared to have an effect on the combination of traits selected for.

81. Pierre J, Mesquida J, Marilleau R, Pham-Delegue MH, Renard M: **Nectar secretion in winter oilseed rape, *Brassica napus* - quantitative and qualitative variability among 71 genotypes.** *Plant Breed* 1999, **118**:471-476.