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**FATORES PREDITIVOS DE RECÉM-NASCIDO EM CICLOS DE
MICROINJEÇÃO INTRACITOPLASMÁTICA COM
ESPERMATOZÓIDES DO EJACULADO**

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ARTIGO

Clinical outcome predictors in ICSI cycles with ejaculated sperm: report on 3844 consecutive treatment cycles

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Abstract

Background: Although numerous studies have been undertaken to establish predictive clinical factors associated with a higher likelihood of attaining a live birth, there is still no consensus, and its generally discussed only specific factors. The aim of the present study was to assess whether any clinical characteristics in intracytoplasmic sperm injection cycles are associated with an increased likelihood of live birth delivery.

Methods: Retrospective evaluation of 3844 treatment cycles with intracytoplasmic sperm injection using homologous ejaculated sperm. The study was conducted during 11 years (from 2003 to 2014). Cycles were divided in two groups according to live birth delivery outcomes: 993 cycles (25.8%) with newborn and 2851 cycles (74.2%) without newborn. Comparisons were performed regarding detailed demographic, stimulation, embryologic and clinical characteristics.

Results: A higher live birth delivery rate was associated to lower age and lower basal follicle stimulating hormone levels; lower total dose of gonadotropins, time of stimulation and dose of human chorionic gonadotropin; higher number of follicles and estradiol levels; higher number of aspirated oocytes and mature oocytes, higher rates of embryo cleavage, high quality embryos and blastocysts.

Conclusions: Several indicators appeared associated with an increase in the likelihood of live birth, eliciting the determination of cut-offs for female age, time of infertility, basal follicle stimulating hormone, estradiol levels, number of follicles, retrieved oocytes, mature oocytes, high quality embryos, embryos transferred and the day of embryo transfer. Antagonist and agonist protocols, as well as recombinant follicle stimulating hormone and human menopausal gonadotropin, gave similar outcomes, and thus the decision for one or another should be patient personalized. Spermogram values did not affect live birth. These findings are thus considered relevant for clinical guidance and patient information regarding the likelihood of live birth.

Keywords: Intracytoplasmic sperm injection, Assisted reproductive treatment predictors, Clinical outcomes, Newborn outcomes

Background

The possibility of fathering for patients with compromised spermogram values became a reality with the development of ICSI (Intracytoplasmic sperm injection) [Swain et al., 1992; van Steirteghem et al., 1993a,b; Tesarik and Sousa, 1995]. Although numerous studies have been undertaken to establish predictive clinical factors associated with a higher likelihood of attaining a live birth, there is still no consensus [van Loendersloot et al., 2010].

Regarding ovarian stimulation, the recent advances with gonadotropin releasing hormone (GnRH) antagonists and agonists elicited the development of personalized stimulation protocols [Huirne et al., 2007]. The subsequent appearance of ovarian hyperstimulation syndrome [Sousa et al., 2015] could later be reduced to near zero with GnRH agonist ovulation triggering, either associated with a strong luteal supplementation, or followed by embryo freezing for later embryo frozen thaw transfer [Humaidan et al., 2011]. In this context, several predictors have been raised, such as estradiol levels [Joo et al., 2010], progesterone levels [Kolibianakis et al., 2004], the number of follicles [Papanikolaou et al., 2006] and the number of oocytes aspirated [Steward et al., 2014]. Other success limiting factors appeared related to the embryo transfer technique [Schoolcraft, 2016], endometrial thickness [Kovaks et al., 2003], the number of embryos transferred [Pandian et al., 2013] and the day of embryo transfer [Glujosky et al., 2016].

Female infertility factors also limit assisted reproductive treatment (ART) outcomes, such as time of infertility [Niinimäki et al., 2015], karyotype anomalies, anatomic defects, tumors, tubar pathology, endocrinological disarrangements, polycystic ovarian syndrome, endometriosis, immunologic and thrombophilic conditions and body mass index [Rizk et al., 2008]. Another female limiting factor is ovary aging. The ovarian reserve can be predicted based on basal FSH (follicle-stimulating hormone) levels [van Loendersloot et al., 2010], anti-Mullerian hormone levels [Reijnders et al., 2016] and antral follicle count [Fleming et al., 2015]. However, ovary aging also conditions oocyte quality as, after follicle retrieval, oocytes may be immature [Beall et al., 2010] or dimorphic [Rienzi et al., 2011; Sousa et al., 2016]. Morphological inspection of the morphological oocyte is also a critical predictive factor, and the use of zona pellucida birefringence and metaphase II plaque positioning, as determined by specific optics [Montag et al., 2011], can improve oocyte selection.

Male infertility factors can also limit ART outcomes, such as karyotype anomalies, Y chromosome microdeletions, anatomic defects, tumors, endocrinological disarrangements, epididymis and vasa deferent occlusion, and spermatogenesis failure [Nieschlag et al., 2010]. In this context, spermogram parameters have critical predictive importance [Mahat et al., 2016; Pereira et al., 2016], and several methods were developed to determine sperm aneuploidy [Nicopoullos et al., 2008], apoptosis [Almeida et al., 2005] and DNA fragmentation rates [Sá et al., 2015] to guide clinical decisions. Additionally, several methods were introduced to selectively isolate the best sperm for microinjection, such as the use of specific optics [Perdrix and Rives, 2013] and magnetic cell sorting [Bucar et al., 2015].

Other critical aspects that can influence ART outcomes include the optimization of laboratory ART rooms and personnel qualification [Magli et al., 2008; De los Santos et al., 2016], ART technical aspects [Rubino et al., 2016], the number of metaphase II oocytes, fertilization, embryo cleavage and the number of high quality embryos at day 3 [Sousa and Tesarik, 1994; Stoop et al., 2012]. Other important factors have been consistently improved, such as those related to embryo cryopreservation, amelioration of embryo culture media, extended embryo culture up to the blastocyst stage [Gardner and Lane, 2000; Swain and Smith, 2011; Sousa et al., 2012] and embryo dynamics screening [Rubio et al., 2014]. Finally, the use of preimplantation genetic diagnosis [Cooper and Jungheim, 2010; Mastenbroek et al., 2011] enabled couples not only to have their children free of diseases but also enabled to find and counteract the causes of implantation failures.

In order to give patients more reliable information and to clinicians better guidelines on the likelihood of attaining a live birth, we analysed 3844 ICSI cycles to uncover critical factors, with suggested cut-offs, using detailed demographic, stimulation and embryological data.

Methods

Ethics approval

According to the National Law on Medically Assisted Procreation and the National Council on Medically Assisted Procreation guidelines, databases were used after patient informed and written consent.

Patients

Retrospective evaluation of 3844 treatment cycles using homologous ejaculated sperm and ICSI. Seropositive, anejaculation, retrograde ejaculation, preimplantation genetic diagnosis and Y chromosome microdeletions cases were excluded. The period of analysis was between 25-09-2003 and 17-01-2014 (11 years).

Karyotype analysis

Karyotypes were evaluated using G-banding with analysis of at least 30 metaphases from peripheral blood lymphocytes, according to general protocols [Rooney and Czepulkowski, 1997]. There were 10 Klinefelter cases (3 NB from 3 distinct cases).

Stimulation Protocol

Women underwent controlled ovarian hyperstimulation with a GnRH antagonist protocol in 92.2% of the cases (Merck Serono, Geneva, Switzerland; Organon, Oss, Netherlands) and with an agonist protocol in 7.8% of the cases (Sanofi Aventis, Frankfurt, Germany). For stimulation, recombinant follicle stimulating hormone (rFSH) was used in 56.4% of the cases (Organon; Merck Serono); human menopausal gonadotropin (HMG) was added to rFSH in 42.9% of the cases (Ferring, Kiel, Germany); and only HMG in 0.7% of the cases. Ovulation trigger was performed with human chorionic gonadotropin (HCG) in 79.8% of the cases (Organon); with recombinant HCG in 18.9% of the cases (Merck-Serono); and with a GnRH agonist in 1.3% of the cases (Sanofis, Aventis, Frankfurt, Germany; Ipsen Pharma Biotech, Signes France). Estradiol serum levels were assayed at the day of HCG or one day before [Huirne et al., 2007; Pinto et al., 2009].

Gamete and Embryo Handling

Media from Medicult (Jyllinge, Denmark) or Vitrolife (Kungsbacka, Sweden) were used. Microinjection was performed in a Hoffman optics inverted microscope (Nikon, Tokyo, Japan) equipped with Narishige micromanipulators (Tokyo, Japan), using Swemed micropipettes (Goteborg, Sweden) and under a described technique [Tesarik and Sousa, 1995]. Embryo quality was evaluated according to described methods [Vandervorst et al., 1998; Gardner et al., 2000]. Ultrasound-guided embryo transfer used a Sure View Wallace Embryo Replacement Catheter or Wallace malleable stylet (Smiths Medical Int, Kent, UK).

Luteal Supplementation

All patients had luteal supplementation with intravaginal administration of natural-micronized progesterone (Jaba, Besins Int, Montrouge, France; Effik, Bievres, France), from the day of oocyte retrieval. Implantation was confirmed by a rise in β -HCG serum twelve days after embryo transfer. Clinical pregnancy was established by ultrasound at seven weeks of gestation. When a GnRH agonist was used for triggering final oocyte maturation, luteal support was associated with oral estradiol (Novo Nordisk, Bagsvaerd, Denmark) and HCG (1500 IU) at the day of oocyte pick-up [Radesic and Tremellen, 2011].

Statistical Analysis

For statistical analysis the *IBM SPSS Statistics 22* program for Windows was used. Means were compared using t-Test for independent samples. Categorical variables were analysed using descriptive statistics and Chi square test, with continuity correction. All statistical tests were two-tailed, with significance level of 0.05 ($p < 0.05$).

Results

Patients and study groups

The 3844 ICSI cycles were divided in two groups according to their outcomes and compared regarding demographic, stimulation, embryologic, clinical and newborn (NB) characteristics: 993 cycles (25.8%) with NB and 2851 cycles (74.2%) without NB.

Demographic characteristics

Regarding demographic characteristics (Table 1), the NB group showed a significant lower mean female and male ages, a similar time of infertility, a significant higher number of cases with pure male factor and a significant lower number of cases with pure female and mixed infertility factors, and a significant lower mean level of basal FSH (bFSH). There were no significant differences regarding female and male karyotypes. The prevalence of chromosome abnormalities in the general population is about 0.5-1% [Turnpenny and Ellard, 2012]. The present results thus confirm that the rate of abnormal karyotypes is higher in infertile individuals [Sousa et al., 2016; Gekas et al., 2001].

Stimulation characteristics

Regarding stimulation characteristics (Table 1), the NB group manifested a significant lower mean total dose of gonadotropins and HCG used; a lower mean time of stimulation; a higher mean number of developed ovarian follicles; and higher mean serum estradiol (E2) levels.

Embryological and clinical outcomes

As expected, regarding embryological and clinical outcomes, the NB group (Table 2) showed significant higher mean values for all parameters analysed, which included the number of oocytes retrieved (COC), mature metaphase II oocytes (MII), fertilization rate (FR), embryo cleavage rate, high grade embryos at day 3, blastocyst rate, implantation rate (IR), biochemical (BP), clinical (CP) and ongoing (OP) pregnancy rates, without any cases of ectopic pregnancy or abortion.

Newborn outcomes

The characteristics of NB are given in Table 2 and Table 3. The rate of NB malformations was 1.3%, 1% major and 0.3% minor (Table 2). These rates are quite lower regarding population studies that revealed 2-5% of major and 10% of minor NB malformations [Turnpenny and Ellard, 2012]. About 80% of the NB had a term gestation and 65% a normal weight (Table 3).

Subgroup analysis

To uncover some thresholds on the likelihood of achieving a live birth deliver (LBD), several characteristics were analysed: female age, time of infertility, stimulation protocol, gonadotropins, follicles, number of retrieved COC, mature MII, high quality embryos, number of embryos transferred, day of embryo transfer, and sperm concentration, rapid progressive motility and normal morphology (Figs. 1-3).

The female age was divided into six stages (Fig. 1). The NB group showed a significant lower number of cases with ≥ 40 years, and no significant differences were observed in the younger stages. Most of the successful cases were found in women with < 35 years (61%) and < 39 years (89%) (Fig. 1 left). Results evidenced a continuous decrease in the likelihood of LBD with advancing female age, with the higher likelihood of LBD belonging to the age stages of < 30 years and 30-35 years (Fig. 1 right).

The time of infertility was divided into four stages (Fig. 1). No significant differences were observed except when the time of infertility was < 5 years (Fig. 1 left). Results evidenced a continuous decrease in the likelihood of getting a successful pregnancy with advancing time of infertility, with the higher likelihood of achieving a LBD being observed with < 5 years of infertility (Fig. 1 right).

Regarding the stimulation protocol the differences found were significant. The antagonist protocol was used in 90% of the cycles in the NB group and in 93% of the cycles in the no-NB group; whereas the agonist protocol was used in 10% of the cycles in the NB group and in 7% of the cycles in the no-NB group 187 (Fig. 1 left). Results evidenced a higher likelihood of attaining a LBD with the agonist protocol (Fig. 1 right).

In relation to gonadotropins, the NB group evidenced a significant higher number of cases that used only rFSH and a significantly lower number of cases where both rFSH and HMG were used, with no significant differences regarding the sole use of HMG (Fig. 1 left). There was a decrease in the likelihood of achieving a LBD from the use of rFSH, to rFSH+HMG and then to HMG, with the highest likelihood of LBD being observed in cases where only rFSH was used (Fig. 1 right).

For the analysis of follicles we divided cycles in 6 intervals (Fig. 1). Most of the successful cases presented >4 follicles (Fig. 1 left). The likelihood of a successful LBD increased with the number of follicles, being significantly lower when <4 follicles and between 4-9 follicles, and higher between 10-13 and 14-18 follicles (Fig. 1 right).

The number of retrieved COC was divided in six intervals (Fig. 2). The NB group showed a significant higher number of cases with 4-9 COC and 4-15 COC, a significant lower number of cases with ≤ 3 COC, and no significant differences when there were >10 COC retrieved, with most of the cases with NB presenting 4-15 COC (Fig. 2 left). There was a lower likelihood of LBD with ≤ 3 COC and a progressive higher likelihood of LBD up to 15 COC retrieved (Fig. 2 right).

The number of MII was divided in six intervals (Fig. 2). The NB group presented 4-9 MII in the majority of the cases, a significant lower number of cases with ≤ 3 MII and a significant higher number of cases with 4-15 MII, with no significant differences in the other intervals (Fig. 2 left). The likelihood of LBD was lower with ≤ 3 MII and progressively higher up to 15 MII (Fig. 2 right).

The number of high quality embryos at day 3 (AB) was divided in two groups (Fig. 2). The NB group showed a significant lower number of cases with <3 AB and a significant higher number of cases with ≥ 3 AB (Fig. 2 left). The likelihood of LBD was lower when <3 high quality embryos were obtained and higher in cases with ≥ 3 AB (Fig. 2 right).

The number of embryos transferred was divided in four groups (Fig. 2). The NB group evidenced a significant lower number of cases with transfer of 1 or 3 embryos, and a significant higher number of cases with transfer of 2 embryos (Fig. 2 left). The likelihood of LBD decreased with the number of embryos transferred, being higher with transfer of 2 embryos (Fig. 2 right).

The day of embryo transfer was divided in five groups (Fig. 2). The NB group showed a significant higher number of cases with embryo transfer performed at day 5, and a significant lower number of cases with embryo transfer performed at day 2 and day 3 (Fig. 2 left). The likelihood of LBD increased with the day of embryo transfer, and was higher at day 5 (Fig. 2 right).

Sperm concentration, rapid progressive motility and normal morphology were divided in five intervals (Fig. 3). For normal morphology, although the NB group presented ≥ 20 M sperm/ml in the majority of the cases, with significance attained with ≥ 5 M sperm/ml (Fig. 3 left), the likelihood of LBD decreased with sperm concentration (Fig. 3 right). For rapid progressive motility, although in the NB group the majority of cases presented $\geq 10\%$, with significance attained when $\geq 5\%$ (Fig. 3 left), the likelihood of LBD decreased with sperm rapid progressive motility (Fig. 3 right). For normal morphology, although the NB presented $\geq 5\%$ in the majority of the cases, with significance observed when <5% (Fig. 3 left), the likelihood of LBD was independent of sperm normal morphology (Fig. 3 right). Regarding clinical subgroups, although the NB group showed a significant higher number of teratozoospermia cases (Fig. 3 left), the likelihood of LBD was independent of clinical spermogram classifications (Fig. 3 right).

Discussion

Several factors can influence pregnancy outcomes in ICSI cycles with ejaculated sperm. In this report of 3,844 cycles the LBD was associated with lower couple ages, time of infertility, bFSH levels, time of stimulation, total dose of gonadotropins and HCG dose, and with a higher mean number of follicles and E2 levels. Regarding embryological and clinical outcomes, LBD was associated with a higher mean number of COC, MII, cleaved embryos, high quality embryos and blastocysts. Bellow, the relative predictive value of the main factors analysed in the present report are discussed.

Regarding **female age**, the present results showed a decline in the likelihood of LBD after 30 years, with significant differences found in women < 35 years, < 39 years and \geq 40 years. Results confirm previous studies, albeit with a lower number of cycles. One study showed that the likelihood of CP decreased progressively with female age [Ottosen et al., 2007]. Another work evidenced that the chance of achieving a CP in women < 30 years is 3.2 higher, between 30-34 years is 2.8 higher and with < 35 years is 2.5 higher [Sabatini et al., 2008]. Another study showed that the CP rate increases up to 30 years, then slowly decreases and finally sharply decreases when \geq 40 years. In that report, women of 30 years were 1.5 times more likely to become pregnant, women with 35 years had a similar chance, and 40 years old women had 4 times lower expectancy [Pinto et al., 2009]. Finally, a meta-analysis indicated that increasing female age was associated with lower CP rates [van Loendersloot et al., 2010]. Taking into account those observations and results of our large series, female age can be used as a predictive factor for LBD, with an upper cut-off of 35 years.

In relation to the **time of infertility**, results showed a decline in the likelihood of LBD after 2 years of infertility. In accordance, though with a lower number of cycles, other authors observed that the likelihood of CP decreased after 1 year of infertility [Ottosen et al., 2007]. A meta-analysis also confirmed that OP rates decreased with the time of infertility [van Loendersloot et al., 2010]. Based on results from our large series and on those cited above, the time of infertility can be used as predictive factor for LBD, with a positive cut-off of < 2 years and a negative cut-off of > 5 years. The levels of **bFSH** were significantly lower in the NB group, which reflects the higher capacity of the ovary for follicle growth in younger women. Several previous studies, regardless of a lower number of cycles, showed that when bFSH levels are > 10 IU/L, the embryological, clinical and NB outcomes are significantly decreased [Ottosen et al., 2007; Sabatini et al., 2008; Abdalla and Thum., 2004; Thum et al, 2009]. Based on the present results, on those studies cited above and on a recent meta-analysis [van Loendersloot et al., 2010], bFSH levels can be used as a predictive factor of LBD, with an upper cut-off of 10 IU/L.

The **GnRH antagonist protocol** was used in the large majority of the cases, but the likelihood of LBD significantly favoured the agonist protocol. Previous studies showed that the antagonist protocol was associated with lower time of stimulation, higher number of COC, MII, high quality embryos and embryo cleavage rate, whereas the agonist protocol exhibited a higher FR, with no significant differences between both protocols regarding the rates of BP, CP, implantation and LBD [Pinto et al., 2009]. Although there is a general consensus that the antagonist protocol is devoid of side effects, enables lower time of stimulation, gonadotrophin dose and risk of ovarian hyperstimulation syndrome (**OHSS**), other reports debate that the agonist protocol is associated with better embryological and clinical outcomes [Kolibianakis et al., 2006; Al-Inany et al, 2016]. Taking into account these previous studies, differences between both protocols should be considered marginal and clinically irrelevant, and the decision of using one or another stimulation protocol should rely on individual patient characteristics.

Regarding **gonadotropin stimulation**, results showed that the use of rFSH is significantly associated with higher LBD than the combination of rFSH with HMG or the use of HMG. Previous studies revealed distinct results. Cochrane reviews indicated that HMG was associated with higher rates of CP and LBD than rFSH [van Wely et al., 2011]. Another report showed that although HMG was associated with higher female age, bFSH levels and total dose of gonadotropins, and lower E2 levels, COC, MII, cleaved embryos and high quality embryos, it exhibited similar rates of implantation, CP and LBD to rFSH [Shavit et al., 2016]. Based on results and those of the above reports, differences between rFSH and HMH should be considered marginal and clinically irrelevant, and the decision of using one or another stimulation protocol should rely on individual patient characteristics.

The **levels of E2** were significantly higher in the NB group, which was associated with a higher number of follicles. Previous studies showed that although the CP rate was not affected by E2 levels of normal responders (<2500 pg/ml) or high responders (>2500 pg/ml) [Papageorgiou et al., 2002], the rates of LBD increased with E2 levels, with the better LBD rates being achieved with 3000-4000 pg/ml E2 for women <38 years and 279 2000-3000 pg/ml E2 for women >38 years [Joo et al., 2010]. This is in line with the defined upper cut-off for an increased risk of OHSS, which is 4500 pg/ml [Sousa et al., 2015]. Thus, E2 levels can be used as a predictive factor for LBD, with a cut-off of 2000-4000 pg/ml. In relation to the **mean number of follicles**, results suggested a lower likelihood of LBD with ≤ 4 follicles and a progressive higher likelihood of LBD from 4 to 18 follicles, with the maximized LBD attained in the interval 14-18 follicles. Although unable to find similar studies for comparisons, an upper limit of 14 follicles and a lower cut-off of 4-9 follicles are suggested. The number of follicles is related to the development of OHSS. In the NB group, of the 4 cases (0.4%) where an agonist was used for oocyte trigger due to risk of OHSS, 2 cases had > 18 follicles. However, in the no-NB group, of the 43 cases (1.5%) that used an agonist, 32 (74.4%) had >18 follicles, and with ≤ 14 follicles this risk decreased to 7 cases (16%). These results are reinforced by previous studies that showed an increased risk of OHSS when the number of follicles was >18 [Sousa et al., 2015; Papanikolaou et al., 2006]. The **mean number of retrieved cumulus-oocyte complexes** has been used to define women as poor (≤ 3 COC), normal (4-9 COC) or high (≥ 15 COC) responders [Polyzos et al., 2014; Polyzos and Sunkara, 2015], with the observance of an increased risk of OHSS when ≥ 15 COC were retrieved [Sousa et al., 2015]. The present results confirmed the low likelihood of LBD when ≤ 3 COC were retrieved and a higher likelihood of LBD from 4 to 15 COC, with the maximized LBD attained in the interval 10-15 COC. This is in accordance with cut-off suggested by other authors: 15 COC [Sunkara et al., 2011], 8-15 COC [Cai et al., 2013] and 10-15 COC [Steward et al., 2014]. Results suggest that the mean number of follicles can be used as a predictive factor for LBD, with a cut-off of 4-15 COC.

In relation to the **number of MII**, the present results confirmed the low likelihood of LBD when ≤ 3 MII were obtained, and a progressive higher likelihood of LBD from 4 to 15 MII, with the maximized LBD attained in the interval 10-15 MII. Although unable to find similar studies for comparison, previous studies evidenced that the number of MII is positively correlated with LBD [Cai et al., 2011]. Results suggest that the mean number of MII can be used as a predictive factor for LBD, with a cut-off of 4-15 MII. Previous studies revealed that the **number of high quality embryos** at day 3 is a critical predictive factor, but a relevant cut-off was not offered. Those authors observed that the transfer of high quality embryos increased the CP rate [Ottoen et al., 2007; Cai et al., 2011], the rates of CP and LBD [Dennis et al., 2006], or the LBD rate [Niinimäki et al., 2015]. The present results showed that the likelihood of LBD increased with the number of high quality embryos, being higher when ≥ 3 , which indicates that the number of high quality embryos can be used as a predictive factor for LBD, with a cut-off of ≥ 3 high quality embryos.

In relation to the **number of embryos transferred**, the present results showed the transfer of 1 or 3 embryos was associated with a significant lower LBD, with the likelihood of LBD being higher when 2 embryos were transferred. Results also revealed that single blastocyst transfer was associated with no multiple pregnancies and 35% of LBD, whereas double blastocyst transfer presented 20% of twin pregnancies and 44% of LBD. On the other hand, single embryo transfer at day 3 was associated with no multiple pregnancies and 17% of LBD, whereas double embryo transfer at day 3 presented 9% of twin pregnancies and 30% of LBD. This confirms previous studies, which indicated that although single embryo transfers decreased the LBD rate it also reduced the risk of multiple pregnancies, and that with the subsequent replacement of a single frozen embryo the LBD rate became comparable to double embryo transfer [Pandian et al., 2013; Baruffi et al., 2009; Martikainen et al., 2001; Pandian et al., 2005]. The present results and those of previous studies evidence that the number of embryos transferred can be used as a predictive factor for LBD, with a cut-off of 1-2 embryos. Results also indicate that in cases of blastocyst transfer a single embryo transfer should be the first choice. The clinical decision on transfer of two embryos at day 2 or 3 is frequently dependent on the lower LBD, as 2-3 subsequent replacements of a single frozen embryo would be needed to give patients an acceptable LBD per cycle.

Regarding the **day of embryo transfer**, the present results showed a significant increase in the likelihood of LBD with the day of embryo transfer, with the maximum being attained at day 5. This confirms previous reports of randomized controlled clinical trials [Papanikolaou et al., 2008] and Cochrane reviews [Glujosky et al., 2016], which showed higher LBD with blastocyst transfer. The present results and those of previous studies evidence

that the day of embryo transfer can be used as a predictive factor for LBD, and suggest that the preferable day for embryo transfer is day 5, as blastocysts elicit a significant higher LBD rate.

Evaluation of **sperm parameters** revealed that sperm concentration, rapid progressive motility and morphology had no clinical impact in the likelihood of achieving a LBD. These observations confirm previous studies, which showed that severe teratozoospermia [French et al., 2010] or decreased sperm concentration and progressive motility [Zheng et al., 2016] had no negative impact on clinical outcomes. Those studies suggested that these apparently strange observations might be due to the strict sperm preparation method and selection for microinjection. Nevertheless, from the present results and those of previous studies it is indicated that sperm parameters have no negative impact on clinical outcomes and cannot be used as a predictive factor for LBD.

Conclusions

The present results, using a large number of cases (3844 cycles) with microinjection of ejaculated sperm, indicated that LBD is associated with several demographic and stimulation variables such as younger age, more pure male infertility factors, lower bFSH, total gonadotropin dose, time of stimulation and HCG dose, and higher number of follicles and estradiol levels. Regarding embryological outcomes, the LBD was associated with significantly higher mean number of COC, MII, cleaved embryos, high quality embryos and blastocysts. Data from the present results and that of previous studies suggest the following indicators (cut-off) that appeared associated with an increased likelihood of LBD: female age ≤ 35 years; time of infertility ≤ 5 years (ideal < 2 years); bFSH ≤ 10 IU/L; both the antagonist and agonist protocols, as well as rFSH and HMG, gave similar outcomes, and thus the choice should be individualised; E2 levels 2000-4000pg/ml; number of follicles ≥ 4 (ideal maximum of 14 follicles); number of COC ≥ 4 (ideal maximum of 15 COC); number of MII ≥ 4 (ideal maximum of 15 MII); number of high quality embryos ≥ 3 ; double embryo transfer at day 3, single embryo transfer at day 5; and no relation between spermiogram values and LBD. These findings are thus considered relevant for clinical guidance and patient information regarding the likelihood of LBD.

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References

- Abdalla H, Thum MY, 2004. An elevated basal FSH reflects a quantitative rather than qualitative decline of the ovarian reserve. *Hum Reprod.*;19:893-8. doi: 10.1093/humrep/deh141.
- Al-Inany HG, Youssef MA, Ayeleke RO, Brown J, Lam WS, Broekmans FJ, 2016. Gonadotrophin-releasing hormone antagonists for assisted reproductive technology (Review). *Cochrane Database of Syst Rev.*;4. Doi: 10.1002/14651858.CD001750.pub4.
- Almeida C, Cardoso MF, Sousa M, Viana P, Gonçalves A, Silva J, 2005. Quantitative study of caspase-3 activity in semen and after swim-up preparation in relation to sperm quality. *Hum Reprod.*;20:1307-13. doi: 10.1093/humrep/deh727.
- Baruffi RLR, Mauri AL, Petersen CG, Nicoletti A, Ponte A, Batista J, 2009. Single-embryo transfer reduces clinical pregnancy rates and live births in fresh IVF and intracytoplasmic sperm injection (ICSI) cycles: a meta-analysis. *Reprod Biol Endocrinol.*;7:36. Doi: 10.1186/1477-7827-7-36.
- Beall S, Brenner C, Segars J, 2010. Oocyte maturation failure: a syndrome of bad eggs. *Fertil Steril.*;94:2507-13. doi: 10.1016/j.fertnstert.2010.02.037.
- Bucar S, Gonçalves A, Rocha E, Barros A, Sousa M, Sá R, 2015. DNA fragmentation in human sperm after magnetic-activated Cell sorting. *J Assist Reprod Genet.*;32:147-54. doi: 10.1007/s10815-014-0370-5.
- Cai Q, Wan F, Huang K, Zhang H, 2013. Does the number of oocytes retrieved influence pregnancy after fresh embryo transfer? *PLoS One*;8:e56189. Doi: 10.1371/journal.pone.0056189.
- Cai QF, Wan F, Huang R, Zhang HW, 2011. Factors predicting the cumulative outcome of IVF/ICSI treatment: a multivariable analysis of 2,450 patients. *Hum Reprod.*;26:2532-40. doi: 10.1093/humrep/der228.
- Cooper AR, Jungheim ES, 2010. Preimplantation genetic testing: indications and controversies. *Clin Lab Med.*;30:519-31. doi: 10.1016/j.cll.2010.04.008.
- De los Santos MJ, Apter S, Coticchio G, Debrock S, Lundin K, Plancha CE, 2016. Revised guidelines for good practice in IVF laboratories (2015). *Hum Reprod.*;31:685-6. doi: 10.1093/humrep/dew016.
- Dennis SJ, Thomas MA, Williams DB, Robins JC, 2006. Embryo morphology score on day 3 is predictive of implantation and live birth rates. *J Assist Reprod Genet.*;23:171-5. doi: 10.1007/s10815-006-9027-3.
- Fleming R, Seifer DB, Frattarelli JL, Ruman J, 2015. Assessing ovarian response: antral follicle count versus anti-Müllerian hormone. *Reprod Biomed Online.*;31:486-96. doi: 10.1016/j.rbmo.2015.06.015.
- French DB, Sabanegh ES, Golfarb J, Desai N, 2010. Does severe teratozoospermia affect blastocyst formation, live birth rate, and other clinical outcome parameters in ICSI cycles? *Fertil Steril.*;93:1097-103. doi: <http://dx.doi.org/10.1016/j.fertnstert.2008.10.051>.
- Gardner DK, Lane M, Stevens J, Schlenker T, Schoolcraft WB, 2000. Blastocyst score affects implantation and pregnancy outcome: towards a single blastocyst transfer. *Fertil Steril.*;73:1155-8. doi: [http://dx.doi.org/10.1016/S0015-0282\(00\)00518-5](http://dx.doi.org/10.1016/S0015-0282(00)00518-5).
- Gardner DK, Lane M, 2000. Embryo culture systems. In: Trounson AO, Gardner DK, eds. *Handbook of in vitro fertilization*. New York. CRS Press. 205-64. ISBN: 0849340020 9780849340024.
- Gekas J, Thepot F, Turleau C, Siffroi JP, Dadoune JP, Wasels R, 2001. Chromosomal factors of infertility in candidate couples for ICSI: an equal risk of constitutional aberrations in women and men. *Hum Reprod.*;16:82-90. doi: 10.1093/humrep/16.1.82.
- Glujovsky D, Farquhar C, Quinteiro Retamar AM, Alvarez Sedo CR, Blake D, 2016. Cleavage stage versus blastocyst stage embryo transfer in assisted reproductive technology (Review). *Cochrane Database of Syst Rev.*;6. Doi: 10.1002/14651858.CD002118.pub5.
- Huirne JA, Homburg R, Lambalk CB, 2007. Are GnRH antagonists comparable to agonists for use in IVF? *Hum Reprod.*;22:2805-13. doi: 10.1093/humrep/dem270.

- Humaidan P, Kol S, Papanikolaou EG, on behalf of the Copenhagen GnRH agonist triggering workshop group, 2011. GnRH agonist for triggering of final oocyte maturation: time for a change of practice? *Hum Reprod Update.*;17:510-24. doi: 10.1093/humupd/dmr008.
- Joo BS, Park SH, An BM, Kim KS, Moon SE, Moon HS, 2010. Serum estradiol levels during controlled ovarian hyperstimulation influence the pregnancy outcome of in vitro fertilization in a concentration-dependent manner. *Fertil Steril.*;93:442-6. doi: 10.1016/j.fertnstert.2009.02.066.
- Kolibianakis EM, Zikopoulos K, Smitz J, Camus M, Tournaye H, Van Steirteghem AC, 2004. Elevated progesterone at initiation of stimulation is associated with a lower ongoing pregnancy rate after IVF using GnRH antagonists. *Hum Reprod.*;19:1525-9. doi: 10.1093/humrep/deh272.
- Kolibianakis EM, Collins J, Tarlatzis BC, Devroey P, Diedrich K, Griesinger G, 2006. Among patients treated for IVF with gonadotrophins and GnRH analogues, is the probability of live birth dependent on the type of analogue used? A systematic review and meta-analysis. *Hum Reprod Update.*;12:651-71. doi: 0.1093/humupd/dml038.
- Kovacs P, Matyas Sz, Boda K, Kaali SG, 2003. The effect of endometrial thickness on IVF/ICSI outcome. *Hum Reprod.*;18:2337-41. doi: 10.1093/humrep/deg461.
- Magli MC, Van den Abbeel E, Lundin K, Royere D, Van der Elst J, Gianaroli L, 2008. Revised guidelines for good practice in IVF laboratories. *Hum Reprod.*;23:1253-62. doi: 10.1093/humrep/den068.
- Mahat RK, Arora M, Bhale DV, Holkar S, Kumar S, Yadav T, 2016. Risk factors and causes of male infertility-a review. *Biochem Anal Biochem.*;5:2. Doi: org/10.4172/2161-1009.1000271.
- Martikainen H, Tiitinen A, Tomás C, Tapanainen J, Orava M, Tuomivaara L, 2001. One versus two embryo transfer after IVF and ICSI: a randomized study. *Hum Reprod.*;16:1900-3. doi: 10.1093/humrep/16.9.1900.
- Mastenbroek S, Twisk M, van der Veen F, Repping S, 2011. Preimplantation genetic screening: a systematic review and meta-analysis of RCTs. *Hum Reprod Update.*;17:454-66. doi: 10.1093/humupd/dmr003.
- Montag M, Köster M, van der Ven K, van der Ven H, 2011. Gamete competence assessment by polarizing optics in assisted reproduction. *Hum Reprod Update.*;17:654-66. doi: 10.1093/humupd/dmr016.
- Nicopoulos JDM, Gilling-Smith C, Almeida PA, Homa S, Nice L, Tempeste H, 2008. The role of sperm aneuploidy as a predictor of the success of intracytoplasmic sperm injection. *Hum Reprod.*;23:240-50. doi: 10.1093/humrep/dem395.
- Nieschlag E, Behre HM, Nieschlag S, 2010, eds. *Andrology. Male reproductive health and dysfunction*. Berlin. Springer-Verlag. ISBN: 978-3-540-78355-8.
- Niinimäki M, Veleva Z, Martikainen H, 2015. Embryo quality is the main factor affecting cumulative live birth rate after elective single embryo transfer in fresh stimulation cycles. *Eur J Obstet Gynecol Reprod Biol.*;194:131-5. doi: 10.1016/j.ejogrb.2015.08.031.
- Ottosen LDM, Kesmodel U, Hindkjaer J, Ingerslev HJ, 2007. Pregnancy prediction models and aSET criteria for IVF patients – do we need more information? *J Assist Reprod Genet.*;24:29-36. doi: 10.1007/s10815-006-9082-9
- Palermo G, Joris H, Devroey P, Van Steirteghem AC, 1992. Pregnancies after intracytoplasmic injection of single spermatozoon into an oocyte. *Lancet*;340:17-18. doi: http://dx.doi.org/10.1016/0140-6736(92)92425-F.
- Pandian Z, Templeton A, Serour G, Bhattacharya S, 2005. Number of embryos for transfer after IVF and ICSI: a Cochrane review. *Hum Reprod.*;20:2681-7. doi: 10.1093/humrep/dei153.
- Pandian Z, Marjoribanks J, Ozturk O, Serour G, Bhattacharya S, 2013. Number of embryos for transfer following in vitro fertilization or intra-cytoplasmic sperm injection (Review). *Cochrane Database of Syst Rev.*;7. Doi: 10.1002/14651858.CD003416.pub4.
- Pageorgiou T, Guibert J, Goffinet F, Patrat C, Fulla Y, Janssens Y, 2002. Percentile curves of serum estradiol levels during controlled ovarian stimulation in 905 cycles stimulated with recombinant FSH show that high estradiol is not detrimental to IVF outcome. *Hum Reprod.*;17:2846-50. doi: 10.1093/humrep/17.11.2846.

- Papanikolaou EG, Pozzobon C, Kolibianakis EM, Camus M, Tournaye H, Fatemi HM, 2006. Incidence and prediction of ovarian hyperstimulation syndrome in women undergoing gonadotropin-releasing hormone antagonist in in vitro fertilization cycles. *Fertil Steril.*;85:112-20. doi: 10.1016/j.fertnstert.2005.07.1292.
- Papanikolaou EG, Kolibianakis EM, Tournaye H, Venetis CA, Fatemi H, Tarlatzis B, 2008. Live birth rates after transfer of equal number of blastocysts or cleavage-stage embryos in IVF: a systematic review and meta-analysis. *Hum Reprod.*;23:91-9. doi: 10.1093/humrep/dem339.
- Perdrix A, Rives N, 2013. Motile sperm organelle morphology examination (MSOME) and sperm head vacuoles: state of the art in 2013. *Hum Reprod Update.*;19:527-41. doi: 10.1093/humupd/dmt021.
- Pereira R, Sá R, Barros A, Sousa M, 2016. Major regulatory mechanisms involved in sperm motility. *Asian J Androl.*;18:1-10. Doi: 10.4103/1008-682X.167716.
- Pinto F, Oliveira C, Cardoso MF, Teixeira-da-Silva J, Silva J, Sousa M, 2009. Impact of GnRH ovarian stimulation protocols on intracytoplasmic sperm injection outcomes. *Reprod Biol Endocrinol.*;7:5. Doi:10.1186/1477-7827-7-5.
- Polyzos NP, Nwoye M, Corona R, Blockeel C, Stoop D, Haentjens P, 2014. Live birth rates in Bologna poor responders treated with ovarian stimulation for IVF/ICSI. *Reprod Biomed Online.*;28:469-74. doi: 10.1016/j.rbmo.2013.11.010.
- Polyzos NP, Sunkara SK, 2015. Sub-optimal responders following controlled ovarian stimulation: an overlooked group? *Hum Reprod.*;30:2005-8. doi: 10.1093/humrep/dev149.
- Radesic B, Tremellen K, 2011. Oocyte maturation employing a GnRH agonist in combination with low-dose hCG luteal rescue minimizes the severity of ovarian hyperstimulation syndrome while maintaining excellent pregnancy rates. *Hum Reprod.* ;26:3437-42. doi: 10.1093/humrep/der333.
- Reijnders IF, Nelen WJDM, Int'Hout J, van Herwaarden AE, Braat DDM, Fleischer K, 2016. The value of Anti-Müllerian hormone in low and extremely low ovarian reserve in relation to live birth after in vitro fertilization. *Eur J Obstet Gynecol Reprod Biol.*;200:45-50. doi: 10.1016/j.ejogrb.2016.02.007.
- Rienzi L, Vajta G, Ubaldi F, 2011. Predictive value of oocyte morphology in human IVF: a systematic review of the Literature. *Hum Reprod Update.*;17:34-45. doi: 10.1093/humupd/dmq029.
- Rizk BRM, Garcia-Velasco JA, Sallam HN, Makrigiannakis A, 2008, eds. *Infertility and assisted Reproduction*. New York. Cambridge Univ Press. ISBN: 13: 9780521873796.
- Rooney DE, Czepulkowski BH, 1997, eds. *Human chromosome preparation. Essential techniques*. New Jersey. Wiley. ISBN: 0471962996.
- Rubino P, Viganò P, Luddi A, Piomboni P, 2016. The ICSI procedure from past to future: a systematic review of the more controversial aspects. *Hum Reprod Update.* 22:194-227. doi: 10.1093/humupd/dmv050.
- Rubio I, Galán A, Larreatequi Z, Ayerdi F, Bellver J, Herrero J, 2014. Clinical validation of embryo culture and selection by morphokinetic analysis: a randomized, controlled trial of the Embryoscope. *Fertil Steril.*;102:1287-94. doi: <http://dx.doi.org/10.1016/j.fertnstert.2014.07.738>.
- Sá R, Cunha M, Rocha E, Barros A, Sousa M, 2015. Sperm DNA fragmentation is related to sperm morphological staining patterns. *Reprod Biomed Online*;31:506-15. doi: 10.1016/j.rbmo.2015.06.019.
- Sabatini L, Zosmer A, Hennessy EM, Tozer A, Al-Shawaf T, 2008. Relevance of basal serum FSH to IVF outcome varies with patient age. *Reprod Biomed Online*;17:10-9. doi: [http://dx.doi.org/10.1016/S1472-6483\(10\)60287-8](http://dx.doi.org/10.1016/S1472-6483(10)60287-8).
- Schoolcraft WB, 2016. Importance of embryo transfer technique in maximizing assisted reproductive outcomes. *Fertil Steril.*;105:855-60. doi: 10.1016/j.fertnstert.2016.02.022.
- Shavit T, Shalom-Paz E, Samara N, Aslih N, Michaeli M, Ellenbogen A, 2016. Comparison between stimulation with highly purified hMG or recombinant FSH in patients undergoing IVF with GnRH antagonist protocol. *Gynecol Endocrinol.*;1-5. Doi: [org/10.3109/09513590.2016.1153058](http://dx.doi.org/10.3109/09513590.2016.1153058).
- Sousa M, Tesarik J, 1994. Ultrastructural analysis of fertilization failure after intracytoplasmic sperm injection. *Hum Reprod.*;9:2374-80.

- Sousa M, Cunha M, Viana P, Silva J, Teixeira da Silva J, Oliveira C, 2012. Outcomes of human blastocyst transfer after slow-freezing using sequential culture: a Clinical report. *Arch Gynecol Obstet.*;285:1473-8. doi: 10.1007/s00404-011-2174-5.
- Sousa M, Cunha M, Teixeira da Silva J, Oliveira C, Silva J, Viana P, 2015. Ovarian hyperstimulation Syndrome: a Clinical report on 4894 consecutive ART treatment cycles. *Reprod Biol Endocrinol.*;13:66. Doi: 10.1186/s12958-015-0067-3.
- Sousa M, Cunha M, Silva J, Oliveira E, Pinho MJ, Almeida C, 2016. Ultrastructural and cytogenetic analyses of mature human oocyte dysmorphisms with respect to Clinical outcomes. *J Assist Reprod Genet.*;33:1041-57. doi: 10.1007/s10815-016-0739-8.
- Steward RG, Lan L, Shah AA, Yeh JS, Price TM, Goldfarb JM, 2014. Oocyte number as a predictor for ovarian hyperstimulation syndrome and live birth: an analysis of 256,381 in vitro fertilization cycles. *Fertil Steril.*;101:967-73. doi: 10.1016/j.fertnstert.2013.12.026.
- Stoop D, Ermini B, Polyzos NP, Haentjens P, De Vos M, Verheyen G, 2012. Reproductive potential of a metaphase II oocyte retrieved after ovarian stimulation: an analysis of 23,354 ICSI cycles. *Hum Reprod.*;27:2030-5. doi: 10.1093/humrep/des131.
- Sunkara SK, Rittenberg V, Raine-Fenning N, Bhattacharya S, Zamora J, Coomarasamy A, 2011. Association between the number of eggs and live birth in IVF treatment: an analysis of 400,135 treatment cycles. *Hum Reprod.*;26:1768-74. doi: 10.1093/humrep/der106.
- Swain JE, Smith GD, 2011. Advances in embryo culture platforms: novel approaches to improve preimplantation embryo development through modifications of the microenvironment. *Hum Reprod Update.*;17:541-557. doi: 10.1093/humupd/dmr006.
- Tesarik J, Sousa M, 1995. Key elements of a highly efficient intracytoplasmic sperm injection technique: Ca²⁺ fluxes and oocyte cytoplasmic dislocation. *Fertil Steril.*;64:770-6. doi: [http://dx.doi.org/10.1016/S0015-0282\(16\)57853-4](http://dx.doi.org/10.1016/S0015-0282(16)57853-4).
- Thum MY, Kalu E, Abdalla H, 2009. Elevated basal FSH and embryo quality: lessons from extended culture embryos. *J Assist Reprod Genet.*;26:313-8. doi: 10.1007/s10815-009-9313-y.
- Turnpenny P, Ellard S, 2012, eds. *Emery's Elements of Medical Genetics*. Philadelphia. Elsevier, Churchill Livingstone. ISBN: 9780702040436.
- van Loendersloot LL, van Wely M, Limpens J, Bossuyt PMM, Repping S, van der Veen F, 2010. Predictive factors in in vitro fertilization (IVF): a systematic review and meta-analysis. *Hum Reprod Update.*;16:577-89. doi: 10.1093/humupd/dmq015.
- Van Steirteghem AC, Liu J, Joris H, Nagy Z, Janssenswillen C, Tournaye H, 1993. Higher success rate by intracytoplasmic sperm injection than by subzonal insemination. Report of a second series of 300 consecutive treatment cycles. *Hum Reprod.*;8:1055-60.
- Van Steirteghem AC, Nagy Z, Joris H, Liu J, Staessen C, Smitz J, 1993. High fertilization and implantation rates after intracytoplasmic sperm injection. *Hum Reprod.*;8:1061-6.
- van Wely M, Kwan I, Burt AL, Thomas J, Vail A, Van der Veen F, 2011. Recombinant versus urinary gonadotrophin for ovarian stimulation in assisted reproductive technology cycles (Review). *Cochrane Database of Syst Rev.*;2. Doi: 10.1002/14651858.CD005354.pub2.
- Vandervorst M, Liebaers I, Sermon K, Staessen C, De Vos A, Van de Velde H, 1998. Successful preimplantation genetic diagnosis is related to the number of available cumulus-oocyte complexes. *Hum Reprod.*;13:3169-76. doi: 10.1093/humrep/13.11.3169.
- Zheng J, Lu Y, Qu X, Wang P, Zhao L, Gao M, 2016. Decreased sperm motility retarded ICSI fertilization rate in severe oligozoospermia but good-quality embryo transfer had achieved the prospective clinical outcomes. *PLoS One.*;11:e0163524. Doi: 10.1371/journal.pone.0163524.

ANEXOS

Legend to Figures

Left panel. Percentage of cycles with NB (dark box) and without NB (light box). This percentage was calculated within each group independently using the total of 993 cycles for the NB group and the total of 2,851 cycles for the no-NB group.

Right panel. Percentage of cycles with NB (dark box) and without NB (light box). This percentage was calculated using the total number of cycles in each grade (with NB + without NB). The linear tendency line and the linear regression (R²) indicate the likelihood of achieving a live birth.

Female age, Time of infertility (years);

Ag: Agonist protocol;

Antag: Antagonist protocol;

COC: Cumulus-oocyte complexes (retrieved oocytes);

MII: Mature metaphase II oocytes;

AB: High quality embryos at day 3;

N° ET: Number of embryos transferred;

ET day: day of embryo transfer;

Conc: sperm concentration (M/ml);

RPM: sperm rapid progressive motility (%);

NM: sperm normal morphology (%);

O: oligozoospermia;

A: asthenozoospermia;

T: teratozoospermia.

Table 1.

TABLE 1 - Demographic and stimulation characteristics. ICSI cycles using ejaculated sperm				
Variable	Total	with NB	without NB	p
Cycles (n)	3844	993	2851	
Female age (y) (mean ± SD)	35.41 ± 4.49	33.46 ± 3.76	36.09 ± 4.52	a-b-c
Male age (y) (mean ± SD)	36.92 ± 5.73	35.46 ± 5.24	37.43 ± 5.81	a-b-c
Time infertility (y) (mean ± SD)	3.25 ± 2.70	3.18 ± 2.52	3.27 ± 2.76	NS
Male factor (rate)	72.4	77.6	70.5	a-c
Female factor (rate)	4.5	2.5	5.2	a-c
Mixed factors (rate)	23.1	19.9	24.3	a-c
Abnormal female karyotype (rate)	2.5	2.7	2.5	NS
Abnormal male karyotype (rate)	2.4	1.7	2.8	NS
bFSH (IU) (mean ± SD)	7.42 ± 3.71	6.59 ± 2.52	7.71 ± 4.01	a-b-c
Follicles (mean ± SD)	10.93 ± 6.45	12.99 ± 5.84	10.22 ± 6.50	a-b-c
Total dose (IU) (mean ± SD)	2195.30 ± 1132.29	1779.37 ± 824.01	2340.27 ± 1187.97	a-b-c
Time stimulation (days) (mean ± SD)	8.53 ± 1.86	8.37 ± 1.50	8.58 ± 1.97	a-c
Estradiol (pg) (mean ± SD)	1281.10 ± 955.29	1365.17 ± 740.14	1250.43 ± 1020.99	a-c
HCG dose (IU) (mean ± SD)	9025.79 ± 2070.44	8552.56 ± 2134.67	9201.69 ± 2018.38	a-b-c
Busereline (IU) (mean ± SD)	0.80 ± 0.00 (n=47)	0.80 ± 0.00 (n=4)	0.80 ± 0.00 (n=43)	b

Significance ($p < 0.05$).

Comparisons as:

a = Total cycles vs cycles with NB.

b = Total cycles vs cycles without NB.

c = cycles with NB vs cycles without NB.

NS: not significant.

NB newborn, *bFSH* basal follicle stimulating hormone, *Total dose* total gonadotropin dose, *HCG* human chorionic gonadotropin.

Table 2.

TABLE 2 - Embryological and clinical outcomes. ICSI cycles using ejaculated sperm

Parameters	Total	with NB	without NB	p
Cycles (n)	3844	993	2851	
Embryo transfer cycles (n)	3334	993	2341	
COC (n, mean \pm SD)	26581 (6.91 \pm 4.83)	8554 (8.61 \pm 4.42)	18027 (6.32 \pm 4.83)	a-b-c
MII (n, rate)	20818 (5.9)	6948 (7.0)	13870 (5.4)	
Maturation rate (MII/COC) (rate)	78.3	81.2	76.9	a-b-c
2PN/2PB (n, rate)	16068 (4.7)	5569 (5.6)	10499 (4.3)	
Fertilization rate (2PN/MII) (rate)	77.2	80.2	75.7	a-b-c
Embryos cleaved-day 2 (n, rate)	15938 (4.6)	5555 (5.6)	10383 (4.2)	
Embryo cleavage rate (d2/2PN) (rate)	99.2	99.7	98.9	a-b-c
Day 3 embryos (n, rate)	11409 (4.2)	4304 (4.8)	7105 (3.9)	
Day 3 grade A/B embryos (n, rate)	10891 (4.0)	4146 (4.7)	6745 (3.7)	
Day 3 grade A/B rate (rate)	95.5	96.3	94.9	a-c
Day-4 embryos (n, rate)	5695 (3.9)	2469 (4.3)	3226 (3.7)	
Day-5 embryos (n, rate)	4355 (3.5)	1917 (3.9)	2438 (3.3)	
Blastocyst rate (rate)	47.6	52.4	44.4	a-b-c
n° of transferred embryos (n, mean \pm SD)	6211 (1.86 \pm 0.51)	1896 (1.90 \pm 0.39)	4315 (1.84 \pm 0.55)	a-c
Biochemical pregnancy (/ETC) (n, rate)	1354 (40.6)	990 (99.7)	364 (15.5)	
Sacs (n)	1534	1304	230	
Implantation rate (n° sacs/n° ET) (rate)	24.7	68.8	5.3	
Clinical pregnancy (/ETC) (n, rate)	1221 (36.6)	993 (100)	228 (9.7)	
Singletons (/CP) (n, rate)	863 (70.7)	685 (69)	178 (78.1)	
Twins (/CP) (n, rate)	331 (27.1)	305 (30.7)	26 (11.4)	
Triplets (/CP) (n, rate)	3 (0.2)	3 (0.8)	0	
Ectopic pregnancy (/CP) (n, rate)	24 (2.0)	0	24 (10.5)	
Abortion (/CP) (n, rate)	204 (16.7)	0	204 (89.5)	
OP (/ETC)-(CP-Ab-Ect) (n, rate)	993 (29.8)	993 (100)	0	
Delivery (/ETC) (n, rate)	984 (29.5)	984 (99.1)	0	
Stillborn (n)	3	3	0	
LBDR (/ETC)-(Delivery-Stillborn) (n, rate)	981 (29.4)	981 (98.8)	0	
Newborn (/ETC): (n, rate):	1234 (37.0)	1234 (100)	0	
Male (/NB)	638 (52.0)	638 (52)		
Female (/NB)	588 (48.0)	588 (48.0)		
M/F ratio	1.2	1.2		
NB malformations (/NB): (n, rate):	16 (1.3)	16 (1.3)		
Major (/NB)	12 (1.0)	12 (1.0)		
Minor (/NB)	4 (0.3)	4 (0.3)		
ENdeath (/ETC) (n)	0	0		

Significance ($p < 0.05$).

Comparisons as:

a = Total cycles vs cycles with NB.

b = Total cycles vs cycles without NB.

c = cycles with NB vs cycles without NB.

NS = not significant.

NB newborn, *COC* cumulus-oocyte complexes (aspirated oocytes), *MII* mature oocytes at metaphase II of meiosis, *2PN-2PB* normal fertilized oocytes (with 2 pronuclei and 2 polar bodies), *OP* ongoing pregnancy, *LBDR* live-birth delivery rate, *EN death* early neonatal death.

Table 3.

TABLE 3 - Newborn outcomes. ICSI cycles using ejaculated sperm

Parameters	Total	with NB	without NB	p
Cycles (n)	3844	993	2851	NA
Embryo transfer cycles (n)	3334	993	2341	
Newborn (n)	1234	1234	-	
Gestation age (weeks) (mean \pm SD) (n)	37.7 \pm 2.8	993		
Term (weeks) (mean \pm SD) (rate, n)	38.7 \pm 1.1 (79.7)	791		
Preterm (PT) (weeks) (mean \pm SD) (rate, n)	33.5 \pm 3.3 (20.0)	198		
Very PT (weeks) (mean \pm SD) (rate, n)	28.0 \pm 3.2 (4.1)	41		
Extremely PT (weeks) (mean \pm SD) (rate, n)	23.80 \pm 2.3 (1.2)	12		
Weight (g) (mean \pm SD) (n)	2739.7 \pm 703.3	1204		
Normal weight (g) (mean \pm SD) (rate, n)	3111.4 \pm 357.5 (65.4)	788		
Low weight (LW) (g) (mean \pm SD) (rate, n)	1943.6 \pm 490.0 (33.1)	399		
Very LW (g) (mean \pm SD) (rate, n)	1102.4 \pm 269.0 (6.3)	76		
Extremely LW (g) (mean \pm SD) (rate, n)	790.6 \pm 172.2 (2.2)	26		

NA not applicable.

Fig. 1

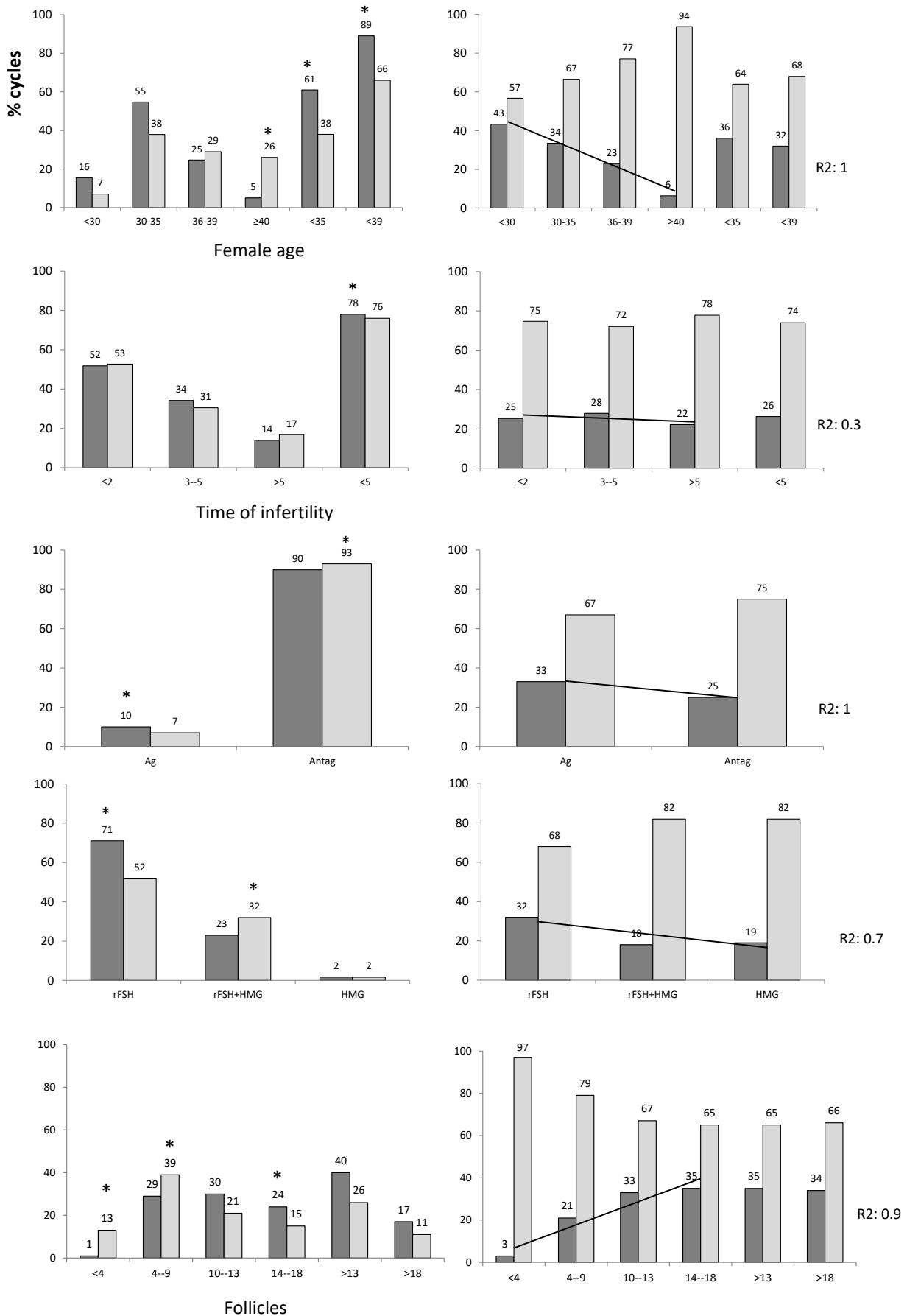


Fig. 2

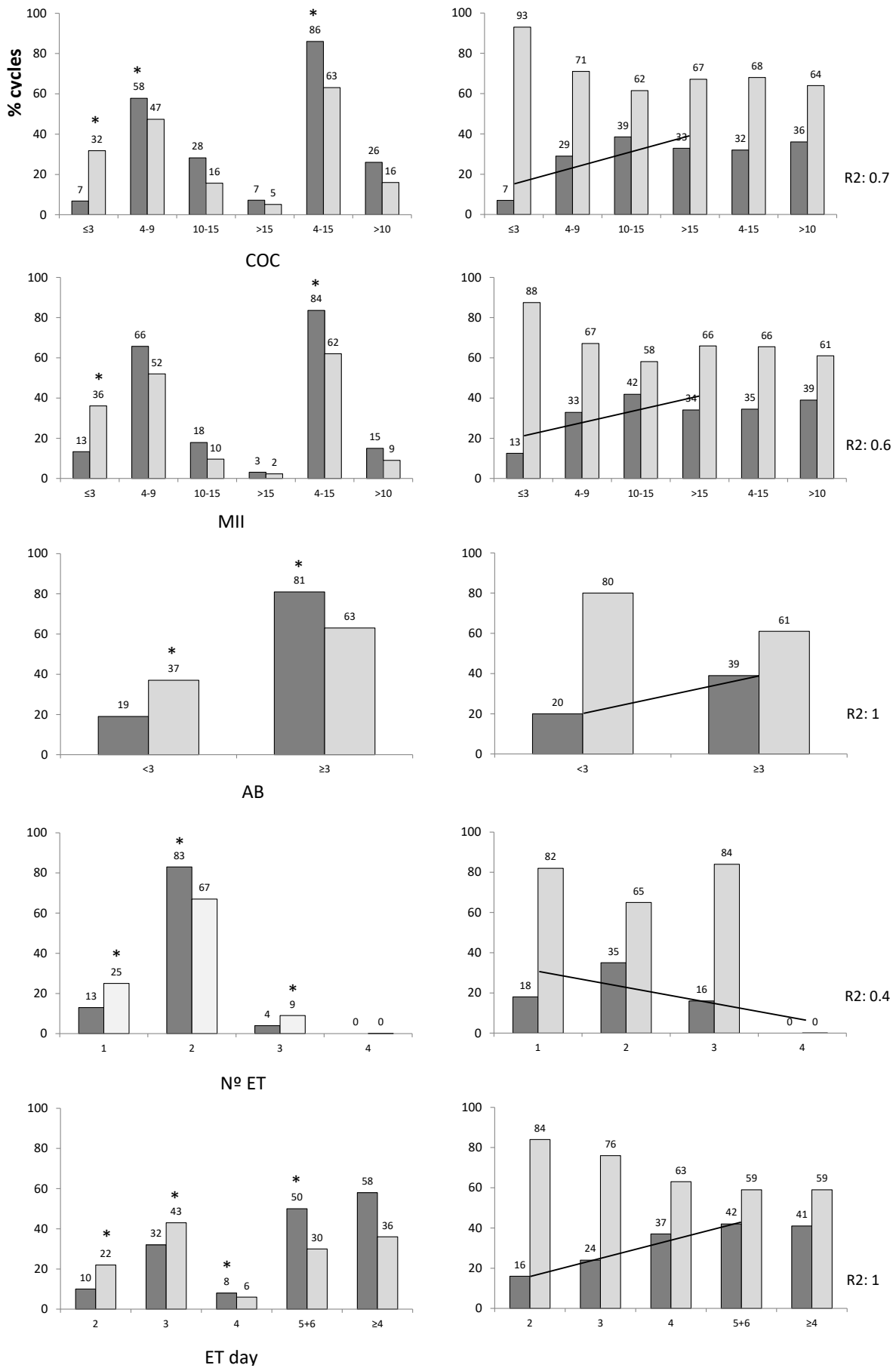


Fig. 3

