

1 The genetic architecture of sexually selected  
2 traits in two natural populations of *Drosophila*  
3 *montana*

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16 Running Title: Intra-specific song and CHC QTL in *Drosophila montana*.

17 Word count: 5027

18 Keywords: sexual selection, QTL, *Drosophila montana*, courtship song

19 **Abstract**

20 We investigated the genetic architecture of courtship song and cuticular  
21 hydrocarbon traits in two phylogenetically distinct populations of *Drosophila*  
22 *montana*. In order to study natural variation in these two important traits, we  
23 analysed within-population crosses among individuals sampled from the  
24 wild. Hence, the genetic variation analysed should represent that available for  
25 natural and sexual selection to act upon. In contrast to previous  
26 between-population crosses in this species, no major QTLs were detected,  
27 perhaps because the between-population QTL were due to fixed differences  
28 between the populations. Partitioning the trait variation to chromosomes  
29 suggested a broadly polygenic genetic architecture of within population  
30 variation, although some chromosomes explained more variation in one  
31 population compared to the other. Studies of natural variation provide an  
32 important contrast to crosses between species or divergent lines, but our  
33 analysis highlights recent concerns that segregating variation within  
34 populations for important quantitative ecological traits may largely consist of  
35 small effect alleles, difficult to detect with studies of moderate power.

36

## 37 **Introduction**

38 Quantitative trait locus (QTL) studies are typically performed by  
39 identifying lines or species that differ in a trait of interest, crossing them and  
40 correlating the phenotypic variation in the resulting offspring with genetic  
41 variation, to pinpoint genomic regions associated with the trait of interest. If  
42 the trait of interest is thought to be important to speciation, the initial cross is  
43 often made between inbred lines from different species or differentially  
44 adapted populations. However, using highly differentiated populations  
45 cannot indicate whether the same differences were involved in the initial  
46 isolation of the populations, because current differences may have arisen  
47 since the populations became isolated. In fact, it seems that the types of  
48 mutations providing an initial response to selection within species may differ  
49 from those eventually fixed between species (Stern and Orgogozo, 2008;  
50 2009). QTL studies should, therefore, be performed within and between  
51 populations representing multiple divergence times.

52 More generally, the range of trait variation captured by the initial cross  
53 will influence both the ability to detect QTL and possibly the nature and the  
54 effect size of QTLs detected. Choosing very different individuals for the initial  
55 cross may identify large effect QTLs, which may not be segregating in wild  
56 populations. The effectiveness of the candidate gene approach is a related  
57 issue. Candidate genes are identified by their large effects and, if their  
58 function is evolutionarily conserved, they should be associated with variation  
59 in the same function in different species. Such loci have been found for a  
60 variety of traits, including behaviour patterns (Fitzpatrick *et al.*, 2005; Martin  
61 and Orgogozo, 2013). In other cases a polygenic genetic architecture is  
62 reported, for example in recent QTL studies of ecologically important traits  
63 (Davies *et al.*, 2011; Rockman, 2012; Travisano and Shaw, 2013). This has led  
64 some to question the value of looking for large effect loci for understanding  
65 ecological adaptation within species. Even where large effect loci are  
66 segregating, they may be at very low frequency leading to only a small  
67 contribution to the population level heritability of such traits (Scoville *et al.*,  
68 2011). It is therefore important to compare the genetic architecture of traits in  
69 different types of crosses, such as within vs between populations or lab- vs  
70 field- reared, and assess how often similar or different genetic effects are

71 found.

72 Studies of the quantitative genetics of ecologically important traits in the  
73 field are proceeding well for some vertebrates with extensive pedigree data  
74 (Kruuk and Hill, 2008; Robinson *et al.*, 2013) and some important segregating  
75 loci have been identified (Johnston *et al.*, 2011) but there are very few studies  
76 for quantitative traits in species such as *Drosophila*, that are the source of much  
77 of the information on species differences. The circum-arctic species *Drosophila*  
78 *montana* has proved to be a good species in which to study adaptive traits,  
79 including cold tolerance, developmental time, juvenile body weight and  
80 reproductive diapause (Vesala and Hoikkala, 2011; Salminen *et al.*, 2012;  
81 Parker *et al.*, 2015) in the wild. Phylogeographic analysis of the species based  
82 on mtDNA and microsatellites suggests two clades, a North-American and  
83 European, that were isolated about 0.5 Myr ago (Mirol *et al.*, 2007).  
84 North-American *D. montana* populations have been classified as Standard,  
85 Giant and Alaskan-Canadian based on their geographical origin, size and  
86 chromosome structure (Throckmorton, 1982), and Eurasian populations  
87 contain several unique inversions (Morales-Hojas *et al.*, 2007). There is  
88 evidence for prezygotic isolation (assortative mating), post-mating,  
89 pre-zygotic isolation (successful sperm transfer but failure to fertilise eggs),  
90 and post-mating post-zygotic isolation (progeny production) between some  
91 populations (Jennings *et al.*, 2011; Jennings, Snook, *et al.*, 2014). The main  
92 reasons to classify these populations as belonging to the same species are that  
93 the strength of these different barriers to gene flow depends on the  
94 population pairs being crossed and that pre-mating isolation is only evident  
95 when females can choose between a male from their own population and  
96 another population. Population differences therefore fall into the rarely  
97 studied early part of the speciation continuum (Safran *et al.*, 2013).

98 There have been extensive studies of within and between species  
99 phenotypic variation in the sexual behaviour of the virilis group to which *D.*  
100 *montana*, belongs. Male courtship song is essential for mating in *D. montana*  
101 (Liimatainen *et al.*, 1992), has a species-specific component, inter-pulse  
102 interval (IPI) (Liimatainen and Hoikkala, 1998; Saarikettu, Liimatainen, and  
103 Hoikkala, 2005a), and crosses between closely related species have indicated  
104 X-linkage for genes influencing some song traits (Hoikkala *et al.*, 2000;

105 Päällysaho *et al.*, 2003). Courtship song is under sexual selection and seems to  
106 be an honest indicator of male quality: one song component, intra-pulse  
107 frequency (FRE), is condition-dependent, correlated with offspring survival  
108 and under strong and contrasting viability and sexual selection in the field  
109 (Aspi and Hoikkala, 1995; Hoikkala *et al.*, 1998). Various song components  
110 influence mating latency and courtship duration (Veltsos *et al.*, 2012), and  
111 populations differ in both song and preference (Klappert *et al.*, 2007; Ritchie *et*  
112 *al.*, 2007). Recent studies of cuticular hydrocarbons (CHCs) in *D. montana*  
113 show that they influence mating decisions independently of song and also  
114 differ between populations (Veltsos *et al.*, 2012; Jennings, Etges, *et al.*, 2014).

115 We previously analysed the influence of song and CHC variation in male  
116 mating success in *D. montana* through no-choice experiments where females  
117 were presented with a male from the same population (Oulanka, Finland, or  
118 Vancouver, Canada) and the mating occurrence, mating latency and female  
119 rejection song production were correlated with various components of male  
120 courtship song and CHCs (Veltsos *et al.*, 2012). In this paper, we complete  
121 quantitative genetic and QTL analyses of the same traits by investigating and  
122 comparing their genetic architecture. The same two independent populations,  
123 representing distinct phylogeographic clades (Oulanka and Vancouver) were  
124 used. The phenotypic data of both populations were jointly analysed with  
125 principal component analysis, so that the populations can be compared. We  
126 compare the within-population QTL analysis of song variation with previous  
127 work that identified QTL influencing courtship song by employing  
128 between-population outbred crosses of *D. montana* (from Oulanka, Finland,  
129 and Colorado, N. America). The detected QTL were on two chromosomes (X  
130 and 2) and one candidate gene was potentially associated with each QTL (*per*  
131 and *fru*, respectively) (Schafer *et al.*, 2010; Lagisz *et al.*, 2012), although it has  
132 not been confirmed if these or other genes under the peaks are responsible for  
133 the phenotypic effects.

134 Here we investigate whether within-population variation in the same  
135 traits involves the same genomic regions as the between-population analysis.  
136 We established two independent pedigrees from wild collected females,  
137 developed SNP markers by transcriptome sequencing and scored individuals  
138 from the pedigrees for SNPs and song and CHC traits. Most traits exhibited

139 moderate heritability. Our QTL and chromosome partitioning results reveal  
140 no large effects in the regions identified by the between-population crosses,  
141 and suggest differences in genetic architecture between the populations in  
142 terms of the proportion of additive genetic variation explained by each  
143 chromosome. Overall, they highlight the difficulty of gaining adequate power  
144 to detect loci of small effect in studies on the genetic architecture of wild  
145 populations.

146

## 147 **Materials and Methods**

### 148 *Sampling and phenotyping*

149 Field sampling, cross design and phenotyping have been described  
150 previously (Veltsos *et al.*, 2012). Briefly, we constructed and collected  
151 phenotypic and genotypic information on 2 or 3 generation pedigrees of  
152 about 500 individuals each from the offspring of wild-caught females from  
153 Vancouver, Canada, and Oulanka, Finland. There were 30 isofemale lines  
154 from Vancouver, and 42 from Oulanka. Each line may have had more than  
155 one father, as the frequency of multiple mating in the wild has been estimated  
156 as  $1.19 \pm 0.31$  (Aspi, 1992). Song recording took place while setting up the  
157 crosses of the next generation. We measured four song parameters:  
158 intra-pulse frequency (FRE), inter-pulse interval (IPI), pulse number (PN) and  
159 cycle number per pulse (CN). We analysed these traits and the principal  
160 components of song variation (using data from both populations so that the  
161 PCs are directly comparable), including recording temperature as a covariate  
162 because it has a strong effect on song (Ritchie *et al.*, 2001).

163 CHCs were extracted from whole individuals of both sexes, after the  
164 females had laid sufficient eggs to provide individuals for the next pedigree  
165 generation (>20 eggs). Principal components of CHC data were obtained from  
166 both sexes simultaneously because they are only moderately sexually  
167 dimorphic in *D. montana*, with no sex-specific compounds (Veltsos *et al.*, 2012;  
168 Jennings, Etges, *et al.*, 2014). The PCA analysis was on relative proportions of  
169 log transformed CHCs (Rundle *et al.* 2009) and used 18 compounds in total.  
170 Principal component analysis was performed on data from both populations  
171 simultaneously, to make them directly comparable in QTL analysis. A full  
172 description of the PCA analysis, characterisation of population differences in  
173 CHCs as well as their association with measures of fitness in the lab are  
174 presented in Veltsos *et al.* (2012).

### 175 *Transcriptome sequencing*

176 RNA was extracted from 120 heads of 2-3 week old virgin adults of both  
177 sexes from offspring of the last generation of each pedigree, using a Qiagen  
178 RNA extraction kit (Qiagen, Hilden, Germany), following the manufacturer's  
179 instructions and including on-column DNA digestion. RNA samples were

180 processed by Genepool in Edinburgh (Vancouver flies) and the Centre for  
181 Genomics Research in Liverpool (Oulanka flies). cDNA libraries were  
182 constructed without normalisation, and were sequenced on half a 454  
183 pyrosequencing plate each (Roche, FLX).

184 The sequencing reads were filtered to remove low quality sequence  
185 (threshold of 20) and trimmed to remove adaptor, primer sequence and  
186 poly-A tails using SeqMan NGen version 1.2 (DNASTAR Madison, WI, USA).  
187 348,267/417,467 and 804,456/1,211,678 assembled/total reads were available  
188 for Vancouver and Oulanka respectively. Significantly more reads were  
189 available for Oulanka because two sequencing runs were combined (the first  
190 run yielded only a few reads). An initial assembly was made by combining all  
191 reads from both populations. The resulting master contigs were used to guide  
192 two assemblies, one for each transcriptome. This ensured easy comparison of  
193 SNP locations from each population. All assemblies were performed with  
194 SeqMan NGen version 1.2 (DNASTAR Madison, WI, USA).

#### 195 *SNP marker development*

196 We aimed to obtain SNP markers that segregated in both populations.  
197 The best 384 SNPs to score were identified through custom scripts (on Dryad).  
198 Briefly, SNPs were called in each population using SeqMan NGen version 1.2  
199 (DNASTAR Madison, WI, USA), and were categorised as being unique to a  
200 population, on the same contig, or identical between populations. Criteria for  
201 shortlisting SNPs were good coverage (at least 8 reads in total and 3 reads or  
202 20% for the minor allele), long distance from other SNPs (at least 25 bp),  
203 scoring primer location outside of intron-exon boundary and, when possible,  
204 presence in both populations. Information on the first blastp hit against the  
205 non-redundant (nr) database of each contig was obtained using Blast2GO  
206 (Conesa *et al.*, 2005) and used along with blast searches on a list of candidate  
207 genes to add SNPs that fell within candidate genes using less restrictive  
208 criteria than other SNPs, for example accepting SNPs unique to one  
209 population. Blastclust  
210 (<http://www.csc.fi/english/research/sciences/bioscience/programs/blast/blastclust>)  
211 was also used to remove contigs aligning to potentially highly  
212 repeated sequences, which could cause SNP scoring problems. For more  
213 information on SNP development see Data Archiving.



## 214 *Genotyping*

215 DNA extractions were performed by squashing individual flies in 50  $\mu$ l  
216 Squishing buffer (10mM TrisHCl pH 8.2, 1mM EDTA, 25mM NaCl,  
217 200mg/ml proteinase K), incubating at 50°C for 2 h and boiling for 2 min to  
218 deactivate the proteinase K. DNA clean-up was performed with standard  
219 ethanol precipitation (Sambrook and Russell, 2001).

220 The 384 SNPs were scored on the Illumina BeadXpress platform  
221 (Illumina Inc.) and analysed with GenomeStudio 2010.1 (Genotypic module  
222 1.7.4, Illumina, San Diego, CA). Individuals were removed if they were  
223 associated with genotyping errors (non-Mendelian inheritance in multiple  
224 markers), as identified by GenotypeChecker (Paterson and Law, 2011).  
225 Genotyping errors cause too many spurious recombination events and inflate  
226 genetic maps. Individuals were also removed if they did not have relatives or  
227 were missing phenotypic data. After cleaning, 214 and 334 individuals  
228 (Vancouver) and 171 and 340 individuals (Oulanka) were included in the QTL  
229 analyses for song and CHC, respectively. SNPs were removed from further  
230 analysis if they resulted in any parent-offspring mismatches, or were obvious  
231 outliers in call rate (<0.80). In total 127 and 130 SNPs were considered of  
232 sufficient quality for genetic map construction for Vancouver and Oulanka,  
233 respectively, and 79 of them were common between the populations. All data  
234 used in the QTL analysis have been submitted to Dryad.

## 235 *Genetic map construction*

236 Linkage mapping was performed as in (Slate, 2008) using CRI-MAP  
237 v2.504a (Green *et al.*, 1990; 2009)  
238 (<http://www.animalgenome.org/tools/share/crimap/>). The pedigrees were  
239 first split into subfamilies using the CRIGEN command, and linkage groups  
240 were initially defined by the AUTOGROUP command. Linked markers were  
241 identified using the TWOPOINT command, with all pairs of markers  
242 producing LOD scores in excess of 3.0 being regarded as linked. For each  
243 linkage group, the most parsimonious marker order was determined using  
244 the BUILD, FLIPS, FLIPS3 and FLIPS5 commands. The linkage groups were  
245 assigned to the five *D. montana* chromosomes by blasting the sequence of their  
246 markers against the *D. virilis* scaffolds which have been mapped to  
247 chromosomes in FlyBase (Altschul *et al.*, 1990; St Pierre *et al.*, 2014). When the

248 linkage maps had a different marker order between the populations  
249 (chromosomes 2 and 4), the difference in log likelihood of the collinear  
250 marker order and the parsimonious marker order was obtained for the shared  
251 markers using CHROMPIC and was always  $>3$  (40.06 and 38.13 respectively),  
252 providing support that the markers are not collinear. The genetic maps were  
253 plotted using MAPCHART v2.2 (Voorrips, 2002).

254 Marker 12716\_321\_id was placed by CRI\_MAP 100 cM away from the  
255 next marker at one end of the Oulanka X chromosome. We retained the  
256 marker, by reducing the distance to 49.5 cM, for three reasons: It mapped  
257 without problems on the Vancouver X, it has a homologue on the *D. virilis* X,  
258 and there was low marker coverage of the X.

### 259 *Chromosome partitioning analysis*

260 Genome-wide relatedness matrices (GWRMs), weighted for expected  
261 relatedness from pedigree information, were constructed using the  
262 methodology of (Robinson *et al.*, 2013): For each autosome, pairs of GWRM  
263 matrices were made for its markers and those of the remaining autosomes.  
264 Each pair of GWRMs was used, in turn, to partition the variance explained by  
265 an autosome. To determine the additive genetic variation explained by an  
266 autosome, we compared the likelihood of a model with both GWRMs and a  
267 model with only the GWRM of the remaining autosomes, for each GWRM  
268 pair. A separate GWRM was constructed for the X chromosome and was only  
269 used to estimate the variance explained by the X, in a similar fashion. The  
270 models included temperature for song and sex for CHC data as fixed effects.  
271 Model fitting was performed in ASReml version 3 (Gilmour *et al.*, 2009). Full  
272 details of the methodology are available in (Santure *et al.*, 2013).

273 If a trait is polygenic, the variance explained by each chromosome should  
274 be proportional to its size. We considered three proxies of gene number per  
275 chromosome: the physical length of the *D. virilis* homologous chromosome  
276 and the genetic map length of the chromosome in either *D. montana* or *D.*  
277 *virilis*. We consider the *D. virilis* chromosome physical length to be the best  
278 proxy because loci on the X chromosome were underrepresented in our  
279 genetic maps, making them shorter than expected. Possible reasons for the  
280 low SNP density on the X include the fact that males, which provided about  
281 half the RNA used in sequencing, are hemizygous (rare SNPs would be less

282 likely to be retained by the SNP filtering pipeline). The *D. virilis* chromosome  
283 lengths were obtained from Flybase (Altschul *et al.*, 1990; St Pierre *et al.*, 2014),  
284 by summing up the lengths of all the *D. virilis* scaffolds, that have been  
285 mapped to each chromosome.

#### 286 *QTL - animal model analysis*

287 Identity by descent (IBD) matrices were constructed at 10 cM intervals  
288 for all individuals of each pedigree, using all SNPs on the relevant  
289 chromosome with an MCMC approach implemented in LOKI v2.4.5 (Heath *et*  
290 *al.*, 1997; Heath, 1997), and a parameter setting of 10,000 iterations for each  
291 position. The X chromosome is hemizygous in males, but LOKI requires two  
292 alleles for all markers. We manually made the male X marker information  
293 compatible with the analysis by adding a novel allele for each X male  
294 genotype (i.e. all males carried one copy of a dummy allele that was absent in  
295 females). The phenotypic variance was partitioned into fixed and random  
296 effects in a restricted maximum-likelihood (REML) framework using ASReml  
297 version 3 (Gilmour *et al.*, 2009), as in Slate (2005). Heritability ( $h^2$ ) was  
298 estimated as the ratio of additive genetic to total phenotypic variance from a  
299 polygenic model. Potential QTL effects were estimated as in Santure *et al.*  
300 (2013). Briefly, a LOD score was calculated by subtracting the log-likelihood  
301 of a polygenic model from the log-likelihood of a polygenic plus QTL model,  
302 the latter also including the IBD matrix, for each genomic interval.  
303 Significance thresholds were adjusted for multiple testing based on genome  
304 size, following Lander and Kruglyak (1995). Two genome-wide linkage  
305 thresholds were calculated for each population (File S1): the suggestive  
306 linkage threshold is expected once per genome scan, while the significant  
307 linkage threshold has a 1/20 false positive rate, i.e. it represents a  
308 genome-wide significance of  $P < 0.05$  (Lander and Kruglyak, 1995; Nyholt,  
309 2000).

310 We could not directly compare the two populations within a single  
311 analysis because not all of their markers were shared, and because the relative  
312 positions of the markers differed. Instead we indirectly compared the  
313 populations by comparing results from the chromosome partitioning analysis,  
314 which is not affected by marker order.

315 *Power analysis*

316 We performed simulations using custom scripts to estimate the  
317 probability of detecting, within the Vancouver pedigree, a QTL of similar  
318 magnitude to the one detected in the inter-population study (Lagisz *et al.*,  
319 2012). We used sex-averaged map distances and simulated a trait with  
320 heritability of 0.30, half the variance of which was caused by a single QTL in  
321 the middle of chromosome 2 ( $V_a = 0.15$ ,  $V_{\text{qtl}} = 0.15$ ,  $V_{\text{residual}} = 0.70$ ). The  
322 simulations were run for 100 replicates, assumed all individuals were  
323 phenotyped and employed the same suggestive and significant linkage  
324 thresholds as the experimental QTL analysis.

## 325 Results

### 326 *Phenotypic variation*

327 Phenotypic variation for the two pedigrees has been described before  
328 and the populations differ in all traits (Veltsos *et al.*, 2012). Means and  
329 standard deviations for all phenotyped traits are presented in Table 1.

### 330 *Genetic maps*

331 The total length of the map from the Vancouver pedigree was 789.8 cM,  
332 and that from the Oulanka pedigree 689.8 cM. Comparison of these maps  
333 clearly shows a different marker order for chromosomes 2 and 4 (Figure 1,  
334 File S2). Chromosomal rearrangements are known to have occurred within *D.*  
335 *montana* and we therefore believe that extensive changes in gene order are not  
336 unlikely. A population on the Western Coast of North America (around  
337 Vancouver) has at least one inversion on the 2nd chromosome and 10  
338 inversions on the 4th chromosome that are not found in Finland, while the  
339 Finnish population has 3 inversions on the 2nd chromosome that are not  
340 found in the Vancouver region (Hsu, 1952; Patterson and Stone, 1952;  
341 Moorhead, 1954; Morales-Hojas *et al.*, 2007). Information on the markers such  
342 as heterozygosities, their top blast hit, potential functions and sequence of the  
343 relevant contig is provided in File S3.

### 344 *Heritability estimates*

345 Heritability and standard error estimates based on the relatedness  
346 matrices calculated from all SNPs are shown in Table 1. The estimates are  
347 broadly consistent between the populations, suggesting some similarities  
348 between the traits (correlation coefficient between heritability estimates for  
349 PCs = 0.83).

### 350 *Power analysis*

351 Simulations suggest there was only moderate power to detect QTLs of an  
352 effect size similar to those detected in between-population crosses. Out of 100  
353 simulations, 55 detected the simulated QTL at the suggestive threshold and  
354 only 13 detected it at the genome-wide significance threshold (File S4a). When  
355 detected, the Beavis Effect (Göring *et al.*, 2001) was observed, i.e. the QTL

356 effect size was usually overestimated (mean effect size was 0.25 instead of  
357 simulated 0.15 proportion of total variance). When the QTL location was  
358 detected, it was accurately mapped (File S4b).

#### 359 *QTL results*

360 Based on the total genetic map length, the suggestive and significant  
361 LOD scores for Vancouver were 1.12 and 2.62 respectively, and for Oulanka  
362 1.05 and 2.56. We did not detect genome-wide significant results in the QTL  
363 analysis but observed suggestive linkage once, on chromosome 5, for CN in  
364 Oulanka (Figure 2). The result had a slightly higher LOD score when song  
365 principal components were used, (song\_PC3 which is strongly influenced by  
366 CN (Veltsos *et al.*, 2012)), but was otherwise qualitatively similar (File S6).  
367 Interestingly the song characters mostly influencing song\_pc3 (FRE, IPI and  
368 CN (Veltsos *et al.*, 2012)) had lower LRT (log likelihood ratio test) values  
369 individually than the LRT for the principal component, suggesting that  
370 multivariate analysis may increase detection power, presumably because a  
371 principal component axis reduces the noise of individual measurements.

#### 372 *Genetic variation partitioning*

373 There was no relationship between percent variance explained (PVE) and  
374 chromosome length, regardless of the proxy of chromosome length used  
375 (Table 2, File S5). It is difficult to detect such a correlation because of the small  
376 number of data points as there are only 5 chromosomes in *D. montana*. In  
377 addition it is difficult to detect differences in PVE between the chromosomes  
378 because they have similar sizes, which may make the between chromosome  
379 variance small, relative to the within chromosome error in estimating  
380 variance. There were 18 cases where a single chromosome explained non-zero  
381 variance (Table 3). No formal test of significance was performed because the  
382 null hypothesis of no variation explained by any chromosome is false for  
383 polygenic heritable traits.

#### 384 *Population comparison*

385 We compared the genetic architecture between the populations by  
386 plotting the percent variance explained by each chromosome for each trait, in  
387 the two populations (Files S7, S9). While the values were often similar, there

388 were occasions where a chromosome clearly explained more variance in one  
389 population than in the other population (e.g. for song PC1).  
390

391 **Discussion**

392 We have attempted to identify the architecture of genetic variation  
393 available for natural and sexual selection to act upon, within natural  
394 populations, by performing heritability and QTL analysis on song and CHC  
395 phenotypes using pedigrees established from two populations of *D. montana*.  
396 Although we found significant heritability, we did not find evidence of large  
397 effect QTL for any trait. Our results are broadly suggestive of polygenic  
398 determination for intrapopulation phenotypic variation, though some  
399 individual chromosomes explained more phenotypic variation than others, in  
400 a population-specific manner.

401 Song and CHCs contribute to mating success in *D. montana*, and CHCs  
402 are probably also involved in ecological adaptation (Veltsos *et al.*, 2012;  
403 Jennings, Etges, *et al.*, 2014). Heritabilities were sometimes low (Table 1),  
404 which may well have militated against our ability to detect QTL (the only  
405 suggestive QTL found was for a song trait with one of the highest  
406 heritabilities, CN in Oulanka). Nevertheless, our heritability estimates for  
407 song were usually larger than previous estimates in *D. montana*, though  
408 sometimes they were not (Aspi and Hoikkala, 1993; Suvanto *et al.*, 1999). The  
409 estimates reported here are probably more robust because of the larger  
410 sample size, the estimation of relatedness from pedigrees and the reduced  
411 measurement noise achieved by using different song analysis methods. The  
412 earlier estimation of heritabilities from overwintered flies and the finding that  
413 they increase compared to non-overwintered flies (Suvanto *et al.*, 1999)  
414 remains important because sexual selection on courtship song often acts only  
415 after the flies overwinter as adults.

416 Simulations suggested that our power to detect QTL was only moderate.  
417 The simulated trait effect size was based on a QTL detected for song variation  
418 in crosses between North American and Finnish *D. montana* (Colorado and  
419 Oulanka, i.e. a different population from North America) (Mirol *et al.*, 2007;  
420 Schafer *et al.*, 2010; Lagisz *et al.*, 2012). We failed to detect QTL for the same  
421 traits and genomic positions as the between-population study. These were for  
422 IPI and PN on the X chromosome, and FRE, CN and IPI on chromosome 2  
423 (Lagisz *et al.*, 2012). We then attempted to implicate the same chromosomes  
424 using the chromosome partitioning analysis. Of the above traits, a large



425 proportion of variance ( $31.8 \pm 17.3 \%$ ) was only found to be predicted for IPI  
426 on chromosome 2 in Oulanka (Files S5, S7). One possible explanation for the  
427 disparity between the studies is that large-effect alleles that differ between  
428 clades are not making a major contribution to genetic variation within  
429 populations. This supports recent studies suggesting that the genetic  
430 architecture of intra-population and inter-clade variation may differ for  
431 *Drosophila song*, (Gleason, 2005; Arbuthnott, 2009) and for sexually selected  
432 traits generally (Chenoweth and McGuigan, 2010). Similarly, some  
433 between-population QTL may be due to Colorado-specific alleles of large  
434 effect, which were not sampled in this study. An alternative explanation for  
435 the disparity between the within- and between- population studies may be  
436 the greater range in phenotypes amongst the crosses between two  
437 populations, compared to within population variation. Colorado flies are  
438 different from those from Vancouver, in both their songs and pheromones  
439 (Klappert *et al.*, 2007; Routtu *et al.*, 2007; Jennings, Etges, *et al.*, 2014), despite  
440 coming from the same phylogeographic clade.

441 A third possibility is that QTL of large effect are difficult to detect using  
442 intra-population analysis. When detected in crosses between lines, QTL may  
443 be caused by variants of large effect that are at low frequency within the  
444 population. Variants of large effect under selection would rapidly fix within a  
445 population during adaptation to new environments and are not likely to  
446 segregate within populations (Scoville *et al.*, 2011). Consequently they may be  
447 readily associated with large QTL in between-population crosses that do not  
448 explain much phenotypic variance at the population level.

449 A fourth possibility is that the between-population QTL seem large  
450 because fixed chromosomal rearrangements between the populations caused  
451 a reduction in recombination in some genomic regions, effectively combining  
452 the effects of many loci of small effect, and causing them to segregate as a  
453 single locus. Loci implicated in repeated evolution of adaptive traits are often  
454 associated with inversions (Martin and Orgogozo, 2013). When the genes  
455 captured by an inversion are involved in reproductive isolation, the inversion  
456 may become important in maintaining isolation in the face of gene flow (Noor  
457 *et al.*, 2001; Kirkpatrick and Barton, 2006). The IPI QTL in an inversion on  
458 chromosome 2 in the between-population study (Lagisz *et al.*, 2012) is

459 compatible with both ideas of capturing multiple genes and being associated  
460 with reproductive isolation, since IPI is under sexual selection and differs  
461 between species (Saarikettu, Liimatainen, and Hoikkala, 2005b; Veltsos *et al.*,  
462 2012). Within-population crosses would not be affected by such inversions,  
463 which could explain the failure to detect QTL.

464 The QTL in sexually selected traits in within- and between- population  
465 studies are usually different (Arbuthnott, 2009; Chenoweth and McGuigan,  
466 2010). One exception, for a pre-mating signal, involves a pheromone QTL in  
467 the moth species *Heliothis subflexa* and *H. virescens* (Groot *et al.*, 2013). Few  
468 studies have compared the magnitude of effect sizes within and between  
469 populations in addition to assessing if the same QTLs are implicated, and it  
470 may be that consistent genetic effects are associated with phenotypes where  
471 large effect loci are more likely to be found, such as pheromone  
472 polymorphisms or genes influencing melanism (Wittkopp *et al.*, 2009).

#### 473 *Comparison between populations*

474 One common method to demonstrate polygenic variation is through a  
475 correlation of percent variance explained (PVE) with chromosome size. The  
476 relatively low number of similarly-sized chromosomes makes this an  
477 ineffective test for *Drosophila*, but most chromosomes explained some  
478 variation which is compatible with polygenic genetic architecture. There were  
479 cases where one chromosome explained more variation than others. This  
480 sometimes happened in a population-specific manner (File S7), providing  
481 evidence of a subtly different genetic architecture between the Oulanka and  
482 Vancouver populations.

483 The X chromosome had the lowest number of markers in our study even  
484 though it is the largest chromosome. The low X coverage would be  
485 particularly unfortunate if there is a large X effect, that is if the X is  
486 disproportionately influencing reproductive isolation (Presgraves, 2008) or  
487 genes under sexual selection (Qvarnstrom and Bailey, 2009; Dean and Mank,  
488 2014), which may have further hindered our ability to detect segregating  
489 genetic variation for these traits.

#### 490 *Conclusion*

491 In performing this study we have developed useful resources, including

492 a comparative transcriptomic data set and a set of SNP markers and  
493 associated genetic maps for this species. Like many studies of wild  
494 populations, our study suffers from limited statistical power. Investigating  
495 the genetic architecture of within-population variation probably requires  
496 sample sizes in the order of a few thousand, in order to ensure sufficient  
497 statistical power (Rockman, 2012; Slate, 2013). Despite significant heritabilities  
498 of the traits we found limited evidence of common genetic architecture in  
499 within- population phenotypic variation and interspecific differences,  
500 extending the trend found in other comparisons (Arbuthnott, 2009;  
501 Chenoweth and McGuigan, 2010). This may suggest that large effect alleles  
502 fixed between populations are not major contributors to variation within  
503 populations. It is important to study within- population variation despite  
504 such difficulties because it is the variation upon which natural and sexual  
505 selection act, making it the most relevant variation for immediate responses to  
506 selection (Prokop *et al.*, 2012). Concentrating on QTL mapping populations  
507 from between-species crosses can reveal genetic mechanisms of species  
508 differences, but such genes may have diverged in frequency or become fixed  
509 after speciation. Understanding sources of genetic variation across the  
510 speciation continuum remains a major challenge of evolutionary genetics (The  
511 Marie Curie SPECIATION Network, 2012).

512

### 513 **Acknowledgments**

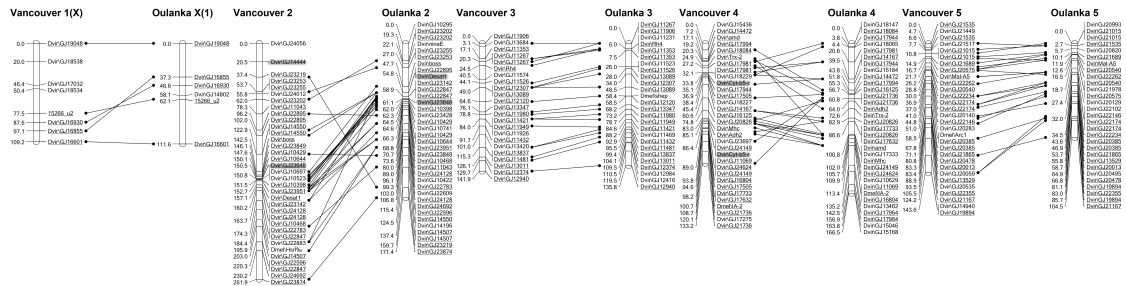
514 The work was supported by the National Environment Research Council  
515 (grant NE/E015255/1 to M.G.R and R.K.B.) and the Academy of Finland  
516 (project 132619 to A.H.). We would like to thank Juan Galindo and Anna  
517 Santure for bioinformatic and statistical help. We are grateful to Dario Beraldi  
518 for providing a script to streamline the pedigree analysis and parse files  
519 between PedigreeChecker, CRIMAP and ASREML, and Matthew Robinson  
520 for providing scripts allowing the chromosome partitioning analysis.

### 521 **Data archiving**

522 Phenotype, genotype and pedigree data as well as raw reads and the  
523 assembled transcriptome, are available from the Dryad Digital Repository  
524 doi:10.5061/dryad.4p9j3.

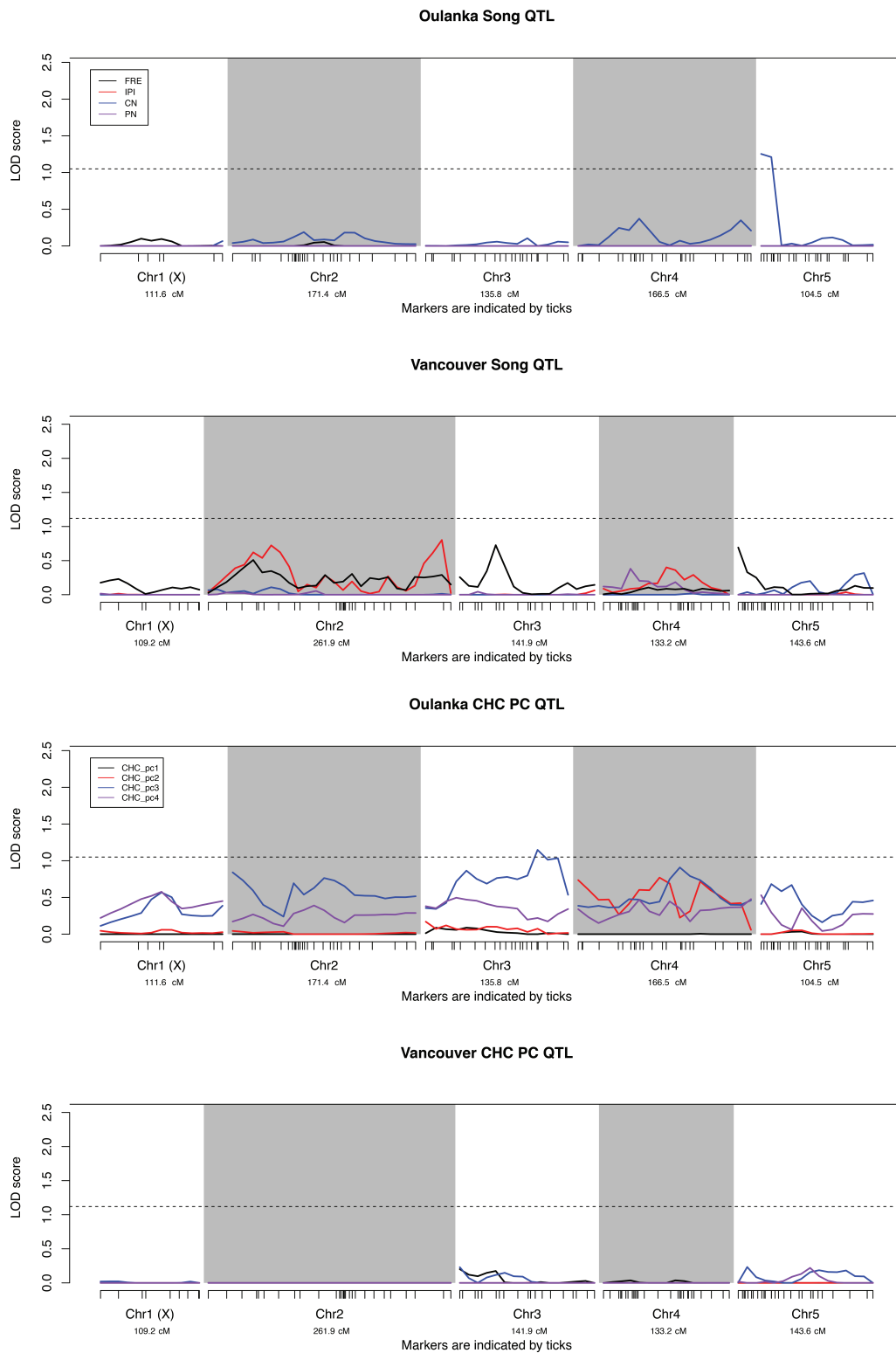
525 **Figures**

526 **Figure 1:** Comparison of genetic maps from the two populations. The  
 527 markers are represented by the name of the top *D. virilis* blast hit of the  
 528 relevant contig. Chromosome 4 is clearly not collinear between the two  
 529 populations. Candidate genes are indicated by shading: *Dvir/GJ14444-Ebony*,  
 530 *Dvir/GJ23648-Slowpoke*, *Dvir/Desat1-DsatF* and *Dvir/GJCyt-b5-r-CytB5*.



531  
532

533 **Figure 2:** QTL maps of song and CHC in the two populations. The dotted  
 534 and uninterrupted horizontal lines indicate suggestive and significant  
 535 linkage, respectively. Chromosomes X and 2, which showed QTL for song in  
 536 the between-population study, have no QTL identified.



537

538 **Tables**

539 **Table 1:** Trait values (genotyped individuals from (Veltso *et al.*, 2012)) and  
 540 heritability estimates from polygenic models of song and CHC traits.

541

Character	value $\pm$ sd		h <sup>2</sup> $\pm$ se	
	Oulanka	Vancouver	Oulanka	Vancouver
FRE (Hz)	253 $\pm$ 35	235 $\pm$ 39	0.08 $\pm$ 0.11	0.11 $\pm$ 0.09
IPI (msec)	40.18 $\pm$ 6.38	43.47 $\pm$ 6.83	0.2 $\pm$ 0.14	0.21 $\pm$ 0.11
PN	4.80 $\pm$ 0.95	4.79 $\pm$ 0.85	0.14 $\pm$ 0.12	0.06 $\pm$ 0.1
CN	9.40 $\pm$ 1.25	10.13 $\pm$ 1.40	0.52 $\pm$ 0.18	0.15 $\pm$ 0.12
Song_pc1	0.39 $\pm$ 1.16	-0.41 $\pm$ 1.34	0.14 $\pm$ 0.13	0.11 $\pm$ 0.10
Song_pc2	-0.02 $\pm$ 1.05	0.14 $\pm$ 1.00	0.28 $\pm$ 0.14	0.17 $\pm$ 0.10
Song_pc3	0.16 $\pm$ 0.93	-0.09 $\pm$ 0.90	0.4 $\pm$ 0.17	0.25 $\pm$ 0.14
Song_pc4	-0.03 $\pm$ 0.46	0.04 $\pm$ 0.48	NA	0.16 $\pm$ 0.10
CHC_MF_pc1	1.19 $\pm$ 2.19	-1.34 $\pm$ 2.91	0.02 $\pm$ 0.06	0.05 $\pm$ 0.05
CHC_MF_pc2	-0.90 $\pm$ 0.93	1.14 $\pm$ 1.47	0.56 $\pm$ 0.08	0.37 $\pm$ 0.09
CHC_MF_pc3	-0.19 $\pm$ 1.08	0.19 $\pm$ 1.42	0.35 $\pm$ 0.09	0.06 $\pm$ 0.05
CHC_MF_pc4	0.38 $\pm$ 0.78	-0.45 $\pm$ 1.04	0.12 $\pm$ 0.07	0.1 $\pm$ 0.06

542 **Table 2:** Correlation between PVE and *D. virilis* chromosome length. Each  
 543 chromosome is represented by the combined PVE values from the two  
 544 pedigrees, since they did not differ significantly (data not shown). P-values of  
 545 negative correlations are labelled as 'NS' because only positive correlations are expected.

character	correlation	p
FRE	-0.52	NS
IPI	0.47	0.17
PN	-0.04	NS
CN	-0.50	NS
song_pc1	0.51	0.13
song_pc2	-0.30	NS
song_pc3	-0.41	NS
song_pc4	-0.53	NS
CHC_pc1	-0.19	0.60
CHC_pc2	0.28	0.43
CHC_pc3	-0.52	NS
CHC_pc4	-0.3	NS

546

547

548

549 **Table 3:** Traits where a chromosome explains non-zero variance (percent  
 550 variance explained - SE > 0). In all cases only one of the populations explains  
 551 > 0 PVE.

Chromosome	Character	PVE $\pm$ SE	Population
2	song_pc1	0.30 $\pm$ 0.17	O
		0.00 $\pm$ 0.00	V
2	IPI	0.32 $\pm$ 0.17	O
		0.00 $\pm$ 0.00	V
2	CHC_pc2	0.10 $\pm$ 0.09	O
		0.07 $\pm$ 0.08	V
3	song_pc3	0.35 $\pm$ 0.16	O
		0.04 $\pm$ 0.07	V
3	PN	0.10 $\pm$ 0.12	O
		0.16 $\pm$ 0.12	V
3	FRE	0.00 $\pm$ 0.00	O
		0.13 $\pm$ 0.11	V
3	CN	0.19 $\pm$ 0.17	O
		0.03 $\pm$ 0.07	V

3	CHC_pc3	0.03 ± 0.07	O
		0.09 ± 0.07	V
4	CN	0.22 ± 0.16	O
		0.04 ± 0.07	V
4	CHC_pc3	0.00 ± 0.00	O
		0.03 ± 0.05	V
4	CHC_pc1	0.00 ± 0.00	O
		0.08 ± 0.07	V
4	IPI	0.15 ± 0.13	O
		0.07 ± 0.08	V
4	song_pc2	0.15 ± 0.14	O
		0.00 ± 0.00	V
5	song_pc2	0.00 ± 0.00	O
		0.29 ± 0.12	V
5	CHC_pc3	0.07 ± 0.07	O
		0.00 ± 0.00	V
5	CHC_pc2	0.07 ± 0.07	O
		0.17 ± 0.08	V
5	IPI	0.04 ± 0.13	O
		0.10 ± 0.09	V
5	song_pc1	0.07 ± 0.12	O
		0.11 ± 0.10	V

552

553

554

### 555 **Supplementary files**

556 **File S1:** QTL significance thresholds per chromosome and genome  
557 (significant and suggestive linkage).

558 **File S2:** Visualisation of the *D. virilis* scaffold associated with each  
559 marker. A different colour was used for different scaffolds in each  
560 chromosome.

561 **File S3:** Marker information. The columns show the SNP name, the name  
562 of the contig it is associated with, the most likely *D. virilis* homologue of the  
563 contig, a description of the top blast hit based on Blast2GO annotation, the *D.*  
564 *virilis* scaffold it maps to, the *D. virilis* chromosome associated with that  
565 scaffold, the map distance estimated from the Oulanka and Vancouver  
566 pedigrees and the number of individuals genotyped and the heterozygosity of  
567 the marker for each of the pedigrees (obtained using Cervus v3.0.3  
568 (Kalinowski *et al.*, 2007)). The last column can be used to make a SNP chip  
569 order from Illumina for the BeadExpress platform.

570 **File S4: A.** Simulated maximum LRT score distribution. Summary of the  
571 maximum LRT score over 100 simulations. Suggestive and significant linkage  
572 correspond to LRT values 5.16 and 12.08, respectively. **B.** Mean values of LRT  
573 across 100 simulations. The simulated QTL is at 80 cM (arrow). LRT values  
574 5.16 and 12.08 correspond to the suggestive and significant LOD scores 1.12  
575 and 2.62 respectively. The plot can be regarded as 'conservative', because the

576 same position does not yield the maximum test stat each time and because it  
577 includes the replicates where the QTL was not found.

578 **File S5:** Chromosome partitioning results. **PVE:** Percent variance  
579 explained. **LRT:** Log likelihood ratio test. Three proxies of chromosome  
580 length are provided. The *D. virilis* physical and genetic map lengths are taken  
581 from Flybase.

582 **File S6:** QTL map of song PCs. The result is qualitatively similar to the  
583 analysis of individual song characters. The PCs are the same as in (Veltsos *et*  
584 *al.*, 2012).

585 **File S7:** Comparison of PVE by chromosome in the two populations.  
586 The scale is the same in all plots. While the values often correlate, there are  
587 clear occasions where one chromosome in one population explains more  
588 variance than in the other population.



589 **References**

590

591 Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ (1990). Basic local alignment search  
592 tool. *Journal of Molecular Biology* **215**: 403–410.

593 Arbuthnott D (2009). The genetic architecture of insect courtship behavior and premating  
594 isolation. *Heredity* **103**: 15–22.

595 Aspi J (1992). Incidence and adaptive significance of multiple mating in females of two boreal  
596 *Drosophila virilis*-group species. *Annales Zoologici Fennici* **29**: 147–159.

597 Aspi J, Hoikkala A (1993). Laboratory and natural heritabilities of male courtship song  
598 characters in *Drosophila montana* and *D. littoralis*. *Heredity* **70** : 400–406.

599 Aspi J, Hoikkala A (1995). Male mating success and survival in the field with respect to size  
600 and courtship song characters in *Drosophila littoralis* and *D. montana* (Diptera,  
601 Drosophilidae). *Journal of Insect Behavior* **8**: 67–87.

602 Chenoweth SF, McGuigan K (2010). The genetic basis of sexually selected variation. *Annu Rev*  
603 *Ecol Evol Syst* **41**: 81–101.

604 Conesa A, Gotz S, Garcia-Gomez JM, Terol J, Talon M, Robles M (2005). Blast2GO: a universal  
605 tool for annotation, visualization and analysis in functional genomics research.  
606 *Bioinformatics* **21**: 3674–3676.

607 Davies G, Tenesa A, Payton A, Yang J, Harris SE, Liewald D, *et al.* (2011). Genome-wide  
608 association studies establish that human intelligence is highly heritable and polygenic.  
609 *Mol Psychiatry* **16**: 996–1005.

610 Dean R, Mank JE (2014). The role of sex chromosomes in sexual dimorphism: discordance  
611 between molecular and phenotypic data. *J Evol Biol*.

612 Fitzpatrick MJ, Ben-Shahar Y, Smid HM, Vet LEM, Robinson GE, Sokolowski MB (2005).  
613 Candidate genes for behavioural ecology. *Trends Ecol Evol (Amst)* **20**: 96–104.

614 Gilmour AR, Gogel MJ, Cullis BR, R T (2009). *ASReml User Guide Release 3.0*. Hemel  
615 Hempstead, UK.

616 Gleason JM (2005). Mutations and natural genetic variation in the courtship song of  
617 *Drosophila*. *Behav Genet* **35**: 265–277.

618 Göring HHH, Terwilliger JD, Blangero J (2001). Large upward bias in estimation of  
619 locus-specific effects from genomewide scans. *The American Journal of Human Genetics* **69**:  
620 1357–1369.

621 Green P, Falls K, Crooks S (1990). *CRI-MAP 2.4 documentation*.

622 Green P, Falls K, Crooks S (2009). *CRI-MAP 2.503 documentation*.

623 Groot AT, Staudacher H, Barthel A, Inglis O, Schofl G, Santangelo RG, *et al.* (2013). One  
624 quantitative trait locus for intra- and interspecific variation in a sex pheromone.  
625 *Molecular Ecology* **22**: 1065–1080.

626 Heath SC (1997). Markov chain Monte Carlo segregation and linkage analysis for oligogenic  
627 models. *Am J Hum Genet* **61**: 748–760.

628 Heath SC, Snow GL, Thompson EA (1997). MCMC segregation and linkage analysis. *Genetic*  
629 *Epidemiology* **14**: 1011–1016.

- 630 Hoikkala A, Aspi J, Suvanto L (1998). Male courtship song frequency as an indicator of male  
631 genetic quality in an insect species, *Drosophila montana*. *Proc Biol Sci* **265**: 503–508.
- 632 Hoikkala A, Päällysaho S, Aspi J, Lumme J (2000). Localization of genes affecting species  
633 differences in male courtship song between *Drosophila virilis* and *D. littoralis*. *Genet Res* **75**:  
634 37–45.
- 635 Hsu TC (1952). Chromosomal variation and evolution in the virilis group of *Drosophila*.  
636 *University of Texas Publication* **52004**: 35–72.
- 637 Jennings J, Etges WJ, Schmitt T, Hoikkala A (2014). Cuticular hydrocarbons of *Drosophila*  
638 *montana*: Geographic variation, sexual dimorphism and potential roles as pheromones. *J*  
639 *Insect Physiol* **61**: 16–24.
- 640 Jennings J, Mazzi D, Ritchie M, Hoikkala A (2011). Sexual and postmating reproductive  
641 isolation between allopatric *Drosophila montana* populations suggest speciation potential.  
642 *BMC Evol Biol* **11**: 68.
- 643 Jennings J, Snook RR, Hoikkala A (2014). Reproductive isolation among allopatric *Drosophila*  
644 *montana* populations. *Evolution* **68**: 3095–3108.
- 645 Johnston SE, McEwan JC, Pickering NK, Kijas JW, Beraldi D, Pilkington JG, *et al.* (2011).  
646 Genome-wide association mapping identifies the genetic basis of discrete and  
647 quantitative variation in sexual weaponry in a wild sheep population. *Molecular Ecology*  
648 **20**: 2555–2566.
- 649 Kirkpatrick M, Barton NH (2006). Chromosome inversions, local adaptation and speciation.  
650 *Genetics* **173**: 419–434.
- 651 Klappert K, Mazzi D, Hoikkala A, Ritchie M (2007). Male courtship song and female  
652 preference variation between phylogeographically distinct populations of *Drosophila*  
653 *montana*. *Evolution* **61**: 1481–1488.
- 654 Kruuk LEB, Hill WG (2008). Introduction. Evolutionary dynamics of wild populations: the  
655 use of long-term pedigree data. *Proc Biol Sci* **275**: 593–596.
- 656 Lagisz M, Wen S-Y, Routtu J, Klappert K, Mazzi D, Morales-Hojas R, *et al.* (2012). Two distinct  
657 genomic regions, harbouring the period and fruitless genes, affect male courtship song in  
658 *Drosophila montana*. *Heredity* **108**: 602–608.
- 659 Lander E, Kruglyak L (1995). Genetic dissection of complex traits: guidelines for interpreting  
660 and reporting linkage results. *Nat Genet* **11**: 241–247.
- 661 Liimatainen JO, Hoikkala A (1998). Interactions of the males and females of three sympatric  
662 *Drosophila virilis*-group species, *D. montana*, *D. littoralis*, and *D. lummei*, (Diptera:  
663 Drosophilidae) in intra- and interspecific courtships in the wild and in the laboratory.  
664 *Journal of Insect Behavior* **11**: 399–417.
- 665 Liimatainen J, Hoikkala A, Aspi J, Welbergen P (1992). Courtship in *Drosophila montana*: the  
666 effects of male auditory signals on the behaviour of flies. *Animal Behaviour* **43**: 35–48.
- 667 Martin A, Orgogozo V (2013). The Loci of repeated evolution: a catalog of genetic hotspots of  
668 phenotypic variation. *Evolution* **67**: 1235–1250.
- 669 Mirol PM, Schafer MA, Orsini L, Routtu J, Schlötterer C, Hoikkala A, *et al.* (2007).  
670 Phylogeographic patterns in *Drosophila montana*. *Molecular Ecology* **16**: 1085–1097.
- 671 Moorhead PS (1954). VIII. Chromosome variation in giant forms of *Drosophila montana*. In:  
672 *University of Texas Publication No 5422*, pp 106–129.
- 673 Morales-Hojas R, Päällysaho S, Vieira C, Hoikkala A, Vieira J (2007). Comparative polytene

- 674 chromosome maps of *D. montana* and *D. virilis*. *Chromosoma* **116**: 21–27.
- 675 Noor MA, Grams KL, Bertucci LA, Reiland J (2001). Chromosomal inversions and the  
676 reproductive isolation of species. **98**: 12084–12088.
- 677 Nyholt D (2000). All LODs are not created equal. *Am J Hum Genet* **67**: 282–288.
- 678 Parker DJ, Vesala L, Ritchie M, Laiho A, Hoikkala A, Kankare M (2015). How consistent are  
679 the transcriptome changes associated with cold acclimation in two species of the  
680 *Drosophila virilis* group? *Heredity*.
- 681 Paterson T, Law A (2011). Genotypechecker: an interactive tool for checking the inheritance  
682 consistency of genotyped pedigrees. *Animal Genetics* **42**: 560–562.
- 683 Patterson JT, Stone WS (1952). *Evolution in the Genus Drosophila*. MacMillan: New York.
- 684 Päälyysaho S, Aspi J, Liimatainen J, Hoikkala A (2003). Role of X chromosomal song genes in  
685 the evolution of species-specific courtship songs in *Drosophila virilis* group species. *Behav*  
686 *Genet* **33**: 25–32.
- 687 Presgraves DC (2008). Sex chromosomes and speciation in *Drosophila*. *Trends Genet* **24**: 336–  
688 343.
- 689 Prokop ZM, Michalczyk Ł, Drobnik SM, Herdegen M, Radwan J (2012). Meta-analysis  
690 suggests choosy females get sexy sons more than ‘good genes’. *Evolution* **66**: 2665–2673.
- 691 Qvarnstrom A, Bailey R (2009). Speciation through evolution of sex-linked genes. *Heredity*  
692 **102**: 4–15.
- 693 Ritchie M, Etges WJ, de Oliveira CC, Gragg E, Ortiz-Barrientos D, Noor M (2007). Genetics of  
694 incipient speciation in *Drosophila mojavensis*. I. Male courtship song, mating success, and  
695 genotype x environment interactions. *Evolution* **61**: 1481–1488.
- 696 Ritchie M, Saarikettu M, Livingstone S, Hoikkala A (2001). Characterization of female  
697 preference functions for *Drosophila montana* courtship song and a test of the temperature  
698 coupling hypothesis. *Evolution* **55**: 721–727.
- 699 Robinson MR, Santure AW, DeCauwer I, Sheldon BC, Slate J (2013). Partitioning of genetic  
700 variation across the genome using multimarker methods in a wild bird population.  
701 *Molecular Ecology* **22**: 3963–3980.
- 702 Rockman MV (2012). The QTN program and the alleles that matter for evolution: All that’s  
703 gold does not glitter. *Evolution* **66**: 1–17.
- 704 Routtu J, Mazzi D, der Linde Van K, Mirol P, Butlin RK, Hoikkala A (2007). The extent of  
705 variation in male song, wing and genital characters among allopatric *Drosophila montana*  
706 populations. *J Evol Biol* **20**: 1591–1601.
- 707 Rundle, H. D., Blows, M. W. & Chenoweth, S. F. The diversification of mate preferences by  
708 natural and sexual selection. *Journal of Evolutionary Biology* **22**, 1608–1615 (2009).
- 709 Saarikettu M, Liimatainen JO, Hoikkala A (2005b). Intraspecific variation in mating behaviour  
710 does not cause sexual isolation between *Drosophila virilis* strains. *Animal Behaviour* **70**:  
711 417–426.
- 712 Safran RJ, Scordato ESC, Symes LB, Rodríguez RL, Mendelson TC (2013). Contributions of  
713 natural and sexual selection to the evolution of premating reproductive isolation: a  
714 research agenda. *Trends Ecol Evol (Amst)*.
- 715 Salminen T, Vesala L, Hoikkala A (2012). Photoperiodic regulation of life-history traits before  
716 and after eclosion: egg-to-adult development time, juvenile body mass and reproductive

- 717 diapause in *Drosophila montana*. *J Insect Physiol* **58**: 1541–1547.
- 718 Sambrook J, Russell D (2001). *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor  
719 Laboratory Press.
- 720 Santure AW, De Cauwer I, Robinson MR, Poissant J, Sheldon BC, Slate J (2013). Genomic  
721 dissection of variation in clutch size and egg mass in a wild great tit (*Parus major*)  
722 population. *Molecular Ecology* **22**: 3949–3962.
- 723 Schafer M, Mazzi D, Klappert K, Kauranen H, Vieira J, Hoikkala A, *et al.* (2010). A  
724 microsatellite linkage map for *Drosophila montana* shows large variation in recombination  
725 rates, and a courtship song trait maps to an area of low recombination. *J Evol Biol* **23**: 518–  
726 527.
- 727 Scoville AG, Lee YW, Willis JH, Kelly JK (2011). Explaining the heritability of an ecologically  
728 significant trait in terms of individual quantitative trait loci. *Biology Letters* **7**: 896–898.
- 729 Slate J (2005). Quantitative trait locus mapping in natural populations: progress, caveats and  
730 future directions. *Molecular Ecology* **14**: 363–379.
- 731 Slate J (2008). Robustness of linkage maps in natural populations: a simulation study. *Proc*  
732 *Biol Sci* **275**: 695–702.
- 733 Slate J (2013). From beavis to beak colour: A simulation study to examine how much QTL  
734 mapping can reveal about the genetic architecture of quantitative traits. *Molecular*  
735 *Ecology*: no–no.
- 736 St Pierre SE, Ponting L, Stefancsik R, McQuilton P, FlyBase Consortium (2014). FlyBase  
737 102--advanced approaches to interrogating FlyBase. *Nucleic Acids Res* **42**: D780–8.
- 738 Stern D, Orgogozo V (2008). The loci of evolution: How predictable is genetic evolution?  
739 *Evolution* **62**: 2155–2177.
- 740 Stern D, Orgogozo V (2009). Is genetic evolution predictable? *Science* **323**: 746–751.
- 741 Suvanto L, Liimatainen JO, Hoikkala A (1999). Variability and evolvability of male song  
742 characters in *Drosophila montana* populations. *Hereditas* **130**: 13–18.
- 743 The Marie Curie SPECIATION Network (2012). What do we need to know about speciation?  
744 *Trends Ecol Evol (Amst)* **27**: 27–39.
- 745 Throckmorton LH (1982). 15. The virilis species group. In: Ashburner M, Carson HL,  
746 Thompson JN (eds) *The Genetics and Biology of Drosophila*, Academic Press: London, pp  
747 227–289.
- 748 Travisano M, Shaw RG (2013). Lost in the map. *Evolution* **67**: 305–314.
- 749 Veltsos P, Wicker-Thomas C, Butlin RK, Hoikkala A, Ritchie M (2012). Sexual selection on  
750 song and cuticular hydrocarbons in two distinct populations of *Drosophila montana*.  
751 *Ecology and Evolution* **2**: 80–94.
- 752 Vesala L, Hoikkala A (2011). Effects of photoperiodically induced reproductive diapause and  
753 cold hardening on the cold tolerance of *Drosophila montana*. *J Insect Physiol* **57**: 46–51.
- 754 Voorrips RE (2002). MapChart: software for the graphical presentation of linkage maps and  
755 QTLs. *J Hered* **93**: 77–78.
- 756 Wittkopp PJ, Stewart EE, Arnold LL, Neidert AH, Haerum BK, Thompson EM, *et al.* (2009).  
757 Intraspecific polymorphism to interspecific divergence: Genetics of pigmentation in  
758 *Drosophila*. *Science* **326**: 540–544.

