



Length Dependent Toxicity of TiO₂ Nanofibers: Mitigation via Shortening

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Keywords:	macrophages, nanofibers, safety by design, titanium, aspect ratio
Abstract:	<p>Length and aspect ratio represent important toxicity determinants of fibrous nanomaterials. We have previously shown that anatase TiO₂ nanofibers (TiO₂ NF) cause a dose-dependent decrease of cell viability as well as the loss of epithelial barrier integrity in polarized airway cell monolayers. Herein we have investigated the impact of fiber shortening, obtained by ball-milling, on the biological effects of TiO₂ NF. Long TiO₂ NF (L-TiO₂ NF) were more cytotoxic than their shortened counterparts (S-TiO₂ NF) towards alveolar A549 cells and bronchial 16HBE cells. Moreover, L-TiO₂ NF affected the trans-epithelial electrical resistance of 16HBE monolayers. This effect was associated with altered distribution of tight-junction proteins and also mitigated by fiber shortening. Macrophages efficiently internalized S-TiO₂ NF but not L-TiO₂ NF, which caused cell stretching and deformation. In macrophages S-TiO₂ NF enhanced the expression of pro-inflammatory genes, NO production and cytokine secretion, which was significantly inhibited by the phagocytosis inhibitor cytochalasin D. <i>In vivo</i> experiments indicated length-dependent toxicity in both the lungs and peritoneal cavity of mice, leading to significant increase in markers of inflammation in animals treated with L-TiO₂ NF. It is concluded that fiber shortening mitigates NF detrimental effects on cell viability and epithelial barrier competence. As far as inflammation is concerned, shortening enhances phagocytosis and macrophage activation <i>in vitro</i> but prevents the increase of inflammatory cytokines upon <i>in vivo</i> exposure. These data suggest that fiber shortening may represent an effective safe-by-design strategy for mitigating TiO₂ NF toxic effects both <i>in vitro</i> and <i>in vivo</i>.</p>

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Length Dependent Toxicity of TiO₂ Nanofibers: Mitigation via Shortening

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A mitigation strategy for the toxicity of TiO₂ nanofibers: high aspect ratio reduction

Length and aspect ratio represent important toxicity determinants of fibrous nanomaterials. We have previously shown that anatase TiO₂ nanofibers (TiO₂ NF) cause a dose-dependent decrease of cell viability as well as the loss of epithelial barrier integrity in polarized airway cell monolayers. Herein we have investigated the impact of fiber shortening, obtained by ball-milling, on the biological effects of TiO₂ NF. Long TiO₂ NF (L-TiO₂ NF) were more cytotoxic than their shortened counterparts (S-TiO₂ NF) towards alveolar A549 cells and bronchial 16HBE cells. Moreover, L-TiO₂ NF affected the trans-epithelial electrical resistance of 16HBE monolayers. This effect was associated with altered distribution of tight-junction proteins and also mitigated by fiber shortening. Macrophages efficiently internalized S-TiO₂ NF but not L-TiO₂ NF, which caused cell stretching and deformation. In macrophages S-TiO₂ NF enhanced the expression of pro-inflammatory genes, NO production and cytokine secretion, which was significantly inhibited by the phagocytosis inhibitor cytochalasin B. *In vivo* experiments indicated length-dependent toxicity in both the lungs and peritoneal cavity of mice, leading to significant increase in markers of inflammation in animals treated with L-TiO₂ NF. It is concluded that fiber shortening mitigates NF detrimental effects on cell viability and epithelial barrier competence *in vitro* as well as the increase of inflammatory cytokines and granulocytes influx upon *in vivo* exposure. These data suggest that fiber shortening may represent an effective safe-by-design strategy for mitigating TiO₂ NF toxic effects both *in vitro* and *in vivo*.

Keywords: aspect ratio; macrophages; nanofibers; safety-by-design; titanium dioxide

1. Introduction

High aspect ratio nanostructures (HARN) such as nanotubes, nanofibers, nanowires, are increasingly used, either pure or in composites, in many industrial applications, such as electronics and photovoltaics. However, whilst nanofibers continue to show their utility in many applications, the morphological similarity of these materials to pathogenic

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3 fibers such as asbestos still represents a serious concern about the potential health
4 implications of exposure. This association, based on morphology, relies upon the
5 structure-activity relationship, known as the “fiber pathogenicity paradigm” (FPP),
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7 which identifies three critical features that are required for a fibrous particle to present a
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9 fiber-type health hazard: i) aspect ratio and length (dimension), ii) persistence of a
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11 particle in the biological environment and its resistance to breakage and dissolution
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13 (durability) and, most crucially for consideration of risk, iii) the exposure to the particle
14
15 in question (dose) (Donaldson and Tran, 2004). Dimension, durability and dose, or the 3
16
17 Ds, provide the cornerstone of the FPP and have been reviewed alongside other
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19 determinants of particle-induced toxicity (Donaldson *et al.*, 2011, Donaldson and
20
21 Poland, 2013, Donaldson *et al.*, 2013).

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29 The focus of this study is the role of length in the pathogenicity of TiO₂ nanofibers
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31 (TiO₂ NF). The role of length in fiber pathogenicity, originally proposed by Stanton
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33 (Stanton *et al.*, 1977) and applied to more recent evaluations of nanofibers (Schinwald
34
35 *et al.*, 2012a), is in part reflected by the World Health Organisation which defines a
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37 minimum length and aspect ratio (5 µm and 3:1, respectively) for respirable fibers. Over
38
39 the years, numerous experimental studies have investigated *in vivo* the contribution of
40
41 length to fiber toxicity by either comparing samples containing different proportions of
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43 long mineral fibers (Davis *et al.*, 1991) or modifying their length (e.g. via ball milling)
44
45 to assess the impact on toxicity (Kuschner and Wright, 1976, Davis *et al.*, 1986b, Davis
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47 and Jones, 1988, Donaldson *et al.*, 1989, Goodglick and Kane, 1990). Those studies
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49 have usually found that fiber shortening causes mitigation or, in some cases, virtual
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51 absence of toxicity.
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58 Although the application of these conclusions to the biological effects of nanomaterials
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3 has been under scrutiny, most studies indicate that length and aspect ratio are important
4 toxicity determinants also for nanofibers (Poland *et al.*, 2008b, Rotoli *et al.*, 2009,
5 Poland *et al.*, 2011, Ji *et al.*, 2012, Schinwald *et al.*, 2012b), with long fibers exerting
6 higher toxicity than shorter ones. A challenge in these studies is the needing of
7 comparing long and short versions of the same material that ideally differ only in terms
8 of length instead of comparing two separate samples obtained from different sources
9 (e.g. batches, processes or manufacturers).

10
11
12 One of the few studies that addresses this issue concerns laboratory-made TiO₂
13 nanobelts of different length (Hamilton *et al.*, 2009). TiO₂ HARN, either pure or as
14 composite components, are increasingly used in a variety of fields in the form of
15 nanotubes, nanofibers or nanobelts. Although the toxic properties of each form have not
16 been characterized in detail, TiO₂ nanobelts have been described to cause cytotoxicity in
17 vitro, inflammasome activation and airway inflammation after respiratory exposure,
18 (Hamilton *et al.*, 2009, Xia *et al.*, 2013, Hamilton *et al.*, 2014, Tilton *et al.*, 2014, Jessop
19 *et al.*, 2017). Short nanobelts have been described to exert milder toxic effects than
20 longer ones (Hamilton *et al.*, 2009, Porter *et al.*, 2013). Recently, some information on
21 the effects of TiO₂ nanofibers after oral administration *in vivo* has been also provided
22 (Gato *et al.*, 2017, Bartel *et al.*, 2018, Hunter *et al.*, 2018), indicating that mild renal and
23 hepatic toxicity is detected along with an increased expression of genes involved in
24 immunity and inflammation, although these studies did not assess the relationship
25 between fiber length and biological effects. We have recently characterized the toxic
26 properties of TiO₂ nanofibers of industrial origin (Allegri *et al.*, 2016a), finding that, at
27 *in vitro* doses (10 µg/cm²) corresponding to high, but plausible, human equivalent
28 exposures (29.3 mg/m³), a significant cytotoxic effect was detected in murine
29 macrophages.

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3 Those results prompted us to evaluate whether fiber shortening mitigates TiO₂ NF
4 toxicity. In particular, we studied the biological effects of long and shortened TiO₂ NF,
5 investigating their effects on epithelial barrier function and macrophage activation *in*
6 *vitro* as well as on airway secretion of cytokines upon respiratory exposure *in vivo*.
7
8 Furthermore, we have also investigated the acute inflammatory effects of these
9 materials in the peritoneal cavity, used here as a surrogate for the pleural space and
10 mesothelial cell exposure. This model has been used widely to investigate fiber-type
11 toxicity of amosite asbestos fibers (Davis *et al.*, 1986a) as well as of fibrous
12 nanomaterials (Poland *et al.*, 2008a, Poland *et al.*, 2011).
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25 **2. Materials and methods**

26 ***2.1 Preparation, characterization and dispersion of the materials***

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28 Titanium dioxide nanofibers (Long fibers, L-TiO₂ NF) obtained through
29 electrospinning, were supplied by Elmarco, Liberec, (Czech Republic). The
30 characteristics of L-TiO₂ NF have been described by Allegri *et al.* (Allegri *et al.*,
31 2016a).
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39 For fiber shortening, the TiO₂ NF were suspended in distilled water and subjected to
40 ball milling using ZrO₂ spheres for 15h. In order to control size homogenization of to
41 favor the milling process, grinding media with different diameters were used (50 wt%
42 ZrO₂ spheres with 3 mm diameter and 50 wt% ZrO₂ spheres with 5 mm diameter). No
43 release of Zr occurred during the milling process, as assessed with ICP analysis.
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50 Both L- and ball-milled TiO₂ nanofibers (Short fibers, S-TiO₂ NF) were dispersed at 3
51 % (wt) in a colloidal suspension (distilled water, pH = 3.8) and characterized as
52 described in our previous work (Allegri *et al.*, 2016a). Characteristics are summarized
53 in Table 1. Aliquots of NF colloidal suspensions were incubated at 40°C, till a complete
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3 evaporation of the water. After that, L- and S-TiO₂ NF were put in glass tubes and
4
5 heated at 240°C for 4h to obtain sterile, LPS-free powders.
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8 Zeta-potential was determined by electrophoretic light scattering (ELS) measurements
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10 using Zetasizer Nanoseries apparatus (Malvern Instruments, Malvern, UK) on both (L-
11
12 and S-TiO₂ NF) 1 g/L nanosuspensions, diluting with distilled water.
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15 The investigation of the nanofiber size distribution was performed by scanning electron
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17 microscopy using FE-SEM (Carl Zeiss Sigma NTS GmbH, Oberkochen, Germany).
18

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20 Nanofibers were dispersed on a standard aluminum support by dripping the diluted
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22 prepared nanosuspensions. Samples were dried in air and then treated at 100°C for 5
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24 minutes to ensure the complete evaporation of water before FE-SEM analysis. Image
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26 analysis (ImageJ, Wayne Rasband, 1997) was performed on more than 150 nanofibers
27
28 to calculate the average diameters, lengths and standard deviation of TiO₂ NF samples.
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31 NF were then suspended, according to a slightly modified Nanogenotox protocol in a
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33 solution of 0.05% Bovine Serum Albumin (BSA, Sigma Aldrich, Milan, Italy) in
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35 Phosphate Buffered Saline without calcium and magnesium (PBS) to obtain 50x
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37 suspensions of the highest dose tested on cells (80 µg/cm²). After vortex mixing (30
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39 sec) and water bath sonication (10 min, 400 W with a Branson 5510), the stock
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41 suspensions were subsequently diluted in the same solvent to obtain 50x stocks of the
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43 other doses.
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47 The multiwalled carbon nanotubes NM-401 (JRC Repository, Ispra, Varese, Italy) were
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49 used as a positive benchmark material (Allegri *et al.*, 2016b) and treated as the NF.
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52 Amosite asbestos was enriched for long fiber component to create the long fiber amosite
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54 asbestos (LFA) sample of which 50.36% fibres >15 µm and 35.25% fibres >20 µm as
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56 previously determined (Poland *et al.*, 2008b). A sub-aliquot of LFA was ball milled,
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3 pulverizing the long fibers to produce a short fiber amosite (SFA) fraction as previously
4 described (Davis *et al.*, 1986a) with 4.46% fibres >15 μm and 0.99% fibres >20 μm .
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9 **2.2 Cell culture**

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11 For the *in vitro* experiments we used mouse peritoneal monocyte-macrophage Raw
12 264.7 cells, human alveolar carcinoma epithelial cells A549 and human normal
13 immortalized 16HBE bronchial cells. Both Raw 264.7 and A549 cell lines were
14 obtained from the Cell Bank of the IZSLER (Istituto Zooprofilattico Sperimentale della
15 Lombardia e dell'Emilia-Romagna, Brescia, Italy), while 16HBE cells were kindly
16 provided by Prof. Peter Hoet (University of Leuven, Belgium). Raw 264.7 cells were
17 cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10%
18 Fetal Bovine Serum (FBS), 4 mM glutamine, and antibiotics (streptomycin 100 $\mu\text{g}/\text{mL}$ /
19 penicillin, 100 U/mL), while A549 cells were cultured in F-12 Ham's medium
20 supplemented with 10% FBS, 1 mM glutamine, and antibiotics. 16HBE cells were
21 grown in DMEM/F12 (1:1) supplemented with 5% of FBS, 2 mM glutamine and
22 antibiotics. All cultures were maintained in a humidified atmosphere of 5% CO_2 in air
23 in 10-cm dishes.
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43 **2.3 Cell viability**

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45 For cell viability analysis, both A549 and 16HBE cells were seeded in complete growth
46 medium at a density of 3.1×10^4 cells/cm² in 96-well plates (Falcon, Corning Inc.,
47 Corning, NY, USA). After 24h in culture, growth medium was replaced with fresh
48 medium supplemented with the materials (dose range from 20 to 80 $\mu\text{g}/\text{cm}^2$). Vehicle
49 (0.05% BSA in PBS) was added to the control cells at the same dilution used for
50 treatment with NF. Cell viability was tested (O'Brien *et al.*, 2000) replacing medium
51 with a solution of resazurin (44 μM) in serum-free medium, after 24, 48 and 72h of
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3 incubation with the materials. Fluorescence was measured at 572 nm with a multimode
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5 plate reader Enspire (Perkin Elmer, Waltham, MA, USA). Possible interference of the
6
7 materials with the test was assessed incubating the dye in the presence of the materials
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9 at the highest dose tested. In the absence of cells, no fluorescence signal was detected
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11 above the background.
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14 15 16 **2.4 Nitrite medium concentration**

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18 Nitrite concentration, as a proxy for NO production, was determined as described in
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20 Bianchi et al. (Bianchi *et al.*, 2015). Briefly, after treatments, 100 μ l of medium were
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22 transferred to black 96-well plates with a clear bottom (Corning, Corning, NY, USA).
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24 20 μ l of a solution of 2,3-diaminonaphthalene (DAN, 0.025 mg/ml in 0.31 M HCl) were
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26 then added to the media and, after 10 min at RT, the reaction was stopped with 20 μ l of
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28 0.7 M NaOH. Standards were performed in the same medium from a solution of 1 mM
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30 sodium nitrite. Fluorescence was determined with a multimode plate reader Perkin
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32 Elmer Enspire
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39 **2.5 Trans-Epithelial Electric Resistance (TEER) and permeability**

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41 For these experiments, 16HBE cells were seeded into cell culture inserts with
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43 membrane filters (pore size 0.4 μ m) for Falcon 24-well-multitrays (Becton, Dickinson
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45 & Company, Franklin Lakes, NJ, USA) at a density of 10×10^4 cells/cm², and grown for
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47 4d until a tight monolayer was formed (TEER > 1000 Ω /cm²). Materials were then
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49 added in the apical chamber without changing the medium, and TEER was measured at
50
51 the indicated times of treatment. Every 3d from NF supplementation, 100 μ l and 300 μ l
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53 of culture medium were replaced with fresh medium in the apical and basolateral side,
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55 respectively. TEER changes were expressed as the percentage of the initial value
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57 adjusted for control cell monolayers according to Equation 1 (Salem *et al.*, 2009):
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$$\% \Delta TEER = \frac{Final TEER_{(treated)}}{Final TEER_{(control)}} \times \frac{Initial TEER_{(treated)}}{Initial TEER_{(control)}} \times 100 \quad (1)$$

At the end of the incubation (19d of treatment) monolayers were used for the determination of viability, incubating cells with resazurin from the apical side, and fluorescein permeability, adding the fluorescent dye (Fluorescein di-sodium salt, MW 376,27, Sigma-Aldrich F6377, 5 μ M) at the apical side and measuring the fluorescence in the baso-lateral medium after 30 min.

2.6 RT-PCR

Total RNA was isolated from Raw 264.7 cells with GenElute Mammalian Total RNA Miniprep Kit (Sigma–Aldrich). After reverse transcription, aliquots of cDNA from each sample were amplified in a total volume of 25 μ l with Go Taq PCR Master Mix (Promega, Italia, Milan, Italy), along with the forward and reverse primers (5 pmol each) reported in Table 1. Real time PCR was performed in a 36-well RotorGeneTM3000, version 5.0.60 (Corbett Research, Mortlake, Australia). For all the cDNA to be quantified, each cycle consisted of a denaturation step at 95 °C for 20s, followed by separate annealing (30s) and extension (30s) steps at a temperature characteristic for each pair of primers (Table 1). Fluorescence was monitored at the end of each extension step. At the end of the amplification cycles, a melting curve analysis was added. Data analysis was made according to the relative standard curve method (Bustin, 2000), while cDNA abundance was expressed as the ratio between each investigated cDNA and *Gapdh* cDNA.

2.7 Confocal microscopy

For the evaluation of NF internalization, confocal microscopy of fixed Raw264.7 cells

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3 was performed as previously described with minor modifications (Bianchi *et al.*, 2015).
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5 Raw264.7 cells were seeded on four-chamber slides at a density of 15×10^4 cells/cm²
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7 and, after 24h, treated for further 24h with TiO₂ NF at the dose of 10 µg/cm². 20 min
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9 before the end of exposure, medium was supplemented with CellTracker™ Red
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11 CMPTX (8 µM, Molecular Probes, Invitrogen, Carlsbad, CA, USA) to label the
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13 cytoplasm and; in the last 5 min, with 1,5-bis[2-(di-methylamino)ethyl]amino-4, 8-
14
15 dihydroxyanthracene-9,10-dione (DRAQ5®, 20 µM, Alexis Biochemicals, San Diego,
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17 CA, USA) to counterstain the nuclei. Cells were then fixed with 3.7%
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19 paraformaldehyde (15 min RT) and observed. Reflectance at 488 nm or excitation at
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21 543 nm and emission recorded through a 580–630 nm band pass barrier filter were used
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23 to visualize TiO₂ NF and the cytoplasm, respectively.
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29 For the analysis of epithelial monolayers, 16HBE monolayers, grown on membrane
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31 filters, were washed with PBS and incubated for 5 min in absolute methanol at -4°C.
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33 Fixed cells were then treated for 10min with 10% of Triton-X100 in PBS, incubated for
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35 further 2h in a solution of 10% BSA and 2% of Normal Goat Serum (DAKO SpA,
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37 Milan, Italy), then incubated overnight in the presence of primary antibodies (anti-
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39 Claudin-7, rabbit polyclonal; anti ZO-1, mouse polyclonal, both at 1:400, R&D
40
41 Systems) in 10% BSA in PBS. The same solution, supplemented with DRAQ5 (5µM)
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43 for nuclei counterstaining, was exploited for the incubation with secondary antibodies
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45 Alexa Fluor 488 goat anti-mouse and Alexa Fluor 543 goat anti-rabbit for the detection
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47 of ZO-1 and claudin-7, respectively. At the end of the incubation, filters were mounted
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49 on glass slides. Confocal images were taken with excitation at 543 nm and emission
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51 recorded through a 580- to 630-nm band pass barrier filter for Alexa Fluor 543 to
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53 visualize claudin-7 and excitation at 488 nm and emission through a 515- to 540-nm
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55 band pass filter for Alexa Fluor 488 to visualize ZO-1.
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Both analyses were performed with a LSM 510 Meta scan head integrated with an inverted microscope (Carl Zeiss, Jena, Germany) using a 63x (1.4 NA) oil objective. In both cases, excitation at 633 nm and emission recorded through a 670 long pass filter were adopted for nuclei visualization with DRAQ5.

2.8 Scanning Electron Microscopy (SEM)

Macrophages were seeded on coverslips in 24-well plates (Thermanox™, Scientific Laboratory Supplies Limited, Hessle, UK). After 24h cells, were treated with cell culture medium with L- or S-TiO₂ NF (both at 10 µg/cm²) or in plain medium for 24h. Cells were then rinsed in NaCl 0.9% and fixed overnight at 4°C in glutaraldehyde (3% in 0.1 M sodium cacodylate buffer). The preparations were gradually dehydrated with subsequent incubation with increasing concentrations of ethanol. Coverslips were then treated for 10 min with hexamethyldisilazane and, after drying, mounted on SEM stub, sputter coated with gold and analysed using a Scanning Electron Microscope (SEM, Oxford Instruments).

2.9 In vivo experiments

For the *in vivo* studies, eight-week-old C57BL/6 male mice were obtained from Charles River Italia (Calco, LC, Italy) and housed in the Tor Vergata Animal Technology Station or The University of Edinburgh under standard conditions (25 °C, 50 % relative humidity) on a 12h light/dark cycle with free access to water and laboratory animal food. All animal procedures were in compliance with the European Legislation (2010/63/EU) and have been approved by the Institutional Animal Care and Use Committee of the University of Tor Vergata or of the University of Edinburgh.

For the pharyngeal aspiration experiments, C57BL/6 mice of 20–22 g were

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3 anaesthetized using a mixture of Ketalar®, 1 mg/mouse (Warner-Lambert, Zaventem,
4 Belgium), and Rompun®, 0.2 mg/mouse (Bayer, Leverkusen, Germany) given
5
6 intraperitoneally. At least 3 animals were included in each experimental group. The
7
8 procedure was performed as previously described (Polimeni *et al.*, 2016). Both L-TiO₂-
9
10 NF and S-TiO₂-NF were dispersed in distilled water containing 1.4 mg/ml endotoxin-
11
12 free BSA and sonicated on ice. Each mouse received 50 µl of a suspension containing
13
14 40 µg of either L-TiO₂-NF or S-TiO₂-NF. At the end of the procedure, animals were
15
16 allowed to recover on a heated pad and then located back in their cages. Control animals
17
18 were treated with an equivalent volume of sterile water containing 1.4 mg/ml BSA.
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20 After 14 days, Bronchoalveolar Lavage (BAL) was performed under terminal
21
22 anaesthesia by cannulating the trachea and lavaging with 1 ml of ice-cold saline
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24 solution. The lavage was then centrifuged at 200 × g for 10 min at 4 °C and the
25
26 supernatant was stored at -80 °C for further biochemical analysis
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33 For the intraperitoneal experiments, mice of 20–22 g were injected
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35 intraperitoneally at a dose of 100 µg/ml (0.5 ml; total dose 50 µg per mouse) with each
36
37 particle suspended in sterile 0.5 % BSA/ saline solution or vehicle control. After 24-hrs,
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39 the mice were sacrificed by cervical dislocation and the peritoneal cavity lavaged using
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41 3 x 2 ml washes of ice-cold sterile saline which were pooled together on ice. The lavage
42
43 fluid was centrifuged to isolate the cellular fraction and separate the cellular fraction.
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45 Cyto-centrifugation preparations were made using the isolated cells and differential cell
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47 counts were performed to determine leukocyte populations.
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53 **2.10 Cytokine secretion**

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55 TNFα and IL6 secretion in the culture media of Raw264.7 cells was determined with
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57 the ELISA RayBio® kit (Ray Biotech, Norcross, GA, USA) as described in Bianchi et
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3 al.(Bianchi *et al.*, 2015).The determination of IL-6, TNF α and TGF β secretion in BAL
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5 samples, derived from mice exposed to L-TiO₂ NF or S-TiO₂ NF, was performed using
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7 the ELISA kit from EIAab[®] Science (Optics Valley, Wuhan, China) following the
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9 manufacturer's instructions.
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14 **2.11 Statistical analysis**

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16 For viability, trans-epithelial electrical resistance and permeability data were analyzed
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18 with ANOVA with Bonferroni *post hoc* test. Data of nitrite concentration, gene
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20 expression and cytokine secretion were analyzed with two-tail t test for unpaired data. A
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22 p value < 0.05 was considered statistically significant. GraphPad Prism5[™] software was
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24 used for statistical analyses.
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30 **2.12 Chemicals**

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32 Sigma-Aldrich was the source of the all the chemicals whenever not stated otherwise.
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39 **3. Results**

40 41 **3.1 Physico-chemical characterization of long and ball milled TiO₂ NF**

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43 TiO₂ nanofibers (NF), produced by an electro-spinning process (see Materials and
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45 Methods), were modified with a “size control” strategy through a ball milling treatment,
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47 which was chosen due to its industrial relevance as well as low cost and easily scalable
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49 characteristics. This strategy was applied in order to decrease length and aspect ratio
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51 (AR) of the fibers and, thus, as a remediation step in the manufacturing line.
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56 The starting pristine NF were found to be very inhomogeneous but showed the
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58 desired nanostructured morphology, as demonstrated by high specific surface area by
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3 BET measurements. As expected, the milling treatment did not affect fiber diameter,
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5 although it produced a modification of the average fiber length with a consequent,
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7 significant decrease of aspect ratio (Table 2).
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10 11 **3.2 Effects of TiO₂ NF on cell viability**

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14 Figure 1 reports data of cell viability obtained in A549 and 16HBE cells treated with 20,
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16 40 and 80 $\mu\text{g}/\text{cm}^2$ of either L- or S-TiO₂ NF for 24, 48 or 72h. In A549 cells (Figure 1,
17
18 A-B-C), L-TiO₂ NF caused a clear-cut dose- and time-dependent decrease of cell
19
20 viability, with a maximal effect of 60% at 80 $\mu\text{g}/\text{cm}^2$ obtained after 72h of treatment.
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22 Fiber shortening almost completely abolished the toxicity of the fibers with marginally
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24 significant viability loss that, however, was not dose- or time-dependent.
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28 16HBE cells (Figure 1, D-E-F) were much less sensitive than A549 cells to the
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30 toxicity of L-TiO₂ NF, with the maximal effect (30% reduction in viability) recorded at
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32 72h of exposure. However, the toxicity of L- and S-TiO₂ NF was low and comparable
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34 except for the 72h-time point, when L-NF were more toxic than S-NF at the higher
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36 doses. Interestingly, the toxicity of S-NF was larger, although mild, for 16HBE cells
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38 than for A549 cells. In both cell models, the benchmark material NM-401 MWCNT
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40 caused a marked time-dependent decrease of cell viability, which was more evident in
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42 A549 than in 16HBE cells.
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46 47 48 **3.3 Impact of TiO₂ NF on airway epithelial barrier integrity**

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51 Figure 2 (Panel A) reports the time course of changes in Trans-Epithelial Electrical
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53 Resistance (TEER) of 16HBE cell monolayers exposed to TiO₂ NF (80 $\mu\text{g}/\text{cm}^2$) up to
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55 19d. L-TiO₂ NF significantly lowered TEER by 30% after 4d of exposure. At longer
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57 times, TEER exhibited a partial recovery but remained significantly lower than control
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3 monolayers. With S-TiO₂ NF, TEER decrease was smaller throughout the experiment
4 and, at 19d of treatment, was no more different from control (Panel B). As expected, the
5 benchmark MWCNT preparation produced a large TEER decrease with no sign of
6 recovery. TEER changes were associated to significant increases in fluorescein
7 permeability in monolayers treated with F-TiO₂ NF and MWCNT (Panel C), while no
8 significant loss of viability was detected in any condition (Panel D).
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17 Changes in trans-epithelial permeability and resistance are associated with
18 alterations in the distribution of tight junction proteins (Figure 3). The confocal images
19 show that, compared with control conditions (Panel A), a 19d-exposure to L-TiO₂ NF
20 (Panel B) caused a marked and widespread perturbation of the distribution of Claudin-7
21 and ZO-1, while only milder changes are detected in monolayers treated with S-TiO₂
22 NF (C). In particular, the orthogonal projections indicated that Claudin-7 was
23 delocalized to the cytoplasm in a high fraction of cells in panel B, while were
24 maintained in close proximity of ZO-1 in most cells of panel C.
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36 37 **3.4 Nanofiber internalization by macrophages**

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39 To investigate macrophage behavior after challenge with TiO₂ NF we used both
40 confocal microscopy (Figure 4, A-C) and SEM (Figure 4, D-F). Confocal microscopy
41 showed that bundles of nanofibers were readily detected in macrophage cultures
42 exposed either to L-TiO₂ NF (Panel B) or to S-TiO₂ NF (Panel C). Only limited
43 internalization of L-TiO₂ NF occurred, while many short nanofibers were detected
44 inside the cells. However, in both cases, no co-localization of internalized NF with the
45 cytoplasmic marker was observed, indicating that the material was sequestered in a
46 compartment other than cytoplasm. In cultures treated with L-TiO₂ fibers, several cells
47 tended to adhere to the same bundle, while macrophages exposed to S-TiO₂ NF
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3 appeared elongated. SEM images revealed morphological deformations in macrophages
4 exposed to L-TiO₂ NF (Panel E), while in cultures exposed to S-TiO₂ NF (Panel F) cells
5 were larger and more elongated than control (Panel D) although no gross alteration was
6 detected.
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13 **3.5 Inflammatory response of murine macrophages exposed to long and short** 14 **TiO₂ NF** 15

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18 Figure 5A reports nitrite concentration in the medium, as a proxy of nitric oxide (NO)
19 production, of Raw 264.7. macrophages exposed to increasing doses of TiO₂ NF (range
20 20-80 µg/cm²). In cells treated with L-TiO₂ NF, NO production was only slightly
21 increased at 40 and 80 µg/cm². Much larger NO production was detected in the
22 Raw264.7 cells incubated with S-TiO₂ NF, with significant NO output already present
23 at the lowest dose tested (20 µg/cm²). These data were consistent with the level of *Nos2*
24 expression (Figure 5B), which was more than 30-fold stimulated in cells treated for 24h
25 with S-TiO₂ NF but only 3-fold induced by L-TiO₂ NF at the same dose (80 µg/cm²). A
26 larger pro-inflammatory activity of S- compared to L- TiO₂ NF was also observed when
27 the expression of *Ptgs2*, the gene which encodes for Cox2, was investigated (Figure
28 5C). At the same experimental time, also the expression of *Il1b* was significantly
29 induced, with a larger effect of S- compared with L-TiO₂ NF (Panel 5D). After a 48h
30 treatment, both TNF-α and IL-6 secretion was markedly stimulated by S-TiO₂ NF in a
31 dose-dependent manner, while L-TiO₂ NF were ineffective.
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51 To investigate the mechanism underlying the higher pro-inflammatory activity
52 of S-TiO₂ NF, Raw264.7 cells were exposed for 48h to the materials (40 µg/cm²) in the
53 presence or in the absence of the cytoskeletal drug cytochalasin B (5 µg/ml), which
54 prevents actin polymerization and, hence, phagocytosis (Figure 6). Under these
55 conditions, nitrite production was almost completely suppressed. On the contrary,
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3 cytochalasin B did not reduce nitrite production in cells exposed to L-TiO₂ NF and only
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5 marginally decreased it in LPS-treated cultures.
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9 **3.6 Airway cytokine secretion after the exposure to L or S nanofibers**

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11 To evaluate *in vivo* effects of TiO₂ NF, C57BL/6 mice were exposed to L- or S-TiO₂ NF
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13 (40 µg/animal) through pharyngeal aspiration and, after 14d, the levels of TGF-β, TNF-
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15 α and IL-6 were measured in the bronchoalveolar lavage (BAL) fluid. L-TiO₂ NF
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17 induced significant increase of the three cytokines (Figure 7), while no significant
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19 changes were detected in BAL cytokine levels between animals treated with S-TiO₂ NF
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21 and control mice.
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26 **3.7 Peritoneal inflammation after the exposure to L or S nanofibers**

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28 The peritoneal and pleural cavities have very defined routes of clearance with pore-like
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30 stomata of 2-8µm in diameter being found in the peritoneal diaphragm and chest wall,
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32 respectively. Materials that cannot penetrate these stomata and access the underlying
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34 draining lymphatic system can be retained at the mesothelial surface where they may
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36 trigger inflammation (Murphy *et al.*, 2011). As shown in Figure 8A, internalized L-
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38 TiO₂ NFs were only occasionally detected (Figure 8A), while S-TiO₂ NF were seen
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40 within several macrophages (Figure 8B). Administration of long but not of short fibers
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42 led to a significant elevation in peritoneal granulocytes (Figure 8C), which was most
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44 notable for the LFA sample (P 0.0004). The influx of neutrophils was significantly
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46 higher in animals treated with L-TiO₂ NF compared with control animals (P 0.0136) or
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48 animals exposed to S-TiO₂ NF (P 0.0166).
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4. Discussion

In this study we compare the biological effects of two preparations of TiO₂ NF derived from an industrial process and characterized by different lengths. The longer preparation (L-TiO₂ NF) is endowed with features consistent with the dimensional criteria of a respirable fiber (length > 5 μm, diameter < 3 μm, length-to-width ratio (aspect ratio) > 3:1 (1997)). S-TiO₂ NF have been obtained from L-TiO₂ NF by milling with ZrO₂ microspheres, thus causing a marked decrease of average fiber length, as well as of their aspect ratio. However, although both the preparations have the aspect ratio of a respirable fiber, only L-TiO₂ NF have an average length > 5 μm.

A previous *in vitro* study (Allegrì *et al.*, 2016a) has demonstrated that L-TiO₂ NF are more cytotoxic than TiO₂ nanoparticles and cause a sizable decrease of the trans-epithelial electrical resistance of airway cell monolayers. On the contrary, the expression of several pro-inflammatory markers was not substantially different in macrophages exposed to L-TiO₂ NF or to TiO₂ nanoparticles.

Here the biological effects L-TiO₂ NF have been compared with those caused by shortened fibers. In two human airway cell lines, which exhibited different sensitivities to the toxic effects of L-TiO₂ NF (alveolar A549 cells > bronchial 16HBE cells), viability studies indicated that L-TiO₂ NF are more cytotoxic than S-TiO₂ NF. Length-dependent toxicity of TiO₂ HARNs was also described in rat alveolar macrophages by Hamilton *et al.* (Hamilton *et al.*, 2009), who found that only the longer nanobelts were significantly cytotoxic at relatively high doses. These authors proposed that the mechanism of long-fiber toxicity was attributable to post-phagocytosis lysosomal destabilisation, resulting in the release of destructive enzymes, such as cathepsin B, into the cytoplasm (Hamilton *et al.*, 2009). Here we demonstrate that length-dependent

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3 toxicity of TiO₂ HARNs is not specific for specialized phagocytic cells but extends to
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5 epithelial airway cells which represent primary targets upon respiratory exposure.
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8 Functional consequences of the exposure of epithelial cells to TiO₂ NF have
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10 been also investigated. Indeed, loss of epithelial barrier competence and monolayer
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12 integrity are important parameters correlated to penetration of the nanomaterials into the
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14 bronchial wall, alterations of epithelial functions and nanomaterial bio-persistence
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16 (Rotoli *et al.*, 2008, Banga *et al.*, 2012). L-TiO₂ NF, but not S-TiO₂ NF, cause a clear-
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18 cut epithelial barrier impairment, indicated by the decrease in TEER, the increase in
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20 fluorescein permeability and an evident alteration in the distribution of the tight-
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22 junction proteins claudin-7 and ZO-1 in correspondence to NF agglomerates. Similar
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24 effects on TEER were observed upon exposure of airway epithelial cells to long,
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26 needle-like MWCNT (Farcas *et al.*, 2015, Rotoli *et al.*, 2015), which are used here as
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28 benchmark materials. In the case of MWCNT, loss of barrier integrity is due to a focal
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30 damage to the epithelial cell monolayer, attributable to the interaction with nanomaterial
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32 agglomerates, in the absence of a generalized decrease in monolayer viability (Rotoli *et*
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34 *al.*, 2015). The confocal images reported here, along with lack of viability loss at the
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36 whole population level, suggest that a similar mechanism underlies L-TiO₂ NF effects.
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38 Interestingly, as observed here for TiO₂ NF, length is an important structural
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40 determinant for epithelial barrier damage also for MWCNT (Rotoli *et al.*, 2009).
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47 In their toxicological assessment of short (0.8 – 4 μm) and long (15-30 μm)
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49 TiO₂ nanobelts (Hamilton *et al.*, 2009), Hamilton et al. noted that only long TiO₂
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51 nanobelts led to a significant increase in IL-1β release (a proxy measure for the NALP3
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53 inflammasome formation) by LPS-primed rat alveolar macrophages, suggesting that
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55 length also influences inflammation. Using TiO₂ nanobelts of intermediate length (7
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57 μm), Tilton et al. performed a transcriptomic and proteomic analysis of treated human
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3 macrophages, demonstrating changes in the expression of inflammatory genes (Tilton *et*
4 *al.*, 2014). Those results were further extended *in vivo* (Porter *et al.*, 2013),
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6 demonstrating that mice, exposed through pharyngeal aspiration to TiO₂ nanobelts,
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8 developed significant dose- and time-dependent pulmonary inflammation and fibrosis,
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10 with much more evident alterations in animals treated with longer nanostructures
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12 (Porter *et al.*, 2013). The results shown here after aspiration of L- and S-TiO₂ nanofibers
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14 are consistent with those data, since only longer preparation caused a persistent cytokine
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16 secretion. Interestingly, among the cytokines tested, besides the typical pro-
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18 inflammatory IL-6 and TNF- α , a significant increase is also detected for the fibrogenic
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20 cytokine TGF- β , suggesting a pro-fibrotic activity of L-TiO₂ NF *in vivo*. Furthermore,
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22 *in vivo* administration through intra-peritoneal injection has demonstrated that retention
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24 at the mesothelial surface of long but not short NF leads to a prominent inflammatory
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26 response as indicated by a larger granulocyte influx.
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33 In contrast with data obtained *in vivo*, the results presented here indicate
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35 however, that shortened NF have a more evident pro-inflammatory activity than long
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37 NF, as evaluated from early activation events in murine macrophages *in vitro*. This
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39 effect cannot be attributed to LPS contamination during the ball milling procedure. LPS
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41 contamination of nanomaterials constitutes a frequently encountered, and often
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43 overlooked, problem in nanotoxicology (Esch *et al.*, 2010, Smulders *et al.*, 2012).
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45 Moreover, recent work from our laboratory has demonstrated that adsorption to
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47 nanomaterials may synergize the pro-inflammatory effects of LPS (Bianchi *et al.*, 2015,
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49 Di Cristo *et al.*, 2016, Bianchi *et al.*, 2017). However, the NF used in this contribution
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51 have been thermally pre-treated just before cell exposure and, therefore, are to be
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53 considered LPS-free. Thus, the effects observed on the induction of pro-inflammatory
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55 genes, inflammatory mediators and cytokine secretion should be attributed to intrinsic
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3 properties of NF rather than to a combined effect of NF and LPS, as in the experiments
4 reported by Hamilton et al. (Hamilton *et al.*, 2009). It may be the case that in the
5 complex and self-regulating environment of the lung, such pro-inflammatory stimuli
6 serve to increase the rate of clearance of particulates through a controlled response.
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8 Such a response noted *in vitro* may not be elevated *in vivo* (at the doses used for more
9 easily cleared low aspect ratio particulates) up to a full inflammatory response and,
10 therefore, would be an adaptive ('dust') response rather than a true inflammation.
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19 The experiments reported here provide some information on the mechanism
20 involved in the higher pro-inflammatory potential of S-TiO₂ NF *in vitro*. Indeed, the
21 inhibition of phagocytosis by the cytoskeletal drug cytochalasin B severely hampers NO
22 production by S-TiO₂ NF but not by L-TiO₂ NF. On the other hand, S-TiO₂ NF are
23 much more efficiently internalized than longer counterparts (see Figures 4 and 8), which
24 would lead to a high intracellular dose. These arguments support the hypothesis that
25 phagocytosis of NF is involved in the brisk, but possibly transient, macrophage
26 activation observed upon the exposure to S-TiO₂ NF, a characteristic of self-limited
27 'dust' response. Interestingly, all the inflammatory parameters tested (NO production,
28 induction of inflammatory genes, TNF- α /IL-6 secretion) are related to the M1 activation
29 type (Mills *et al.*, 2000, Murray *et al.*, 2014).
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45 Confocal microscopy and SEM showed that most cells incubated with S-TiO₂
46 internalized the material, displaying a nearby-normal rounded morphology or a
47 symmetrically elongated morphology, indicative of cell activation, but no severe
48 distortion or stretching. On the contrary, the incubation with long fibers caused marked
49 cell stretching along the fiber surface, with several cells aligned along or penetrated by
50 the same fiber but with very few cells exhibiting clear cut internalization. This situation
51 of attempted, partial or failed internalization is indicative of frustrated phagocytosis, as
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3 reported for other fibrous nanomaterials (Hamilton *et al.*, 2009, Sweeney *et al.*, 2015).
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5 As suggested by Schinwald *et al.* (Schinwald *et al.*, 2012b), this situation can cause
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7 hindered macrophage motility, occurs at fiber lengths greater than 5 μm and, hence,
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9 would be relevant to the L- but not to the S TiO₂ NF. Within the *in vivo* context,
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11 impaired macrophage-mediated clearance of NF from the lung could lead to increased
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13 lung burden over time. On the contrary, exposure to S-TiO₂ NF is expected to trigger
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15 efficient phagocytosis with enhanced clearance of the nanomaterial and, consistently,
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17 such efficient uptake was noted in the peritoneal model.
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22 Data obtained after *in vivo* respiratory exposure are also consistent with this hypothesis,
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24 since the more prolonged inflammatory reaction observed with L-TiO₂ NF, documented
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26 by the higher cytokine levels in BAL (Figure 7), could be associated with a higher bio-
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28 persistence of this preparation. In accordance, Porter *et al.* reported that, after more
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30 than 100 d from exposure, a sizable proportion of long TiO₂ nanobelts was still present
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32 in the lung (Porter *et al.*, 2013). This substantial retention may be the result of both
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34 impaired macrophage-mediated clearance and increased permeation of the altered
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36 epithelial barrier. Importantly, the results presented here indicate that L-, but not S-TiO₂
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38 NF, are able to produce a sustained secretion of pro-inflammatory and fibrogenic
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40 cytokines in the lung as well as a marked leukocyte influx in the peritoneal model.
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42 Thus, the increased bio-persistence of long NF is expected to trigger more chronic
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44 responses (Figure 9).
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56 Department of Medicine and Surgery of the University of Parma.
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Figure Legends

Table 1. Primers and temperatures of annealing adopted for RT-PCR experiments

Table 2. Properties of L-TiO₂ NF and S-TiO₂ NF

Figure 1. Effect of TiO₂ nanofibers on cell viability. A549 (A,C,E) and 16HBE cells (B,D,F) were exposed for up to 72h to the indicated doses of long (L-) or short (S-) TiO₂ NF or to the benchmark material NM-401. Cell viability was assessed with the resazurin assay at 24, 48, 72h. Data are means \pm S.D of six independent determinations. ** $p < 0.01$, *** $p < 0.001$ vs. control (untreated cultures), as assessed with one-way ANOVA; ## $p < 0.01$, ### $p < 0.001$ between L- and S-TiO₂ NF, as assessed with two-tail t-test for unpaired data.

Figure 2. Effect of TiO₂ nanofibers on the barrier integrity of human bronchial cell monolayers. 16HBE airway epithelial cells were cultured on membrane filters and, once reached confluence, L- or S-TiO₂ NF were added to the apical chamber at the dose of 80 $\mu\text{g}/\text{cm}^2$. The Trans-Epithelial Electrical Resistance (TEER) was then measured up to 19d of treatment (A). NM-401 MWCNT, used as benchmark material, were added to other monolayers in parallel. At the end of the experiment TEER (B), fluorescein permeability (C) and cell viability (D) were measured in the same monolayers. Data are means \pm S.D of 4 independent determinations. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, ns, not significant vs. control, untreated cultures; # $p < 0.05$, ## $p < 0.01$ vs. monolayers treated with NM-401 MWCNT.

Figure 3. At the end of the experiment shown in Figure 2, cell monolayers, kept adherent to the filters, were fixed and immunostained for claudin-7 (red) and ZO-1 (green). DRAQ5 (blue) was used to stain nuclei. (A), After the staining procedure,

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3 filters were trimmed from the transwells, laid down and mounted on the coverslip for
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5 observation. Control, untreated monolayers; (B) monolayers treated for 19d with L-
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7 TiO₂ NF (80 µg/cm²); (C) monolayers treated for 19d with S-TiO₂ NF (80 µg/cm²). For
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9 each condition, a single horizontal confocal section of a representative field is shown,
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11 with orthogonal projections. Areas where signals are not detected are due to changes in
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13 observation plan in correspondence of filter deformations. Arrows, cells with gross
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15 claudin-7 delocalization. The experiment has been performed twice with similar results.
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19 Bar = 20 µm.

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22 **Figure 4.** Characterization of nanofiber internalization by Raw264.7 macrophages.

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24 Macrophage were seeded on coverslips and, after 24h, exposed for 24h to L-TiO₂ NF
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26 (B, E) or S-TiO₂ NF (C, F), both at a dose of 10 µg/cm², or incubated in control medium
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28 (A, D). Cells were then labelled, fixed, mounted and observed in confocal microscopy
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30 (A-C, see Methods) Single-plan horizontal sections of representative fields are shown
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32 where TiO₂ NF are pseudo-colored in white, the cytoplasm in red and nuclei in blue.
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34 For SEM, monolayers were fixed and dehydrated before being mounted (D-F, see
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36 Methods). Bars, 20 µm (A, B) or 10 µm (C, D).

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39 **Figure 5.** Comparative effects of long and short TiO₂ NF on the expression of pro-
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41 inflammatory markers in Raw 264.7 cells. Macrophages were treated with L-TiO₂ NF or
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43 S-TiO₂ NF at the indicated doses or with 10 ng/ml of LPS as a positive control and
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45 several inflammatory markers were assessed. (A,B,C) After 24h of treatment at the
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47 indicated doses of NF, medium nitrite concentration (A), *Nos2* expression (B) or *Ptgs2*
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49 expression (C) were assessed. (D) At the same experimental time, the effects of NF (at
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51 the dose of 80 mg/cm²) on the expression of *Il1b* were assessed. (E,F) After 48h of
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53 treatment, the levels of TNF-α and IL-6 were determined in the extracellular medium.
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3 Data are means \pm S.D of 3 (B-F) or 4 (A) independent determinations. * $p < 0.05$, ** p
4 < 0.01 , *** $p < 0.001$ vs. control, untreated cultures; \$\$ $p < 0.01$, \$\$\$ $p < 0.001$ vs. the
5 same dose of L-TiO₂ NF, as assessed with two-tail t test for unpaired data.
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11 **Figure 6.** Role of TiO₂ nanofiber internalization in NO production in Raw 264.7 cells.

12 Nitrite concentration was measured in the culture medium of macrophages exposed for
13 48h to L-TiO₂ NF or S-TiO₂ NF (40 $\mu\text{g}/\text{cm}^2$) or to LPS (10 ng/ml) with or without
14 cytochalasin B (5 $\mu\text{g}/\text{ml}$). Data are means \pm SD of 8 independent determinations in two
15 experiments. *** $p < 0.001$ vs. control, untreated cultures; # $p < 0.05$, ### $p < 0.001$ vs. the
16 same condition without cytochalasin B.
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26 **Figure 7.** BAL cytokines in C57BL/6 mice exposed to TiO₂ NF. C57BL/6 mice were
27 exposed through pharyngeal aspiration to 40 μg of L- (red bars) or S-TiO₂ NF (blue
28 bars) suspended in sterile water containing 1.4 mg/ml BSA or in vehicle alone (control,
29 empty bars). After 14d, BAL were collected and TGF- β , TNF- α and IL-6 were
30 measured with ELISA. * $p < 0.05$, ** $p < 0.01$ vs. control; ns, not significant vs. control;
31 # $p < 0.05$ vs. S-TiO₂ NF, as assessed with ANOVA.
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42 **Figure 8.** Peritoneal lavage granulocytes in C57BL/6 mice exposed to TiO₂ NF or long
43 fibre (LFA) or short fibre (SFA) amosite asbestos. C57BL/6 mice were exposed through
44 intra-peritoneal injection to L-TiO₂ NF, S-TiO₂ NF, long fibre amosite, short fibre
45 amosite asbestos (50 mg of each material) suspended in sterile water containing 0.5%
46 BSA or to vehicle alone. After 24h, the peritoneal cavity was lavaged and cells
47 determined via a differential cell count. (A) L-TiO₂ (chevron) containing macrophage.
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(B). Several macrophages containing S-TiO₂ (chevrons). (C) Percentage granulocytes in

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3 the peritoneal fluid *p < 0.05, ***p < 0.001 vs. control; \$ p < 0.05 vs. S-TiO₂ NF, as
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5 assessed with ANOVA (n = 3).
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11 **Figure 9.** Differential outcomes of exposure to long and shortened TiO₂ nanofibers.
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For Peer Review Only

Gene	Protein	Forward	Reverse	T (°C)	Amplicon Size (bp)
<i>Nos2</i>	Inducible Nitric oxide synthase (Nos2)	5'-GTT CTCAGC CCA ACAATA CAA GA-3'	5'-GTG GACGGG TCGATG TCA C-3'	57	127
<i>Ptgs2</i>	Cyclooxygenase-2 (Cox2)	5'-GCTCAGCCAGGCAGCAAATC-3'	5'-ATCCAGTCCGGGTACAGTCA-3'	56	107
<i>Gapdh</i>	Glyceraldehyde 3-phosphate dehydrogenase	5'-TGT TCCTAC CCC CAATGT GT-3'	5'-GGT CCTCAG TGTAGC CCA AG-3'	57	137
<i>Il1b</i>	IL-1 β	5'-GCA ACTGTT CCT GAACTC AAC T-3'	5'-ATC TTTTGG GGTCCG TCAACT-3'	58	76

Table 1. Primers and temperatures of annealing adopted for RT-PCR experiments

	Average Length (μm)	Average Diameter (μm)	Average Aspect Ratio	SSA (m^2/g)	Zeta potential (mV)
L-TiO₂ NF	9.9 \pm 5.8	0.3 \pm 0.1	29:1	144.5	-35.0 \pm 1.1
S-TiO₂ NF	2.1 \pm 2.4	0.4 \pm 0.1	5.7:1	138.8	-22.5 \pm 1.4

Table 2. Properties of L-TiO₂ NF and S-TiO₂ NF

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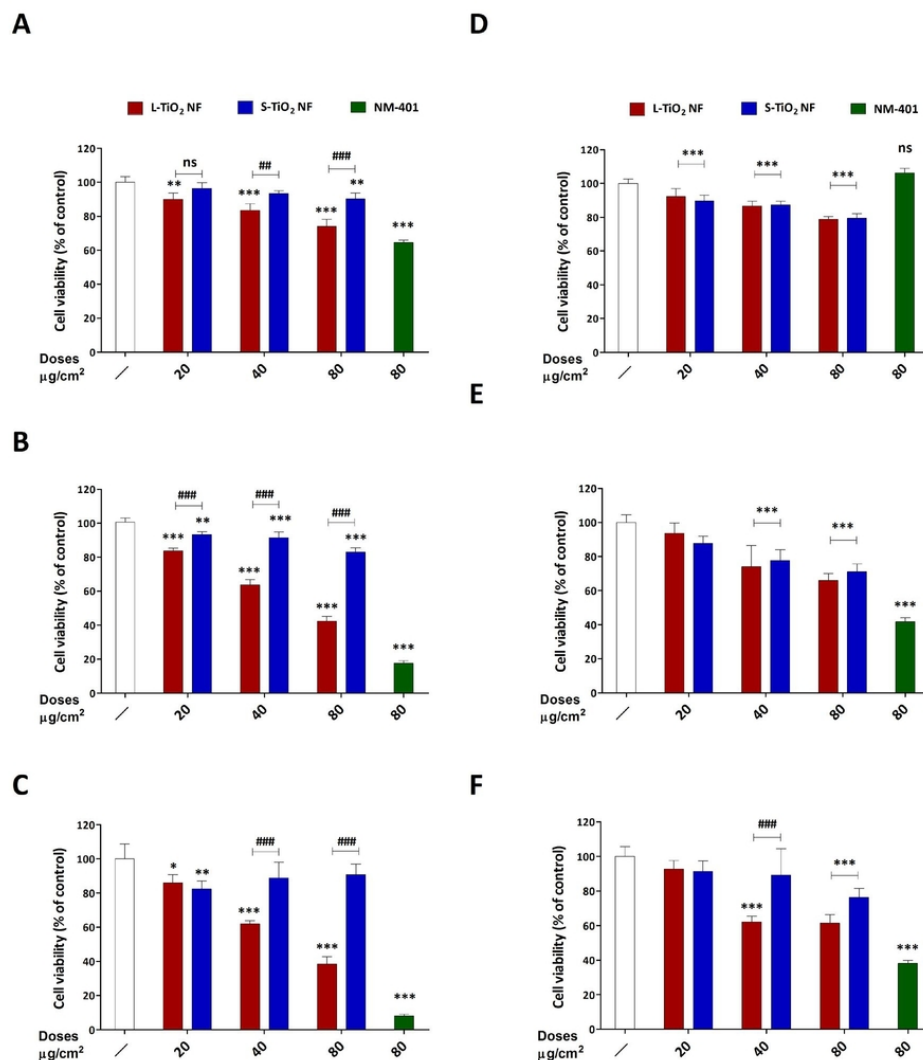


Figure 1. Effect of TiO₂ nanofibers on cell viability. A549 (A,C,E) and 16HBE cells (B,D,F) were exposed for up to 72h to the indicated doses of long (L-) or short (S-) TiO₂ NF or to the benchmark material NM-401.

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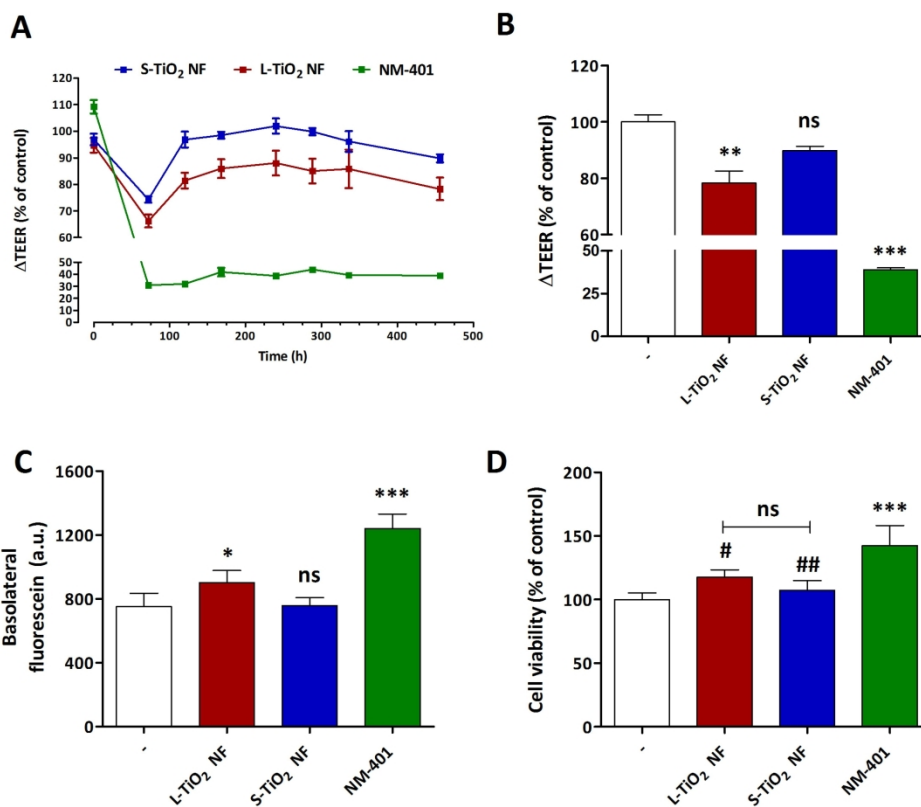


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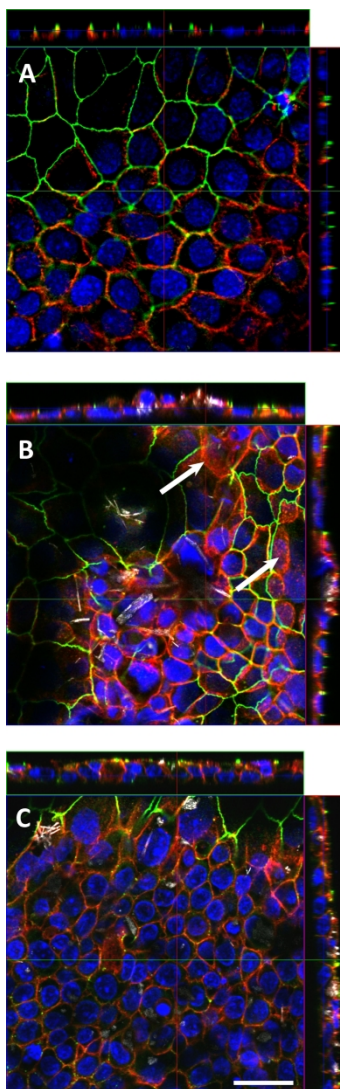


Figure 3. At the end of the experiment shown in Figure 2, cell monolayers, kept adherent to the filters, were fixed and immunostained for claudin-7 (red) and ZO-1 (green). DRAQ5 (blue) was used to stain nuclei. (A), After the staining procedure, filters were trimmed from the transwells, laid down and mounted on the coverslip for observation. Control, untreated monolayers; (B) monolayers treated for 19d with L-TiO₂ NF (80 µg/cm²); (C) monolayers treated for 19d with S-TiO₂ NF (80 µg/cm²). For each condition, a single horizontal confocal section of a representative field is shown, with orthogonal projections. Areas where signals are not detected are due to changes in observation plan in correspondence of filter deformations. Arrows, cells with gross claudin-7 delocalization. The experiment has been performed twice with similar results. Bar = 20 µm.

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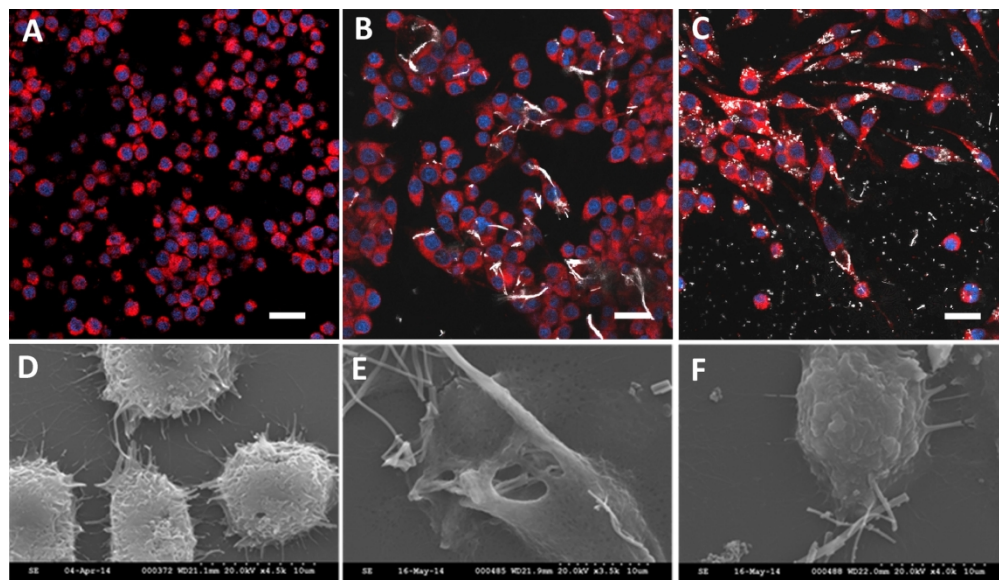


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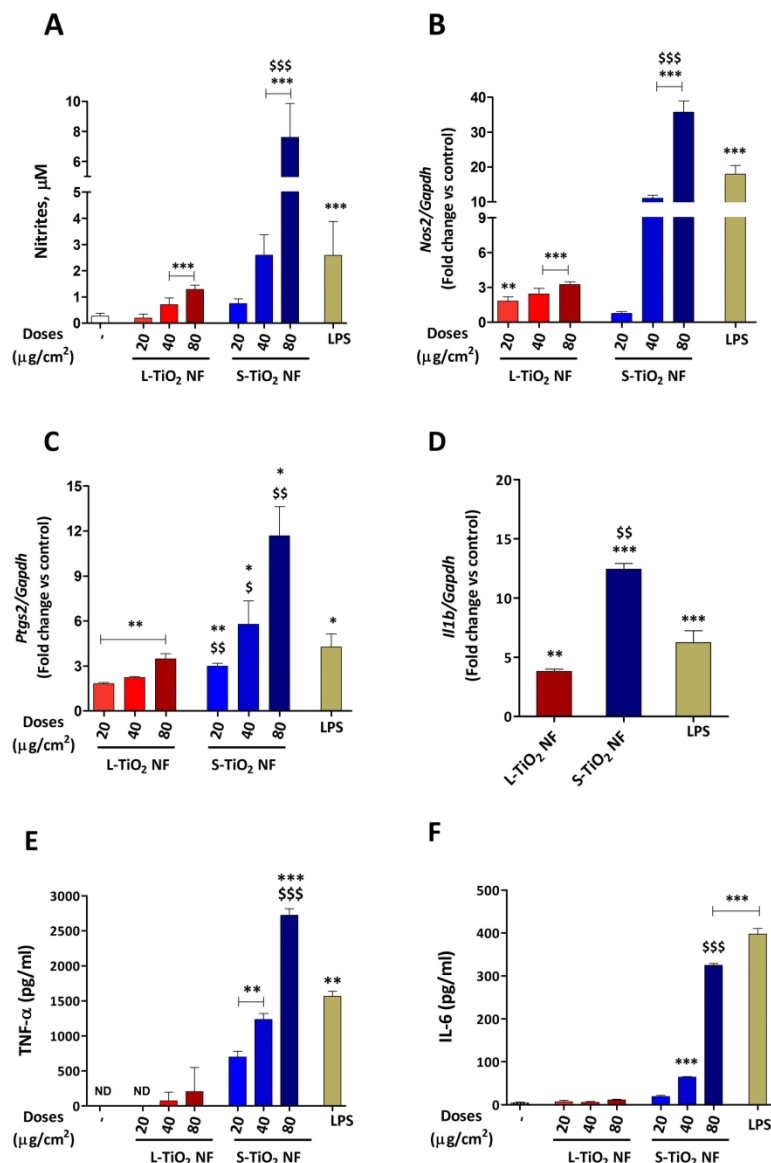


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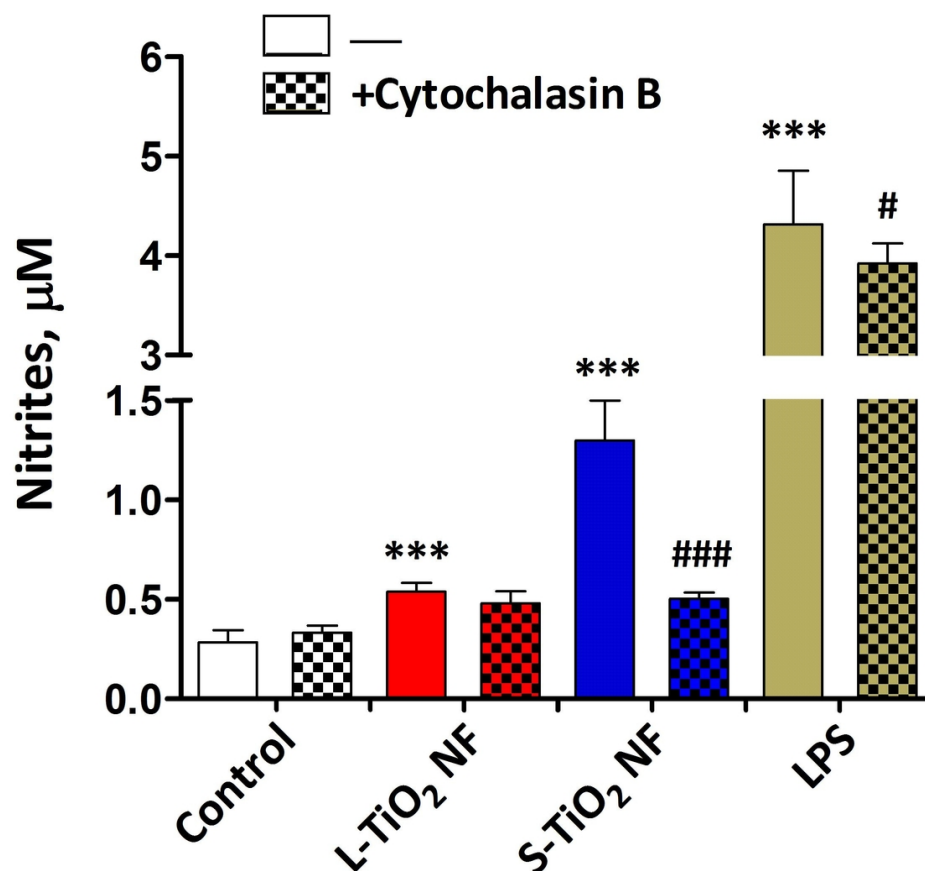


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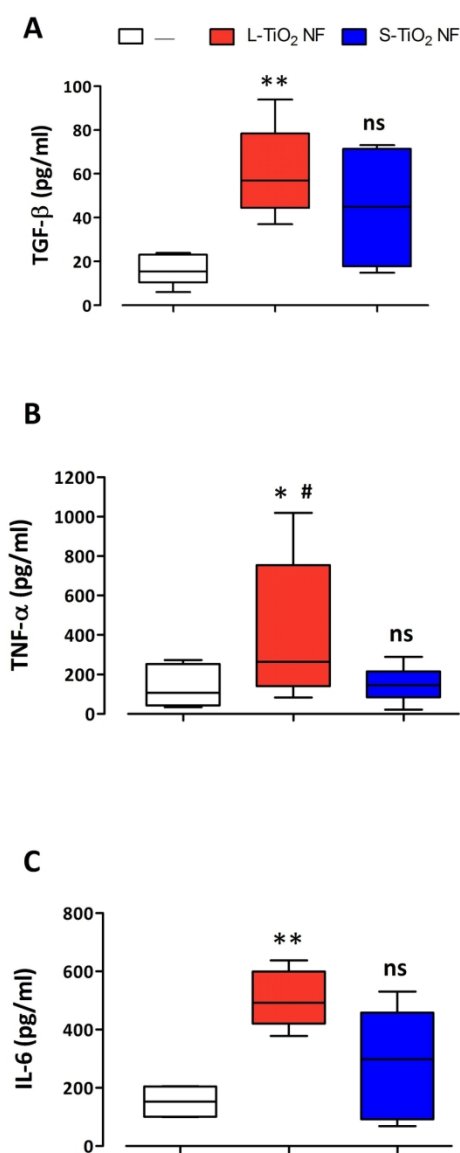


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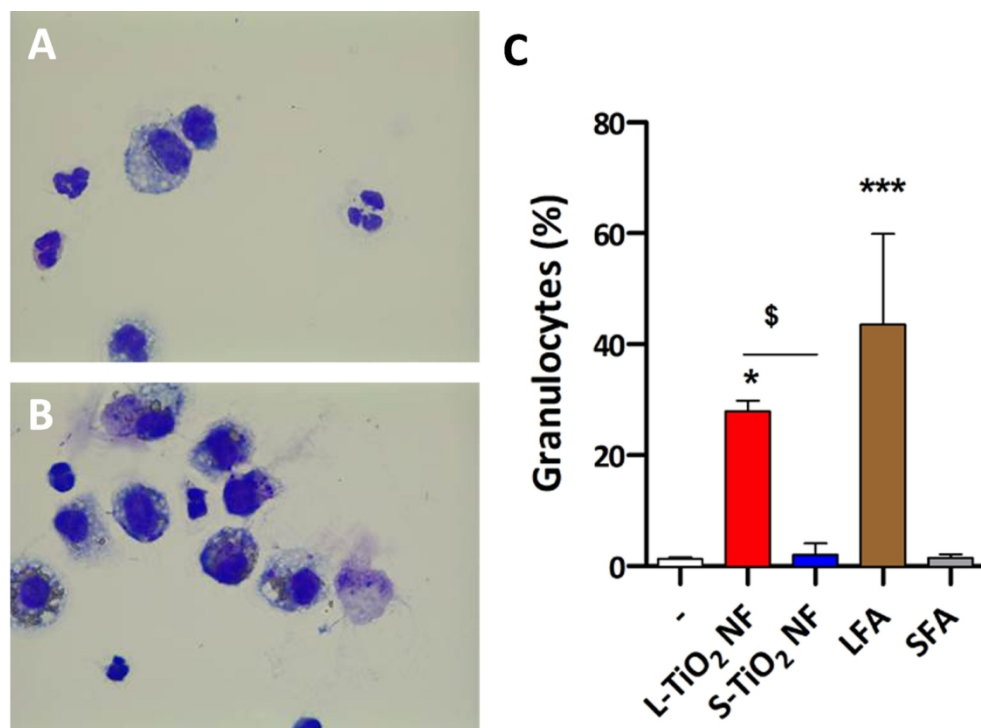


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132x95mm (300 x 300 DPI)

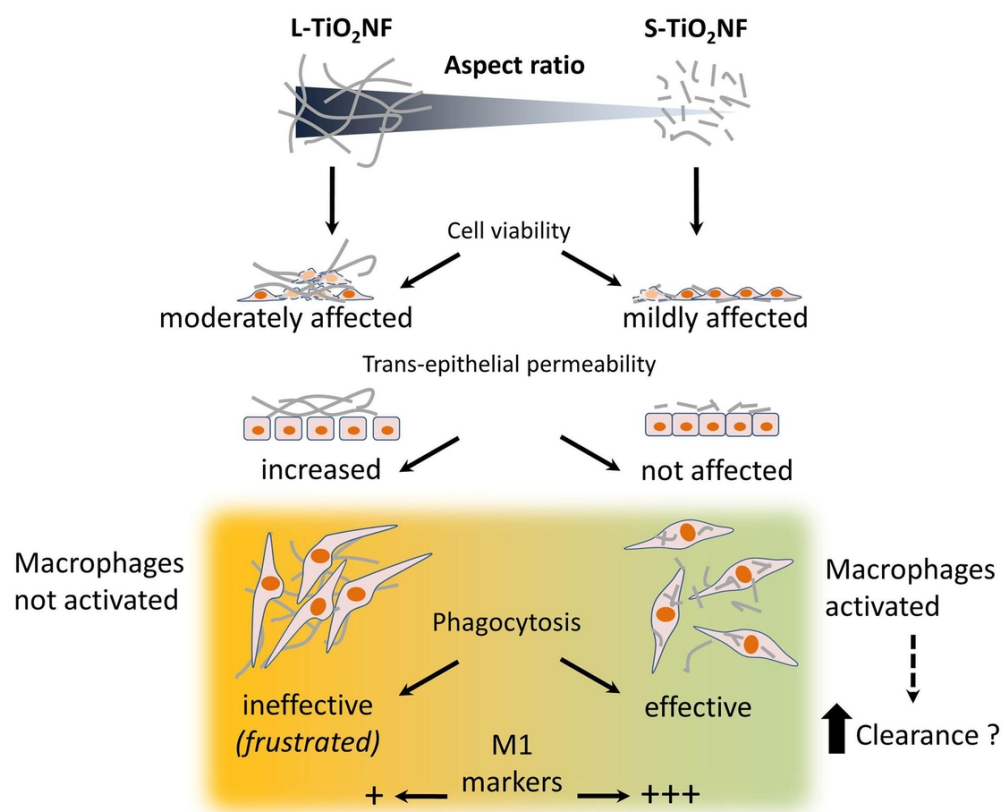


Figure 9. Differential outcomes of exposure to long and shortened TiO₂ nanofibers.

94x90mm (300 x 300 DPI)