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Alpha-Enolase (*ENO1*), a potential target in novel immunotherapies

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1. ABSTRACT

Alpha-enolase (*ENO1*) is a metabolic enzyme involved in the synthesis of pyruvate. It also acts as a plasminogen receptor and mediates the activation of plasmin and extracellular matrix degradation. In tumor cells, *ENO1* is up-regulated and supports the Warburg effect; it is expressed at the cell surface, where it promotes cancer invasion, and is subjected to a specific array of post-translational modifications, namely acetylation, methylation and phosphorylation. *ENO1* overexpression and post-translational modifications could be of diagnostic and prognostic value in many cancer types. Information on the biochemical, proteomics and immunological characterization of *ENO1*, and particularly its ability to trigger a strong specific humoral and cellular immune response, make this ubiquitous protein an interesting tumor target; DNA vaccination with *ENO1* in preclinical models efficiently delays the development of very aggressive tumors such as pancreatic cancer. This review aims to analyze the main stages by which the tumor associated antigen (TAA) *ENO1* has become a promising target that opens potential avenues for cancer immunotherapy.

2. INTRODUCTION

Tumor immunotherapy is mostly based on the overexpression of tumor associated antigens (TAAs) in cancer, compared to normal tissues and on the ability of the immune system to recognize them and to induce a specific immune response (1). In 1943, the pioneering study of Gross and colleagues proved that tumors induced by oncogenic viruses were rejected through the recognition of tumor antigens, and that chemically-induced tumors were able to immunize mice to recognize a second exposure of the same tumor cells (2). Further studies in the late 70s also demonstrated the presence of tumor antigens in a mouse teratocarcinoma cell line (3) and in spontaneous mouse tumors (4), indicating that tumor antigens were not strictly artifacts induced by chemical treatment and that they are also likely to be present in human tumors (5). As TAAs have been shown to be increasingly important as immunotherapy targets, many groups have looked for not yet characterized or more immunogenic new TAAs, by developing different approaches. Some strategies were based on the reactivity of cytotoxic T lymphocytes (CTL) isolated from cancer patients against the autologous tumor (6-9), while others focused on the antibody response of cancer patients.

3. IMMUNE RESPONSE TO TUMOR ASSOCIATED ANTIGENS IN CANCER PATIENTS

A methodology known as serological analysis of recombinant cDNA expression libraries (SEREX) (10) has been useful to identify several hundreds of TAAs (11, 12). A similar technique, exploiting the presence of antibodies in the sera of cancer patients, coupled with a proteomic approach, is SERological Proteome Analysis (SERPA), which allowed to identify TAAs in many kind of tumors (13, 14), and in particular against pancreatic ductal adenocarcinoma (PDA), which is one of most aggressive solid tumors (15). Among these antigens, alpha-enolase (*ENO1*) was identified as a promising TAA due to its ability to induce a humoral and/or cellular immune response in cancer patients (16). Moreover, overexpression of *ENO1* at mRNA and protein level was observed in different tumor types including brain, breast, cartilage, cervix, colon, eye, gastric, head and neck, kidney, leukemia, liver, lung, muscle, ovary, pancreas, prostate, skin and testis cancers (17-50) (Table 1).

ENO1 has been shown to induce autoantibody production in many types of cancer patients. More in detail with the SERPA technique has been found the presence of anti-*ENO1* antibodies in cholangiocarcinoma, breast cancer, head and neck cancer, leukemia, lung cancer, pancreatic cancer and melanoma patients (39, 42, 50-62). In late stage of lung and breast cancers, anti-*ENO1* autoantibodies have been found decreased suggesting that they may serve as a prognostic marker to monitor disease progression (63). An explanation could be that tumor cells reduce the circulating levels of anti-*ENO1* antibodies through physical absorption and neutralization with surface-expressed and secreted *ENO1*, as suggested by *in vivo* experiment (64). Interestingly in lung cancer the higher presence of anti-*ENO1* antibodies after surgery correlated with a lower hazard ratio and a better progression-free survival (64). A spontaneous immune responses to *ENO1* has also manifested in patients with primary and metastatic melanomas (65) (Table 1).

Circulating anti-*ENO1* antibodies have been found in several autoimmune disease such as lupus nephritis (66, 67) and autoimmune retinopathy (68) as well as in cancer-associated retinopathy (69, 70). In breast cancer patients with associated retinopathy an increased incidence of autoantibodies in general has been observed (71, 72). Moreover, antibodies against citrullinated *ENO1* epitopes were observed in rheumatoid arthritis patients (73, 74). The correlation between cancer and autoimmunity could be due to the production of immunogenic and pro-inflammatory stimuli by tumor cell death and to the resulting activation of the inflammatory process within the tumor microenvironment, which concurs to increase the presentation of self-antigens to the immune system (75).

Almost two-thirds of PDA patients display an antibody response against *ENO1*, which is absent or present at a very low frequency in healthy donors, non-PDA cancer and chronic pancreatic patients (15) suggesting its diagnostic value in PDA (16). Autoantibodies from PDA patients specifically recognize two highly phosphorylated acid isoforms of *ENO1* (identified as *ENO1*,2). *ENO1*,2 isoforms are up-regulated in PDA compared to normal pancreas and display phosphorylation of serine 419, which is absent in other tumor cell lines (57, 76). Importantly, the presence of autoantibodies against *ENO1*,2 discriminate PDA patients with normal levels of CA19.9. and complement the diagnostic performance of serum CA19.9., increasing the sensitivity from 62% to 95%. The presence of anti-*ENO1*,2 antibodies correlate with a longer progression-free survival and a better clinical outcome in PDA patients treated with standard gemcitabine-based chemotherapy (57). In general, phosphorylation of a protein is associated with a higher affinity of its peptides for Major Histocompatibility Complex (MHC) molecules (77), suggesting that peptides from phosphorylated *ENO1*,2 could be better or more frequently presented by specific MHC complex to T cells. The detection of specific autoantibodies for phosphorylated isoforms of *ENO1* indeed correlates with a higher frequency of the allele HLA-DRB1*8 among PDA patients. Furthermore, repeated *in vitro* stimulation of peripheral blood mononuclear cells (PBMC) from HLA-DRB1*8 healthy subjects with phosphorylated *ENO1* peptide (predicted by a bioinformatics algorithm) elicits a significant CD4+ T cell proliferative response compared to the unphosphorylated *ENO1* peptide (78). An *ENO1* natural antigenic HLA-DRB1*8-restricted peptide from a squamous cell carcinoma cell line (OSC-20), which is specifically recognized by the CD4 cytotoxic T cell line TcOSC-20 has been also identified (79). Many TAAs stimulate an integrated humoral and cellular response by activating both T and B cells (16), and this coordinate response is one of the main effector mechanism exploitable by tumor immunotherapy.

To date, a coordinate specific response to *ENO1* has only been demonstrated in head and neck cancer (79, 80), melanoma (65) and pancreatic cancer patients (47, 81-83) (Table 1). PDA patients with anti-*ENO1* circulating antibodies display peripheral T cells that secreted IFN-γ when they are activated *in vitro* with recombinant *ENO1* (47). *In vitro, ENO1* is also able to elicit specific proliferation and activation of T cells and differentiation of specific CTL from healthy donor PBMC. *ENO1*-specific CTL spare *in vitro* normal skin Human Leukocyte Antihen (HLA)-matched fibroblasts from killing, but induce the inhibition of HLA-matched PDA cells *in vitro* and *in vivo* (47).

The response of T cells against *ENO1* does not always lead to an effector function as, in some circumstances, *ENO1*-specific T cells display a T regulatory (Treg) phenotype. The presence of *ENO1*-specific Treg cells in mice with *ENO1*-overexpressed lung tumors has been demonstrated. In particular, Treg cells isolated from tumors suppressed the proliferation of *ENO1*-specific CD4+ T cells, and mice bearing *ENO1*-overexpressed tumors showed a reduced production of anti-*ENO1* antibodies

(64). These results indicate that the presence of anti-*ENO1* antibodies correlates with the anti-tumor effector responses and that these are impaired by anti-*ENO1* Treg cells (64). In an extensive *ex vivo* phenotypic characterization of PDA infiltrating-tumor T cells, it has been found a statistically significant increase in the percentage of *ENO1*-specific Treg T cell clones (TCC) generated from T cells that infiltrate cancer tissue compared to TCC generated from T cells that infiltrate the healthy pancreatic tissue (82). These Treg TCC also inhibit the proliferation and cytotoxic activity of *ENO1*-specific effector Th1 and Th17, TCC generated from PDA tissue-infiltrating T cells (82). Th1 and Th17 TCC from PDA tissue-infiltrating T cells have an anti-tumor effector function, and are increased in healthy pancreatic mucosa compared to PDA tissue (82), suggesting that recruitment of anti-tumor Th17 cells to the tumor site is impaired by the immunosuppressive microenvironment, in which Treg cells play an important role (84-86). The presence of *ENO1*-specific T cells in the peripheral blood of many PDA patients, is triggered by *in vitro* re-stimulation with recombinant *ENO1*. Moreover, *ENO1*-specific TCC generated from PDA patient PBMC display a more pronounced Th1 than Treg phenotype, as observed in the tumor (83). These data suggest that although the recruitment of *ENO1*-specific T cells from peripheral blood to the tumor is repressed by many immunosuppressive mechanisms (83), their presence in peripheral blood is relevant to prevent metastasis through the removal of cancer-circulating cells (87, 88). Indeed PDA patients with a higher number of ENO-specific TCC from PBMC show significantly longer survival, underlying the importance of *ENO1*-specific response to improve PDA patient anti-tumor immunity (83).

4. ROLE OF *ENO1* IN CANCER

ENO1 is a multi-functional protein, which mainly acts as an enzyme and a plasminogen receptor, thus playing a critical role in cancer proliferation, metastasis and spreading (16, 89). Its enzymatic function is carried out by the conversion reaction of dehydration of 2-phospho-D-glycerate to phosphoenolpyruvate in the final step of the glycolytic pathway, while in tumors *ENO1* implicated in the maintenance of "*aerobic glycolysis*" (90). At the surface of cancer cells, *ENO1* acts as a plasminogen receptor and, by promoting plasminogen activation into plasmin, a serine-protease involved in extracellular matrix degradation, *ENO1* favors cell invasion and metastasis (89, 91). Due to *ENO1* upregulation in several tumors and the correlation with shorter overall survival in cancer patients (16, 91), several groups have focused on the study of the perturbation of *ENO1* expression in tumors.

4.1. Metabolic functions of *ENO1* in tumors

Reprogramming of metabolism is a well-known mechanism carried out by cancer cells, which demand a high rate of glycolysis to satisfy the increasing requirements for nucleotide, lipid and protein synthesis and to maintain rapid proliferation (92). Although the Tricarboxylic Acid (TCA) cycle and Oxidative Phosphorylation (OXPHOS) would generate more ATP, cancer cells tend to utilize the less efficient method of glycolysis, producing large quantities of pyruvate and lactate, even in the presence of abundant oxygen; this phenomenon is known as "*aerobic glycolysis*" or the "*Warburg effect*" (93). The most relevant data concerning the metabolic function of *ENO1* in tumor cells are related to its role in maintaining the *Warburg effect*; inhibition of *ENO1* has been shown to increase reactive oxygen species (ROS) that are mainly generated through the sorbitol and NADPH oxidase pathways (94). In particular, *ENO1*-silenced cells from breast, lung and pancreatic cancers display increased glucose uptake and consequently leads to an excess of intracellular glucose, which is forced towards alternative pathways, such as the pentose phosphate pathway (PPP) and the polyol pathway (PP), with a consequent decrease in lactate levels (94). Similar results in terms of inhibition of glycolysis are obtained after *ENO1* silencing in endometrial carcinoma cells (EC); the mRNA level of lactate dehydrogenase A (LDHA) and the protein level of cell glycolysis-associated LDHA is decreased in *ENO1*-silenced EC as well as the amount of extruded lactate into the media (95). These metabolic changes increase the oxidative stress induced-autophagy, the fatty acid oxidation and the amino acid catabolism, resulting in less growth and senescent phenotype of cancer cells (94). Taken together, these data confirm that *ENO1* is essential for maintaining tumor metabolism and suggest that therapies targeting the metabolic function of *ENO1* are effective in blocking tumor progression.

4.1.1. Potential approaches for targeting the metabolic function of *ENO1*

The metabolic switch that occurs in cancer cells may provide promising novel targets for cancer therapy. There is growing evidence to support the potential role of many enzymes, transporters or transcription factors as suitable candidate targets for cancer treatment (96, 97). In particular, there are at least four different approaches: i) inhibition of glycolytic pathway enzymes, ii) inhibition of pentose phosphate pathway enzymes, iii) promotion of the OXPHOS process and iv) attenuation of HIF-1 activity (97). In the glucose metabolic pathway there are multiple therapeutic targets, which could be potential targets for anti-cancer strategies and offer promising clinical potential (98-103). Among this, the inhibition of a key glycolytic enzyme, *ENO1* might represent an interesting approach. Chemical enolase inhibitors are sodium fluoride (104), D-tartonate and 3-aminoenolpyruvate 2-phosphate (102, 105), but none of these are appropriate for use in therapeutic protocols. The translational relevance of *ENO1* targeting was

demonstrated by the use of phosphonoacetohydroxamic acid (PhAH), a pan-enolase transition-state analogue inhibitor (106), for the treatment of glioblastoma (107), and recently for pancreatic, breast and lung cancers (94). PhAH inhibits enolase both enzymatic activity and proliferation in cancer cells (94, 107). The small molecule, named "*ENOblock*" (AP-III-a4), which is the first non-substrate analogue that directly binds to *ENO1* and inhibits its activity (108) decreases cancer cell viability under hypoxic conditions. Under normoxic conditions, ENOblock reduces cancer cell invasion/migration *in vitro* and *in vivo* without inducing cytotoxicity and synergizes with microtubule-destabilizing drugs, suggesting that this ENO inhibitor is suitable for biological assays. In tumors such as Non-Hodgkin's Lymphomas (NHLs) and breast cancer, inhibition of *ENO1* decreased tolerance to hypoxia while increasing sensitivity to radiation therapy, thus indicating that *ENO1* may favor chemoresistance (109, 110).

All these *ENO1* inhibitors are very attractive candidate compounds for pharmacokinetic and pharmacodynamic studies to assess their potential as anti-cancer drugs, but still require concerted efforts to develop suitable drugs for use *in vivo* without affecting normal cells.

4.2. Pro-invasive function of *ENO1*

Overexpression of *ENO1* has been correlated with size, disease stage, metastasis and prognosis for many tumors (16, 91). The pro-invasive function of *ENO1* is mainly linked to its role as a plasminogen receptor; *ENO1* facilitates the binding of elevated concentrations of plasminogen which, after conversion into the serine protease plasmin, promotes extracellular matrix (ECM) degradation. At the cell surface, *ENO1* is part of a multi-protein complex including the uPA receptor (uPAR), integrins and cytoskeletal proteins, responsible for adhesion, migration and proliferation (91), while in the cytoplasm, *ENO1* interacts with the cytoskeleton to promote migration of tumor cells by providing ATP (111, 112). The spreading and invasion of cancer cells *in vivo* is strictly related to the high expression of *ENO1*. *ENO1* silencing in tumor cells not only reduces glycolysis but also migration and *in vitro* invasion, as well as tumorigenesis and metastasis *in vivo* (18, 48, 94, 95, 113), mirroring the different functions of *ENO1* in tumor cells. These effects are mediated by inactivation of PI3K/AKT pathway and its downstream signals including glycolysis, cell cycle progression, and epithelial-mesenchymal transition (EMT)-associated genes in non-small cell lung cancer (113), glioma (18, 113) and endometrial cancer (95).

4.2.1. Potential approaches for targeting the pro-invasive function of *ENO1*

The prognostic value of high *ENO1* expression and its correlation with worse survival has been confirmed in several tumors (18, 19, 25, 33, 34, 36, 37, 42, 57) (Table 1). In parallel, the protective role of anti-*ENO1* antibodies in cancer patients has been also highlighted, suggesting that their use in cancer therapy could represent a good strategy.

In pancreatic cancer patients, the presence of anti-phosphorylated-*ENO1* antibodies correlates with a longer response to therapy as well as overall survival (57). By contrast, in the late-stage of disease in lung and breast cancer patients, there is a marked decrease in basal levels of anti-*ENO1* autoantibodies as a common event, which correlates with a better prognosis and longer survival (63). This is due to physical absorption and neutralization of anti-*ENO1* Ab to surface-expressed and secreted *ENO1*, respectively (64). In fact, *in vivo* adoptive transfer of anti-*ENO1* specific antibodies to mice results in accumulation of antibodies in subcutaneous tumors that expressed high levels of *ENO1*, and a consequent reduction of free circulating anti-*ENO1* antibodies. In addition, patients who underwent surgery display an increase of anti-*ENO1* Ab, a lower hazard ratio, and better progression-free survival (64). *In vivo* adoptive transfer of anti-*ENO1* antibody in mice previously injected with tumor cells results in a strong inhibition of tumor metastasis in lungs and bone (43). A monoclonal antibody against *ENO1* blocks the interaction between plasminogen and *ENO1*, which leads to the inhibition of *in vitro* and *in vivo* migration and invasion (48). However, targeting of the surface *ENO1* by the monoclonal antibody (mAb) did not affect *in vitro* cell proliferation (48). Of note, the *in vivo* passive immunotherapy using a single administration of Adeno-Associated Virus (AAV)-expressing cDNA coding for anti-*ENO1* mAb greatly reduces tumor spreading in the lungs of immunosuppressed mice injected with PDA cells, to a much greater extent than soluble IgG injected twice a week (48).

To define surface molecules targetable by peptides to develop nanocarriers for specific delivery of chemotherapy into tumor cells, *ENO1* has been identified as a specific target of a 12-mer peptide (114). An *in vitro* panning of a phage-displayed peptide library against the colorectal cancer cell line HCT116 identified 36 phages displaying peptides capable of stronger binding compared to the control phage. Of these, three phage clones were subsequently validated *in vivo* for the increased ability to accumulate at the tumor mass compared to normal organs such as brain, lungs and heart. These three phage clones are highly specific for surface binding of different tumor cell lines as well as HCT116, such as A498 (renal cell carcinoma), B16-F10 (melanoma), H640 (lung carcinoma), HTB-10 (neuroepithelioma), MDA-MB-231 (breast carcinoma), Mia-Pa-Ca2 (pancreatic adenocarcinoma), PC-3 (prostatic adenocarcinoma) and SKOV3 (ovarian carcinoma). Two phages were chosen for *in vitro* and *in vivo* pre-clinical studies (114). Synthetic peptide conjugates were incorporated into liposomes filled with chemotherapeutic drugs and assessed for their *in vitro* and *in vivo* cytotoxic ability against colorectal cancer cell line. One peptide in particular was very efficient in delivering drugs into tumor cells and decreasing tumor growth compared to liposomes filled with drugs that were not covered with peptide (114). The LC-MS/MS analysis of the sequences bound by this peptide revealed *ENO1* as its target (114). Overall, these data indicate that targeting of *ENO1* represents a potential strategy for gene-based therapy, immunotherapy and chemical cancer treatment.

5. ANTI-*ENO1* VACCINE AND MECHANISMS OF PROTECTION

Over the last decade, many scientists have invested great efforts in developing approaches for eliciting anti-tumor responses by priming a novel or boosting an existent immune response against tumor cells. These have included tumor-targeting mAbs, oncolytic viruses, dendritic cell (DC)-based therapies, cytokines, immunomodulatory mAbs, pattern recognition receptor (PRR) agonists, peptides, and mRNA- or DNA-based vaccines (5, 115-122). The huge amount of pre-clinical and clinical results led to the approval of their use by the US Food and Drug Administration (FDA) agency and the European Medicines Agency as immunotherapy in cancer patients. The great clinical success of immunotherapy has earned the title "Breakthrough of the Year" in the prestigious Science journal (123).

DNA-based vaccines may represents a suitable and efficient option for immunotherapy. They display several advantages in that they are stable, do not contain viral proteins that could down-regulate the immune system or elicit neutralizing antibodies, and are safe, as several studies have shown that mutations arising from a putative integration event are extremely rare (124). On the other hand, DNA vaccination usually fails to mount a strong immune response and requires additional adjuvant strategies. However, DNA fusion gene vaccine offers the opportunity to include different genes encoding a range of immunostimulatory molecules or short hairpin RNA to switch off suppressive molecules, either into the vaccine vector or by a separate vector (121). A necked plasmid, pVax vector, approved by the FDA for clinical use, was used to express full length human *ENO1* and to vaccinate mice that had been genetically engineered to develop autochthonous pancreatic adenocarcinoma (called KC and KPC) (81). The identity of human and mouse *ENO1* is up to 95% while the homology is up to 99%, and some CD8-specific epitopes that are shared between human HLA-A02 and mouse H-2b molecule tasks were also found (NetMHC 3.0.). In this setting, KC and KPC mice were vaccinated when Pancreatic Intraepithelial Neoplasia (PanINs) lesions were already present and received a total of three and four rounds of immunization every 3 and 2 weeks, respectively. The *ENO1* vaccine induces a specific integrated humoral and cellular response that efficiently prolonged mouse survival from 48 to 68 weeks of age for KC mice, and from 29 to 35 weeks of age for KPC (81).

There are several protective immunological mechanisms induced by the *ENO1* vaccine, namely high levels of anti-*ENO1* IgG, activation of specific Th1 and Th17 cells, as well a large recruitment of CD3 cells into the tumor, and an important decrease of circulating myeloid-derived suppressor cells (MDSC) and both circulating and intra-tumoral Treg cells (125). Notably, the *ENO1* vaccine-induced IgG are able to mediate the complement-dependent cytotoxicity of PDA cells, and it was assumed that the cytokines released by activated Th1/Th17 cells promoted the isotype switching necessary to activate the complement. The crucial role of anti-*ENO1* antibodies was confirmed by the observation that *ENO1*-vaccinated mice showed B cells organized in dense aggregates that displayed a distinct structure, the so-called tertiary lymphoid tissue (TLT), which was not found in normal pancreas and only sporadically in PDA of untreated mice or those vaccinated with an empty-vector (126). B cells organized into TLT, namely CD20-TLT, are shown to correlate with a better prognosis and with a greater infiltration of CD8^{+} T cells in a cohort of 104 PDA patients. To assess the role of tumor infiltrating-CD20^{+} B cells (CD20-TIL) compared to CD20-TLT, mice orthotopically injected with

syngeneic PDA cells were depleted of B cells by a single injection of an anti-CD20 Ab. No TLT is observed in this implantable tumor model, probably because of the absence of a chronic inflammatory response, but CD20-TIL are dramatically reduced in those receiving the Ab. The anti-CD20 treatment induces a significant increase in genes related to T and NK cell recruitment as well as genes involved in lymphoid tissue structure development and CD8+ T cell differentiation and maintenance, suggesting a dual role of B cells in PDA progression (126). *ENO1*-vaccinated mice not only show more TLT than control mice but also a higher number of tumor-infiltrating CD3 cells but not Treg cells (81).

Another effect elicited by the *ENO1*-vaccine is the reduction of MDSC. Due to the expression of *ENO1* in activated monocytes, its presence on the surface of both human and mouse MDSC has been assessed. In PDA patients, myeloid cells express higher levels of *ENO1* and this was further increased by LPS stimulation, as observed on MDSC purified from spleens of tumor-bearing mice (127). An anti-*ENO1* mAb is able to limit MDSC adhesion and migration on and through a monolayer of pancreatic endothelial cells, respectively. In addition, the anti-*ENO1* Ab reduces the *in vivo* migration of MDSC from the footpad to the draining lymph node. Antibodies induced by the *ENO1*-vaccine also limit the infiltration of MDSC into the tumor area of an orthotopic transplantable model of PDA (127). However, the *in vitro* targeting of surface *ENO1* on MDSC does not affect their suppressive function in terms of T cell proliferation, although T cells co-cultured with *ENO1*-targeted MDSC secrete much more IFN-γ and IL17 and less IL10 and TGF-b compared to those co-cultured with MDSC treated with a control Ab (127). *ENO1*-targeting does not affect co-stimulatory molecule expression on MDSC, with the exception of CD80 expression, which is up-regulated, but it decreases arginase activity compared to that of control MDSC (127). Overall, these results demonstrate that the *ENO1*-DNA vaccine elicits an integrated humoral and cellular response to counteract tumor growth, which not only affects the tumor cells themselves but also stromal and reactive cells. Unfortunately, these effects don't last beyond 6 months after immunization, when the mice died; therefore the aim is to combine the vaccination with other strategies to enhance the specific anti-*ENO1* integrated response. The *ENO1*-DNA vaccine, however, is very promising as it efficiently decreased the tumor size in KC mice, which were therapeutically vaccinated when adenocarcinomas were well established, at 8 months of age (81).

6. CONCLUSIONS

The evidence obtained so far demonstrate the role of *ENO1* in tumor progression and the concept that *ENO1* vaccination effectively induces an integrated immune response, which is able to significantly enhance the survival of genetically-engineered mice (GEM) that spontaneously develop PDA. In addition, blocking *ENO1* by a specific monoclonal antibody, or its functional silencing by chemical inhibitors could complement and integrate the mechanisms that are employed, to target *ENO1* and counteract cancer progression and spreading. Metabolic inhibition of *ENO1* blocks tumor cell proliferation and induces changes that can be perturbed to definitively kill cells (Figure 1). Surface blockade of *ENO1* by mAb, instead, may efficiently inhibit PDA spreading and accumulation of myeloid cells in the primary tumor, which can suppress the anti-tumor response (Figure 1). The approach of targeting metabolic and immunological functions in tumors could be further strengthened by combining them with pharmacological inhibitors of immune suppressor cells. For example, recent data have shown that targeting of phosphoinositide-3-kinase (PI3K) γ and δ isoforms is an effective way to unleash the suppressive activity of tumor-associated macrophages (TAM) and regulatory T cells, respectively, thus reinforcing the anticancer immune response (128-130). Inhibitory targeting of PI3Kγ, in fact, stimulates anti-tumor immune responses, leading to improved survival and responsiveness to standard-of-care chemotherapy in animal models of PDA. PI3Kγ selectively drives immunosuppressive transcriptional programming in macrophages, which inhibits adaptive immune responses and promotes tumor cell invasion and desmoplasia in PDA. Inhibition of PI3Kγ in PDA-bearing mice reprograms TAM to stimulate CD8+ T cell-mediated tumor suppression and to inhibit tumor cell invasion, metastasis and desmoplasia (130). These results suggest that the combination of the *ENO1*-DNA vaccine and the PI3Kγ inhibitor enhances the anti-tumor response. Studies are currently ongoing to verify the hypothesis that the targeting of myeloid suppressive cells, via genetic or pharmacological PI3Kγ inhibition synergizes with *ENO1*-DNA vaccination by inducing the sustained immune response that can effectively counteract PDA. Of note, all these approaches can be easily translated into clinical practice as most inhibitors, as well as the AVV vectors, are already used in the clinic, and the pVAX used for the DNA vaccination has been approved by the FDA.

7. ACKNOWLEDGMENTS

8. REFERENCES

1. D. Pardoll: Cancer and the Immune System: Basic Concepts and Targets for Intervention. *Semin Oncol*, 42(4), 523-38 (2015)

2. L. Gross: Intradermal immunization of C3H mice against a sarcoma that originated in an animal of the same line. *Cancer Res*, 3, 326–333 (1943)

3. T. Boon and A. Van Pel: Teratocarcinoma cell variants rejected by syngeneic mice: protection of mice immunized with these variants against other variants and against the original malignant cell line. *Proc Natl Acad Sci U S A*, 75(3), 1519-23 (1978)

4. A. Van Pel and T. Boon: Protection against a nonimmunogenic mouse leukemia by an immunogenic variant obtained by mutagenesis. *Proc Natl Acad Sci U S A*, 79(15), 4718-22 (1982)

5. P. G. Coulie, B. J. Van den Eynde, P. van der Bruggen and T. Boon: Tumour antigens recognized by T lymphocytes: at the core of cancer immunotherapy. *Nat Rev Cancer*, 14(2), 135-46 (2014)

6. P. O. Livingston, H. Shiku, M. A. Bean, C. M. Pinsky, H. F. Oettgen and L. J. Old: Cell-mediated cytotoxicity for cultured autologous melanoma cells. *Int J Cancer*, 24(1), 34-44 (1979)

7. A. Knuth, B. Danowski, H. F. Oettgen and L. J. Old: T-cell-mediated cytotoxicity against autologous malignant melanoma: analysis with interleukin 2-dependent T-cell cultures. *Proc Natl Acad Sci U S A*, 81(11), 3511-5 (1984)

8. S. A. Rosenberg, B. S. Packard, P. M. Aebersold, D. Solomon, S. L. Topalian, S. T. Toy, P. Simon, M. T. Lotze, J. C. Yang, C. A. Seipp and *et al.*: Use of tumor-infiltrating lymphocytes and interleukin-2 in the immunotherapy of patients with metastatic melanoma. A preliminary report. *N Engl J Med*, 319(25), 1676-80 (1988)

9. M. Herin, C. Lemoine, P. Weynants, F. Vessiere, A. Van Pel, A. Knuth, R. Devos and T. Boon: Production of stable cytolytic T-cell clones directed against autologous human melanoma. *Int J Cancer*, 39(3), 390-6 (1987)

10. U. Sahin, O. Tureci, H. Schmitt, B. Cochlovius, T. Johannes, R. Schmits, F. Stenner, G. Luo, I. Schobert and M. Pfreundschuh: Human neoplasms elicit multiple specific immune responses in the autologous host. *Proc Natl Acad Sci U S A*, 92(25), 11810-3 (1995)

11. U. Sahin, O. Tureci and M. Pfreundschuh: Serological identification of human tumor antigens. *Curr Opin Immunol*, 9(5), 709-16 (1997)

12. L. J. Old and Y. T. Chen: New paths in human cancer serology. *J Exp Med*, 187(8), 1163-7 (1998)

13. F. Le Naour: Contribution of proteomics to tumor immunology. *Proteomics*, 1(10), 1295-302 (2001)

14. O. Tureci, U. Sahin and M. Pfreundschuh: Serological analysis of human tumor antigens: molecular definition and implications. *Mol Med Today*, 3(8), 342-9 (1997)

15. B. Tomaino, P. Cappello, M. Capello, C. Fredolini, A. Ponzetto, A. Novarino, L. Ciuffreda, O. Bertetto, C. De Angelis, E. Gaia, P. Salacone, M. Milella, P. Nistico, M. Alessio, R. Chiarle, M. G. Giuffrida, M. Giovarelli and F. Novelli: Autoantibody signature in human ductal pancreatic adenocarcinoma. *J Proteome Res*, 6(10), 4025-31 (2007)

16. M. Capello, S. Ferri-Borgogno, P. Cappello and F. Novelli: alpha-Enolase: a promising therapeutic and diagnostic tumor target. *FEBS J*, 278(7), 1064-74 (2011)

17. B. Altenberg and K. O. Greulich: Genes of glycolysis are ubiquitously overexpressed in 24 cancer classes. *Genomics*, 84(6), 1014-20 (2004)

18. Y. Song, Q. Luo, H. Long, Z. Hu, T. Que, X. Zhang, Z. Li, G. Wang, L. Yi, Z. Liu, W. Fang and S. Qi: Alpha-enolase as a potential cancer prognostic marker promotes cell growth, migration, and invasion in glioma. *Mol Cancer*, 13, 65 (2014)

19. S. H. Tu, C. C. Chang, C. S. Chen, K. W. Tam, Y. J. Wang, C. H. Lee, H. W. Lin, T. C. Cheng, C. S. Huang, J. S. Chu, N. Y. Shih, L. C. Chen, S. J. Leu, Y. S. Ho and C. H. Wu: Increased expression of enolase alpha in human breast cancer confers tamoxifen resistance in human breast cancer cells. *Breast Cancer Res Treat*, 121(3), 539-53 (2010)

20. R. I. Somiari, A. Sullivan, S. Russell, S. Somiari, H. Hu, R. Jordan, A. George, R. Katenhusen, A. Buchowiecka, C. Arciero, H. Brzeski, J. Hooke and C. Shriver: High-throughput proteomic analysis of human infiltrating ductal carcinoma of the breast. *Proteomics*, 3(10), 1863-73 (2003)

21. M. Kabbage, K. Chahed, B. Hamrita, C. L. Guillier, M. Trimeche, S. Remadi, J. Hoebeke and L. Chouchane: Protein alterations in infiltrating ductal carcinomas of the breast as detected by nonequilibrium pH gradient electrophoresis and mass spectrometry. *J Biomed Biotechnol*, 2008, 564127 (2008)

22. L. Malorni, G. Cacace, M. Cuccurullo, G. Pocsfalvi, A. Chambery, A. Farina, A. Di Maro, A. Parente and A. Malorni: Proteomic analysis of MCF-7 breast cancer cell line exposed to mitogenic concentration of 17beta-estradiol. *Proteomics*, 6(22), 5973-82 (2006)

23. A. Hennipman, B. A. van Oirschot, J. Smits, G. Rijksen and G. E. Staal: Glycolytic enzyme activities in breast cancer metastases. *Tumour Biol*, 9(5), 241-8 (1988)

24. A. Hennipman, J. Smits, B. van Oirschot, J. C. van Houwelingen, G. Rijksen, J. P. Neyt, J. A. Van Unnik and G. E. Staal: Glycolytic enzymes in breast cancer, benign breast disease and normal breast tissue. *Tumour Biol*, 8(5), 251-63 (1987)

25. H. Zhou, C. B. Chen, J. Lan, C. Liu, X. G. Liu, L. Jiang, F. Wei, Q. J. Ma, G. T. Dang and Z. J. Liu: Differential proteomic profiling of chordomas and analysis of prognostic factors. *J Surg Oncol*, 102(7), 720-7 (2010)

26. S. M. Bae, H. J. Min, G. H. Ding, S. Y. Kwak, Y. L. Cho, K. H. Nam, C. H. Park, Y. W. Kim, C. K. Kim, B. D. Han, Y. J. Lee, D. K. Kim and W. S. Ahn: Protein expression profile using two-dimensional gel analysis in squamous cervical cancer patients. *Cancer Res Treat*, 38(2), 99-107 (2006)

27. S. M. Bae, C. H. Lee, Y. L. Cho, K. H. Nam, Y. W. Kim, C. K. Kim, B. D. Han, Y. J. Lee, H. J. Chun and W. S. Ahn: Two-dimensional gel analysis of protein expression profile in squamous cervical cancer patients. *Gynecol Oncol*, 99(1), 26-35 (2005)

28. M. Katayama, H. Nakano, A. Ishiuchi, W. Wu, R. Oshima, J. Sakurai, H. Nishikawa, S. Yamaguchi and T. Otsubo: Protein pattern difference in the colon cancer cell lines examined by two-dimensional differential in-gel electrophoresis and mass spectrometry. *Surg Today*, 36(12), 1085-93 (2006)

29. C. S. Wong, V. W. Wong, C. M. Chan, B. B. Ma, E. P. Hui, M. C. Wong, M. Y. Lam, T. C. Au, W. H. Chan, W. Cheuk and A. T. Chan: Identification of 5-fluorouracil response proteins in colorectal carcinoma cell line SW480 by two-dimensional electrophoresis and MALDI-TOF mass spectrometry. *Oncol Rep*, 20(1), 89-98 (2008)

30. Y. Qi, J. F. Chiu, L. Wang, D. L. Kwong and Q. Y. He: Comparative proteomic analysis of esophageal squamous cell carcinoma. *Proteomics*, 5(11), 2960-71 (2005)

31. J. Zhao, A. C. Chang, C. Li, K. A. Shedden, D. G. Thomas, D. E. Misek, A. P. Manoharan, T. J. Giordano, D. G. Beer and D. M. Lubman: Comparative proteomics analysis of Barrett metaplasia and esophageal adenocarcinoma using two-dimensional liquid mass mapping. *Mol Cell Proteomics*, 6(6), 987-99 (2007)

32. R. B. Govekar, A. K. D'Cruz, K. Alok Pathak, J. Agarwal, K. A. Dinshaw, R. F. Chinoy, N. Gadewal, S. Kannan, R. Sirdeshmukh, C. S. Sundaram, S. A. Malgundkar, S. V. Kane and S. M. Zingde: Proteomic profiling of cancer of the gingivo-buccal complex: Identification of new differentially expressed markers. *Proteomics Clin Appl*, 3(12), 1451-62 (2009)

33. S. T. Tsai, I. H. Chien, W. H. Shen, Y. Z. Kuo, Y. T. Jin, T. Y. Wong, J. R. Hsiao, H. P. Wang, N. Y. Shih and L. W. Wu: *ENO1*, a potential prognostic head and neck cancer marker, promotes transformation partly via chemokine CCL20 induction. *Eur J Cancer*, 46(9), 1712-23 (2010)

34. N. M. White-Al Habeeb, A. Di Meo, A. Scorilas, F. Rotondo, O. Masui, A. Seivwright, M. Gabril, A. H. Girgis, M. A. Jewett and G. M. Yousef: Alpha-enolase is a potential prognostic marker in clear cell renal cell carcinoma. *Clin Exp Metastasis*, 32(6), 531-41 (2015)

35. C. Lopez-Pedrera, J. M. Villalba, E. Siendones, N. Barbarroja, C. Gomez-Diaz, A. Rodriguez-Ariza, P. Buendia, A. Torres and F. Velasco: Proteomic analysis of acute myeloid leukemia: Identification of potential early biomarkers and therapeutic targets. *Proteomics*, 6 Suppl 1, S293-9 (2006)

36. T. Hamaguchi, N. Iizuka, R. Tsunedomi, Y. Hamamoto, T. Miyamoto, M. Iida, Y. Tokuhisa, K. Sakamoto, M. Takashima, T. Tamesa and M. Oka: Glycolysis module activated by hypoxia-inducible factor 1alpha is related to the aggressive phenotype of hepatocellular carcinoma. *Int J Oncol*, 33(4), 725-31 (2008)

37. M. Takashima, Y. Kuramitsu, Y. Yokoyama, N. Iizuka, M. Fujimoto, T. Nishisaka, K. Okita, M. Oka and K. Nakamura: Overexpression of alpha enolase in hepatitis C virus-related hepatocellular carcinoma: association with tumor progression as determined by proteomic analysis. *Proteomics*, 5(6), 1686-92 (2005)

38. L. S. Li, H. Kim, H. Rhee, S. H. Kim, D. H. Shin, K. Y. Chung, K. S. Park, Y. K. Paik and J. Chang: Proteomic analysis distinguishes basaloid carcinoma as a distinct subtype of nonsmall cell lung carcinoma. *Proteomics*, 4(11), 3394-400 (2004)

39. C. Li, Z. Xiao, Z. Chen, X. Zhang, J. Li, X. Wu, X. Li, H. Yi, M. Li, G. Zhu and S. Liang: Proteome analysis of human lung squamous carcinoma. *Proteomics*, 6(2), 547-58 (2006)

40. L. J. Huang, S. X. Chen, W. J. Luo, H. H. Jiang, P. F. Zhang and H. Yi: Proteomic analysis of secreted proteins of non-small cell lung cancer. *Ai Zheng*, 25(11), 1361-7 (2006)

41. A. Rubporn, C. Srisomsap, P. Subhasitanont, D. Chokchaichamnankit, K. Chiablaem, J. Svasti and P. Sangvanich: Comparative proteomic analysis of lung cancer cell line and lung fibroblast cell line. *Cancer Genomics Proteomics*, 6(4), 229-37 (2009)

42. G. C. Chang, K. J. Liu, C. L. Hsieh, T. S. Hu, S. Charoenfuprasert, H. K. Liu, K. T. Luh, L. H. Hsu, C. W. Wu, C. C. Ting, C. Y. Chen, K. C. Chen, T. Y. Yang, T. Y. Chou, W. H. Wang, J. Whang-Peng and N. Y. Shih: Identification of alpha-enolase as an autoantigen in lung cancer: its overexpression is associated with clinical outcomes. *Clin Cancer Res*, 12(19), 5746-54 (2006)

43. K. C. Hsiao, N. Y. Shih, H. L. Fang, T. S. Huang, C. C. Kuo, P. Y. Chu, Y. M. Hung, S. W. Chou, Y. Y. Yang, G. C. Chang and K. J. Liu: Surface alpha-enolase promotes extracellular matrix degradation and tumor metastasis and represents a new therapeutic target. *PLoS One*, 8(7), e69354 (2013)

44. L. Cao, X. Li, Y. Zhang, F. Peng, H. Yi, Y. Xu and Q. Wang: Proteomic analysis of human ovarian cancer paclitaxel-resistant cell lines. *Zhong Nan Da Xue Xue Bao Yi Xue Ban*, 35(4), 286-94 (2010)

45. J. Shen, M. D. Person, J. Zhu, J. L. Abbruzzese and D. Li: Protein expression profiles in pancreatic adenocarcinoma compared with normal pancreatic tissue and tissue affected by pancreatitis as detected by two-dimensional gel electrophoresis and mass spectrometry. *Cancer Res*, 64(24), 9018-26 (2004)

46. K. Mikuriya, Y. Kuramitsu, S. Ryozawa, M. Fujimoto, S. Mori, M. Oka, K. Hamano, K. Okita, I. Sakaida and K. Nakamura: Expression of glycolytic enzymes is increased in pancreatic cancerous tissues as evidenced by proteomic profiling by two-dimensional electrophoresis and liquid chromatography-mass spectrometry/mass spectrometry. *Int J Oncol*, 30(4), 849-55 (2007)

47. P. Cappello, B. Tomaino, R. Chiarle, P. Ceruti, A. Novarino, C. Castagnoli, P. Migliorini, G. Perconti, A. Giallongo, M. Milella, V. Monsurro, S. Barbi, A. Scarpa, P. Nistico, M. Giovarelli and F. Novelli: An integrated humoral and cellular response is elicited in pancreatic cancer by alpha-enolase, a novel pancreatic ductal adenocarcinoma-associated antigen. *Int J Cancer*, 125(3), 639-48 (2009)

48. M. Principe, P. Ceruti, N. Y. Shih, M. S. Chattaragada, S. Rolla, L. Conti, M. Bestagno, L. Zentilin, S. H. Yang, P. Migliorini, P. Cappello, O. Burrone and F. Novelli: Targeting of surface alpha-enolase inhibits the invasiveness of pancreatic cancer cells. *Oncotarget*, 6(13), 11098-113 (2015)

49. I. Rehman, A. R. Azzouzi, J. W. Catto, S. Allen, S. S. Cross, K. Feeley, M. Meuth and F. C. Hamdy: Proteomic analysis of voided urine after prostatic massage from patients with prostate cancer: a pilot study. *Urology*, 64(6), 1238-43 (2004)

50. A. Suzuki, A. Iizuka, M. Komiyama, M. Takikawa, A. Kume, S. Tai, C. Ohshita, A. Kurusu, Y. Nakamura, A. Yamamoto, N. Yamazaki, S. Yoshikawa, Y. Kiyohara and Y. Akiyama: Identification of melanoma antigens using a Serological Proteome Approach (SERPA). *Cancer Genomics Proteomics*, 7(1), 17-23 (2010)

51. R. Rucksaken, C. Pairojkul, P. Pinlaor, N. Khuntikeo, S. Roytrakul, C. Selmi and S. Pinlaor: Plasma autoantibodies against heat shock protein 70, enolase 1 and ribonuclease/angiogenin inhibitor 1 as potential biomarkers for cholangiocarcinoma. *PLoS One*, 9(7), e103259 (2014)

52. Z. Mojtahedi, A. Safaei, Z. Yousefi and A. Ghaderi: Immunoproteomics of HER2-positive and HER2-negative breast cancer patients with positive lymph nodes. *OMICS*, 15(6), 409-18 (2011)

53. S. Shukla, R. B. Govekar, R. Sirdeshmukh, C. S. Sundaram, A. K. D'Cruz, K. A. Pathak, S. V. Kane and S. M. Zingde: Tumor antigens eliciting autoantibody response in cancer of gingivo-buccal complex. *Proteomics Clin Appl*, 1(12), 1592-604 (2007)

54. S. Shukla, A. Pranay, A. K. D'Cruz, P. Chaturvedi, S. V. Kane and S. M. Zingde: Immunoproteomics reveals that cancer of the tongue and the gingivobuccal complex exhibit differential autoantibody response. *Cancer Biomark*, 5(3), 127-35 (2009)

55. J. W. Cui, W. H. Li, J. Wang, A. L. Li, H. Y. Li, H. X. Wang, K. He, W. Li, L. H. Kang, M. Yu, B. F. Shen, G. J. Wang and X. M. Zhang: Proteomics-based identification of human acute leukemia antigens that induce humoral immune response. *Mol Cell Proteomics*, 4(11), 1718-24 (2005)

56. L. Zou, Y. Wu, L. Pei, D. Zhong, M. Gen, T. Zhao, J. Wu, B. Ni, Z. Mou, J. Han, Y. Chen and Y. Zhi: Identification of leukemia-associated antigens in chronic myeloid leukemia by proteomic analysis. *Leuk Res*, 29(12), 1387-91 (2005)

57. B. Tomaino, P. Cappello, M. Capello, C. Fredolini, I. Sperduti, P. Migliorini, P. Salacone, A. Novarino, A. Giacobino, L. Ciuffreda, M. Alessio, P. Nistico, A. Scarpa, P. Pederzoli, W. Zhou, E. F. Petricoin Iii, L. A. Liotta, M. Giovarelli, M. Milella and F. Novelli: Circulating autoantibodies to phosphorylated alpha-enolase are a hallmark of pancreatic cancer. *J Proteome Res*, 10(1), 105-12 (2011)

58. M. Forgber, U. Trefzer, W. Sterry and P. Walden: Proteome serological determination of tumor-associated antigens in melanoma. *PLoS One*, 4(4), e5199 (2009)

59. P. He, T. Naka, S. Serada, M. Fujimoto, T. Tanaka, S. Hashimoto, Y. Shima, T. Yamadori, H. Suzuki, T. Hirashima, K. Matsui, H. Shiono, M. Okumura, T. Nishida, I. Tachibana, N. Norioka, S. Norioka and I. Kawase: Proteomics-based identification of alpha-enolase as a tumor antigen in non-small lung cancer. *Cancer Sci*, 98(8), 1234-40 (2007)

60. R. Jankowska, D. Witkowska, I. Porebska, M. Kuropatwa, E. Kurowska and W. A. Gorczyca: Serum antibodies to retinal antigens in lung cancer and sarcoidosis. *Pathobiology*, 71(6), 323-8 (2004)

61. K. Ueda: (Proteome analysis of autoantibodies in sera of patients with cancer). *Rinsho Byori*, 53(5), 437-45 (2005)

62. T. Nakanishi, T. Takeuchi, K. Ueda, H. Murao and A. Shimizu: Detection of eight antibodies in cancer patients' sera against proteins derived from the adenocarcinoma A549 cell line using proteomics-based analysis. *J Chromatogr B Analyt Technol Biomed Life Sci*, 838(1), 15-20 (2006)

63. N. Y. Shih, H. L. Lai, G. C. Chang, H. C. Lin, Y. C. Wu, J. M. Liu, K. J. Liu and S. W. Tseng: Anti-alpha-enolase autoantibodies are down-regulated in advanced cancer patients. *Jpn J Clin Oncol*, 40(7), 663-9 (2010)

64. K. C. Hsiao, N. Y. Shih, P. Y. Chu, Y. M. Hung, J. Y. Liao, S. W. Chou, Y. Y. Yang, G. C. Chang and K. J. Liu: Anti-alpha-enolase is a prognostic marker in postoperative lung cancer patients. *Oncotarget*, 6(33), 35073-86 (2015)

65. P. L. Triozzi, W. Aldrich, J. W. Crabb and A. D. Singh: Spontaneous cellular and humoral tumor antigen responses in patients with uveal melanoma. *Melanoma Res*, 25(6), 510-8 (2015)

66. M. Bruschi, R. A. Sinico, G. Moroni, F. Pratesi, P. Migliorini, M. Galetti, C. Murtas, A. Tincani, M. Madaio, A. Radice, F. Franceschini, B. Trezzi, L. Bianchi, A. Giallongo, R. Gatti, R. Tardanico, A. Scaloni, C. D'Ambrosio, M. L. Carnevali, P. Messa, P. Ravani, G. Barbano, B. Bianco, A. Bonanni, F. Scolari, A. Martini, G. Candiano, L. Allegri and G. M. Ghiggeri: Glomerular autoimmune multicomponents of human lupus nephritis *in vivo*: alpha-enolase and annexin AI. *J Am Soc Nephrol*, 25(11), 2483-98 (2014)

67. M. Bruschi, M. Galetti, R. A. Sinico, G. Moroni, A. Bonanni, A. Radice, A. Tincani, F. Pratesi, P. Migliorini, C. Murtas, F. Franceschini, B. Trezzi, F. Brunini, R. Gatti, R. Tardanico, G. Barbano, G. Piaggio, P. Messa, P. Ravani, F. Scolari, G. Candiano, A. Martini, L. Allegri and G. M. Ghiggeri: Glomerular Autoimmune Multicomponents of Human Lupus Nephritis *In vivo* (2): Planted Antigens. *J Am Soc Nephrol*, 26(8), 1905-24 (2015)

68. A. Magrys, T. Anekonda, G. Ren and G. Adamus: The role of anti-alpha-enolase autoantibodies in pathogenicity of autoimmune-mediated retinopathy. *J Clin Immunol*, 27(2), 181-92 (2007)

69. G. Adamus, N. Aptsiauri, J. Guy, J. Heckenlively, J. Flannery and P. A. Hargrave: The occurrence of serum autoantibodies against enolase in cancer-associated retinopathy. *Clin Immunol Immunopathol*, 78(2), 120-9 (1996)

70. C. Dot, J. Guigay and G. Adamus: Anti-alpha-enolase antibodies in cancer-associated retinopathy with small cell carcinoma of the lung. *Am J Ophthalmol*, 139(4), 746-7 (2005)

71. M. Ejma, M. Misiuk-Hojlo, W. A. Gorczyca, R. Podemski, S. Szymaniec, M. Kuropatwa, J. Rogozinska-Szczepka and W. Bartnik: Antibodies to 46-kDa retinal antigen in a patient with breast carcinoma and cancer-associated retinopathy. *Breast Cancer Res Treat*, 110(2), 269-71 (2008)

72. G. Adamus: Latest updates on antiretinal autoantibodies associated with vision loss and breast cancer. *Invest Ophthalmol Vis Sci*, 56(3), 1680-8 (2015)

73. A. Kinloch, V. Tatzer, R. Wait, D. Peston, K. Lundberg, P. Donatien, D. Moyes, P. C. Taylor and P. J. Venables: Identification of citrullinated alpha-enolase as a candidate autoantigen in rheumatoid arthritis. *Arthritis Res Ther*, 7(6), R1421-9 (2005)

74. K. Lundberg, A. Kinloch, B. A. Fisher, N. Wegner, R. Wait, P. Charles, T. R. Mikuls and P. J. Venables: Antibodies to citrullinated alpha-enolase peptide 1 are specific for rheumatoid arthritis and cross-react with bacterial enolase. *Arthritis Rheum*, 58(10), 3009-19 (2008)

75. R. Bei, L. Masuelli, C. Palumbo, M. Modesti and A. Modesti: A common repertoire of autoantibodies is shared by cancer and autoimmune disease patients: Inflammation in their induction and impact on tumor growth. *Cancer Lett*, 281(1), 8-23 (2009)

76. W. Zhou, M. Capello, C. Fredolini, L. Piemonti, L. A. Liotta, F. Novelli and E. F. Petricoin: Mass spectrometry analysis of the post-translational modifications of alpha-enolase from pancreatic ductal adenocarcinoma cells. *J Proteome Res*, 9(6), 2929-36 (2010)

77. F. Mohammed, M. Cobbold, A. L. Zarling, M. Salim, G. A. Barrett-Wilt, J. Shabanowitz, D. F. Hunt, V. H. Engelhard and B. E. Willcox: Phosphorylation-dependent interaction between antigenic peptides and MHC class I: a molecular basis for the presentation of transformed self. *Nat Immunol*, 9(11), 1236-43 (2008)

78. M. Capello, C. Caorsi, P. J. Bogantes Hernandez, E. Dametto, F. E. Bertinetto, P. Magistroni, S. Rendine, A. Amoroso and F. Novelli: Phosphorylated alpha-enolase induces autoantibodies in HLA-DR8 pancreatic cancer patients and triggers HLA-DR8 restricted T-cell activation. *Immunol Lett*, 167(1), 11-6 (2015)

79. N. Sato, Y. Nabeta, H. Kondo, H. Sahara, Y. Hirohashi, K. Kashiwagi, T. Kanaseki, Y. Sato, S. Rong, I. Hirai, K. Kamiguchi, Y. Tamura, A. Matsuura, S. Takahashi, T. Torigoe and H. Ikeda: Human CD8 and CD4 T cell epitopes of epithelial cancer antigens. *Cancer Chemother Pharmacol*, 46 Suppl, S86-90 (2000)

80. H. Kondo, H. Sahara, A. Miyazaki, Y. Nabeta, Y. Hirohashi, T. Kanaseki, A. Yamaguchi, N. Yamada, K. Hirayama, M. Suzuki, J. Hamuro, T. Torigoe, N. Takahashi, G. I. Kohama, H. Ikeda and N. Sato: Natural antigenic peptides from squamous cell carcinoma recognized by autologous HLA-DR8-restricted CD4+ T cells. *Jpn J Cancer Res*, 93(8), 917-24 (2002)

81. P. Cappello, S. Rolla, R. Chiarle, M. Principe, F. Cavallo, G. Perconti, S. Feo, M. Giovarelli and F. Novelli: Vaccination with *ENO1* DNA prolongs survival of genetically engineered mice with pancreatic cancer. *Gastroenterology*, 144(5), 1098-106 (2013)

82. A. Amedei, E. Niccolai, M. Benagiano, C. Della Bella, F. Cianchi, P. Bechi, A. Taddei, L. Bencini, M. Farsi, P. Cappello, D. Prisco, F. Novelli and M. M. D'Elios: *Ex vivo* analysis of pancreatic cancer-infiltrating T lymphocytes reveals that ENO-specific Tregs accumulate in tumor tissue and inhibit Th1/Th17 effector cell functions. *Cancer Immunol Immunother*, 62(7), 1249-60 (2013)

83. E. Niccolai, P. Cappello, A. Taddei, F. Ricci, M. M. D'Elios, M. Benagiano, P. Bechi, L. Bencini, M. N. Ringressi, A. Coratti, F. Cianchi, L. Bonello, P. F. Di Celle, D. Prisco, F. Novelli and A. Amedei: Peripheral *ENO1*-specific T cells mirror the intratumoral immune response and their presence is a potential prognostic factor for pancreatic adenocarcinoma. *Int J Oncol* (2016)

84. E. Bettelli, Y. Carrier, W. Gao, T. Korn, T. B. Strom, M. Oukka, H. L. Weiner and V. K. Kuchroo: Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature*, 441(7090), 235-8 (2006)

85. D. C. Linehan and P. S. Goedegebuure: CD25+ CD4+ regulatory T-cells in cancer. *Immunol Res*, 32(1-3), 155-68 (2005)

86. U. K. Liyanage, T. T. Moore, H. G. Joo, Y. Tanaka, V. Herrmann, G. Doherty, J. A. Drebin, S. M. Strasberg, T. J. Eberlein, P. S. Goedegebuure and D. C. Linehan: Prevalence of regulatory T cells is increased in peripheral blood and tumor microenvironment of patients with pancreas or breast adenocarcinoma. *J Immunol*, 169(5), 2756-61 (2002)

87. K. Tjensvoll, O. Nordgard and R. Smaaland: Circulating tumor cells in pancreatic cancer patients: methods of detection and clinical implications. *Int J Cancer*, 134(1), 1-8 (2014)

88. F. C. Bidard, F. Huguet, C. Louvet, L. Mineur, O. Bouche, B. Chibaudel, P. Artru, F. Desseigne, J. B. Bachet, C. Mathiot, J. Y. Pierga and P. Hammel: Circulating tumor cells in locally advanced pancreatic adenocarcinoma: the ancillary CirCe 07 study to the LAP 07 trial. *Ann Oncol*, 24(8), 2057-61 (2013)

89. V. Pancholi: Multifunctional alpha-enolase: its role in diseases. *Cell Mol Life Sci*, 58(7), 902-20 (2001)

90. O. Warburg, F. Wind and E. Neglers: The metabolism of tumors. *London: Constable & Co* (1930)

91. P. Ceruti, M. Principe, M. Capello, P. Cappello and F. Novelli: Three are better than one: plasminogen receptors as cancer theranostic targets. *Exp Hematol Oncol*, 2(1), 12 (2013)

92. L. M. Phan, S. C. Yeung and M. H. Lee: Cancer metabolic reprogramming: importance, main features, and potentials for precise targeted anti-cancer therapies. *Cancer Biol Med*, 11(1), 1-19 (2014)

93. O. Warburg, F. Wind and E. Negelein: The Metabolism of Tumors in the Body. *J Gen Physiol*, 8(6), 519-30 (1927)

94. M. Capello, S. Ferri-Borgogno, C. Riganti, M. S. Chattaragada, M. Principe, C. Roux, W. Zhou, E. F. Petricoin, P. Cappello and F. Novelli: Targeting the Warburg effect in cancer cells through *ENO1* knockdown rescues oxidative phosphorylation and induces growth arrest. *Oncotarget*, 7(5), 5598-612 (2016)

95. M. Zhao, W. Fang, Y. Wang, S. Guo, L. Shu, L. Wang, Y. Chen, Q. Fu, Y. Liu, S. Hua, Y. Fan, X. Deng, R. Luo, Z. Mei, Q. Jiang and Z. Liu: Enolase-1 is a therapeutic target in endometrial carcinoma. *Oncotarget*, 6(17), 15610-27 (2015)

96. E. Bobrovnikova-Marjon and J. B. Hurov: Targeting metabolic changes in cancer: novel therapeutic approaches. *Annu Rev Med*, 65, 157-70 (2014)

97. M. Vander Heiden: Targeting cancer metabolism: a therapeutic window opens. *Nat Rev Drug Discov*, 10(9), 671-84 (2011)

98. L. Zhao, Y. Mao, Y. Zhao, Y. Cao and X. Chen: Role of multifaceted regulators in cancer glucose metabolism and their clinical significance. *Oncotarget* (2016)

99. Y. Zhao, E. B. Butler and M. Tan: Targeting cellular metabolism to improve cancer therapeutics. *Cell Death Dis*, 4, e532 (2013)

100. Y. Zhao, H. Liu, A. I. Riker, O. Fodstad, S. P. Ledoux, G. L. Wilson and M. Tan: Emerging metabolic targets in cancer therapy. *Front Biosci (Landmark Ed)*, 16, 1844-60 (2011)

101. H. Pelicano, D. S. Martin, R. H. Xu and P. Huang: Glycolysis inhibition for anticancer treatment. *Oncogene*, 25(34), 4633-46 (2006)

102. A. K. Chan, J. Bruce and A. K. Siriwardena: Glucose metabolic phenotype of pancreatic cancer. *World J Gastroenterol*, 22(12), 3471-85 (2016)

103. M. Sanzey, S. A. Abdul Rahim, A. Oudin, A. Dirkse, T. Kaoma, L. Vallar, C. Herold-Mende, R. Bjerkvig, A. Golebiewska and S. P. Niclou: Comprehensive analysis of glycolytic enzymes as therapeutic targets in the treatment of glioblastoma. *PLoS One*, 10(5), e0123544 (2015)

104. R. Scatena, P. Bottoni, A. Pontoglio, L. Mastrototaro and B. Giardina: Glycolytic enzyme inhibitors in cancer treatment. *Expert Opin Investig Drugs*, 17(10), 1533-45 (2008)

105. T. G. Spring and F. Wold: Studies on two high-affinity enolase inhibitors. Chemical characterization. *Biochemistry*, 10(25), 4649-54 (1971)

106. A. S. N. M. V. de, S. M. Gomes Dias, L. V. Mello, M. T. da Silva Giotto, S. Gavalda, C. Blonski, R. C. Garratt and D. J. Rigden: Structural flexibility in Trypanosoma brucei enolase revealed by X-ray crystallography and molecular dynamics. *Febs J*, 274(19), 5077-89 (2007)

107. F. L. Muller, S. Colla, E. Aquilanti, V. E. Manzo, G. Genovese, J. Lee, D. Eisenson, R. Narurkar, P. Deng, L. Nezi, M. A. Lee, B. Hu, J. Hu, E. Sahin, D. Ong, E. Fletcher-Sananikone, D. Ho, L. Kwong, C. Brennan, Y. A. Wang, L. Chin and R. A. DePinho: Passenger deletions generate therapeutic vulnerabilities in cancer. *Nature*, 488(7411), 337-42 (2012)

108. D. W. Jung, W. H. Kim, S. H. Park, J. Lee, J. Kim, D. Su, H. H. Ha, Y. T. Chang and D. R. Williams: A unique small molecule inhibitor of enolase clarifies its role in fundamental biological processes. *ACS Chem Biol*, 8(6), 1271-82 (2013)

109. X. Zhu, X. Miao, Y. Wu, C. Li, Y. Guo, Y. Liu, Y. Chen, X. Lu, Y. Wang and S. He: *ENO1* promotes tumor proliferation and cell adhesion mediated drug resistance (CAM-DR) in Non-Hodgkin's Lymphomas. *Exp Cell Res*, 335(2), 216-23 (2015)

110. J. Gao, R. Zhao, Y. Xue, Z. Niu, K. Cui, F. Yu, B. Zhang and S. Li: Role of enolase-1 in response to hypoxia in breast cancer: exploring the mechanisms of action. *Oncol Rep*, 29(4), 1322-32 (2013)

111. J. L. Walsh, T. J. Keith and H. R. Knull: Glycolytic enzyme interactions with tubulin and microtubules. *Biochim Biophys Acta*, 999(1), 64-70 (1989)

112. K. Liu and N. Y. Shih: The role of Enolase in tissue invasion and metastasis of pathogens and tumor cells. *J Cancer Mol*, 3, 45–48 (2007)

113. Q. F. Fu, Y. Liu, Y. Fan, S. N. Hua, H. Y. Qu, S. W. Dong, R. L. Li, M. Y. Zhao, Y. Zhen, X. L. Yu, Y. Y. Chen, R. C. Luo, R. Li, L. B. Li, X. J. Deng, W. Y. Fang, Z. Liu and X. Song: Alpha-enolase promotes cell glycolysis, growth, migration, and invasion in non-small cell lung cancer through FAK-mediated PI3K/AKT pathway. *J Hematol Oncol*, 8, 22 (2015)

114. C. H. Wu, Y. H. Kuo, R. L. Hong and H. C. Wu: alpha-Enolase-binding peptide enhances drug delivery efficiency and therapeutic efficacy against colorectal cancer. *Sci Transl Med*, 7(290), 290ra91 (2015)

115. I. Melero, S. Hervas-Stubbs, M. Glennie, D. M. Pardoll and L. Chen: Immunostimulatory monoclonal antibodies for cancer therapy. *Nat Rev Cancer*, 7(2), 95-106 (2007)

116. P. E. Hughes, S. Caenepeel and L. C. Wu: Targeted Therapy and Checkpoint Immunotherapy Combinations for the Treatment of Cancer. *Trends Immunol* (2016)

117. M. K. Callahan, M. A. Postow and J. D. Wolchok: Targeting T Cell Co-receptors for Cancer Therapy. *Immunity*, 44(5), 1069-78 (2016)

118. D. N. Khalil, E. L. Smith, R. J. Brentjens and J. D. Wolchok: The future of cancer treatment: immunomodulation, CARs and combination immunotherapy. *Nat Rev Clin Oncol*, 13(6), 394 (2016)

119. L. Vandenberk, J. Belmans, M. Van Woensel, M. Riva and S. W. Van Gool: Exploiting the Immunogenic Potential of Cancer Cells for Improved Dendritic Cell Vaccines. *Front Immunol*, 6, 663 (2015)

120. F. Aranda, E. Vacchelli, A. Eggermont, J. Galon, C. Sautes-Fridman, E. Tartour, L. Zitvogel, G. Kroemer and L. Galluzzi: Trial Watch: Peptide vaccines in cancer therapy. *Oncoimmunology*, 2(12), e26621 (2013)

121. J. Rice, C. H. Ottensmeier and F. K. Stevenson: DNA vaccines: precision tools for activating effective immunity against cancer. *Nat Rev Cancer*, 8(2), 108-20 (2008)

122. L. M. Kranz, M. Diken, H. Haas, S. Kreiter, C. Loquai, K. C. Reuter, M. Meng, D. Fritz, F. Vascotto, H. Hefesha, C. Grunwitz, M. Vormehr, Y. Husemann, A. Selmi, A. N. Kuhn, J. Buck, E. Derhovanessian, R. Rae, S. Attig, J. Diekmann, R. A. Jabulowsky, S. Heesch, J. Hassel, P. Langguth, S. Grabbe, C. Huber, O. Tureci and U. Sahin: Systemic RNA delivery to dendritic cells exploits antiviral defence for cancer immunotherapy. *Nature* (2016)

123. J. Couzin-Frankel: Breakthrough of the year 2013. Cancer immunotherapy. *Science*, 342(6165), 1432-3 (2013)

124. Z. Wang, P. J. Troilo, X. Wang, T. G. Griffiths, S. J. Pacchione, A. B. Barnum, L. B. Harper, C. J. Pauley, Z. Niu, L. Denisova, T. T. Follmer, G. Rizzuto, G. Ciliberto, E. Fattori, N. L. Monica, S. Manam and B. J. Ledwith: Detection of integration of plasmid DNA into host genomic DNA following intramuscular injection and electroporation. *Gene Ther*, 11(8), 711-21 (2004)

125. P. Cappello and F. Novelli: A self antigen reopens the games in pancreatic cancer. *Oncoimmunology*, 2(6), e24384 (2013)

126. G. F. Castino, N. Cortese, G. Capretti, S. Serio, G. Di Caro, R. Mineri, E. Magrini, F. Grizzi, P. Cappello, F. Novelli, P. Spaggiari, M. Roncalli, C. Ridolfi, F. Gavazzi, A. Zerbi, P. Allavena and F. Marchesi: Spatial distribution of B cells predicts prognosis in human pancreatic adenocarcinoma. *Oncoimmunology*, 5(4), e1085147 (2016)

127. P. Cappello, E. Tonoli, R. Curto, D. Giordano, M. Giovarelli and F. Novelli: Anti-α-enolase antibody limits the invasion of myeloid-derived suppressor cells and attenuates their restraining effector T cell response. *Oncoimmunology*, 5(5) (2016)

128. E. Hirsch and F. Novelli: Cancer: natural-born killers unleashed. *Nature*, 510(7505), 342-3 (2014)

129. K. Ali, D. R. Soond, R. Pineiro, T. Hagemann, W. Pearce, E. L. Lim, H. Bouabe, C. L. Scudamore, T. Hancox, H. Maecker, L. Friedman, M. Turner, K. Okkenhaug and B. Vanhaesebroeck: Inactivation of PI(3)K p110delta breaks regulatory T-cell-mediated immune tolerance to cancer. *Nature*, 510(7505), 407-11 (2014)

130. M. M. Kaneda, P. Cappello, A. V. Nguyen, N. Ralainirina, C. R. Hardamon, P. Foubert, M. C. Schmid, P. Sun, E. Mose, M. Bouvet, A. M. Lowy, M. A. Valasek, R. Sasik, F. Novelli, E. Hirsch and J. A. Varner: Macrophage PI3Kgamma drives pancreatic ductal adenocarcinoma progression. *Cancer Discov* (2016)

Abbreviations: Adeno-Associated Virus: AAV; Adenosine Triphosphate: ATP; Alpha-enolase: *ENO1*; Carbohydrate Antigen 19.9.: CA19.9.; Cytotoxic T lymphocytes: CTL; Dendritic cells: DC; Epithelial-Mesenchymal Transition: EMT; Extracellular Matrix: ECM; genetically engineered mice: GEM; Interferon-γ: IFN-γ; Interleukin 10: IL10; Interleukin 17: IL17; Human Leukocyte Antingens: HLA; lactate dehydrogenase A: LDHA; Liquid chromatography tandem-mass spectrometry: LC-MS/MS; Hypoxia Inducible Factor 1: HIF-1; Lipopolysaccharide: LPS; Kras-mutated-genetically engineered mice that spontaneously develop pancreatic cancer: KC; Kras and p53-mutated genetically engineered mice that spontaneously develop pancreatic cancer: KPC; Myeloid Derived Suppressor Cells: MDSC; Major Histocompatibility Complex: MHC; monoclonal antibody: mAb; reduced Nicotinamide Adenine Dinucleotide Phosphate: NADPH; Oxidative Phosphorylation: OXPHOS; Pancreatic Ductal Adenocarcinoma: PDA; Pancreatic Intraepithelial Neoplasia: PanIN; Peripheral Blood Mononuclear Cells: PBMC; Phosphonoacetohydroxamic acid: PhAH; phosphoinositide-3-kinase: PI3K; Radical Oxygen Species: ROS; Serological Analysis of Recombinant cDNA Expression Libraries:SEREX; SERological Proteome Analysis: SERPA; tertiary lymphoid tissue: TLT; T cell clones: TCC; Th1 cells: T helper 1 cells; Th17 cells: T helper 1 cells; Transforming Growth Factor b: TGF-b; T regulatory cells: Treg cells; Tricarboxylic Acid :TCA; Tumor Associated Antigens: TAAs; Tumor Associated Macrophages: TAM, Urokinase-type Plasminogen Activator (uPA) Receptor: uPAR.

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Table 1. *ENO1* expression and the immune response in cancers and any clinical correlation

Cancer	*ENO1* overexpression	Immune response	Clinical correlation	References
Brain	mRNA		DP	(17, 18)
Breast	mRNA, protein	Antibody	DP, DFS, M	(19-24,52, 63, 71)
Cartilage	Protein		DFS	(25)

Cervix	mRNA, protein			(17, 26, 27)
Cholangiocarcinoma		Antibody		(51)
Colon	mRNA, protein			(17, 28, 29)
Eye	mRNA			(17)
Gastric	mRNA, protein			(17, 30, 31)
Head and neck	mRNA, protein	Antibody, T cell	OS, PFS	(32, 33,53, 54, 79, 80)
Kidney	mRNA, protein		OS, DFS	(17, 34)
Leukemia	Protein	Antibody		(35,55, 56)
Liver	mRNA, protein		M	(17, 36, 37)
Lung	mRNA, protein	Antibody	DP, OS, PFS, M	(17, 38-43,59-64, 70)
Muscle	mRNA			(17)
Ovary	mRNA, protein			(17, 44)
Pancreas	mRNA, protein	Antibody, T cell	OS, PFS	(17, 45-48,57, 78, 81)
Prostate	mRNA, protein			(17, 49)
Skin	mRNA	Antibody, T cell		(50,58, 65)
Testis	mRNA			(17)

DP: disease progression, DFS: disease-free survival, M: malignancy, OS: overall survival, PFS: progression-free survival.

Figure 1. Cartoon shows multiple localizations of *ENO1* in tumor cells (yellow square), myeloid cells (green square) and potential effects of *ENO1* targeting on tumor (red and blue squares). Inhibition of cytoplasmic *ENO1* by chemical inhibitors leads to senescence and blocking of cell cycle (yellow square). *ENO1* targeting on cell surface of myeloid cells inhibits their endothelial adhesion, invasion and migration (green square) and modulates their restraining functions. Anti-*ENO1*antibodies induced by *ENO1*-DNA vaccine elicit complement-dependent cytotoxicity of tumor cells and together Th1/Th17 cells significantly delay PDA progression (red square). Lastly, anti-*ENO1* antibodies impair PDA cell invasion and migration and ultimately metastasis (blue square). Target *ENO1* with mAb could have a multiple effects. Surface blockade of *ENO1* by mAb inhibits PDA cells spreading and prevents the migration of myeloid cells to the primary tumor.

Running Title: Targeting *ENO1* in immunotherapy

Figure 1.