View metadata, citation and similar papers at core.ac.uk

brought to you by CORE

provided by Institutional Research Information System University of Turin

UNIVERSITÀ DEGLI STUDI DI TORINO

This is an author version of the contribution published on:

Questa è la versione dell'autore dell'opera:

[Journal of Hepatology, 70(3), 2019, 10.1016/j.jhep.2018.10.021]

*ovvero [*Mayada Metwally[1], Ali Bayoumi[1], Manuel Romero-Gomez[2], Khaled Thabet[1,3], Miya John[1], Leon A. Adams[4], Xiaoqi Huo[1], Rocio Aller[5], Carmelo García-Monzón[6], María Teresa Arias-Loste[7], Elisabetta Bugianesi[8], Luca Miele[9], Rocio Gallego-Durán[2], Janett Fischer[10], Thomas Berg[10], Christopher Liddle[1], Liang Qiao[1], Jacob George[1], Mohammed Eslam[1], *70, ELSEVIER, 2019, pagg.494-500]*

The definitive version is available at:

La versione definitiva è disponibile alla URL:

[http://www.sciencedirect.com/science/journal/01688278]

A polymorphism in the Irisin-encoding gene (*FNDC5*) associates with hepatic steatosis by differential miRNA binding to the 3′UTR

Mayada Metwally[1], Ali Bayoumi[1], Manuel Romero-Gomez[2], Khaled Thabet[1,3], Miya John[1], Leon A. Adams[4], Xiaoqi Huo[1], Rocio Aller[5], Carmelo García-Monzón[6], María Teresa Arias-Loste[7], Elisabetta Bugianesi[8], Luca Miele[9], Rocio Gallego-Durán[2], Janett Fischer[10], Thomas Berg[10], Christopher Liddle[1], Liang Qiao[1], Jacob George[1], Mohammed Eslam[1]

Highlights

• Irisin, the cleaved extra-cellular fragment of FNDC5 is a myokine thought to have favorable metabolic activity.

• The role of variants in the *FNDC5* gene in NAFLD is not defined.

• Genetic variants in *FNDC5* confer risk of human severe hepatic steatosis.

• Functional studies reveal that this variant mediates this effect via a miRNA-mediated control of *FNDC5* mRNA stability.

• Irisin is likely to have a favourable metabolic impact on NAFLD.

Background & Aims

Irisin, the cleaved extra-cellular fragment of the Fibronectin type III domain-containing protein 5 (FNDC5) is a myokine that is proposed to have favorable metabolic activity. We aimed to elucidate the currently undefined role of variants in the *FNDC5* gene in non-alcoholic fatty liver disease (NAFLD).

Methods

We prioritized single nucleotide polymorphisms in *FNDC5* on the basis of their putative biological function and identified rs3480 in the 3′ untranslated region (3′UTR). We studied the association of rs3480 with liver disease severity and the metabolic profile of 987 Caucasian patients with NAFLD. Functional investigations were undertaken using luciferase reporter assays of the 3′UTR of human FNDC5, pyrosequencing for allele-specific expression of FNDC5 in liver, measurement of serum irisin, and bioinformatics analysis.

Results

The rs3480 (G) allele was associated with advanced steatosis (OR 1.29; 95% CI 1.08–1.55; $p = 0.004$), but not with other histological features. This effect was independent but additive to *PNPLA3* and *TM6SF2*. The rs3480 polymorphism influenced *FNDC5* mRNA stability and the binding of miR-135a-5P. Compared with controls, hepatic expression of this microRNA was upregulated while FNDC5 expression was downregulated. Elevated serum irisin was associated with reduced steatosis, and an improved metabolic profile.

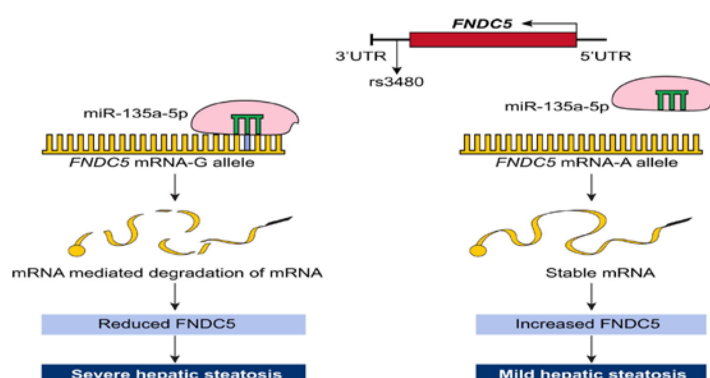
Conclusions

Carriage of the *FNDC5* rs3480 minor (G) allele is associated with more severe steatosis in NAFLD through a microRNA-mediated mechanism controlling *FNDC5* mRNA stability. Irisin is likely to have a favorable metabolic impact on NAFLD.

Lay summary

Irisin is a novel protein produced mainly by muscle, which is known to be released into the circulation, with an unclear role in liver fat deposition. This study demonstrates that genetic variants in the gene encoding the irisin protein modulate the risk of liver fat in patients with fatty liver disease. Interestingly, these effects are independent of, but additive to those of other recently described genetic variants that contribute to liver fat. In functional studies, we have deciphered the detailed molecular mechanisms by which this genetic variant mediates its effects.

Graphical abstract



Introduction

Non-alcoholic fatty liver disease (NAFLD) is a principal liver disorder in Western countries and is on trajectory to become the leading cause of end-stage liver disease and liver transplantation.[1], [2], [3] Notably, NAFLD is part of a multisystem metabolic disturbance, as its impact is not limited to the liver but also affects extra-hepatic sites such as the cardiovascular system and kidneys.[4] In NAFLD, there is pathological hepatic accumulation of fat that over time can lead to inflammation and progress to cirrhosis, end-stage liver disease and hepatocellular carcinoma. The mechanisms underlying the accumulation of liver fat are complex and gene × environment interactions play a critical role.[5], [6], [7]

The heritable component of hepatic steatosis has been estimated at ~50% based on a prospective twin study.[8] To date, the major inherited determinants of hepatic fat accumulation based on genome wide association studies are the patatin-like phospholipase domain-containing 3 (*PNPLA3*) I148M and the transmembrane 6 superfamily member 2 (*TM6SF2*) E167K gene variants.[9], [10]. However, these polymorphisms explain only 10–20% of the heritability.[6], [11] Thus, other as yet unidentified genetic and epigenetic variations likely exist to explain the missing heritability. Adipose-derived hormones, collectively termed adipokines are established drivers/contributors to the pathogenesis of NAFLD.[12], [13], [14] More recently, the myokine irisin was isolated from

muscle by Boström *et al.* 2012.[15] Irisin is a 12 kD, 112 amino acid fragment that is proteolytically processed from the fibronectin type III domain-containing protein 5 (FNDC5) and released into the circulation.[16] The functions of irisin and even its existence however have been a matter of debate,[16], [17] with some reports suggesting that the irisin polypeptide is a "myth".[17] Human *FNDC5* has an atypical ATA translation start codon rather than the ATG sequence. Hence, it has been argued that the human ATA codon represents a null mutation without irisin production and that reports measuring human irisin are an artefact from using an FNDC5 antibody with poor specificity.[17] However, recent work using quantitative mass spectrometry has confirmed the existence of circulating irisin.[18] The 3 published reports have shown inconsistent associations of NAFLD with irisin levels;[19], [20], [21] in addition, the role of genetic variants in *FNDC5* and the mechanisms involved have not been well defined.

In this work, after single nucleotide polymorphism (SNP) prioritization, we explored the role of the *FNDC5* polymorphism rs3480 on hepatic steatosis and other histological features in a cohort of 987 Caucasian patients with NAFLD and investigated the mechanisms underlying the association.

Patients and methods

Patient cohort

The study comprised 987 Caucasian patients with biopsy-proven NAFLD. The details of the cohort and their clinical and laboratory assessment are provided in the supplementary information. Ethics approval was obtained from the Human Research Ethics Committees of the Western Sydney Local Health District and the University of Sydney. All other sites had ethics approval from their respective ethics committees.

Prioritization of FNDC5 SNPs

To prioritize SNPs for genotyping, we followed the scheme illustrated in Fig. S1. In brief, common SNPs (minor allele frequency [MAF] >1%) within the *FNDC5* gene were tested for their status as expression quantitative trait loci (eQTLs) using the GTEx (Genotype-Tissue Expression) project,[22] as well as eQTLs from the MuTHER study using the exSNP database.[23] SNPs that were eQTLs for *FNDC5* were then investigated for p values for the eQTL. Following this filter, 2 SNPs (rs3480 and rs10753269) in tight LD ($r^2 = 0.99$) were found; the latter is upstream of *FNDC5*, while rs3480 is located in the 3′UTR. From this scheme, rs3480 was determined to be the SNP with the highest priority for genotyping in our cohort.

Genotyping

Genotyping for *FNDC5* rs3480, *PNPLA3* rs738409 and *TM6SF2* rs58542926 was undertaken using the TaqMan SNP genotyping allelic discrimination method (Applied Biosystems, Foster City, CA, USA). All genotyping was blinded to clinical variables. Some of the *PNPLA3* rs738409 and *TM6SF2* rs58542926 genotyping data were extracted from a recent report.[24]

Liver histopathology

Liver biopsies were scored by an expert liver pathologist in each participating center who was unaware of clinical or genetic data. Histological scoring was based on the system proposed by Kleiner *et al.*[25] Steatosis was graded from 0 to 3, lobular inflammation from 0 to 3 and hepatocellular ballooning from 0 to 2. Fibrosis was staged from 0 to 4 with 4 representing cirrhosis. The NAFLD activity score (NAS) was calculated to quantify disease activity.[25] The inter-observer agreement between pathologists was studied previously and was excellent for steatosis ($\kappa = 0.85$) and good for fibrosis ($\kappa = 0.78$).[26]

Allele-specific expression

Allelic ratios quantified by pyrosequencing were performed at the Australian Genome Research Facility. Full details are provided in supplementary methods, Table S1 and Fig. 2.

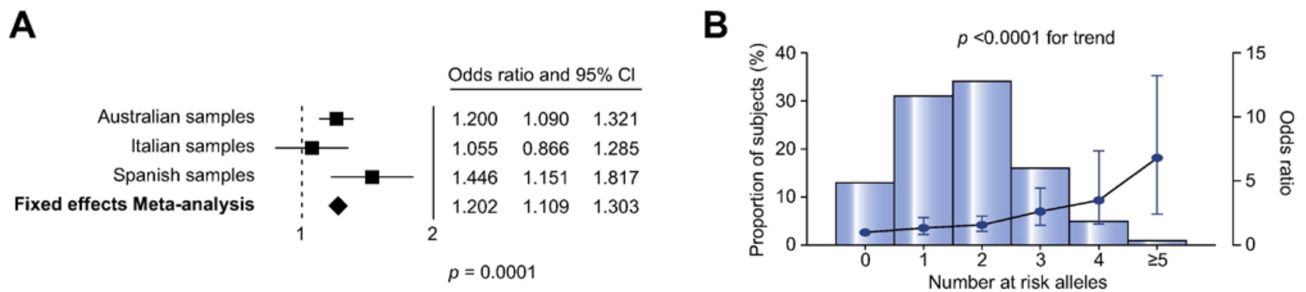


Fig. 1. ***FNDC5* alleles association with the severity of hepatic steatosis.** (A) Forest plot of associations of rs3480 with the severity of hepatic steatosis in individual samples and in the combined, fixed-effects meta-analysis (n = 987). (B) Association between the number of *PNPLA3*, *TM6SF2*, and *FNDC5* risk alleles and the severity of hepatic steatosis. The bar plots show the ORs (95% CI) for each number of risk alleles using zero risk alleles as the reference (right y axis). The histogram denotes the proportion of individuals in each genotype score category (left y axis). ORs, Odds ratios.

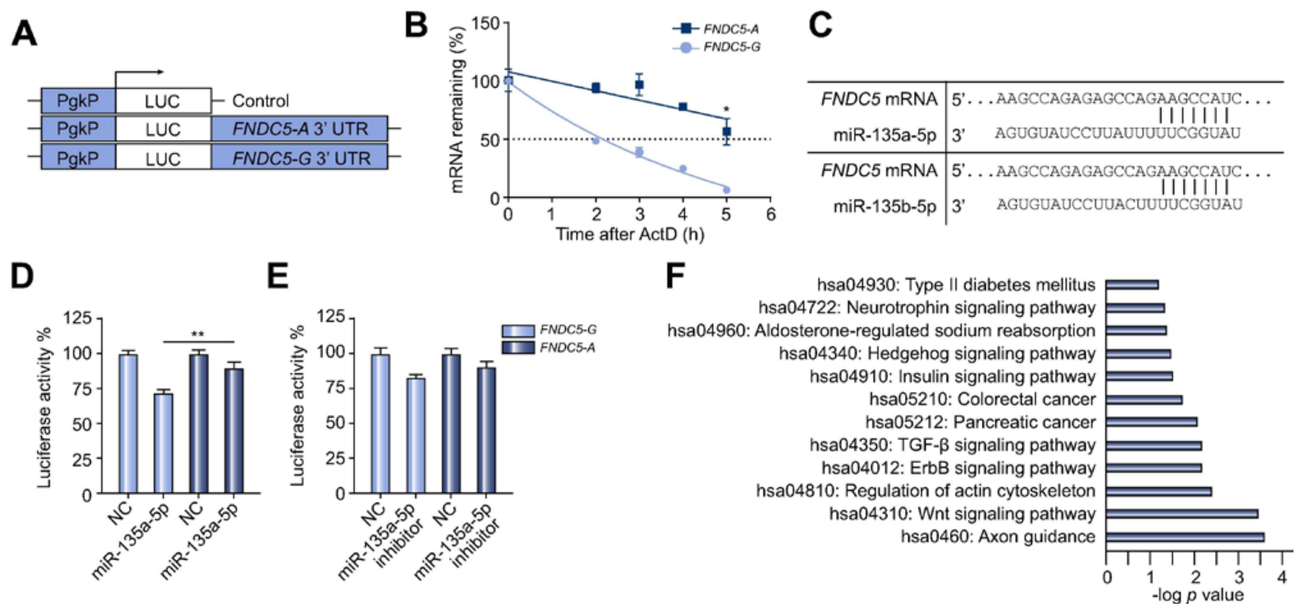


Fig. 2. **Allele-specific regulation of *FNDC5* by miRNA-135a-5P.** (A) *FNDC5* dual luciferase reporter constructs. The 3′UTR of *FNDC5* was cloned downstream of luciferase, which is driven by the constitutive Pgk promoter. (B) Stability of luciferase mRNA in Huh-7 cells transfected with the *FNDC5*-A or *FNDC5*-G luciferase construct and treated with actinomycin D (ActD) to arrest new transcription, presented as mRNA remaining over time relative to that at 0 h, set as 100%. Half-lives (50% mRNA remaining (dashed line)): *FNDC5*-A 3′UTR, 6.2 h; *FNDC5*-G 3′UTR, 3.7 h. (C) Bioinformatic analyses showing that rs3480 is the predicted target gene for miR-135a-5p and miR-135b-5p. (D) The miRNA mimics or miR inhibitor were co-transfected with the reporter gene constructions into Huh-7 cells. Luciferase assay showing that miR-135a-5p binds preferentially to the G allele, while there was less binding to the A allele. (E) Depletion of miR-135a-5p by miR inhibitor abrogated the difference in luciferase activity between the 2 alleles. (F) Gene set enrichment analysis of pathways targeted by miR-135a-5p showing −log *p* value (Table S5). **p* <0.05, ***p* <0.01, ****p* <0.001 by 2-tailed Student's *t* test. Error bars represent standard error of the mean.

Enzyme-linked immunosorbent assay (ELISA)

Serum irisin level was measured using a recently released irisin kit from Phoenix Pharmaceuticals (EK-067-29)[27] in a sub-cohort of 152 patients with available serum samples and with characteristics similar to the entire cohort.

Bioinformatics analyses

We queried the online miRdSNP database which is based on TargetScan and PicTar databases[28] to determine whether rs3480 (the SNP derived from our prioritization approach) in the 3′UTR of FNDC5 could be in miRNA target sites. Secondary structures of the mRNA and minimum folding energy (MFE) of FNDC-G and FNDC5-A were extrapolated using the RNAfold online server (Vienna RNA package, version 2.0.0, with option '-p').[29]

Gene set enrichment analysis

The target genes of MiR135a predicted by TargetScan. Gene set enrichment analysis used the Database for Annotation, Visualization and Integrated Discovery (DAVID)[30] to ask which Kyoto Encyclopedia of Genes and Genomes (KEGG) were enriched within the genes that contained binding sites for miR-135a.

Luciferase reporter assay

The full-length 3′UTR of human *FNDC5,* with genetic variation corresponding to A or G (of rs3480) was cloned into the PmeI/SalI restriction site downstream of the gene encoding firefly luciferase in the pmirGLO reporter vector (Promega).

Human liver sample miRNA extraction and Droplet Digital™ PCR

RNA was extracted from 5 μm of liver tissue (obtained from resections and stored at −80 °C) using the miRNeasy kit (Qiagen) from 20 individuals (n = 10 controls and n = 10 with NAFLD). The first-strand complementary synthesis reaction (cDNA) was performed using the TaqMan MicroRNA Reverse Transcription kit (Life Technologies) for miRNAs.

For miRNAs and FNDC5 copy number quantification, droplet digital PCR was performed, as previously described.[31], [32]

Statistical methods

Results are expressed as mean ± SD (standard deviation), median and interquartile range or number (percentage) of patients. All tests were 2-tailed and p values <0.05 were considered significant. For further details regarding the materials and methods used, please refer to the CTAT table and supplementary information.

Results

FNDC5 rs3480 and the degree of hepatic steatosis

Baseline characteristics of the cohort are summarized in Table S2. After SNP prioritization (described in detail in Fig. S2 and in methods), we found that rs3480 satisfied our criteria. *FNDC5* rs3480 was confirmed to be in Hardy–Weinberg equilibrium and showed a MAF of 0.41; 0.403, 0.41 and 0.404 in Australian, Italian and Spanish samples, respectively, similar to that observed in other European populations (MAF = 0.41), http://browser.1000genomes.org). No differences in clinical, anthropometric and biochemical indices including aminotransferases were found between the *FNDC5* rs3480 genotypes (Table S3).

Fixed-effects meta-analysis of the association of rs3480 with steatosis (S2-S3) was significant (OR 1.20; 95% CI 1.01–1.3; meta-p = 0.0001; Fig. 1A). In multivariate logistic regression analysis incorporating age, gender, body mass index (BMI), type 2 diabetes mellitus, recruiting center and

PNPLA3 rs738409 and *TM6SF2* rs58542926 genotype, the *FNDC5* rs3480 (G) allele was associated with an increased risk of steatosis (S2-S3) (OR 1.29; 95% CI 1.08–1.55; $p = 0.004$) (Table S4).

PNPLA3, TM6SF2 and *FNDC5* polymorphisms have an additive effect

We next examined the proportion of attributable risk conferred by the *FNDC5*, *PNPLA3*, and *TM6SF2* genetic variants. As expected, the *PNPLA3* I148M variant had the largest impact on steatosis (S2-S3); the estimated population-attributable risk (PAR) was 25.1%. Of interest, the *FNDC5* rs3480 variant had a greater PAR compared to the E167K *TM6SF2* variant (10.6% *vs.* 6.7%, respectively).
We then examined the interaction between the 3 variants but did not observe an impact on hepatic steatosis ($p > 0.05$). Instead, the 3 variants appeared to have an additive impact with a stepwise increase in odds ratio for steatosis (S2-S3) for each additional risk allele ($p < 0.0001$ for trend) (Fig. 1). In total, *FNDC5* rs3480 is a novel risk variant for steatosis, independent, but additive to the known risk variants, *PNPLA3* and *TM6SF2*.

FNDC5 rs3480 and the severity of hepatic inflammation and fibrosis

We tested the association of *FNDC5* rs3480 with the severity of lobular inflammation and ballooning. *FNDC5* rs3480 was not associated with more severe lobular inflammation (A2-A3) (OR 1.3; 95% CI 0.9–1.87; $p = 0.1$) and remained the same after adjustment for age, sex, BMI, diabetes, and severity of steatosis ($p = 0.2$). Similarly, no association with fibrosis (F2-F4) was observed (OR 1.05; 95% CI 0.74–1.47; $p = 0.7$), which remained the same after adjustment for the variables previously mentioned ($p = 0.79$).

Impact of SNP at rs3480 on the 3′UTR of FNDC5

To investigate the mechanisms for the genetic association, we evaluated the influence of the 3′UTR SNP rs3480 on the post-transcriptional regulation and stability of *FNDC5* mRNA. To examine whether mRNA folding of *FNDC5* could be altered by rs3480, bioinformatic analysis using the RNAfold web server was used. This demonstrated that rs3480 SNPs altered mRNA structure, suggesting that it may affect the stability of *FNDC5* mRNA, with a marginal variation in the MFE, from $\Delta G = -1102.20$ to -1102.10 kcal mol^{-1} (Fig. S2).
To validate these findings, we generated full-length 3′UTRs of *FNDC5* containing either the G or the A allele and cloned them into the pmirGLO vector (Fig. 2A), then transfected human hepatoma (Huh-7) cells with the constructs and measured luciferase activity. The *FNDC5*-G 3′ UTR conferred significantly lower luciferase activity compared to the *FNDC5*-A 3′UTR (Fig. S3). We next assessed the effect of rs3480 on mRNA stability. Analysis of the luciferase–encoding mRNA that remained in Huh-7 cells after treatment with actinomycin D revealed that mRNA bearing the *FNDC5*-G 3′UTR decayed significantly faster than that bearing the *FNDC5*-A 3′UTR (Fig. 2B). This implies that rs3480 alleles affect the stability of the mRNA transcript.
Next, we performed allele-specific expression analysis using pyrosequencing of gDNA (blood) and cDNA from 10 human liver samples from rs3480 heterozygous individuals. As demonstrated in Fig. S4, modest but significant allelic differences in gene expression were observed, with lower expression in the G allele ($p = 0.003$, nonparametric Wilcoxon's paired test). Together these data demonstrated that rs3480 influences the stability and expression of *FNDC5*.

Regulation of FNDC5 by miRNA

We investigated whether the rs3480 SNP could influence recruitment of miRNAs to the 3′UTR of *FNDC5*. Bioinformatics analysis of the 3′UTR sequence around the site of rs3480 identified potential binding sites for miR-135a-5P and miR-135b-5P (Fig. 2C). To investigate the preferential

binding of the miRNAs to the rs3480 alleles, we co-transfected Huh-7 cells with miR-135a-5P and miR-135-5P and negative control mimics with luciferase reporter constructs containing FNDC-A or FNDC5-G. The luciferase assay shows that miR-135a-5p binds to both A and G alleles, but preferentially to the G allele, demonstrated by significantly lower luciferase activity between G+ miR-135a-5p compared to A+ miR-135a-5p (Fig. 2D). Notably, the difference between alleles was blunted in the presence of the miR-135a-5P inhibitor (Fig. 2E), suggesting the specificity of decreased luciferase activity by miR-135a-5P. In contrast, miR-135b-5P binds to both alleles with no preferential binding (Fig. S5).

Gene set enrichment analysis

We then undertook pathway analysis of the common predicted targets of miR-135a-5P using KEGG pathway mapping. Collectively miR-135a-5P targets mapped onto different pathways (Fig. 2F and Table S5). Notably, they included overrepresentation of insulin signaling and type 2 diabetes pathways.

Hepatic miRNA and *FNDC5* levels in NAFLD

We next interrogated hepatic levels of miR-135a-5P, miR-135b-5P and *FNDC5* in liver from patients with NAFLD and controls by ddPCR. Compared to controls, patients with NAFLD had significantly higher levels of miR-135a-5P but not miR-135b ($p < 0.05$, Fig. 3A and Fig. S6). *FNDC5* levels tended to be lower in NAFLD, but not significantly (Fig. 3B). A statistically significant ($r^2 = 0.80$ $p \leq 0.002$) positive correlation was observed between miR-135a-5P and *FNDC5* in liver from normal control individuals, while they were negatively correlated in NAFLD ($r^2 = -0.36$) (Fig. S7). There was no difference in hepatic miR-135a and miR-135b levels according to rs3480 genotype (data not shown).

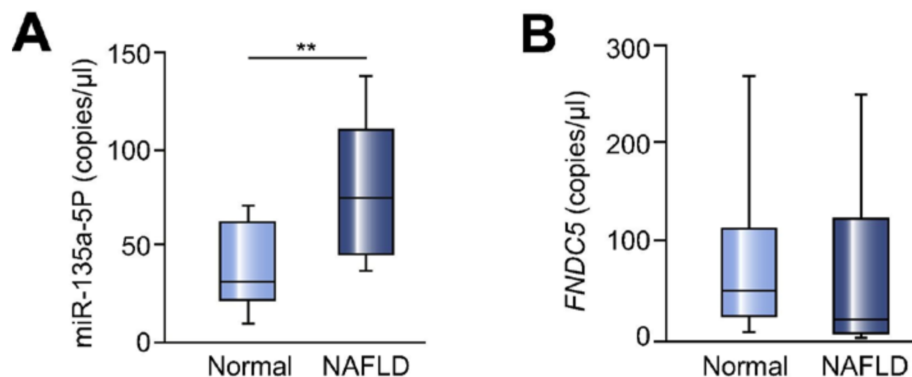


Fig. 3. **Absolute quantification of hepatic miR-135a-5p levels and *FNDC5* levels.** (A) miR-135a-5p levels are upregulated in NAFLD compared to controls in liver samples. (B) *FNDC5* levels are downregulated in the livers of patients with NAFLD compared to controls (n = 10 controls, n = 10 NAFLD). *$p < 0.05$, **$p < 0.01$, ***$p < 0.001$ by Mann-Whitney U test. Boxplots are median, minimum and maximum.

Elevated circulating irisin is associated with reduced steatosis and a favorable metabolic profile

Finally, we examined which clinical parameters were associated with serum irisin level in a sub-cohort of patients with NAFLD (n = 152) having available serum. Irisin levels correlated positively with high-density lipoprotein cholesterol levels (p <0.001), and negatively with HOMA-IR (p = 0.04). No correlations were observed with other clinical parameters (Table S6). Median irisin levels were higher in patients less than 40 years old compared to those ≥40 years old (p <0.01) (Fig. 4A) and in individuals with HOMA-IR <3 compared to those with HOMA-IR ≥3 (p <0.001) (Fig. 4B). Irisin serum levels were similar between females and males (p = 0.5). Finally, serum irisin levels (Fig. 4C) were lower in those with greater steatosis compared to those with mild steatosis (p <0.03), but did not differ by other histological features. Patients with the GG genotype had a trend towards lower median irisin levels compared with individuals carrying at least one A allele, though it was not significant (p = 0.07, Fig. S7). No difference in irisin level according to PNPLA3 or TM6SF2 genotypes was observed. Collectively, elevated serum irisin levels are associated with reduced steatosis and an improved metabolic profile.

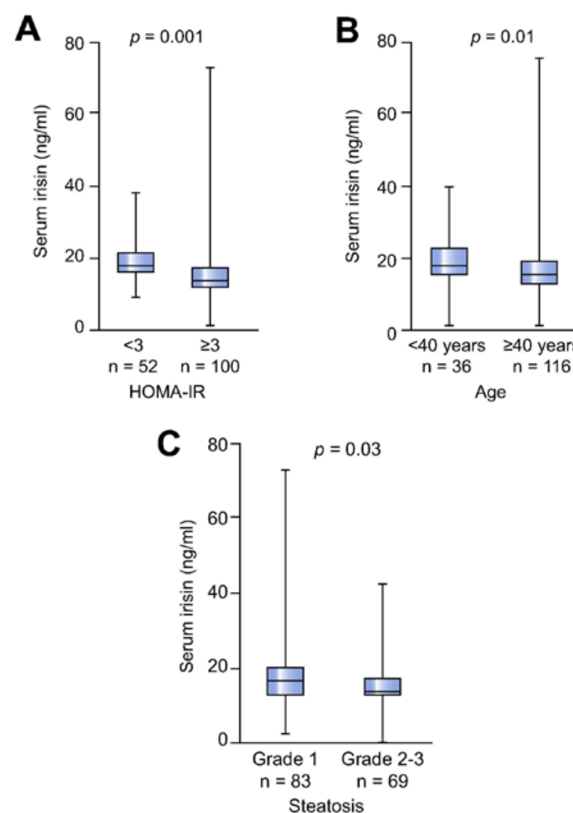


Fig. 4. Serum irisin levels. Serum irisin levels by (A) HOMA-IR, (B) age and (C) steatosis in a sub-cohort of 152 patients with NAFLD. Boxplots are median, minimum and maximum. Statistical significance was analyzed by the Mann-Whitney U test. The number of samples tested in each group is shown.

Discussion

In this study, we demonstrate that *FNDC5* rs3480 is a novel additional and possibly hitherto overlooked genetic variant affecting the risk of hepatic steatosis in NAFLD. The influence of the risk genotype was independent of, but additive to *PNPLA3* and *TM6SF2* in Caucasian patients. We show a 1.4- to 6.8-fold increase in hepatic steatosis risk based on the cumulative association of the 3 variants among individuals who had at least one copy of the risk allele at each of the 3 regions, compared with individuals without any risk alleles. The G allele associated with greater *FNDC5* mRNA degradation compared to A allele, had a modest but significant effect on liver *FNDC5* allele-specific expression and also created a binding site for miR-135a-5P. Compared with controls, patients with NAFLD had significantly higher levels of hepatic miR-135a-5p and lower *FNDC5*. Concomitantly, this miRNA can regulate several pathways linked to liver disease including those for insulin signaling and type 2 diabetes. Lastly, elevated serum irisin levels were associated with reduced steatosis and with improved indices of the circulating metabolic profile. Collectively, these data suggest that miRNA binding to an SNP in the 3′UTR, impairs *FNDC5* expression and thereby increases the severity of hepatic steatosis (Fig. 5).

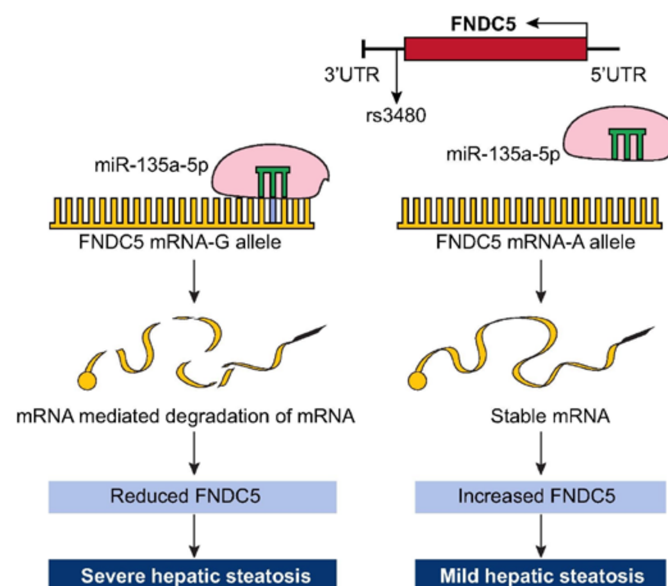


Fig. 5. **Proposed model for the regulation of *FNDC5* and hepatic steatosis.** miR-135a-5p binds preferentially to the G allele of rs3480 in the 3′UTR of the *FNDC5* gene. Consequently, miRNA mediated decay of *FNDC5* is enhanced and this results in less *FNDC5* mRNA. As FNDC5 has favorable metabolic impacts, this results in greater hepatic steatosis.

While carriage of the *FNDC5* rs3480 (G) minor allele was associated with an increased risk of steatosis, it did not impact other histologic features. The influence on steatosis was also independent of known steatosis risk confounders including age, gender, BMI, type 2 diabetes mellitus and recruitment center. Since no interaction was observed between *FNDC5* rs3480, *PNPLA3* rs738409 and *TM6SF2* rs58542926 genotypes, our data suggests that the 3 genes regulate different pathways in steatosis pathogenesis.

Serum irisin levels were reduced in individuals with greater hepatic steatosis (S2-S3). Both in a study of 296 obese Chinese adults in whom hepatic triglyceride content was determined by ^{1}H MRS spectroscopy[21] and another of 84 Koreans with NAFLD in whom steatosis was measured by

abdominal ultrasonography,[19] irisin levels declined with increased liver fat. However, in another small study (n = 31) of patients with biopsy-proven NAFLD, no difference in serum irisin according to steatosis grade was observed.[20] These studies used the same ELISA kit and thus differences are likely due to sample size considerations. While there has been skepticism on the quality of commercial ELISAs to quantify irisin, in the present work it was measured using the recently released Phoenix Pharmaceuticals (EK-067-29) kit that has been validated by comparison with data from Western blot and MALDI-TOF mass spectrometry.[27]

The mechanisms for the protective effects of irisin on hepatic steatosis in NAFLD are not clear, but evidence suggests both direct and indirect beneficial effects. In humans, *FNDC5* is expressed in skeletal muscle and adipose tissues serving as both a myokine and adipokine, though the principle source appears to be muscle.[33] In the liver, hepatocyte-released irisin functions as a paracrine/autocrine hormone.[34] It has been reported that adipose tissue *FNDC5* mRNA and circulating irisin levels negatively correlate with hyperglycemia, elevated triglycerides, visceral adiposity and extramyocellular lipid deposition, features that are concordant with the metabolic abnormalities observed in NAFLD.[35] Moreover, treating muscle cells *in vitro* with palmitate and glucose lowered *FNDC5* mRNA suggesting peripheral effects on metabolic and glucose homeostasis.[35] This data is supported by animal studies that FNDC5 improves hepatic steatosis, protects mice against genetic- and diet-induced obesity, and improves hepatic and peripheral insulin resistance via regulating AMPK signaling in myocytes and hepatocytes.[34], [36], [37], [38]

miRNAs are small non-coding RNA transcripts of ∼22 nucleotides that regulate gene expression at the post-transcriptional level.[39] They do so by targeting sites of complementarity in the 3′UTR of mRNAs, thereby mediating mRNA decay or translational repression. miRNAs can repress multiple targets within the same pathway resulting in amplification of their biological effects. SNPs can create, destroy or modify sites for miRNA binding. Our results reveal that rs3480 with an A-to-G substitution in the 3′UTR of *FNDC5* results in decreased expression of *FNDC5* through two post-transcriptional mechanisms. Firstly, *FNDC5*-G suffered more degradation than did *FNDC5*-A; secondly, it also differentially bound mir-135a-5P. Hepatic levels of mir-135a-5P and *FNDC5* as measured by ddPCR were upregulated and downregulated, respectively, in patients with NAFLD compared with controls. Not surprisingly, a positive correlation was present between mir-135a-5P and *FNDC5* in normal individuals, while this correlation was reversed in NAFLD. This suggests that miRNA modulation of *FNDC5* might be a form of “compensatory adaptation“ in normal liver, whereby the miRNAs do not downregulate the expression of *FNDC5* and might indicate a signal for transcript downregulation. When fatty liver develops, the correlation becomes negative as adaptive mechanisms fail. mir-135a-5P is modulated more than *FNDC5* in NAFLD, suggesting that induction of mir-135a-5P might be an early event in NAFLD development. Consistently, miR-135a levels were elevated in serum, plasma, renal and skeletal muscle in patients with diabetes compared to controls.[40], [41], [42] Notably, our pathway analysis also suggested that miR-135a maps to different pathways, including an overrepresentation of insulin signaling and type 2 diabetes pathways. In this regard, mir-135a has been shown to inhibit insulin signaling and glucose uptake by targeting insulin receptor substrate 2 (IRS2). Of relevance, we demonstrated that serum irisin level was negatively correlated with HOMA-IR.

We found no association between *FNDC5* rs3480 genotype or serum irisin levels with histological disease activity or fibrosis, or with liver tests as indices of liver injury. According to our data, genotypic variation at rs3480 has a modest effect size on steatosis and thus is likely insufficient to influence the transition to steatohepatitis and fibrosis. Our data however do not exclude the possibility of effects on inflammation and fibrosis in larger cohorts powered to detect such effects. This is relevant, as we have shown that serum irisin levels demonstrate an inverse correlation with measures of insulin resistance and dyslipidemia that are associated with inflammation and fibrosis.[12], [43] However, the alternate possibility is that irisin disentangles steatosis from fibrosis. In this context, we have previously shown that serum leptin correlates with steatosis but

not fibrosis in NAFLD,[14] suggesting both shared and differential pathways between susceptibility to steatosis and risk of disease progression.
A recent smaller study (n = 593) reported that the rs3480 (G) allele was associated with the severity of hepatic steatosis only in univariate analysis, consistent with our finding, while the association was lost after multiple logistic regression.[44] Surprisingly, in that study, the same allele was associated with less severe hepatic fibrosis. The reasons for this discrepancy are unclear.
In conclusion, in this study we characterized a functional *FNDC5* variant as an additional risk variant associated with hepatic steatosis independently of *PNPLA3* and *TM6SF2*. This polymorphism influenced transcript stability, as well as the binding of miRNAs, suggesting a model whereby rs3480 modulates steatosis by regulating *FNDC5* expression.

Financial support

ME and JG are supported by the Robert W. Storr Bequest to the Sydney Medical Foundation, University of Sydney; a National Health and Medical Research Council of Australia (NHMRC) Program Grant (APP1053206, APP1149976) and Project grants (APP1107178 and APP1108422). AB is supported by an Australian Government Research Training Program (RTP) scholarship. EB is supported by Horizon 2020 under grant 634413 for the project EPoS.

Conflict of interest

The authors declare no conflicts of interest that pertain to this work.

Please refer to the accompanying ICMJE disclosure forms for further details.

Authors' contributions

M.M., M.E. and J.G. conceived the research; patient enrollment, clinical phenotype data collation and sample acquisition/DNA preparation, genotyping was performed; M.M., M.RG., L.A.D, R.A, C.G.M., M.T.A., E.B., L.M., R.G.D., J.F., T.B, C.L., L.Q., M.E, J.G., statistical analysis and interpretation of results was performed by M.M., A.B., M.E. and J.G, ddPCR: M.M., A.B; Tissue culture and luciferase assay: M.M, A.B and M.J, ELISA: K.T, Bioinformatics analysis: X.H., the manuscript was written and revised by M.M, M.E. and J.G. All authors critically reviewed the manuscript and approved the final submitted manuscript.

Acknowledgements

We would like to thank all the patients for their participation in this study.

References

Author names in bold designate shared co-first authorship

[1]

Z. Younossi, Q.M. Anstee, M. Marietti, T. Hardy, L. Henry, M. Eslam, *et al.***Global burden of NAFLD and NASH: trends, predictions, risk factors and prevention**
Nat Rev Gastro Hepat, 15 (2018), pp. 11-20

[2]

L.A. Adams, V. Ratziu**Non-alcoholic fatty liver – perhaps not so benign**

J Hepatol, 62 (2015), pp. 1002-1004

[3]

V. Ratziu, M. Voiculescu, T. Poynard**Touching some firm ground in the epidemiology of NASH**
J Hepatol, 56 (2012), pp. 23-25

[4]

C.D. Byrne, G. Targher**NAFLD: a multisystem disease**
J Hepatol, 62 (2015), pp. S47-64

[5]

M. Eslam, J. George**Genetic and epigenetic mechanisms of NASH**
Hepatol Int, 10 (2016), pp. 394-406

[6]

T.H. Karlsen, F. Lammert, R.J. Thompson**Genetics of liver disease: from pathophysiology to clinical practice**
J Hepatol, 62 (2015), pp. S6-S14

[7]

V. Zimmer, F. Lammert**Genetics in liver disease: new concepts**
Curr Opin Gastroenterol, 27 (2011), pp. 231-239

[8]

R. Loomba, N. Schork, C.H. Chen, R. Bettencourt, A. Bhatt, B. Ang, *et al.***Heritability of hepatic fibrosis and steatosis based on a prospective twin study**
Gastroenterology, 149 (2015), pp. 1784-1793

[9]

J. Kozlitina, E. Smagris, S. Stender, B.G. Nordestgaard, H.H. Zhou, A. Tybjaerg-Hansen, *et al.***Exome-wide association study identifies a TM6SF2 variant that confers susceptibility to nonalcoholic fatty liver disease**
Nat Genet, 46 (2014), pp. 352-356

[10]

S. Romeo, J. Kozlitina, C. Xing, A. Pertsemlidis, D. Cox, L.A. Pennacchio, *et al.***Genetic variation in PNPLA3 confers susceptibility to nonalcoholic fatty liver disease**
Nat Genet, 40 (2008), pp. 1461-1465

[11]

M. Eslam, L. Valenti, S. Romeo**Genetics and epigenetics of NAFLD and NASH: clinical impact**
J Hepatol, 68 (2) (2018), pp. 268-279

[12]

K.L. Milner, D. van der Poorten, A. Xu, E. Bugianesi, J.G. Kench, K.S. Lam, *et al.***Adipocyte fatty acid binding protein levels relate to inflammation and fibrosis in nonalcoholic fatty liver disease**
Hepatology, 49 (2009), pp. 1926-1934

[13]

J.M. Hui, A. Hodge, G.C. Farrell, J.G. Kench, A. Kriketos, J. George**Beyond insulin resistance in NASH: TNF-alpha or adiponectin?**
Hepatology, 40 (2004), pp. 46-54

[14]

S. Chitturi, G. Farrell, L. Frost, A. Kriketos, R. Lin, C. Liddle, *et al.***Serum leptin in NASH correlates with hepatic steatosis but not fibrosis: a manifestation of lipotoxicity?**
Hepatology, 36 (2002), pp. 403-409

[15]

P. Bostrom, J. Wu, M.P. Jedrychowski, A. Korde, L. Ye, J.C. Lo, *et al.***A PGC1-alpha-dependent myokine that drives brown-fat-like development of white fat and thermogenesis**
Nature, 481 (2012)
463-U472

[16]

J.Y. Huh, G. Panagiotou, V. Mougios, M. Brinkoetter, M.T. Vamvini, B.E. Schneider, *et al.***FNDC5 and irisin in humans: I. Predictors of circulating concentrations in serum and plasma and II. mRNA expression and circulating concentrations in response to weight loss and exercise**
Metabolism, 61 (2012), pp. 1725-1738

[17]

E. Albrecht, F. Norheim, B. Thiede, T. Holen, T. Ohashi, L. Schering, *et al.***Irisin – a myth rather than an exercise-inducible myokine**
Sci Rep, 5 (2015), p. 8889

[18]

M.P. Jedrychowski, C.D. Wrann, J.A. Paulo, K.K. Gerber, J. Szpyt, M.M. Robinson, *et al.***Detection and quantitation of circulating human irisin by tandem mass spectrometry**
Cell Metab, 22 (2015), pp. 734-740

[19]

E.S. Choi, M.K. Kim, M.K. Song, J.M. Kim, E.S. Kim, W.J. Chung, *et al.***Association between serum irisin levels and non-alcoholic fatty liver disease in health screen examinees**
PLoS One, 9 (10) (2014)
e110680

[20]

S.A. Polyzos, J. Kountouras, A.D. Anastasilakis, E.V. Geladari, C.S. Mantzoros**Irisin in patients with nonalcoholic fatty liver disease**
Metabolism, 63 (2014), pp. 207-217

[21]

H.J. Zhang, X.F. Zhang, Z.M. Ma, L.L. Pan, Z. Chen, H.W. Han, *et al.***Irisin is inversely associated with intrahepatic triglyceride contents in obese adults**
J Hepatol, 59 (2013), pp. 557-562

[22]

J. Lonsdale, J. Thomas, M. Salvatore, R. Phillips, E. Lo, S. Shad, *et al.***The genotype-tissue expression (GTEx) project**
Nat Genet, 45 (2013), pp. 580-585

[23]

E. Grundberg, K.S. Small, A.K. Hedman, A.C. Nica, A. Buil, S. Keildson, *et al.***Mapping cis- and trans-regulatory effects across multiple tissues in twins**
Nat Genet, 44 (2012), pp. 1084-1089

[24]

M. Eslam, A. Mangia, T. Berg, H.L.Y. Chan, W.L. Irving, G.J. Dore, *et al.***Diverse impacts of the rs58542926 E167K variant in TM6SF2 on viral and metabolic liver disease phenotypes**
Hepatology, 64 (2016), pp. 34-46

[25]

D.E. Kleiner, E.M. Brunt, M. Van Natta, C. Behling, M.J. Contos, O.W. Cummings, *et al.***Design and validation of a histological scoring system for nonalcoholic fatty liver disease**

Hepatology, 41 (2005), pp. 1313-1321

[26]

K. Kazankov, F. Barrera, H.J. Moller, C. Rosso, E. Bugianesi, E. David, *et al.***The macrophage activation marker sCD163 is associated with morphological disease stages in patients with non-alcoholic fatty liver disease**

Liver Int, 36 (10) (2016), pp. 1549-1557

[27]

S.A. Polyzos, C.S. Mantzoros**An update on the validity of irisin assays and the link between irisin and hepatic metabolism**

Metabol Clin Exp, 64 (2015), pp. 937-942

[28]

A.E. Bruno, L. Li, J.L. Kalabus, Y. Pan, A. Yu, Z. Hu**miRdSNP: a database of disease-associated SNPs and microRNA target sites on 3'UTRs of human genes**

BMC Genomics, 13 (2012), p. 44

[29]

I.L. Hofacker**Vienna RNA secondary structure server**

Nucleic Acids Res, 31 (2003), pp. 3429-3431

[30]

D.W. Huang, B.T. Sherman, R.A. Lempicki**Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists**

Nucleic Acids Res, 37 (2009), pp. 1-13

[31]

M. Eslam, D. McLeod, K.S. Kelaeng, A. Mangia, T. Berg, K. Thabet, *et al.***IFN-lambda 3, not IFN-lambda 4, likely mediates IFNL3-IFNL4 haplotype-dependent hepatic inflammation and fibrosis**

Nat Genet, 49 (2017), p. 795

[32]

C. Chai, M. Rivkin, L. Berkovits, A. Simerzin, E. Zorde-Khvalevsky, N. Rosenberg, *et al.***Metabolic circuit involving free fatty acids, microRNA 122, and triglyceride synthesis in liver and muscle tissues**

Gastroenterology, 153 (2017), pp. 1404-1415

[33]

J.M. Moreno-Navarrete, F. Ortega, M. Serrano, E. Guerra, G. Pardo, F. Tinahones, *et al.***Irisin is expressed and produced by human muscle and adipose tissue in association with obesity and insulin resistance**

J Clin Endocr Metab, 98 (2013), pp. E769-E778

[34]

L. Mo, J. Shen, Q.H. Liu, Y.W. Zhang, J.Y. Kuang, S.Y. Pu, *et al.***Irisin is regulated by CAR in liver and is a mediator of hepatic glucose and lipid metabolism**

Mol Endocrinol, 30 (2016), pp. 533-542

[35]

T. Kurdiova, M. Balaz, M. Vician, D. Maderova, M. Vlcek, L. Valkovic, *et al.***Effects of obesity, diabetes and exercise on Fndc5 gene expression and irisin release in human skeletal muscle and adipose tissue: in vivo and in vitro studies**

J Physiol-London, 592 (2014), pp. 1091-1107

[36]

T.Y. Liu, X.Q. Xiong, X.S. Ren, M.X. Zhao, C.X. Shi, J.J. Wang, *et al.***FNDC5 alleviates hepatosteatosis by restoring AMPK/mTOR-mediated autophagy, fatty acid oxidation, and lipogenesis in mice**
Diabetes, 65 (2016), pp. 3262-3275

[37]

C. Xin, J. Liu, J. Zhang, D. Zhu, H. Wang, L. Xiong, *et al.***Irisin improves fatty acid oxidation and glucose utilization in type 2 diabetes by regulating the AMPK signaling pathway**
Int J Obes (Lond), 40 (2016), pp. 443-451

[38]

Z. Yang, X. Chen, Y. Chen, Q. Zhao**Decreased irisin secretion contributes to muscle insulin resistance in high-fat diet mice**
Int J Clin Exp Pathol, 8 (2015), pp. 6490-6497

[39]

M.A. Saunders, H. Liang, W.H. Li**Human polymorphism at microRNAs and microRNA target sites**
Proc Natl Acad Sci U S A, 104 (2007), pp. 3300-3305

[40]

F. He, F.F. Peng, X. Xia, C. Zhao, Q.M. Luo, W.M. Guan, *et al.***MiR-135a promotes renal fibrosis in diabetic nephropathy by regulating TRPC1**
Diabetologia, 57 (2014), pp. 1726-1736

[41]

P. Agarwal, R. Srivastava, A.K. Srivastava, S. Ali, M. Datta**miR-135a targets IRS2 and regulates insulin signaling and glucose uptake in the diabetic gastrocnemius skeletal muscle**
BBA-Mol Basis Dis, 1832 (2013), pp. 1294-1303

[42]

M.R. Sarookhani, M. Honardoost, F. Foroughi, Y.K. Monfared**Plasma miR-135a; a potential biomarker for diagnosis of new type 2 diabetes (T2DM)**
Bali Med J, 7 (2018), pp. 296-301

[43]

S. Chitturi, S. Abeygunasekera, G.C. Farrell, J. Holmes-Walker, J.M. Hui, C. Fung, *et al.***NASH and insulin resistance: insulin hypersecretion and specific association with the insulin resistance syndrome**
Hepatology, 35 (2002), pp. 373-379

[44]

S. Petta, L. Valenti, G. Svegliati-Baroni, M. Ruscica, R.M. Pipitone, P. Dongiovanni, *et al.***Fibronectin type III domain-containing protein 5 rs3480 A > G polymorphism, irisin, and liver fibrosis in patients with nonalcoholic fatty liver disease**
J Clin Endocr Metab, 102 (2017), pp. 2660-2669