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Limited Zn and Ni mobility during simulated iron

formation diagenesis

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ABSTRACT

Iron formations (IF) are iron- and silica-rich chemical precipitates that were deposited during the Precambrian. Several recent studies have demonstrated how the trace metal abundances in IF can be used as proxies for the bioavailability of trace metals in ancient seawater; with the ultimate goal being to understand first-order controls on the composition of the ancient biosphere. However, the utility of IF as proxies depends on the immobilization of trace metals during diagenesis. Here, we assess the mobility of Zn and Ni from ferric oxyhydroxides (ferrihydrite) in the absence and presence of organic matter (glucose) during simulated diagenesis (170°C, 1.2 kbar); similar to what some Precambrian IF experienced. Quantitative concentration data, coupled with x-ray diffraction analysis and electron microprobe element mapping, demonstrate that both metals are relatively immobile during simulated diagenesis. Additionally, the mechanism for initial Ni sorption is examined using x-ray adsorption spectroscopy. For the initial sorption of trace elements in abiotic ferrihydrite experiments, 93.38% Zn and 65.95% Ni were initially sorbed. In experiments utilizing biogenic ferrihydrite, 97.03% of Zn and 93.38% of Ni were initially sorbed. Following the diagenetic capsule treatments, more than 99% of Zn and more than 91.9% of Ni were retained under the varied conditions considered here. Capsule experiments suggest the strong retention of Zn and Ni following the diagenesis of either abiotic or biogenic ferrihydrite. Overall, our results indicate that paleomarine Zn and Ni concentrations are likely to be faithfully recorded in wellpreserved IF deposits.

Keywords: iron formations; Precambrian; zinc; nickel; diagenesis; iron oxyhydroxides; trace metal mobility; X-ray Absorption Spectroscopy

1. Introduction

Iron formations (IF) are a key sedimentary proxy record for tracking the chemical history of seawater and reconstructing trace metal abundances in the Precambrian oceans. Many trace metals, such as zinc (Zn) and nickel (Ni), are critical components of a variety of eukaryotic and prokaryotic metalloenzymes, respectively (Williams and da Silva, 1996). As a result, reconstructing their concentrations in ancient seawater may provide crucial information about the constraints imposed on the evolving early biosphere (e.g., Dupont et al., 2010).

Iron formations contain iron (Fe) minerals with various oxidation states, such as hematite (Fe₂O₃), magnetite (Fe₃O₄) and siderite (FeCO₃), often in a cherty matrix. Additionally, some IF may contain significant Fe(II)- and/or Fe(III)-silicates, such as greenalite, stilpnomelane, minnesotaite, chamosite, riebeckite, and ferri-annite (Klein, 2005). It is important to note, however, that these are all secondary minerals, and that the primary iron precipitates probably comprised ferric oxyhydroxides, such as ferrihydrite (Fe(OH)₃) (see Bekker et al., 2010 for review). Although the actual mechanisms of Fe(II) oxidation remain uncertain, they appear to have been biologically mediated (e.g., Posth et al., 2013a, 2014; Pecoits et al., 2015). The organic biomass deposited with Fe(OH)₃ would subsequently serve as a reductant during diagenesis and metamorphism (e.g., Perry et al., 1973; Konhauser et al., 2005; Johnson et al., 2008; Craddock and Dauphas, 2011). Fe(III) reduction by organic matter is indicated by (i) the presence of ferrous iron-

containing minerals, such as magnetite (Fe₃O₄) and siderite (FeCO₃), (ii) the low organic carbon content of IF today (<0.5%; Gole and Klein, 1981), and (iii) Fe, C and O isotope compositions in Fe-carbonates associated with IF (Johnson et al., 2008; Heimann et al., 2010; Craddock and Dauphas, 2011). In addition to Fe(III) reduction coupled to organic carbon oxidation (i.e., anaerobic chemoheterotrophy), when a solution containing ferrous iron (Fe(II)) is present, redox-driven recrystallization of the primary Fe(OH)₃ minerals may cause loss of Ni to solution (Frierdich et al., 2011). Therefore, constraining the mobility of elements, such and Zn and Ni, under realistic diagenetic scenarios is critical for our interpretation of the IF record, and the implications for paleomarine systems which are drawn from it.

There are indications from Archean IF that Ni and Zn may relatively enriched, and therefore, available as key nutrients on the early Earth. These include relatively high Ni and Zn concentrations found in the 3.75 Ga Nuvvuagittiq IF, Canada (Mloszewska et al., 2012) and Ni enrichments considered to be primary in origin in the IF of the Mesoarchean Witwatersrand Supergroup in South Africa (Smith et al., 2013). Furthermore, compilations of both Ni and Zn in the IF record have recently been presented (Konhauser et al., 2009; Robbins et al., 2013), as well as for Zn in the shale record (Scott et al., 2013). Distribution coefficients (K_D) between seawater and Fe(OH)₃ have been applied to the IF record to extrapolate paleomarine Ni and Zn concentrations (Konhauser et al., 2009; Robbins et al., 2013). These studies have shown dissimilar temporal trends throughout the Precambrian, with seawater Ni concentrations appearing to have decreased rapidly from ~400 to 100 nM between 2.7 and 2.5 Ga (Konhauser et al., 2009). The IF record for Ni indicates a sudden, dramatic decrease in marine Ni levels

immediately prior to the Great Oxidation Event (GOE) (Konhauser et al., 2009). This crash in Ni may have been a contributing factor leading to the GOE and had significant implications for the biosphere ~2.7 Ga. However, the marine Zn reservoir appears to have remained constant with an estimated seawater concentration of ~10 nM (Robbins et al., 2013). When taken together, the IF and shale records for Zn through time effectively and independently point to near modern levels of marine Zn from the Archean through to the modern (Robbins et al., 2013; Scott et al., 2013). This is in contradiction to previous estimates based on thermodynamic models (e.g., Saito et al., 2003) and the idea that Zn would have been biolimiting to early eukarytoes and delayed their diversification (Dupont et al., 2010). Accordingly, studying the IF record provides a key window into evaluating paleomarine conditions, especially with regards to trace metal abundances. As such, understanding the potential diagenetic impacts is critical to such evaluations.

However, the extent to which Zn and Ni are mobilized when the precursor ferrihydrite underwent reduction and dehydration during diagenesis remains unclear. Our work builds on methodologies of previous simulated IF diagenesis experiments that focused on Fe-mineral transformations (Posth et al., 2013b; Köhler et al., 2013), in order to evaluate trace element mobilization during diagenesis and the robustness of previous paleomarine Zn and Ni estimates. Additionally, due to the role of organic matter as a diagenetic reductant, we consider reactions between organic carbon and Fe(III) in our diagenetic experiments; with Zn or Ni sorbed to either synthetic 2-line ferrihydrite with added glucose in the capsule or sorbed directly to biogenic ferrihydrite. We couple these results with micro X-ray diffraction (μ XRD) analysis of the post-diagenetic mineralogy for selected samples and electron microprobe mapping (EMPA) of Zn and Ni to examine diagenetic trace element redistribution.

2. Methods

2.1 Simulated diagenesis experiments

Diagenetic capsule experiments on synthetic Fe oxyhydroxide precipitates (Posth et al., 2013b) demonstrate that the Fe mineralogy observed in IF today can be reproduced after incubation of synthetic 2-line ferrihydrite in the presence or absence of glucose (as a proxy for biomass). Capsules were subjected for 14 days at 170°C and 1.2 kbar, approximately the diagenetic pressure and temperature (P-T) conditions of one of the largest and most studied iron formations on Earth, the Transvaal Supergroup BIF (Miyano and Klein, 1984; Miyano, 1987). Furthermore, these conditions mimic previous diagenetic capsule experiments (Posth et al., 2013b; Köhler et al., 2013), and therefore, facilitate a ready comparison between experimental conditions. Our experimental duration and P-T parameters are obviously much abbreviated compared to natural burial diagenesis and metamorphism that takes thousands to millions of years; and are less than experienced by some IF which were subjected to amphibolite facies. However, we still effectively capture the crucial step of the transformation of Fe(OH)₃ to the minerals typically observed in IF, i.e., hematite (Fe_2O_3), some of which is thought to have occurred during early diagenesis at temperatures below 140°C (Becker and Clayton, 1976).

Diagenetic experiments and analysis were performed at the University of Tübingen using similar procedures and apparatus as per Posth et al. (2013b). Our experiments consider diagenetic mobilization of Zn and Ni from (1) 2-line ferrihydrite (Fh), as a proxy for primary Fe-oxyhydroxides formed from the chemical oxidation of Fe(II) with O₂, and (2) biogenic ferrihydrite (BF) precipitated by a marine, Fe(II)-oxidizing phototrophic bacterial culture (*Rhodovulum iodosum*, see Wu et al., 2014; Eickhoff et al., 2014). We utilize the BF as a proxy for primary Fe-oxyhydroxides formed by Fe(II)-oxidizing bacteria. Our experiments consisted of three general steps: (i) the synthesis of Fh, either as abiotic or cell-mineral aggregates, and sorption of Ni and Zn, (ii) treatment of the trace element sorbed minerals, with or without organic material, under simulated diagenetic P-T conditions, and (iii) quantification of the concentration of Ni and Zn mobilized.

Capsules were named according to a consistent scheme, i.e. ZnFh-1, where Zn or Ni refers to the trace element sorbed, and Fh or BF indicates abiotic 2-line ferrihydrite or the biogenic cell-mineral aggregates, respectively. The numbers refer to the experimental replicates, where for the abiotic Fh experiments, replicates 1 and 2 contained glucose added as an organic carbon source, and replicates 3 and 4 lacked glucose. In experiments with BF, no glucose was added because cell material served as a source of organic carbon. In these samples, the numbers only serve to identify the respective capsule among replicates. For example, NiFh-1 indicates a capsule containing Ni sorbed to Fh with glucose, while ZnBF3 indicates Zn sorbed to BF. Capsules ZnFh-1, NiFh-3 and ZnBF1 were excluded from further analysis as the integrity of the seals were compromised during diagenetic treatments. Finally, the prefix C- denotes a control capsule (e.g., C-ZnFh1). Control capsules were set up to mirror experimental capsules but were stored in the dark in Eppendorf tubes and subjected to room temperature and pressure for the same period of time as experimental capsules.

2.1.1 Synthesis of ferrihydrite and sorption of Zn or Ni

Synthetic Fh used in abiotic experiments was prepared via methods previously described (c.f. Cornell and Schwertmann, 2003) and freeze-dried prior to sorption of Ni or Zn. The marine photoferrotroph, *Rhodovulum iodosum*, was cultivated in artificial seawater medium as described by Wu et al. (2014); the phosphate content was reduced to 1 mM to minimize FePO₄ precipitation, with additional NaCl added to maintain the same ionic strength as per Eickhoff (2013). FeCl₂ was added to a concentration of 10 mM, but the medium was not filtered to remove initial Fe carbonate and Fe phosphate that precipitated. The strain was then inoculated and grown at 12.82 µmol photons/m²/s² until all Fe²⁺ was oxidized, as determined by the Ferrozine assay (Stookey, 1970). The cell-mineral aggregates were then collected by centrifugation and washed four times with the growth medium lacking bicarbonate buffer and harvested by centrifugation at 4000 rpm for 10 minutes before freeze-drying.

Sorption of Zn and Ni to both Fh and BF was accomplished by introducing ~50 μ M of Zn or Ni (from Merck ICP-MS standards) in the presence of 1g/L Fh or BF in solution prepared to seawater ionic strength with 0.5M NaNO₃ and buffered by 0.1 M H₃BO₃ to a pH value of 8. Sorption occurred over a 24-hour period, before the solution was passed through a 0.2 μ M Millipore filter, and the solid particles were recovered. Filtrates were analyzed for Zn and Ni by Inductively Coupled Plasma Optical Emission Spectroscopy (ICP-OES), on a Perkin Elmer Optima 5300 at the University of Tübingen, to determine the amount of Zn or Ni lost due to sorption. For ICP-OES measurements, the percent relative standard deviation (%RSD) was better than ±8% and ±5% for Zn and

Ni, respectively. Calculations of the amount of Zn and Ni added normalized to the weight of solid particles, as well as the percentage of Zn or Ni sorbed, are provided in Table 1. In all cases the majority of Zn and Ni was sorbed. Solid particles were then air-dried at room temperature prior to being loaded in experimental gold capsules.

2.1.2 Mobilization experiments

Three experiments were conducted with Zn and Ni sorbed independently to: (1) Fh only, (2) Fh with added glucose, and (3) BF. Experiments followed the protocols of Posth et al. (2013b) in which Fh is transformed to hematite via dry heating. Experimental gold capsules with Fh were set-up such that 100 mg of ferrihydrite with Zn or Ni sorbed was loaded into the capsules. For capsules with Fh and glucose, 4 mg of glucose were physically mixed with the Fh to achieve a ratio of electron reducing equivalents of ~ 0.6 that could be transferred from glucose to Fh during diagenetic treatment (as per Posth et al., 2013b). This approximates the low organic carbon content of IF, which would limit the number of electrons available for diagenetic Fe(III) reduction (Konhauser et al., 2005). Capsules with BF contained approximately 35 mg of BF cell-mineral aggregate. Gold capsules were crimped closed and then sealed using an arc welder. During welding the capsule remained cool to the touch, precluding any temperature reaction prior to capsule incubation. The capsules were weighed before and after a sonication treatment in a water bath to ensure that the contents were sealed. Capsules were then incubated at 170°C and 1.2 kbar for 14 days in a high-pressure autoclave (Sitec Sieber Engineering Ag, Zürich, Switzerland). In tandem with experimental capsules, control capsules were incubated at room temperature for 14 days to account for any reactions under ambient conditions.

2.1.3 Quantification of Zn or Ni mobilized

Following incubation, capsule exteriors were cleaned with acetone, and then capsules were cut into smaller sections and placed into Falcon tubes. To each Falcon tube 5 mL of pH-buffered seawater ionic strength solution (0.5 M NaNO₃ and 0.1 M H₃BO₃, pH 8) was added and then the capsules were sonicated for 30 minutes. The solution was then removed and the sonication repeated with fresh solutions. Filters for selected samples were retained for subsequent analysis by μ XRD and EMPA, following an 18-month period of oxidation (see section 2.3).

The combined rinse solutions for each sample were filtered (0.22 μ M Millipore filters), acidified (with trace metal grade HNO₃ to a final concentration of 1-2%) and then diluted prior to ICP-OES analysis. The purpose of this step was to rinse the post-diagenetic particles in a solution that resembled seawater (ionic strength, pH) in order to recover any Zn or Ni mobilized during diagenetic reactions. This allowed for the recovery of the resultant diagenetic waters (i.e., to collect the overpressure liquid produced by the dehydration of Fh to hematite), without leaching immobilized Zn or Ni from the particles (i.e., Zn or Ni strongly bound to hematite). Additionally, we consider that porewaters would have likely remained circumneutral – although some CO₂ would have been released during organic mineralization, any anaerobic respiration, as would be realistic for Archean sedimentary systems, would have led to an increase in pH (c.f. Konhauser, 2007). Zn or Ni concentrations were used to determine the percentage of

initially sorbed Zn or Ni to Fh or BF that was retained by the minerals in each capsule (Table 2). Errors on the percentage of metal retained (Table 2) were calculated using the two-sigma level ICP-OES analysis standard deviations (%RSD was better than \pm 8% for Zn and \pm 5% for Ni, when the analyte was above detection limits). The percent error in the trace metal retained (\pm 2 σ TM_{retained} (%)) is equal to the absolute value of the maximum variation from the trace metal retained, relative to the initial trace metal in the capsule (equation 1). Where TM_{capsule} is the initial amount of Zn or Ni in each capsule, TM_{released} is the amount of Zn or Ni released and 2 σ is the variation in ICP-OES measurements.

$$\pm 2\sigma TM_{\text{retained}}(\%) = \left| \left((TM_{\text{capsule}} - TM_{\text{released}} \pm 2\sigma) / TM_{\text{capsule}} \right) - 1 \right| \times 100$$
(1)

2.1.4 Calculation of C:Fe ratios

Molar ratios of C:Fe were calculated for each sample containing organic matter. For capsules with Fh we utilized an ideal formula of $Fe(OH)_3$, molar mass = 106.87 g/mol and glucose (180.15 g/mol). This allowed for the determination of the moles of glucose based on the percentage of the molecular weight that carbon accounts for in glucose and the mass of glucose added (see equation 2). In equation 2, *m* is the mass in grams added to the capsule, and the ratio of 72.06 to 180.15 represents the molar mass of carbon in glucose to the total molar mass. The calculated moles of carbon added as glucose were then normalized to the moles of Fe in the ferrihydrite added (equation 3).

mol C =
$$(m_{glucose} * (72.06/180.15))/(12.01g/mol)$$
 (2)

mol Fe =
$$(m_{Fe-particles}*(55.85/106.87))/(55.85g/mol)$$
 (3)

For experiments with BF the total carbon content was measured using an Elementar Vario EL at the University of Tübingen. Total inorganic carbon was not determined because the acid treatment required would have dissolved some of the BF as well. Inorganic carbon was present in the growth medium as NaHCO₃ buffer in equilibrium with N₂/CO₂ gas. However, the cell-mineral aggregates were washed four times in a basal medium without bicarbonate (see above for description) prior to freeze-drying and this should have removed any inorganic carbon in solution. The moles of carbon were calculated by equation 4 and normalized to the moles of Fe in the BF added to each capsule as calculated in equation 5. For the BF, the cell-mineral aggregates are a combination of Fe(OH)₃ and organic carbon (CH₂O) and these exist in a 4:1 molar ratio, resulting in a cell mineral aggregate molar mass of ~458 g/mol. A 5% variation in the molar mass of the cell-mineral aggregates would result in a corresponding ±5% change in the molar C to Fe ratios for the BF.

$$mol C = ((%C_{organic}/100)*m_{BF})/(12.01g/mol)$$
(4)
$$mol Fe = (m_{BF}*((4*55.85)/458))/(55.85g/mol)$$
(5)

The average molar C:Fe ratio for capsules with Fh and glucose was determined to be $\sim 0.1446 \pm 0.0008$, which is close to that for capsules with BF measured as ~ 0.1792 (Table 2).

2.2 X-ray Absorption Spectroscopy

X-ray absorption spectroscopy (XAS) of the initial Ni-sorbed Fh sample was collected at the Ni K-edge (8.333 keV) at the Diamond Light Source in the UK. During data collection, storage ring energy was 3.0 GeV and the beam current was approximately 200 mA. The sample was presented to the X-ray beam as a thick paste held in a Teflon slide and sealed either side of the sample slot with Kapton tape. XAS data were collected from 16 scans in fluorescence mode using a Ge 9-element detector. A series of test scans were performed prior to data collection to confirm that there was no photo-redox or visible drying of the sample during data collection. Energy calibration was achieved by assigning the first inflection point of Au (L3) to 11.919 keV.

XAS data reduction was performed using ATHENA (Ravel and Newville, 2005) and the spectrum was fit using DL_EXCURV (Tomić et al., 2005). The spectrum was fit in k-space over 3 - 12 Å⁻¹, with no Fourier filtering during data analysis, and the fitting included full multiple scattering as coded in EXCURV98 (Binsted, 1998). Multiple scattering calculations require specification of the full three dimensional structure of the Ni coordination environment (i.e., bond angles in addition to bond lengths). This was done using hypothetical model clusters with C1 symmetry, for various different Ni surface complexation coordination geometries (including mononuclear monodenatate, mononuclear bidentate edge-sharing and mononuclear bidentate corner-sharing).

The number of independent data points (N_{ind}) was determined using Stern's rule (Stern, 1993) as $2\Delta k\Delta R/\pi + 2$ (Booth and Hu, 2009) where Δk and ΔR are the range in k and R-space actually fitted; as such, N_{ind} = 14. The number of fitted parameters (N_{par}) was determined as the total number of parameters optimized during the various model refinements and was always less than N_{ind}. Typical errors associated with EXAFS modeling over the k-range used here are 15% and 25% for first and second shell distances, coordination numbers, respectively, ±0.02 and 0.05Å for first and second shell distances,

respectively, and 15% and 25% for first and second shell Debye–Waller factors (DWF's), respectively (Binsted, 1998). The quality of the fits provided by the different model clusters was assessed using the EXAFS R-factor and Fit Index, with an absolute index of goodness of fit (used for comparing fit quality between clusters where N_{pars} was not equivalent) given by the reduced Chi² function (all as coded in EXCURV98 (Binsted et al., 1996; Binsted, 1998)).

2.3 Micro X-ray Diffraction and Electron Microprobe mapping

The work of Posth et al. (2013b) established that the post diagenetic mineralogy of these capsules consists of hematite when no organic carbon is present, and a mixture of hematite, magnetite, and siderite in the presence of organic carbon. As such, we examined the mineralogy, and Zn and Ni distribution of several key samples, in order to determine the mineralogy and Zn or Ni distributions following the re-oxidation of the post-diagenetic material. As described above, these samples had been removed from capsules after incubation, rinsed, and then filtered, and the filters retained. Following an 18-month re-oxidation period, the material from the filter was sonicated and then rinsed once in water (MilliQ, pH ~6.1) and dried at 30° C.

Dried samples of NiFh-1 and NiBF1 for mineral identification were loaded directly onto a Si single crystal silicon wafer for μ XRD. Data was collected with a Bruker D8 Discover XRD instrument (Bruker, Germany) equipped with a Co K α X-ray tube, (λ =0.17902 nm, 30 kV, 30 mA) and GADDS area detector. Minerals were identified with the International Center for Diffraction Data (ICDD) database. For spatially-resolved concentration analysis, dried particles from capsules ZnFh-2, NiFh-2, ZnBF2, and NiBF2

were cast in epoxy resin and polished to 1 μ m. Elemental concentration maps for Fe, Ni, and Zn were obtained using a Cameca SX100 electron probe micro-analyzer (EMPA), operating in quantitative mode with background subtraction at the facilities in Microsonde Ouest (Université of Brest/IFREMER). Beam operating conditions were 15 kV and 20 nA. Mineral standards for instrument calibration were andradite (Ca₃Fe₂Si₃O₁₂) for Fe, ZnS for Zn, and NiO for Ni. Detection limits for Fe, Zn, and Ni were 0.0825%, 0.22% and 0.063%, respectively. The precision on our EMPA analysis is estimated to be ±1%.

3. Results

3.1 Sorption of Zn and Ni

Trace metals sorbed to the Fh at molar values of 4.856 x 10^{-3} Zn/Fe and 3.918 x 10^{-3} Ni/Fe. This corresponds to 93.98 ± 0.54% of initial Zn and 65.95 ± 1.29% of initial Ni being sorbed. For BF, molar ratios of 5.410 x 10^{-3} Zn/Fe and 6.067x 10^{-3} Ni/Fe were sorbed, corresponding to 97.03 ± 0.60% of initial Zn and 93.38 ± 0.52% of initial Ni (see Table 1 for summary).

For the XAS spectrum of Ni initially sorbed to Fh (Fig. 1) we find that the first shell coordination environment consists of 6.0 O at ~ 2.0 - 2.09 Å, consistent with the octahedral NiO₆H_nⁿ⁻¹⁰ ion (Table 3). Beyond the O shells, we observe a peak in the Fourier transform at ~ 3.2 Å. Based on the literature to date, trace metals may sorb to Fe oxyhydroxides via outer-sphere and/or inner-sphere surface complexation, structural incorporation and/or be precipitated at the sorbent surface as a discrete metal oxyhydroxide phase (e.g., for a review see Brown and Parks, 2001). Because of the small

difference in atomic number, distinguishing between Ni and Fe during EXAFS fitting is difficult. However, Ni structurally incorporated into Fh or present as a precipitate of poorly crystalline hydrated α -Ni(OH)₂ has unique features in k-space, including a shoulder at ~ 5 Å⁻¹, a discrete peak at ~ 5.5 Å⁻¹, and a pronounced splitting of the peak at ~ 8 Å (e.g., Xu et al., 2007; Pandya et al., 1990). We do not observe these features and rather our spectrum is visually similar to those for Ni surface complexed with Fh, where Ni forms inner-sphere mononuclear bidentate complexes (Xu et al., 2007). The presence of the Fourier transform peak at ~ 3.2 Å also rules out the likelihood of Ni outer sphere surface complexation where typically no next-nearest neighbours are observed. On refining model clusters for mononuclear monodentate, mononuclear bidentate edgesharing and mononuclear bidentate corner-sharing, we find that Ni adsorbed in a mononuclear bidentate corner-sharing configuration produces the best fit to the spectrum, with 6.0 O at 2.0 - 2.09 Å and 2.0 Fe at 3.01 and 3.15 Å (Table 3). With a single Ni-Fe distance at ~ 3.05 Å, Xu et al. (2007) invoked mononuclear bidentate edge-sharing surface complexation. However, within the constraints of a Ni-Fh mononuclear bidentate edge-sharing complex, the maximum Ni-Fe distance can be no longer that \sim 3 Å (assuming no significant distortion of the Fe and/or Ni octahedra) and this complex is, therefore, unable to provide a satisfactory fit to our data. In summary, in agreement with Xu et al. (2007), we find that Ni is initially sorbed to Fh via mononuclear bidentate surface complexation.

3.2 Diagenetic experiments

In the post-diagenetic experiments, >99% of initially sorbed Zn is retained by the solid fractions in all scenarios, including controls (Table 2). Uncertainties are $\leq 4.3\%$ for Fh capsules and $\leq 7.2\%$ for BF capsules. Ni mobility is also low with >91% of initially sorbed Ni retained on (1) Fh only, (2) Fh with added glucose, and (3) BF (Table 2). However, ICP-OES standard deviations were higher for Ni, and associated uncertainties for the amount of Ni retained are up to 41.7%, but generally <17%. This is likely due to Ni concentrations being close to detection limits following dilution prior to analysis. For the Fh only experiments (in the absence of organic carbon), effectively all Ni was retained (NiFh-4). However, replicate capsules with Fh and glucose retained the least amount of Ni after incubation, and had the highest uncertainties, 94.7±26.6% and 91.9±41.7% (NiFh-1 and NiFh-2, respectively), while in BF capsules, 96.8±16.8% and 96.4±16.6% of Ni was retained (NiBF-1 and NiBF-2, respectively). Controls for Fh and BF capsules retained 99.8% (with $\leq 1.2\%$ uncertainty) and 98.9% (with $\leq 7.9\%$ uncertainty) of the initial Ni, respectively. It should be noted that the lowest Ni values by percentage of Ni retained were associated with the highest uncertainties and it is possible that 100% of Ni was retained in these experiments. However, in our discussion, we take the lower values as a conservative representation of Ni mobility.

3.3 µXRD and Electron Microprobe Analysis

Bulk mineralogy was analyzed by μ XRD on the two selected samples (NiFh-1 and NiBF1) from this study 18 months after the P-T incubations; during which time samples remained hydrated at 4°C in fully oxic conditions. In this regard, storage of the samples under oxic conditions for 18 months allowed us to determine mineralogy under post-

depositional conditions reminiscent of secondary oxygenated fluids moving through the BIF.

The bulk mineralogy for the selected sample NiFh-1, after P-T incubation, and an 18-month oxidation period, revealed a well-crystalline hematite phase (see Fig. 2). A similar phase was also detected in identical incubations that did not include added trace elements (Posth et al., 2013b). However, in that study, peaks for siderite and magnetite, absent here, were also clearly visible. Any siderite and magnetite present initially after incubation would have been oxidized during subsequent storage, leaving only stable hematite. Interestingly, the NiBF1 sample bulk mineralogy included both hematite and lepidocrocite; the hematite peaks were also broader in NiBF1 than in NiFh-1, likely indicating a more poorly crystalline hematite phase. The persistence of poorly crystalline phases in the biogenic experiments suggests that the presence of organic carbon during mineral precipitation can stabilize poorly ordered iron oxyhydroxides (e.g., Schwertmann, 1966; Jambor and Dutrizac, 1998; Toner et al., 2012).

Samples subjected to EMPA analysis had similarly been stored under the oxic conditions. For all samples subjected to EMPA analysis, except ZnFh-2, data indicate that Ni and Zn appear to be distributed as a function of Fe and no Ni- or Zn-rich zones or discrete Ni or Zn mineral phases were observed (Figs. 3-4). In other words, despite the likely initial presence of siderite or magnetite, as observed by Posth et al. (2013b), those reduced phases were subsequently re-oxidized to hematite but without corresponding mobility of Ni or Zn. In the case of ZnFh-2 (Fig. 3A, B), Zn-rich (up to 3.5 wt% Zn) inclusions of 5-15 µm in diameter were observed encased in the hematite matrix. The Zn-rich inclusions were also significantly more Fe-poor than the surrounding hematite matrix

(but still contained 30-40 wt% Fe), suggesting the possibility of siderite (which is known to form a solid-solution series with smithsonite ($ZnCO_{3}$), prior to re-oxidation.

4. Discussion

4.1 Sorption of Zn and Ni

In the Fh sorption experiments 93.38% Zn and 65.95% Ni were initially sorbed, while in the BF sorption experiments 97.03% of Zn and 93.38% of Ni were initially sorbed (Table 1). For both Zn and Ni in the Fh and BF experiments this is to be expected based on the sorption pH edges of these metals on Fh or other ferric oxyhydroxides (i.e., at pH 8 sorption is essentially complete; e.g., Trivedi and Axe, 2001). It appears from our work that Zn has a somewhat higher affinity than Ni for both Fh and BF. This is in overall agreement with previous work on the general behavior of Zn and Ni when they sorb to ferrihydrite and hematite at ionic strength < 1.5 M (e.g., Trivedi and Axe, 2001; Jeon et al., 2003). Although, the initial sorption of Ni to Fh here is less than expected, at only 65.95%, this may reflect variable Fh particle surface reactivity (e.g., as a result of aggregation and/or freeze-drying prior to metal sorption). However, this does not appear to have affected its retention in the later diagenetic phases of the experiment. Critically, all Fh and BF were treated similarly in their preparation and sorption of Zn or Ni, and overall the diagenetic mobility results are consistent across all the conditions considered. The sorption behavior of Zn and Ni likely affects their retention following diagenesis and will be discussed further in the context of the diagenetic mobility of Zn and Ni.

4.2 Diagenetic mobility of Zn and Ni

Understanding diagenetic trace element mobility is critical for the interpretation of IF paleomarine proxies. Posth et al. (2013b) showed that pressure and temperature treatments of Fh and organic carbon produces a mineralogy similar to IF, dominated by hematite, magnetite and siderite. Accordingly, that approach formed the basis for our diagenetic experiments. Our results suggest that Zn is essentially immobile during Fe(III) mineral diagenesis. Specifically, there is no significant desorption in the unincubated controls (consistent with the observations of Lee and Anderson (2005) who showed Zn was immobile on Fh over six months). Furthermore, it is not significantly desorbed from Fh, Fh with added glucose or BF during simulated diagenesis (Table 2). This is evident as there was effectively no Zn in the fluids generated during diagenesis and recovered in the post-diagenetic rinse. The presence of organic matter did not affect Zn mobility and the lack of mobility in BF capsules is consistent with results from Fh capsules with added glucose (Table 2). Similarly, Ni shows no significant desorption in the unincubated controls or the Fh experiments (Table 2). Nickel may be slightly more mobile when organic carbon was present, but when uncertainties are considered, Ni is likely immobile during diagenesis (Table 2). Nickel was slightly desorbed from BF controls with $\sim 1.1\%$ mobilized, whereas in Fh controls desorption was <0.25% regardless of glucose.

The limited mobility of Zn and Ni likely relates to their behaviour during sorption. In agreement with Xu et al. (2007), we show that Ni strongly adsorbs to Fh via inner-sphere mononuclear bidentate surface complexation. It is also known that Zn can similarly adsorb strongly to the surfaces of amorphous and crystalline iron oxyhydroxides via inner-sphere surface complexes (e.g., for Zn on hydrous ferric oxide, Lee and Anderson, 2005; ferrihydrite, Waychunas et al., 2002, 2003; and goethite, Trivedi et al...

2001). Both Zn and Ni sorb more strongly to amorphous vs. crystalline Fe oxyhydroxide phases, such as ferrihydrite (Trivedi and Axe, 2001). For Fh, the percent sorbed for both Zn and Ni approaches 100% (5 x10⁻⁵ M Zn or Ni and 1 g/L ferrihydrite at 25°C) as the pH approaches values of 6-8 (Trivedi and Axe, 2001). To the authors' knowledge there are no studies that determine the mechanism of Zn or Ni sorption on BF. However, Moon and Peacock (2012, 2013) showed that, similar to Ni, Cu(II) adsorbs to Fh via innersphere mononuclear bidentate surface complexation, and adsorbs to the Fh fraction in ferrihydrite-bacteria composites via the same molecular mechanism as the isolated endmember Fh. Jeon et al. (2003) also showed that Zn and Ni sorbed rapidly to the surface of hematite, with a secondary stage of relatively slow sorption occurring over several days; where Zn sorbed more strongly to hematite than Ni. The stronger sorption of Zn to Fe oxyhydroxides may be seen in the initial phase of this experiment (93.98% Zn vs. 65.95% Ni sorbed on Fh, and 97.03% Zn vs. 93.38% Ni sorbed on BF; Table 1) and may account for its slightly greater retention following diagenetic capsule treatments (Table 2).

It is also known that during the transformation of Fh to goethite and hematite, Ni can be sequestered into the more crystalline mineral products (Ford et al., 1997). As well, Zn initially sorbed to Fh shows decreased solubility after aging and heat-induced phase transformation of the Fh to more crystalline Fe oxyhydroxides (Martínez and McBride, 1998). In both of the studies above, lead (Pb) was excluded from the more crystalline, neo-formed phases following transformation from Fh, possibly because it was not structurally incorporated into the neo-formed products (Ford et al., 1997; Martínez and McBride, 1998). More recently, several studies have directly demonstrated that trace-metals, including Ni, can be structurally incorporated into crystalline Fe oxyhydroxides

by isomorphic substitution with structural Fe (e.g., Carvalho-E-Silva et al., 2003). Work to date, therefore, suggests that following initial inner-sphere adsorption to Fh, both Zn and Ni will be strongly bound to the neo-forming, more crystalline Fe oxyhydroxides. This may be the result of either similar inner-sphere surface adsorption complexes and/or via structural incorporation into the neo-forming mineral phases. Although the precise molecular mechanisms responsible for Zn and Ni retention during diagenesis will require further study, the importance of our preliminary assessment lies with the empirical observation of strong metal retention during simulated IF diagenesis.

4.3 Considerations on the diagenetic mobility of Zn and Ni and IF diagenesis

Potentially increased Ni mobility from Fh plus glucose and BF (Table 2) may result from reactions involving electron transfer, such as the reduction of solid-phase Fe(III) to soluble Fe(II) coupled to the oxidation of organic carbon (e.g., Frierdich et al., 2011; Posth et al., 2013b). The potential presence of siderite in the post-diagenetic sample ZnFh-2 (see also Köhler et al., 2013; Posth et al., 2013b) would indicate that both Fe(III) and Fe(II) are present for the diagenetic formation of more crystalline mineral phases. This scenario would effectively represent a situation where there is dissimilatory Fe(III) reduction occurring due to the interaction of buried organic carbon with initial ferric oxyhydroxides (Konhauser et al, 2005; Johnson et al., 2008; Craddock and Dauphas, 2011), and suggests that organic carbon in IF depositional settings could lead to slight increases in diagenetic Zn and Ni mobility.

Indeed, a recent focus has been placed on the importance of the interaction of aqueous Fe(II) with the surface of ferric oxyhydroxides. Such reactions have been shown

to cause phase changes, from initial Fe(OH)₃ to secondary lepidocrocite, goethite and magnetite, and this process is generally rapid, occurring within days (e.g., Hansel et al, 2005). Similar studies utilizing Fe isotope tracers show a complete exchange of Fe atoms between the aqueous Fe(II) and Fe(III) oxide pools (e.g., Handler et al., 2009). The effect of these reactions on trace-metal mobility, thus far, are best represented by Frierdich et al. (2011) and Frierdich and Catalano (2012) who demonstrated that trace-metal-free Fe(II) solutions mobilize Zn and Ni from the Fe(III) surface pools of Fe(III) oxides. However, only $\sim 9\%$ and $\sim 2\%$ of incorporated Ni is lost from goethite and hematite, respectively, during a rapid approach to equilibrium conditions (Frierdich and Catalano, 2012). This metal loss during re-equilibration is generally on the order of the largest relative remobilization values we observe for our simulated diagenesis experiments. Furthermore, this mobility is relatively minor and likely to be secondary with respect to uncertainties associated with K_D values predicted for the Precambrian. Moreover, the experiments above on trace metal mobilization by aqueous Fe(II) were performed on pure iron-oxide phases. It has subsequently been shown that the inclusion of insoluble elements (e.g., Al, Cr and Sn) within the Fe oxide lattice can impede Fe(II) induced phase changes (e.g., Al, Hansel et al., 2011) and significantly reduce the release of Ni and Zn by inhibiting Fe atom exchange with Fe(II) (Frierdich et al., 2012). Given these considerations, this is a scenario likely to be more relevant for naturally occurring Fe oxide phases comprising IF.

We also consider that diagenetic re-equilibration will most likely not occur under open system conditions. IF are often considered to behave as a closed system during diagenesis, because the pore-waters are insulated from overlying seawater by layers of chert and iron oxides with vastly different surface charges (see McConchie, 1987).

Essentially, neither cations nor anions would be able to freely diffuse back into the overlying water column. Instead, any Zn or Ni mobilized during the dehydration or reduction of primary ferric oxyhydroxides would most likely be rapidly sorbed by the neo-forming mineral phases (see section 4.2) or at least remain in the IF precursor sediment to be sorbed by recrystallized minerals during later burial diagenesis (e.g., Raiswell et al., 2011; Posth et al., 2013b; Köhler et al., 2013). Indeed, paleomarine reconstructions of seawater constituents from IF often rely on the preservation of marinelike rare earth element (REE) patterns as confirmation that authigenic seawater signatures were retained (e.g., Bau and Möller, 1993; Bolhar et al., 2004). These include the preservation of distinct Eu/Eu* anomalies observed in the cm-scale bands of the Kuruman IF (e.g., Bau and Dulski, 1996). IF mesobands also retain inter- and intra-band variability in trace metal abundances, including Zn and Ni (e.g., Pecoits et al., 2009), and small-scale variability in inter- and intra-layer Fe isotope composition of IF (e.g., Frost et al., 2007; Whitehouse and Fedo, 2007; Steinhoefel et al., 2010). Collectively, these studies indicate limited post depositional alteration and that IF, to at least some extent, behaved as closed systems during diagenesis. To this end Konhauser et al. (2009) showed Ni concentrations were very similar in co-occurring hematite and magnetite grains despite the necessity for magnetite to have formed via diagenetic reactions.

During the transformation of ferrihydrite to hematite it is difficult to determine whether transformation occurred via a solid-state pathway instigated via heating ferrihydrite in the dry state (as performed here) or via an aqueous pathway technically achieved with added water present. This is because water produced from the dry state dehydration may provide sufficient water to instigate the aqueous pathway (Cornell and

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Schwertmann, 2003). Indeed, in our capsule experiments with ~100 mg Fh (estimated formula Fe(OH)₃) about 25 mg of water were released during dehydration, and when our capsules were opened, there was a liquid overpressure that was collected along with the buffered rinse solution for assessing the mobility of Zn and Ni. This liquid overpressure, the 25 mg of water produced by the dehydration of Fh, should provide sufficient water to at least mimic IF diagenesis in a system that would almost certainly have been 'wet' to some extent. Additionally, within a very short reaction time (<14 days) there is up to 10% Fe(II) formed when organic carbon is present (Posth et al., 2013b). Considering our Fh with added glucose experiments, and conservatively assuming only 1% of the generated Fe(II) will be in aqueous form, we can estimate just how much Fe(II) would be enriched in the liquid phase. This would translate as follows: for ~1 mM of initial Fe(III) present per capsule (100 mg Fh), about 100 µmol could potentially dissolve as Fe(II) in the water formed during Fh dehydration, leaving 1 µmol of Fe(II) dissolved in 25 mg of water (i.e. 25μ l) - a concentration of 0.04 M Fe(II). As the capsules are a closed system this would be free to react during ongoing diagenesis, and as such, represent an Fe(II)-rich system where electron exchange could occur. This relatively conservative estimate for the Fe(II) concentrations available for diagenetic reaction in our capsule experiments are an order of magnitude greater than those used by Frierdich et al. (2011) of $2x10^{-4}$ to 10^{-3} M. Such conditions should provide more than adequate Fe(II) concentrations to promote redox driven recrystallization (as described in Frierdich et al., 2011), and potential subsequent trace metal mobility. However, overall Zn and Ni mobility in the presence of glucose and with BF is still low in our experiments, indicating IF should largely retain authigenic signatures and not depart strongly from initially sorbed paleomarine abundances. From an

empirical perspective, we emphasize that almost all Zn and Ni in our experiments was retained, even in capsules with organic carbon where magnetite and/or siderite are likely to have formed diagenetically (Posth et al., 2013b). Indeed, exposing the diagenetic mineral products to oxygenated conditions for 18 months, and thus inducing their re-oxidation to hematite, we still observed minimal mobilization of Ni or Zn. In other words, the trace metals sorbed to the sedimenting ferrihydrite particles in the water column remained in the sediment irrespective of secondary iron mineral. The observation of Zn-rich inclusions in sample ZnFh-2 further supports the stability of Zn and Ni following the secondary oxidation of reduced mineral phases.

5. Future work

We have expanded on the studies of Posth et al. (2013b) and Köhler et al. (2013) here as an important first step aimed at constraining our understanding of IF diagenesis and trace metal mobility. A current limitation of this work that restricts its absolute applicability to the Precambrian is the use of closed system conditions. Although IF were not "open" systems, dewatering of the initially hydrous sediments did indeed occur (Krapež et al., 2003). Favorably, here we observe limited mobility of both Zn and Ni during the dehydration of Fh and BF to hematite. Accordingly, future studies on IF diagenesis and the diagenetic mobility of trace metals should examine the effects of more open system conditions. Furthermore, many IF were altered during metamorphism to P-T conditions up to amphibolite facies (500-600°C and >3 kbar), and as such, an assessment of metal mobility under these conditions will ultimately be required. Future experiments may also include other trace metals, such as Cr, Co, and REE, and varied types of organic carbon to assess what role more recalcitrant carbon may have on Fe(III) reduction and potential trace metal mobility. A number of studies have been published with regards to molecularlevel trace metal complexation by Fe oxyhydroxides (e.g., for a review see Brown and Parks, 2001); for Zn sorbed on ferrihydrite see Waychunas et al., 2002, 2003; for Ni sorbed on ferrihydrite see Xu et al., 2007, and for Ni incorporated into goethite see Carvalho-E-Silva et al., 2003). Increasingly, these studies have also looked at the trace metal complexation by bacteria (e.g., Boyanov et al., 2003; Toner et al., 2005; Moon and Peacock, 2011). Given the utility of such studies, future work should employ similar synchrotron radiation-based techniques (e.g., XAS) to help track the mobility of trace metals during Fe oxyhydroxide mineralogical transformation. This approach would help determine the mechanism(s) underpinning trace metal retention in the final neo-formed phases. Despite the multitude of potential work, we emphasize that our study here is a necessary first step, and the finding of Zn and Ni being immobile during simulated diagenesis is promising for the continued use of IF as paleomarine proxies.

6. Conclusions

Pressure and temperature treatments of Fh and carbon at conditions realistic for IF diagenesis (170°C and 1.2 kbar) demonstrated that Zn and Ni remained largely immobile. This suggests that IF retain authigenic signatures, and that paleomarine Zn and Ni values derived from the IF record (Konhauser et al., 2009; Robbins et al., 2013) are likely to be within the correct order of magnitude. Overall, it is consistent with indications from REE patterns and other geochemical data that IF faithfully record authigenic enrichments

derived from ambient seawater, thus providing further impetus for exploration of IF as an important archive of the chemical history of seawater and early Earth evolution.

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References

- Bau, M., Möller, P., 1993. Rare earth element systematics of the chemically precipitated component in Early Precambrian iron formations and the evolution of the terrestrial atmosphere-hydrosphere-lithosphere system. Geochim. Cosmochim. Acta 57, 2239-2249.
- Bau, M., Dulski, P., 1996. Distribution of yttrium and rare-earth elements in the Penge and Kuruman iron-formations, Transvaal Supergroup, South Africa. Precambrian Res. 79, 37-55.

- Becker, R.H., Clayton, R.N., 1976. Oxygen isotope study of a Precambrian banded ironformation, Hamersley Range, Western Australia. Geochim. Cosmochim. Acta 40, 1153-1165.
- Bekker, A., Slack, J.F, Planavsky, N., Krapež, B., Hofmann, A., Konhauser, K.O., Rouxel, O.J., 2010. Iron formation: the sedimentary product of a complex interplay among mantle, tectonic, oceanic, and biospheric processes. Econ. Geol. 105, 467-508.
- Binsted, N., Pack, M.J., Weller, M.T., Evans, J., 1996. Combined EXAFS and Powder Diffraction Analysis. J. Am. Chem. Soc. 118, 10200-10210.
- Binsted, N., 1998. EXCURV98 The Manual. CLRC Daresbury Laboratory Program., Warrington, United Kingdom.
- Bolhar, R., Kamber, B.S., Moorbath, S., Fedo, C.M., Whitehouse, M.J., 2004.Characterisation of early Archaean chemical sediments by trace element signatures.Earth. Planet. Sci. Lett. 222, 43-60.
- Booth, C.H., Hu, Y.-J., 2009. Confirmation of standard error analysis techniques applied to EXAFS using simulations. J. Phys. Conf. Ser. 190, 1-6.
- Boyanov, M.I., Kelly, S.D., Kemner, K.M., Bunker, B.A., Fein, J.B., Fowle, D.A., 2003. Adsorption of cadmium to *Bacillus subtilis* bacterial cell walls: A pH-dependent Xray absorption fine structure spectroscopy study. Geochim. Cosmochim. Acta 67, 3299-3311.
- Brown, G.E., Jr., Parks, G.A., 2001. Sorption of Trace Elements on Mineral Surfaces:Modern Perspectives from Spectroscopic Studies, and Comments on Sorption in theMarine Environment. Int. Geol. Rev. 43, 963-1073.

- Carvalho-E-Silva, M.L., Ramos, A.Y., Tolentino, H.C.N., Enzweiler, J., Netto, S.M., Alves, M.C.M., 2003. Incorporation of Ni into natural goethite: An investigation by X-ray absorption spectroscopy. Am. Mineral. 88, 876-882.
- Cornell, R.M., Schwertmann, U., 2003. The iron oxides: structure, properties, reactions, occurrences and uses, 2nd edition. Weinheim, Viley-VCH Verlag, pp. 1-664.
- Craddock, P.R., Dauphas, N., 2011. Iron and carbon isotope evidence for microbial iron respiration throughout the Archean. Earth Planet. Sci. Lett. 303, 121-132.
- Dupont, C.L., Butcher, A., Valas, R.E., Bourne, P.E., Caetano-Anollés, G., 2010. History of biological metal utilization inferred through phylogenomic analysis of protein structures. Proc. Natl. Acad. Sci. 107, 10567-10572.
- Eickhoff, M., 2013. Biomarker, Trace Metal and Diagenesis Studies with Fe-Metabolizing Bacteria to Better Understand Their Role on Early Earth. PhD Thesis. University of Tübingen, Germany.
- Eickhoff, M., Obst, M., Schröder, C., Hitchcock, A.P., Tyliszczak, T., Martinez, R.E.,
 Robbins, L.J., Konhauser, K.O., Kappler, A., 2014. Nickel partitioning in biogenic
 and abiogenic ferrihydrite: The influence of silica and implications for ancient
 environments. Geochim. Cosmochim. Acta 140, 65-79.
- Ford, R.G., Bertsch, P.M., Farley, K.J., 1997. Changes in Transition and Heavy Metal Partitioning during Hydrous Iron Oxide Aging. Environ. Sci. Technol. 31: 2028-2033.
- Frierdich, A.J., Luo, Y., Catalano, J.G., 2011. Trace element cycling through iron oxide minerals during redox driven dynamic recrystallization. Geology 39, 1083-1086.

- Frierdich, A.J., Catalano, J.G., 2012. Controls on Fe(II)-Activated Trace Element Release from Goethite and Hematite. Environ. Sci. Technol. 46, 1519-1526.
- Frierdich, A.J., Scherer, M.M., Bachman, J.E., Engelhard, M.H., Rapponotti, B.W., Catalano, J.G., 2012. Inhibition of trace element release during Fe(II)-activated recrystallization of Al-, Cr-, and Sn-substituted goethite and hematite. Environ. Sci. Technol. 46, 10031-10039.
- Frost, C.D., von Blanckenburg, F., Schoenberg, R., Frost, B.R., Swapp, S.M., 2007.
 Preservation of Fe isotope heterogeneities during diagenesis and metamorphism of banded iron formation. Contrib. Mineral. Petrol. 153, 211-235.
- Gole, M.J., Klein, C., 1981. Banded Iron-Formations though much of Precambrian Time.J. Geol. 89, 169-183.
- Handler, R.M., Beard, B.L., Johnson, C.M., Scherer, M.M., 2009. Atom Exchange between Aqueous Fe(II) and Goethite: An Fe Isotope Tracer Study. Environ. Sci. Technol. 43, 1102-1107.
- Hansel, C.M., Benner, S.G., Fendorf, S., 2005. Competing Fe(II)-Induced Mineralization Pathways of Ferrihydrite. Environ. Sci. Technol. 39, 7147-7153.
- Hansel, C.M., Learman, D.R., Lentini, C.J., Ekstrom, E.B., 2011. Effect of adsorbed and substituted Al on Fe(II)-induced mineralization pathways on ferrihydrite. Geochim. Cosmochim. Acta 75, 4653-4666.
- Heimann, A., Johnson, C.M., Beard, B.L., Valley, J.W., Roden, E.E., Spicuzza, M.J., Beukes, N.J., 2010. Fe, C, and O isotope compositions of banded iron formation carbonates demonstrate a major role for dissimilatory iron reduction in ~2.5 Ga marine environments. Earth Planet. Sci. Lett. 294, 8-18.

- Jambor, J.L., Dutrizac, J.E., 1998. Occurrence and constitution of natural and synthetic ferrihydrite, a wide-spread iron oxyhydroxides. Chem. Rev. 98, 2549-2585.
- Jeon, B.-H., Dempsey, B.A., Burgos, W.D., Royer, R.A., 2003. Sorption kinetics of Fe(II), Zn(II), Co(II), Ni(II), Cd(II) and Fe(II)/Me(II) onto hematite. Water Res. 37, 4135-4142.
- Johnson, C.M., Beard, B.L., Klein, C., Beukes, N.J., Roden, E.E., 2008. Iron isotopes constrain biologic and abiologic processes in banded iron formation genesis: Geochim. Cosmochim. Acta 72, 151-169.
- Klein, C., 2005. Some Precambrian banded iron-formations (BIFs) from around the world: Their age, geologic setting, mineralogy, metamorphism, geochemistry and origins. Am. Mineral. 90: 1473-1499.
- Köhler, I., Konhauser, K.O., Papineau, D., Bekker, A., Kappler, A., 2013. Biological carbon precursor to diagenetic siderite with spherical structures in iron formations. Nat. Commun. doi. 10.1038/ncomms2770.
- Konhauser, K.O., 2007. Introduction to Geomicrobiology. Oxford, United Kingdom, Blackwell Publishing. pp. 1-425.
- Konhauser, K.O., Newman, D.K., Kappler, A., 2005. The potential significance of microbial Fe(III) reduction during deposition of Precambrian banded iron formations: Geobiology 3, 167-177.
- Konhauser, K.O., Pecoits, E., Lalonde, S.V., Papineau, D., Nisbet, E.G. Barley, M.E., Arndt, N.T., Zahnle, K., Kamber, B.Z., 2009. Oceanic nickel depletion and a methanogen famine before the Great Oxidation Event. Nature 458, 750-753.

- Krapež, B., Barley, M.E., Pickard, A.L., 2003. Hydrothermal and resedimented origins of the precursor sediments to banded iron formation: sedimentological evidence from the Early Palaeoproterozoic Brockman Supersequence of Western Australia. Sedimentology 50: 979-1011.
- Lee, S., Anderson, P.R., 2005. EXAFS study of Zn sorption on hydrous ferric oxide over extended reaction time. J. Colloid Interf. Sci. 286, 82-89.
- Martínez, C.E., McBride, M.B., 1998. Coprecipitates of Cd, Cu, Pb, and Zn in iron oxides: solid phase transformation and metal solubility after aging and thermal treatment. Clay. Clay. Miner. 46: 537-545.
- McConchie, D., 1987. The geology and geochemistry of the Joffre and Whaleback shale members of the Brockman Iron Formation, Western Australia. In: Appel, P.W.U., LaBerge, G.L. (Eds.), Precambrian iron-formations. Athens, Theophrastus, pp. 541-597.
- Miyano, T., Klein, C., 1984. Evaluation of the stability relations of amphibole asbestos in metamorphosed iron formations. Min. Geol. 33, 213-222.
- Miyano, T., 1987. Diagenetic to low-grade metamorphic conditions of Precambrian iornformations. In: Appel, P.W.U., LaBerge, G.L. (Eds.), Precambrian iron-formations. Athen, Theophrastus, pp. 155-186.

Mloszewska, A.M., Pecoits, E., Cates, N.L., Mojzsis, S.J., O'Neil, J., Robbins, L.J.,
Konhauser, K.O., 2012. The composition of Earth's oldest iron formations: The
Nuvvuagittuq Supracrustal Belt (Québec, Canada). Earth. Planet. Sci. Lett. 317-318:
331-342.

- Moon, E.M., Peacock, C.L., 2011. Adsorption of Cu(II) to *Bacillus subtilis*: A pHdependent EXAFS and thermodynamic modelling study. Geochim. Cosmochim. Acta 75, 6705-6719.
- Moon, E.M., Peacock, C.L., 2012. Adsorption of Cu(II) to ferrihydrite and ferrihydritebacteria composites: Importance of the carboxyl group for Cu mobility in natural environments. Geochim. Cosmochim. Acta 92, 203-219.
- Moon, E.M., Peacock, C.L., 2013. Modelling Cu(II) adsorption to ferrihydrite and ferrihydrite-bacteria composites: Deviation from additive adsorption in the composite sorption system. Geochim. Cosmochim. Acta 104, 148-164.
- Pandya, K.I., O'Grady, W.E., Corrigan, D.A., McBreen, J., Hoffman, R.W., 1990.
 Extended X-ray Absorption Fine Structure Investigations of Nickel Hydroxides. J.
 Phys. Chem. 94, 21-26.
- Pecoits, E., Gingras, M.K., Barley, M.E., Kappler, A., Posth, N.R., Konhauser, K.O.,
 2009. Petrography and geochemistry of the Dales Gorge banded iron formation:
 Paragenetic sequence, source, and implication for palaeo-ocean chemistry.
 Precambrian Res. 172, 163-187.
- Pecoits, E., Smith, M.L., Catling, D.C., Philippot, P., Kappler, A., and Konhauser, K.O.,
 2015. Atmospheric hydrogen peroxide and Eoarchean iron formations. Geobiology,
 13, 1-14.
- Perry, E.C., Tan, F.C., Morey, G.B., 1973. Geology and Stable Isotope Geochemistry of the Biwabik Iron Formation, Northern Minnesota: Econ. Geol. 68, 1110-1125.

- Posth, N.R., Konhauser, K.O., and Kappler, A., 2013a. Microbiological processes in banded iron formation deposition. Sedimentology 60, 1733-1754.
- Posth, N.R., Köhler, I., Swanner, E.D., Schröder, C., Wellmann, E., Binder, B.,
 Konhauser, K.O., Neumann, U., Berthold, C., Nowak, M., Kappler, A., 2013b.
 Simulating Precambrian banded iron formation diagenesis. Chem. Geol. 362, 66-73.
- Posth, N.R., Canfield, D.E., Kappler, A., 2014. Biogenic Fe(III) minerals: from formations to diagenesis and preservation in the rock record. Earth-Sci. Rev. 135, 103-121.
- Raiswell, R., Reinhard, C.T., Derkowski, A., Owens, J., Bottrell, S.H., Anbar, A.D., Lyons, T.W., 2011. Formation of syngenetic and early diagenetic iron mineral in the late Archean Mt. McRae Shale, Hamersley Basin, Australia: New insights on the patterns, controls, and paleoenvironmental implications of authigenic mineral formation. Geochim. Cosmochim. Acta 75, 1072-1087.
- Ravel, B., Newville, M., 2005. ATHENA, ARTEMIS, HEPHAESTUS: data analysis for Xray absorption spectroscopy using IFEFFIT. J. Synchrotron Radiat. 12, 537-541.
- Robbins, L.J., Lalonde, S.V., Saito, M.A., Planavsky, N.J., Mloszewska, A.M., Pecoits,
 E., Scott, C., Dupont, C.L., Kappler, K.A., Konhauser, K.O., 2013. Authigenic iron oxide proxies for marine zinc over geological time and implications for eukaryotic metallome evolution. Geobiology 11, 295-306.
- Saito, M.A., Sigman, D.M., Morrel, F.M.M., 2003. The bioinorganic chemistry of the ancient ocean: the co-evolution of cyanobacterial metal requirements and biogeochemical cycles at the Archean-Proterozoic boundary? Inorg. Chim. Acta 356: 308-318.

- Schwertmann, U., 1966. Inhibitory Effect of Soil Organic Matter on the Crystallization of Amorphous Ferric Hydroxide. Nature 212, 645-646.
- Scott, C., Planavsky, N.J., Dupont, C.L., Kendall, B., Gill, B., Robbins, L.J., Husband,
 K.F., Arnold, G.L., Wing, B., Poulton, S.W., Bekker, A., Anbar, A.D., Konhauser,
 K.O., Lyons, T.W., 2013. Bioavailability of zinc in marine systems through time:
 Nature Geosci. 6, 125-128.
- Smith, A.J.B., Beukes, N.J., Gutzmer, J., 2013. The composition and depositional environments of Mesoarchean Iron Formations of the West Rand Group of the Witwatersrand Supergroup, South Africa. Econ. Geol. 108: 111-134.
- Steinhoefel, G., von Blanckenburg, F., Horn, I., Konhauser, K.O., Beukes, N.J., Gutzmer, J., 2010. Deciphering formation processes of banded iron formations from the Transvaal and the Hamersley successions by combined Si and Fe isotope analysis using femtosecond laser ablation. Geochim. Cosmochim. Acta 74, 2677-2696.
- Stern, E.A., 1993. Number of relevant independent points in x-ray-absorption finestructure spectra. Phys. Rev. B 48, 9825-9827.
- Stookey, L.L., 1970. Ferrozine a new spectrophotometric reagent for iron. Anal. Chem. 42, 79-781.
- Tomić, S., Searle, B.G., Wander, A., Harrison, N.M., Dent, A.J., Mosselmans, J.F.W.,
 Inglesfield, J.E., 2005. New Tools for the Analysis of EXAFS: The DL_EXCURV
 Package. CCLRC Technical Report DL-TR-2005-001. Daresbury, United Kingdom.
- Toner, B., Manceau, A., Marcus, M.A., Millet, D.B., and Sposito, G., 2005. Zinc Sorption by a Bacterial Biofilm. Environ. Sci. Technol. 39, 8288-8294.

- Toner, B.M., Berquó, T.S., Michel, F.M., Sorensen, J.V., Templeton, A.S., Edwards, K.J., 2012. Mineralogy of Iron Microbial Mats from Loihi Seamount. Front. Microbiol. 3, 1-18.
- Trivedi, P., Axe, L., 2001. Ni and Zn Sorption to Amorphous versus Crystalline Iron Oxides: Macroscopic Studies. J. Colloid Interf. Sci. 224, 221-229.
- Trivedi, P., Axe, L., Tyson, T.A., 2001. An Analysis of Zinc Sorption to Amorphous versus Crystalline Iron Oxide Using XAS. J. Colloid. Interf. Sci. 244, 230-238.
- Waychunas, G.A., Fuller, C.C., Davis, J.A., 2002. Surface complexation and precipitate geometry for aqueous Zn(II) sorption on ferrihydrite I: X-ray absorption extended fine structure spectroscopy analysis. Geochim. Cosmochim. Acta 66: 1119-1137.
- Waychunas, G.A., Fuller, C.C., Davis, J.A., Rehr, J.J., 2003. Surface complexation and precipitate geometry for aqueous Zn(II) sorption on ferrihydrite: II. XANES analysis and simulation. Geochim. Cosmochim. Acta 67: 1031-1043.
- Whitehouse, M.J., Fedo, C.M., 2007. Microscale heterogeneity of Fe isotopes in >3.71 Ga banded iron formation from the Isua Greenstone Belt, southwest Greenland. Geology 35, 719-722.
- Williams, R.J.P., da Silva, J.J.R.F., 1996. The Natural Selection of the Chemical Elements. Bath Press Ltd., Great Britain.
- Wu, W., Swanner, E.D., Hao, L., Zeitvogel, F., Obst, M., Pan, Y., Kappler, A., 2014.
 Characterization of the physiology and cell-mineral interactions of the marine anoxygenic phototrophic Fe(II) oxidizer *Rhodovulum iodosum* - implications for Precambrian banded iron formation deposition. FEMS Microbiol. Ecol. 88, 503-515.

Xu, Y., Axe, L., Boonfueng, T., Tyson, T.A., Trivedi, P., Pandya, K., 2007. Ni(II) complexation to amorphous hydrous ferric oxide: An X-ray absorption spectroscopy study. J. Colloid Interf. Sci. 314, 10-17.

Table and Figure Captions

Table 1. Adsorption of Zn and Ni to 2-line synthetic, and biogenic ferrihydrite. Errors are presented as percentages of two standard deviations of the ICP-OES analyses; molar values are presented without errors.

Table 2. Amount of Zn or Ni mobilized from Fe minerals and percent retained following 14 days incubation at 170°C and 1.2 kbar. Unincubated controls are denoted by a precursor C. The trace metal (TM) in column names refers to either Zn or Ni; a description of capsule contents is provided including the TM adsorbed (Zn or Ni), type of Fe particle (Fh or BF) and if glucose was added (G). Errors in TM retained are presented as percentages based on two standard deviations of the ICP-OES analyses. For all samples where the respective plus or minus errors in TM retained differed, we have reported the larger error value.

Table 3. Ni K-edge EXAFS fit for Ni-sorbed ferrihydrite. Where N is the number of atoms in a shell, R is the distance of the atom in a shell from the Ni central absorber, σ is the Debye-Waller factor (DWF), and θ and ϕ are the spherical coordinates of each atom in a shell. EF is the correction to the Fermi energy value set in ATHENA. During

optimization, values in italics were held constant and DWF's for the oxygen shells were constrained to be equal; N_{ind} (number of independent data points) = 14, N_{pars} (number of parameters optimized) = 12.

Figure 1. (a) EXAFS and (b) Fourier transform of the EXAFS for Ni sorbed to ferrihydrite. Solid lines are data, dotted lines are fit. (c) Molecular cluster used to fit the EXAFS data, where O_1 - O_6 = octahedral oxygen atoms in Ni first shell coordination sphere and Fe₁-Fe₂ = iron in ferrihydrite Fe octahedra.

Figure 2. Micro-XRD patterns for NiBF1 and NiFh-1, collected after 18 months of sample storage at 4°C in oxic conditions. Both samples have reflections characteristic of hematite, but the different in peak broadness suggests hematite in NiBF1 is a less crystalline phase. NiBF1 additionally has reflections consistent with lepidocrocite (γ -FeOOH), an Fe(III) oxyhydroxide mineral.

Figure 3. Electron microprobe maps showing the distribution and weight percent for (A) Fe and (B) Zn in sample ZnFh-2, and (C) Fe and (D) Ni in sample NiFh-2. Color bars indicate the weight percent ($\pm 1\%$) of the corresponding element.

Figure 4. Electron microprobe maps showing the distribution and weight percent for (A) Fe and (B) Zn in sample ZnBF2, and (C) Fe and (D) Ni in sample NiBF2. Color bars indicate the weight percent (±1%) of the corresponding element.

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Mineral + TM Combination	Mass of Fe particles (g)	Fe (mol)	Initial Solution [TM] (mg/L)	Final Solution [TM] (mg/L)	Volume of TM solution (L)	TM adsorbed (x10 ⁻⁵ mol)	% TM adsorbed	TM/Fe-particles (x10 ⁻⁵ mol/g)	mol Zn/mol Fe (x10 ⁻³)	mol Ni/ mol Fe (x10 ⁻³)
Zn on 2-line synthetic										
ferrihydrite	1.0224	0.0096	3.24 ± 0.08	0.20 ± 0.01	0.9987	4.646	93.98 ± 0.54	4.544	4.856	-
Ni on 2-line synthetic										
ferrihydrite	0.9965	0.0093	3.25 ± 0.07	1.11 ± 0.02	1.0019	3.653	65.95 ± 1.29	3.666	-	3.918
Zn on biogenic										
ferrihydrite	0.5023	0.0044	3.23 ± 0.16	0.10 ± 0.01	0.4952	2.373	97.03 ± 0.60	4.725	5.410	-
Ni on biogenic										
ferrihydrite	0.4769	0.0042	3.20 ± 0.14	0.21 ± 0.01	0.4964	2.527	93.38 ± 0.52	5.299	-	6.067

Table 2.

Capsule	Mass Fe particles (g)	Mass Glucose (g)	TM/Fe- particels (x10 ⁻⁵ mol/g)	TM/capsule (x10 ⁻⁶ mol)	Zn released (x10 ⁻⁹ mol)	Ni released (x10 ⁻⁹ mol)	%TM retained	TM ICP-OES σ (mg/L)	±2σ TM ICP-OES (x10 ⁻⁹ mol)	±2σ TM retained (%)	Capsule Contents	C:Fe ratio
ZnFh-2	0.1000	0.0041	4.544	4.543	0.000	11.498	100.0	0.0031	94.79	2.1	Zn+Fh+G	0.1449
ZnFh-3	0.1000	-	4.544	4.543	0.000	0.327	100.0	0.0031	94.79	2.1	Zn+Fh	N/A
ZnFh-4	0.1000	-	4.544	4.546	0.000	0.000	100.0	0.0024	73.38	1.6	Zn+Fh	N/A
C-ZnFh1	0.1000	0.0041	4.544	4.542	0.000	0.000	100.0	0.0064	195.7	4.3	Zn+Fh+G	0.1446
C-ZnFh2	0.1000	0.0041	4.544	4.543	0.000	0.000	100.0	0.0030	91.73	2.0	Zn+Fh+G	0.1453
C-ZnFh3	0.1000	-	4.544	4.544	0.000	0.000	100.0	0.0033	100.9	2.2	Zn+Fh	N/A
C-ZnFh4	0.1000	-	4.544	4.546	0.000	0.000	100.0	0.0029	88.67	2.0	Zn+Fh	N/A
ZnBF2	0.0341	-	4.725	1.610	0.000	0.499	100.0	0.0036	110.1	6.8	Zn+BF	0.1792
ZnBF3	0.0363	-	4.725	1.713	5.724	0.164	99.7	0.0020	61.15	3.9	Zn+BF	0.1792
C-ZnBF1	0.0353	-	4.725	1.666	14.863	1.338	99.1	0.0029	88.67	6.2	Zn+BF	0.1792
C-ZnBF2	0.0351	-	4.725	1.658	0.000	0.000	100.0	0.0039	119.2	7.2	Zn+BF	0.1792
NiFh-1	0.1000	0.0041	3.666	3.666	0.000	193.557	94.7	0.0229	780.3	26.6	Ni+Fh+G	0.1456
NiFh-2	0.1000	0.0041	3.666	3.665	0.000	298.692	91.9	0.0361	1230	41.7	Ni +Fh+G	0.1445
NiFh-4	0.1001	-	3.666	3.669	0.000	0.166	100.0	0.0003	10.22	0.3	Ni+Fh	N/A
C-NiFh1	0.1001	0.0041	3.666	3.669	0.000	6.847	99.8	0.0022	74.97	2.2	Ni+Fh+G	0.1440
C-NiFh2	0.1001	0.0040	3.666	3.669	0.000	6.279	99.8	0.0012	40.89	1.3	Ni+Fh+G	0.1433
C-NiFh3	0.1001	-	3.666	3.668	0.000	8.665	99.8	0.0010	34.08	1.2	Ni+Fh	N/A
C-NiFh4	0.1000	-	3.666	3.667	0.000	5.856	99.8	0.0011	37.48	1.2	Ni+Fh	N/A
NiBF1	0.0355	-	5.299	1.881	0.000	61.090	96.8	0.0075	255.6	16.8	Ni+BF	0.1792
NiBF2	0.0354	-	5.299	1.877	0.000	66.660	96.4	0.0072	245.3	16.6	Ni+BF	0.1792
C-NiBF1	0.0355	-	5.299	1.883	0.000	19.771	98.9	0.0009	30.67	2.7	Ni+BF	0.1792
C-NiBF2	0.0352	-	5.299	1.866	0.000	20.701	98.9	0.0037	126.1	7.9	Ni+BF	0.1792

Tab	le 3.
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$ \begin{matrix} N_{O} \\ R(Ni- O_{1}) \\ 2\sigma^{2} \\ \theta, \phi \end{matrix} $	$\begin{array}{c} N_{O} \\ R(Ni- O_{2}) \\ 2\sigma^{2} \\ \theta, \phi \end{array}$	$ \begin{array}{c} N_{O} \\ R(Ni- O_{3}) \\ 2\sigma^{2} \\ \theta, \phi \end{array} $	$\begin{array}{c} N_{O} \\ R(Ni- \\ O_{4}) \\ 2\sigma^{2} \\ \theta, \phi \end{array}$	$\begin{matrix} N_{0} \\ R(Ni-O_{5}) \\ 2\sigma^{2} \\ \theta, \phi \end{matrix}$	$ \begin{array}{c} N_{0} \\ R(\text{Ni-} O_{6}) \\ 2\sigma^{2} \\ \theta, \phi \end{array} $	$\begin{array}{c} N_{Fe} \\ R(Ni-Fe_1) \\ 2\sigma^2 \\ \theta, \phi \end{array}$	$\begin{array}{c} N_{Fe} \\ R(Ni-Fe_2) \\ 2\sigma^2 \\ \theta, \phi \end{array}$	EF	R (%)	Fit Index	Reduced Chi ²
1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0				
2.07 0.009	2.00 0.009	2.09 0.009	2.05 0.009	2.08 0.009	2.04 0.009	3.01 0.010	3.15 0.006	3.73	32.8	1.06	39
90, 0	90, 90	90, 180	90, 270	0, 0	180, 0	55, 100	55, 170				



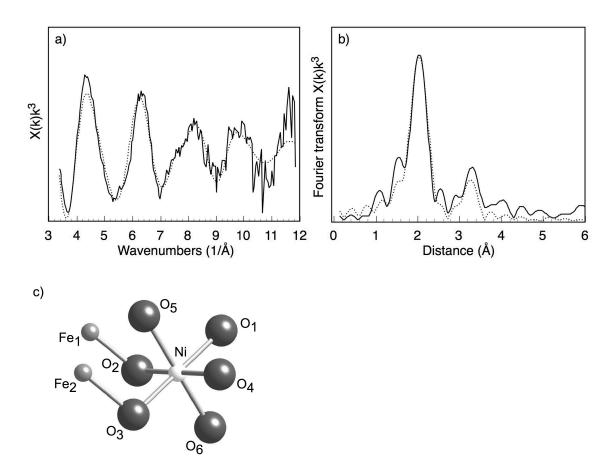


Figure 2.

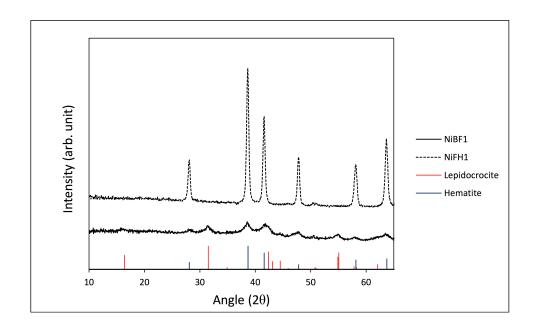


Figure 3.

