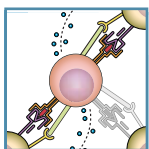


ACQUIRED RESISTANCE TO CLINICAL CANCER THERAPY: A TWIST IN PHYSIOLOGICAL SIGNALING

Andreas Wicki, Mario Mandalà, Daniela Massi, Daniela Taverna, Huifang Tang, Brian A. Hemmings, and Gongda Xue

Department of Biomedicine, University Hospital Basel, Basel, Switzerland; Department of Oncology and Hematology, Papa Giovanni XXIII Hospital, Bergamo, Italy; Department of Surgery and Translational Medicine, University of Florence, Florence, Italy; Department of Molecular Biotechnology and Health Sciences, University of Turin, Torino, Italy; Department of Pharmacology, Zhejiang University School of Medicine, Hangzhou, China; and Department of Mechanisms of Cancer, Friedrich Miescher Institute for Biomedical Research, Basel, Switzerland



Wicki A, Mandalà M, Massi D, Taverna D, Tang H, Hemmings BA, Xue G. Acquired Resistance to Clinical Cancer Therapy: A Twist in Physiological Signaling. *Physiol Rev* 96: 805–829, 2016. Published May 3, 2016; doi:10.1152/physrev.00024.2015.—Although modern therapeutic strategies have brought significant progress to cancer care in the last 30 years, drug resistance to targeted monotherapies has emerged as

a major challenge. Aberrant regulation of multiple physiological signaling pathways indispensable for developmental and metabolic homeostasis, such as hyperactivation of pro-survival signaling axes, loss of suppressive regulations, and impaired functionalities of the immune system, have been extensively investigated aiming to understand the diversity of molecular mechanisms that underlie cancer development and progression. In this review, we intend to discuss the molecular mechanisms of how conventional physiological signal transduction confers to acquired drug resistance in cancer patients. We will particularly focus on protooncogenic receptor kinase inhibition-elicited tumor cell adaptation through two major core downstream signaling cascades, the PI3K/Akt and MAPK pathways. These pathways are crucial for cell growth and differentiation and are frequently hyperactivated during tumorigenesis. In addition, we also emphasize the emerging roles of the deregulated host immune system that may actively promote cancer progression and attenuate immunosurveillance in cancer therapies. Understanding these mechanisms may help to develop more effective therapeutic strategies that are able to keep the tumor in check and even possibly turn cancer into a chronic disease.

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I. RECEPTOR TYROSINE KINASES AND THEIR DOWNSTREAM SIGNALING AXES

Cancer is one of the major life-threatening diseases that continuously attract tremendous social attention. Latest epidemiological statistics highlight the global increase of cancer burden (290). In the United Kingdom, it is estimated that 50% of people will suffer from cancer disease at a certain stage in their lifetime (2). In 2014, out of 41 new drugs approved by the Food and Drug Administration (FDA), 22% (9 drugs) were designated for cancer therapy (186). Encouragingly, new anti-cancer drugs have shown

great success in clinical cancer therapy with significantly improved survival rate over the past 30 years (266). However, a relevant shortcoming of targeted therapies is the quick emergence of acquired drug resistance. This is particularly frequent for small-molecule inhibitors that target receptor tyrosine kinase (RTK)-mediated oncogenic signaling pathways. Cancer cells resistant to these drugs usually exhibit a higher degree of genomic instability and show more aggressive phenotypes, such as accelerated metastasis to distant organs and tissues. Thus drug resistance becomes the major challenge in clinical cancer therapies. Development of novel therapeutic strategies towards overcoming drug resistance is a critical issue in clinical cancer therapy.

RTKs are a group of membrane proteins that are activated through tyrosine phosphorylation within their intracellular kinase domain. In the human genome, there are ~58 genes encoding RTK proteins (235). Although they differ in their patterns of expression and activation, such as the abundance on the cell membrane, the discrepant expression between cell and tissue types as well as at different develop-

mental stages, these RTK proteins are evolutionally conserved and are structurally similar on a molecular level, with an extracellular ligand-binding domain, a single transmembrane α -helix, and an intracellular kinase domain that mediates downstream signaling. RTKs are one of the most important molecular sensors that perceive extracellular signals and evoke cell responses through orchestrating intracellular signaling networks. Under physiological conditions, activation of RTKs regulates cell fate in many aspects, including proliferation, differentiation, migration, and metabolic homeostasis (108, 246).

The multiple functionalities of RTKs are achieved through two principal signaling axes: mitogen-activated protein kinase (MAPK) (35) and phosphoinositide 3-kinase (PI3K)/Akt (75, 296, 317). Mechanistic studies on molecular structure reveal that physiological activation of RTKs is initiated by binding of growth factors to the extracellular domain (ECD), which subsequently triggers RTK homo- or heterodimerization. Oligomerized RTKs undergo conformational changes that rapidly induce trans-autophosphorylation on key tyrosine residues within the COOH-terminal kinase domain. This domain not only stabilizes the active state of the RTK but also provides essential docking sites for other regulatory proteins that contain phosphotyrosine-binding motives such as SH2 (301). Once the signalosome is assembled, downstream signaling modules such as MAPK and PI3K/Akt are recruited and activated. These two pathways act as essential processors to direct cellular response at both transcriptional and translational levels in a context-dependent manner (FIGURE 1). However, to maintain a metabolic homeostasis, developmental patterns require spatiotemporal controls of RTK activation. Indeed, activated RTKs not only integrate positive signaling loops but also modulate feedback to terminate their activities. Although other parallel mechanisms exist, MAPK and PI3K/Akt/mTOR cascades are capable of self-limiting excessive activation of RTKs through direct phosphoinhibition of the key adaptor proteins, which results in the interruption of the link between RTKs and their downstream targets (127, 162).

RTKs are frequently hyperactivated in malignant cells and play important roles in the maintenance of tumorigenic phenotypes in various cancers. Due to their substantial contributions to cell growth, RTKs are natural anticancer targets in the clinic. Aberrant constitutive activation of RTKs can be triggered by gene amplification (16, 278), genetic activating mutation (44, 289), gene rearrangement (258), and overexpression of the respective ligands in the tumor stroma (FIGURE 2). Deregulated RTK activation exponentially amplifies downstream signals released from MAPK and PI3K/Akt which leads to uncontrolled cancer cell proliferation and tumor growth. Strategies to inhibit RTK (hyper-) activation have been developed, including blocking antibodies to neutralize the extracellular ligand-binding

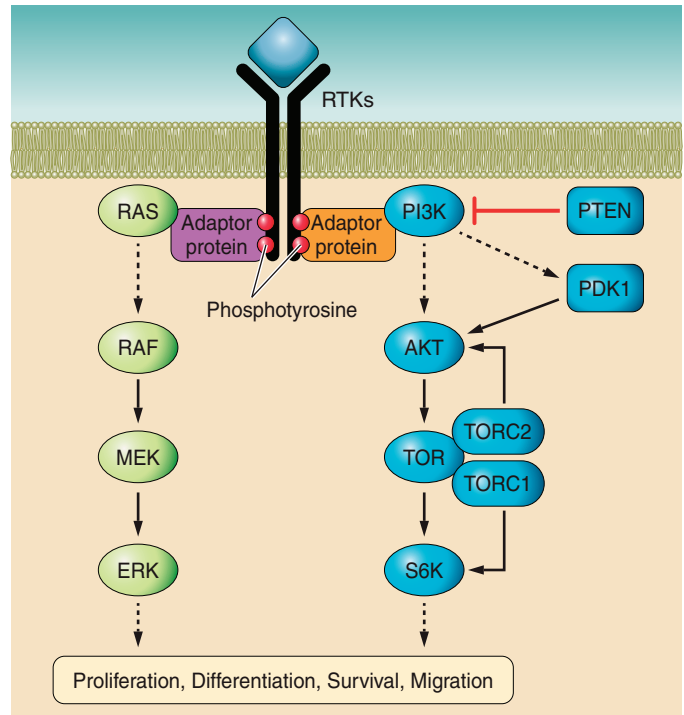


FIGURE 1. Signaling transduction of receptor tyrosine kinase (RTK). The two essential proliferative intracellular signaling pathways, PI3K/Akt/mTOR and RAS/MAPK, are activated downstream of RTKs. Under physiological conditions, these two pathways coordinate with each other to determine proper cell proliferation and organ size. AP, adaptor protein; red ball, phosphotyrosine.

moiety and small molecular compounds (RTK inhibitors, RTKi) to suppress the function of the intracellular kinase domain or prevent RTK dimerization. Since activation of RAS/MAPK and PI3K/Akt/mTOR is either the result of RTK dysfunctions or correlates with mutations further downstream (80, 120, 250), almost all key components along these two signaling axes, such as BRAF, MEK, PI3K, Akt, and mTOR, have been therapeutically targeted to allow for serial and parallel blockade of these two pathways.

ERBB family members, an important member of the RTKs, are frequently hyperactivated (gene amplification and active mutation) during oncogenic progression in many types of cancer, including head and neck squamous cell carcinoma (HNSCC), non-small-cell lung cancer (NSCLC), breast cancer, ovarian cancer, prostate cancer, glioblastoma multiforme (GBM), colorectal cancer, and bladder cancer (256, 283). We will use this family of RTKs as a representative model to discuss the mechanisms of acquired drug resistance in NSCLC and metastatic breast cancer patients undergoing targeted therapies because tumor progression is tightly correlated with therapy-induced drug resistance. In addition, we will also address recent exciting advances in understanding how resistance is developed in metastatic melanomas harboring mutant BRAF, another tumor model that is representative of both self-activating and bypassing mechanisms of therapy-induced resistance.

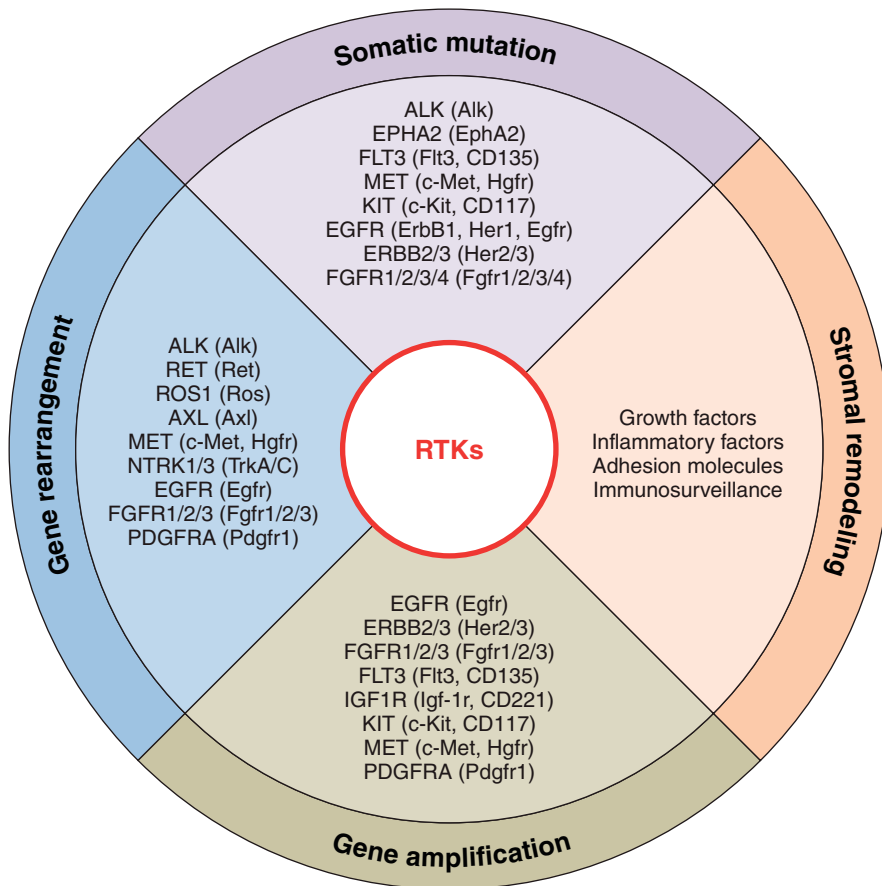


FIGURE 2. Aberrant activation of RTK signaling. Four major factors contribute to elevated activation of major RTK signaling that are frequently detected in human cancer: gene amplification, somatic mutation, gene rearrangement, and remodeling of cancer stroma. These are the predominant players that often lead to pathological activation of many RTK kinases that eventually promote uncontrolled cell proliferation. Persistent activation of RTKs amplifies downstream proliferative signaling, such as PI3K/Akt/mTOR and RAS/MAPK, and inhibits pro-apoptotic signaling like the Hippo pathway. Furthermore, RTK activation can also be caused by drug resistance through interference with signaling cross-talk. Such genetic remodeling encourages genomic instability to overcome cell cycle arrest, and therefore, the cells become cancerous with malignant proliferative and migratory potential.

A. Physiological Signaling of the ERBB Family

The human homolog of the erythroblastic leukemia viral oncoprotein v-erbB, known as the ERBB (also called EGFR or HER) family, is composed of four closely related members (EGFR and ERBB2-4) which localize on chromosomes 7, 17, 12, and 2, respectively (235). Despite the lack of an intracellular kinase domain on ERBB3, the family members share a high degree of similarities in their molecular structure, with a tandem cysteine-rich cascade in the ECD, one single transmembrane helix, and a classical intracellular kinase domain (303). A variety of extracellular ligands including epidermal growth factor (EGF), heparin-binding EGF (HB-EGF), transforming growth factor- α (TGF- α), epiregulin (EpRG), amphiregulin (AmRG), betacellulin (BTC), and neuregulin (NRG) are capable of binding to individual ERBB members and inducing unique homo- and/or hetero-dimerization (109). Dimerized ERBBs subsequently undergo conformational changes and trigger autophosphorylation on specific tyrosine residues within the intracellular kinase domain, which simultaneously switches on the ERBB signaling pathway. This activation pattern with a broad range of functional stimuli possibly reflects the versatility of ERBB signaling in different tissues and organs or at different developmental stages.

The signaling cascades downstream of the ERBB family include RAS/MAPK, PI3K/Akt, JAK/Src/Stat, PLC/DAG/PKC, and CDC42/Rac/Pak (FIGURE 3). Depending on the patterns of dimerization, distinct pathways are activated and functionally regulate cellular transcriptional and translational programs that in turn direct cell cycle progression, proliferation, differentiation, angiogenesis, immunomodulation, polarity, migration, and inflammation in a cell- or tissue type-dependent manner (31). The physiological roles of the ERBB family have been broadly explored using genetic mouse models. Full-body knockout of Egfr leads to embryonic and perinatal lethality (175, 264, 265). Multiple organs and tissues including lung, heart, liver, brain, skin, and bone undergo immature development which accounts for the complex phenotypes seen upon genetic ablation of ERBB family members. For example, severely abnormal placental development and immaturity of the lung were observed in Egfr-knockout mice, leading to spontaneous embryonic or perinatal death. Impaired neural development with progressive neurodegeneration resulting from elevated apoptosis of neural cells in the brain was also observed in these mice (129, 145, 203). These genetic studies demonstrate that Egfr is indispensable during development. The observation of distinct patterns of lethality at different developmental stages of Egfr-knockout mice with different genetic backgrounds (265, 286) implies a partial compen-

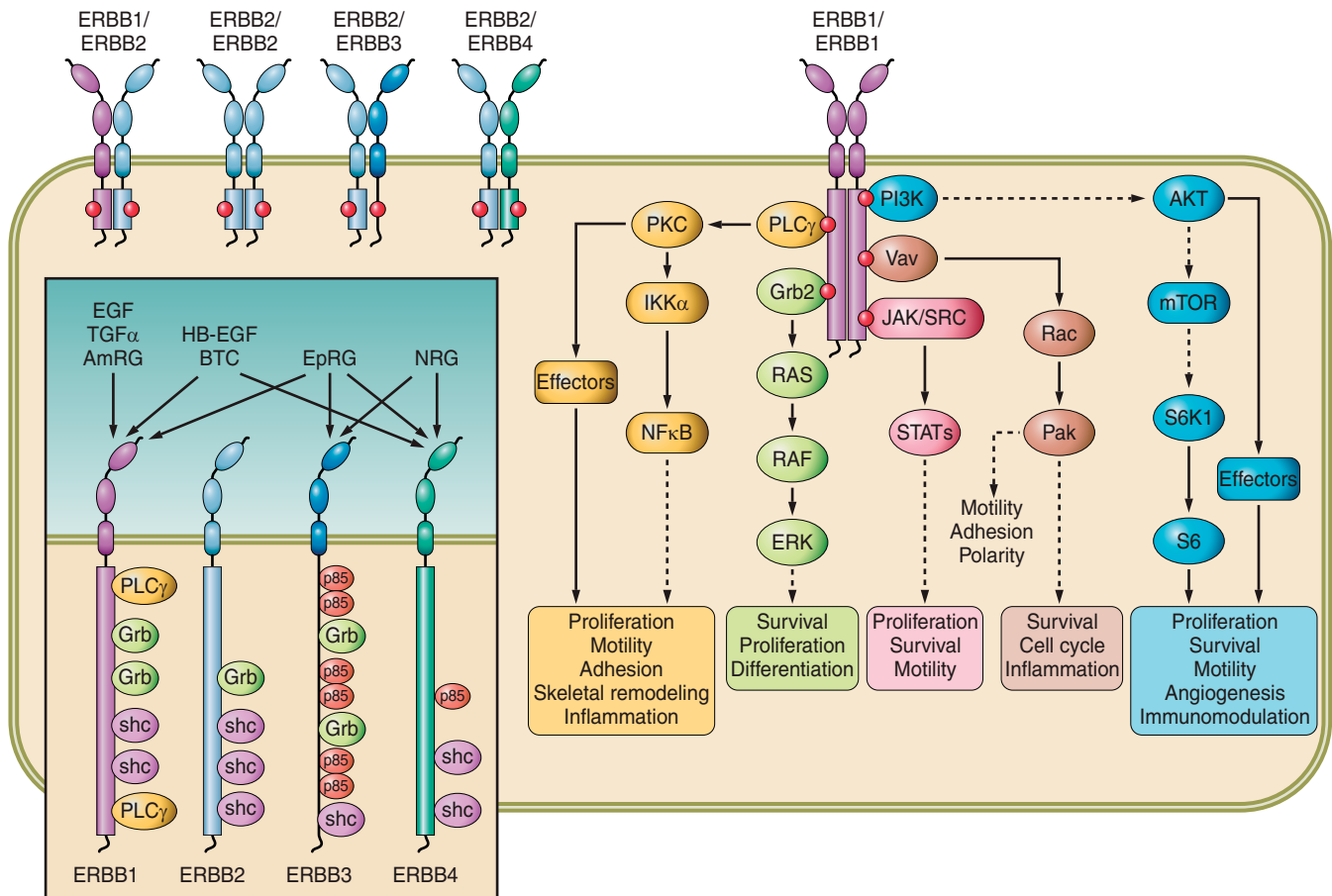


FIGURE 3. Physiological signaling pathway of ERBB family. ERBB signaling is initiated from induced dimerization between four ERBB family members. Five dimerization patterns are reported: ERBB1/ERBB1, ERBB1/ERBB2, ERBB2/ERBB2, ERBB2/ERBB3, and ERBB2/ERBB4. Among the four ERBB isoforms, ERBB3 does not have an intracellular kinase domain, but instead, it enhances ERBB signaling through partnering with ERBB2. In addition, ERBB3 preferentially activates PI3K/Akt as it can directly bind to the regulatory subunit p85 and activate PI3K. Thus ERBB3 activation is widely reported in many types of cancer. Downstream of ERBB are five major axes: PKC, MAPK, PI3K/Akt, and Rac and Jak signaling. In a context-dependent manner, activation of individual pathways contributes to many aspects of cell functionalities through genetic or epigenetic reprogramming.

sation by other ERBB family members in overcoming developmental defects attributed to loss of Egfr. This is indirectly supported by the fact that ablation of certain Egfr ligands, such as EGF and TGF- α , does not result in evident defects (155, 156). In contrast to Egfr, the loss of any other ERBB family member does not necessarily lead to embryonic lethality in mice, which indicates nonredundant roles of Erbb2, Erbb3, and Erbb4 during development. Erbb2 mainly impacts on mammalian neural (140, 146) and cardiac cell development (194, 215), which was confirmed in recent studies on the role of NRG in cardiomyocytes (48, 217) and directly links Erbb2 to cardiac regeneration (49). As expected, due to the direct interaction with Erbb3 and Erbb4, functional loss of NRG mimics Erbb3 or Erbb4 deficiency. Both Erbb3- or Erbb4-knockout mice show neuronal degenerative phenotypes and cardiac maldevelopment (71, 83, 88, 232, 287) similar to NRG-knockout mice (174). Nonetheless, it should be pointed out that functional interference with any single Erbb family member may trig-

ger mixed phenotypic abnormalities with other isoforms, since the specificity of downstream signaling depends on dimerizing partners and a broad spectrum of ligands. Therefore, it is sometimes difficult to correctly define the biological roles of individual ERBB kinases (109).

B. Physiology Downstream of ERBB Signaling: RAF and PI3K/Akt

1. RAF signaling

The mammalian RAF (rapidly accelerated fibrosarcoma) family has three members, ARAF, BRAF, and CRAF (RAF-1), that are intracellular serine/threonine kinases. All three isoforms are ubiquitously expressed in developing embryos but have distinct expression patterns in adulthood. Araf and Crsf are widely expressed in almost all tissues of adult mice, although Araf expression seems to be relatively higher

in the organs of the urogenital tract such as the kidney, bladder, testis, and ovary, as well as in lymphoid organs and lymphatic tissues such as thymus and spleen (277), whereas Braf expression is mostly restricted to the brain and testis (74, 312). Targeted deletion of Araf in mice leads to partially postnatal lethality, growth retardation, defective neurological and gastrointestinal development, and abnormality of limb development (225). In contrast to Araf, knockout of either Braf or Craf results in embryonic death (313, 314). These three mammalian RAF isozymes are evolutionarily conserved and share sequence identities of ~45% in the regulatory domain and ~80% in the kinase domain (333). The deficient phenotypes seen in KO mice and the differences in sequences of functionally defined protein domains indicate nonredundant regulatory roles during development. Although they have a common downstream signaling node, MAPK, their activating capacity is differentially regulated. For example, Braf and Craf differentially respond to NGF or cAMP stimulation (70, 112) and also show considerable differences of their activation driven by Ras (167). Moreover, Src can directly activate Araf and Craf but not Braf (163), and Braf is the major activator of MAPK pathway due to its higher binding capacity to MEK (a MAPKK) (207, 226, 311). This is not due to their expression level (33, 183) but their functional specificity in individual tissues (199). On the other hand, biological interactions between Raf family members are also indispensable for specific extracellular signaling triggers, for example, EGF (312) and downstream signal transmissions (173). Taken together, similar to the ERBB family, RAF family members have an extensive crosstalk, thus differentially regulating cell proliferation, differentiation, and

survival through MAPK signaling across the entire development.

2. PI3K/Akt signaling

Similar to the activation pattern of RAF signaling, intracellular tyrosine phosphorylation on RTKs generates binding pockets for the SH2-containing subunit p85 and subsequently induces catalytic activity of the p110 subunit. These two subunits assemble the functional PI3K protein (PI3K class I) (8). Activated PI3K catalyzes phosphatidylinositol 4,5-bisphosphate (PIP₂) to phosphatidylinositol 3,4,5-trisphosphate (PIP₃), which promotes membrane targeting of pleckstrin homology (PH) domain-containing proteins such as Akt and phosphoinositide-dependent kinase 1 (PDK1) (98, 142). In response to RTK signaling, PI3K transduces physiological signals through intracellular kinases like Akt, protein kinase C (PKC) and phospholipase C (PLC). Among those, the PI3K/Akt axis plays a vital role in regulating cell growth, anti-apoptosis, and metabolic homeostasis. Subsequent Akt-directed phosphoregulation of a large number of specific substrates determines the cell fate depending on tissue type, developmental stage, and environmental stress. During embryonic development and postnatal organ/tissue formation and maturation, all three Akt isoforms play crucial roles in growth and metabolism (TABLE 1). Akt1 is ubiquitously expressed in mammalian cells, and Akt2 is mainly detected in insulin-responsive tissues such as skeletal muscle and adipose tissue, whereas Akt3 is restricted to the brain and testis. Although Akt1 KO mice are viable, they have a smaller body size (~20 and ~25% reduction at birth and 14 mo after birth, respectively) than their littermates,

Table 1. Tissue-specific distribution of *ErbB*, *Raf*, and *Akt* kinases in mice and the individual knockout phenotypes

Genes	Tissue Distribution	Knockout Phenotypes
Egfr	Ubiquitous	Perinatal lethality; defects in the development of multiple tissues/organs including skin, lung, bone, heart
ErbB2	Intestine, stomach, epidermis, uterus, kidney, prostate	Embryonic lethality
ErbB3	Breast, epidermis, stomach, CNS, intestine, prostate, kidney, brain	Embryonic lethality
ErbB4	Brain, liver, heart, eyes, CNS	Embryonic lethality
Araf	Heart, brain, liver, kidney, lung	Partially postnatal lethality; defective development of multiple organs
Braf	Testis, brain	Embryonic lethality
Craf	Ubiquitous	Embryonic lethality
Akt1	Ubiquitous	Increased postnatal lethality; growth retardation; defect of development of placenta
Akt2	Muscle, fat, liver	Diabetic phenotype: hyperglycemia; hyperinsulinemia; insulin resistance; growth retardation; loss of adipose tissue
Akt3	Brain, testis	Neural degeneration; defect of brain development
Akt1/Akt2		Perinatal lethality; severe growth defect
Akt1/Akt3		Embryonic lethality (~E12.5)
Akt2/Akt3		Diabetic phenotype; defect of brain development

defects of testis development and abnormal spermatogenesis associated with elevated spontaneous apoptosis, and a shorter lifespan when challenged with genotoxic substances, probably due to a lack of pro-survival signals (41). In fact, mice without Akt1 show partial neonatal lethality (43), possibly due to a functional deficiency of the placenta (325). Unlike Akt1, Akt2 is mainly detected in muscle, fat, and liver, those tissues which are more sensitive to insulin or play a central role in glucose homeostasis. Akt2 null mice are also viable but exhibit diabetic phenotypes such as hyperglycemia, hyperinsulinemia, and insulin resistance (81), indicating that Akt2 is a key regulator of glucose metabolism (42). This is demonstrated in further studies showing that Akt2 phosphoregulates membrane targeting and activation of glucose transporter (GLUT4) in response to insulin signaling (100, 132). Similar to Akt1, Akt2 deficiency is also associated with moderate growth retardation that results from early metabolic disorders through reduced glycolysis and mitochondrial dysfunction (89). On the other hand, mice lacking Akt3 develop a smaller brain (20-25% reduction) (64, 292). Interestingly, these unique phenotypes governed by individual Akt isoforms are unlikely to be compensatory, suggesting nonredundant physiological functionalities of each Akt isoform across the entire lifespan. Although mice may survive the loss of a single Akt isoforms (63), double-knockout (dKO) of Akt1/Akt3 is embryonically lethal (324) with Akt1/Akt2-dKO mice dying shortly after birth due to severely mixed defective phenotypes of cell proliferation and differentiation (213). Similarly, Akt2/Akt3-dKO mice exhibit aberrant brain development and diabetic phenotypes (63), further demonstrating the specific roles of individual Akt proteins in different cell types and developmental stages.

II. TARGETED STRATEGIES IN CLINICAL CANCER THERAPIES

The latest report on the global cancer burden shows that the incidence of cancer continuously increased from 2002 to 2012 (209, 290). Remarkably, in last 15 years, lung cancer is the leading cause of cancer-related death for both males and females; in addition, breast cancer results in an equally high rate of death among females. Recent advances in the mechanistic understanding of tumor biology rationalize future therapeutic strategies by targeting the fundamental hallmarks of cancer (94).

In many types of cancer, including NSCLC and metastatic breast cancer, gene amplification and/or mutation-driven constitutive activation of the ERBB family facilitates uncontrolled cancer cell proliferation and invasion as well as the evasion of programmed cell death (130, 196). Therefore, ERBB family members have emerged as key therapeutic targets (TABLE 2). Apart from the ERBB family, distinct deregulation of anaplastic lymphoma kinase (ALK) through oncogenic protein fusion with echinoderm microtubule-associated protein-like 4 (EML4) has been found to cause ~5% of NSCLC in clinic (271). Interestingly, using next generation sequencing (NGS), a recent study identified a novel oncogenic fusion of ALK with KIF5B-RET in NSCLC (147), indicating that ALK deregulation is more frequent in NSCLC than it was assumed before. These clinical observations are the rationale for suppressing tumor growth by blocking ERBB and ALK kinase activities. Thus targeting ERBB and ALK families with either small molecular inhibitors or blocking antibodies has become the first line of therapy for lung cancer and Her2-positive breast

Table 2. FDA-approved major kinase (pathway) inhibitors

	Generic Name	Brand Name	Target	Producer
Lung cancer	Gefitinib	Iressa	EGFR	AstraZeneca
	Erlotinib	Tarceva	EGFR	Roche
	Crizotinib	Xalkori	ALK	Pfizer
	Afatinib	Gilotrif/Giotrif	EGFR, ERBB2	Boehringer Ingelheim
	Ceritinib	Zykadia	ALK	Novartis
	Cetuximab	Erbix	EGFR	Yeda & Sanofi-Aventis
	Panitumumab	Vectibix	EGFR	Amgen
Breast cancer	Bevacizumab	Avastin	VEGF-A	Roche
	Lapatinib	Tykerb	ERBB2, EGFR	GSK
	Trastuzumab	Herceptin	ERBB2	Roche
Melanoma	Pertuzumab	Perjeta	ERBB2	Roche
	Vemurafenib	Zelboraf	BRAF (V600E)	Roche
	Dabrafenib	Tafinlar	BRAF (V600E)	Novartis
	Trametinib	Mekinist	MEK1/2	GSK

Food and Drug Administration (FDA)-approved major kinase (pathway) inhibitors (including both small molecular compounds and therapeutic antibodies) in three types of cancer are shown. The inhibitors approved for therapeutic applications in other cancers, or still in clinical trials, are not included.

cancer patients. In fact, with the response rate above 50%, these targeted therapies significantly improve progression-free survival of NSCLC patients (204, 216) and breast cancer patients (11, 17, 24, 52, 86).

In contrast to other types of cancer, the incidence rate for melanoma has been steadily increasing for both men and women (266). When melanoma develops metastases, the five-year survival rate decreases dramatically from 98 to 15%. One of the hallmarks of metastatic melanoma is the constitutive activation of the MAPKKK kinase BRAF that harbors an oncogenic mutation (BRAF_V600E, V600K, or V600R) in ~50% of melanoma patients (20). Constitutively active BRAF promotes cancer cell proliferation through hyperactivated ERK signaling. Compared with the chemotherapeutic agent dacarbazine, treatment with one of the available small molecule inhibitors of BRAF (vemurafenib, Zelboraf, Roche and dabrafenib, Tafinlar, Novartis) significantly improves overall survival of melanoma patients (36, 272).

III. ADAPTIVE RESISTANCE TO TARGETED THERAPIES

Unfortunately, most of the targeted monotherapies against cancer eventually result in resistance. Cancer cells bypass proliferative inhibition through alternative activation of other survival pathways as functional compensation. Activation of these signaling pathways is mediated by various mechanisms on both a transcriptional and translational level, including induction of novel oncogenic mutations, inactivation of negative-feedback signaling loops, aberrant protein-protein interaction/oligomerization, oncogenic gene amplifications, suppressive gene deletions, conversion of apoptotic signaling to survival signaling, and deregulated immunosurveillance. Resistant cancer cells often exhibit an accelerated cell cycle, enhanced metabolism, and increased migration/invasion that ultimately leads to higher malignancy. Therefore, a better understanding of the underlying mechanisms of resistance will contribute to the development of novel therapeutic approaches that may help to turn cancer into a chronic disease.

A. Resistance to Targeted EGFR and ALK Signaling in NSCLC

Since EGFR was discovered and validated as a major drugable target in NSCLC, specific inhibition of EGFR signaling was shown to prolong both PFS and OS compared with conventional platinum-based chemotherapy and is now the accepted standard in the clinic. Although a subset of NSCLC patients treated with gefitinib or erlotinib benefit from a longer lifespan for a longer period of time (131, 157), including those patients with primary somatic mutation L858R on EGFR (178, 201, 205), two large cohort

studies revealed rapid development of acquired resistance in the majority of the monotherapeutically treated patients like these (181, 239). Several independent studies reported that a secondary somatic mutation of EGFR, T790M, was emerging in relapsed NSCLC (126, 206). The decreased inhibitory efficacy of gefitinib against EGFR T790M was confirmed in genetically engineered cell lines expressing T790M mutants (206). Further studies indicated that T790M could also be detected in some patients before treatment, which was possibly responsible for their primary resistance to gefitinib (261) and erlotinib (238, 328). Indeed, with the help of next-generation-sequencing technology, it becomes clearer that T790M is a frequent primary mutation that is observed in a small number of malignant cell clones in NSCLC patients (281). Under selective pressure with an EGFR inhibitor, the T790M clone expands quickly and leads to resistance in ~50% of NSCLC patients. Although it is not fully understood mechanistically, studies from structural biology predict that the T790M mutation mediates steric hindrance in the ATP-binding pocket to limit the access of small molecular inhibitors (134) and increases ATP-binding affinity in the kinase domain of EGFR (332). Functionally, persistent activation of EGFR with the T790M mutation maintains hyperactivation of its downstream pro-survival signaling axes, including PI3K/Akt, JAK/Stat3, and MAPK.

In addition to the dominant gatekeeper mutation, amplification of other oncogenic RTKs in EGFRi-resistant NSCLC was also observed. C-Met, also known as hepatocyte growth factor receptor (HGFR), is a receptor tyrosine kinase that plays essential roles during embryonic development and wound healing and is overexpressed in EGFRi-resistant lung cancer (69). Mechanistically, overexpressed c-Met may heterodimerize with HER3 and subsequently mediates PI3K/Akt activation bypassing EGFR inhibition. Similarly, recent studies also identified gefitinib/erlotinib-mediated overexpression of AXL (337), amplification of HER2/HER3 (161), and activation of an FGFR autocrine signaling loop (284, 304). All of these mechanisms contribute to acquired resistance in NSCLC.

Although the specific tissue distribution pattern suggested a role of ALK in brain and neuronal development (110, 227), the lack of abnormal phenotypes in ALK knockout mice throughout their lifespan (305) makes it difficult to define the physiological role of this kinase. Despite these uncertainties in physiology, numerous reports confirmed its importance in driving tumorigenic progression in many types of cancer, most frequently as rearrangement/translocation-elicited oncogenic fusion proteins. In NSCLC patients, ALK is commonly fused with EML4, a microtubule-stabilizing protein guarding correct formation of cellular skeleton network (220). The resulting chimeric protein functions as an intracellular kinase promoting cancer cell proliferation, invasion, and anti-apoptosis through activating PI3K/Akt,

MAPK, and JAK/Stat signaling. Although inhibitors targeting ALK, such as crizotinib and ceritinib, showed immediate benefits in ALK-positive NSCLC patients (133), the short duration of response indicated an acquired resistance (122, 259). Sequencing analysis of crizotinib/ceritinib-resistant tumors revealed multiple self-activating mutations on ALK (25, 59, 78) and bypassing activation of alternative oncogenic drivers such as KIT (122), IGF-IR (153), EGFR (320), and the GPCR family member P2Y (310). The resulting acquired resistance does not seem to be restricted to TKIs, since the use of blocking antibodies targeting EGFR, such as cetuximab and panitumumab, leads to similar outcomes (10, 309). Clearly, these resistant phenotypes are closely related to and possibly driven by the heterogeneity of NSCLC (FIGURE 4).

B. Resistance to HER2-Targeting in Breast Cancer

The oncogenic role of HER2 has been extensively investigated in human breast cancer. Approximately 25% of invasive breast tumors overexpress HER2 (269, 270). Overexpression of HER2 is associated with a poor prognosis and survival rate (223, 224), and it is also observed as a response to chemotherapies (189, 202, 285). Trastuzumab (Herceptin, Roche) is a humanized monoclonal antibody that targets HER2 and has been approved for clinical breast cancer therapy by the FDA for 15 years. Trastuzumab treatment induces tumor regression through interference with HER2 signaling, in particular HER2 internalization/degradation (47), inactivation of proteolysis of ECD of HER2 (73, 182),

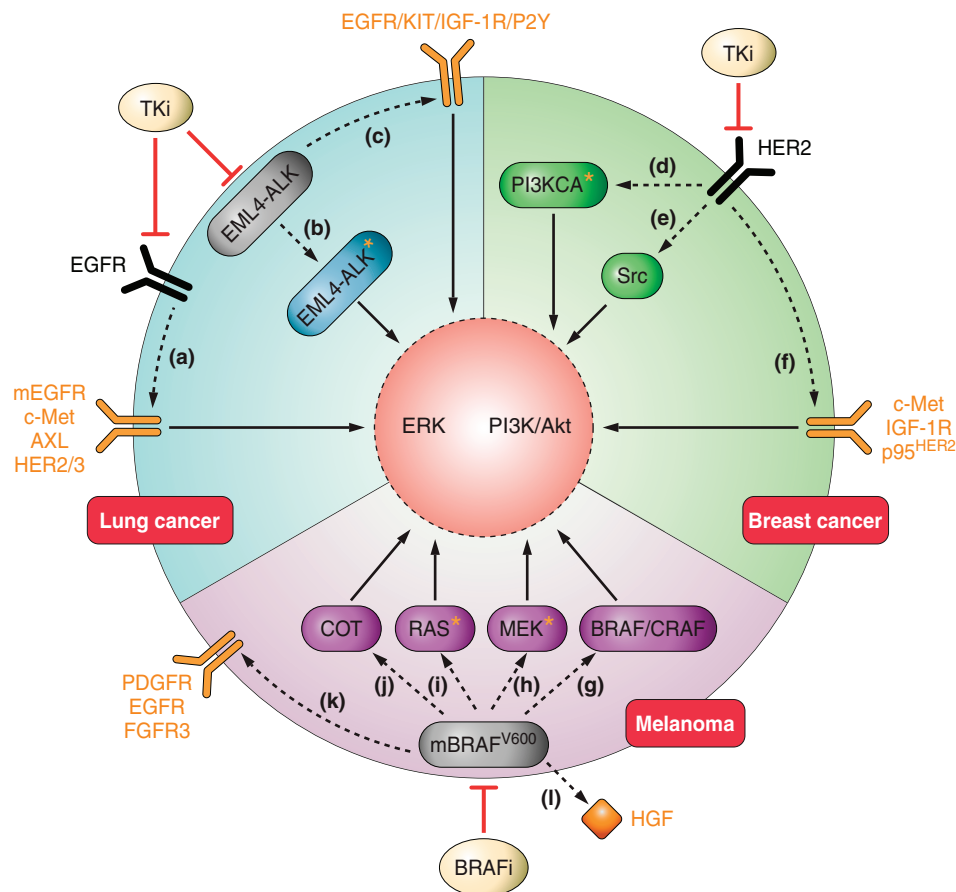


FIGURE 4. Representative mechanisms of drug resistance to clinical therapies in human lung cancer, breast cancer, and melanoma. Several molecular signatures are discovered that contribute to resistance. In lung cancer, *a*) inhibition of EGFR induces not only overexpression of alternative RTKs, such as c-Met, Axl, ERBB2/3, but also activating mutations of EGFR itself; *b*) targeted inhibition of oncogenic ALK fusion protein leads to ALK mutations and *c*) oncogenic activation of EGFR, Kit, IGF-1R, and P2Y kinases. In metastatic breast cancer, blocking HER2 activity frequently results in *d*) activating mutations on catalytic subunit of PI3K, *e*) activation of Src kinase family, and *f*) activation of RTK including c-Met, IGF-1R, and activating truncation form of HER2. In BRAF^{V600E/K} melanomas, blocking kinase activity of mutant BRAF with Zelboraf or Tafenlar can *g*) induce active dimerization between CRAF and kinase-dead mutant BRAF that drives ERK activation, *h*) trigger activating mutation on MEK and *i*) RAS, or upregulate *j*) MAPK3K kinase COT and *k*) RTKs including PDGFR, EGFR, and FGFR. In the cancer environment, growth factor HGF overexpression is also reported to contribute to BRAFi resistance (*l*). All these genetic and/or epigenetic remodeling can empower cancer cells to activate the proliferative and survival pathways, MAPK and PI3K/Akt, to overcome kinase inhibitor-induced apoptosis.

and HER2-dependent angiogenesis (111). HER2-positive patients initially respond well to trastuzumab, but similar to other targeted therapies they often relapse within ~1 year of treatment, suggesting an acquired resistance promoting escape from HER2 blockade. Similarly, resistance to lapatinib, an FDA approved Her2 small molecule inhibitor, occurs rather fast (40, 302, 308). Several potential mechanisms of acquired resistance have been investigated.

In HER2-positive breast tumors including both primary and metastatic lesions that do not respond to trastuzumab, downstream activating mutations of the catalytic subunit of PI3K have been discovered (15). Mutant PI3K constitutively activates Akt-dependent cell proliferation and survival. Such hot spot mutations of PI3K are not only responsible for acquired resistance but also intrinsic primary resistance to trastuzumab. In fact, PI3K was identified as the most important mediator downstream of HER2 signaling in resistant breast tumors, in which PTEN, the negative regulator of PI3K, is simultaneously downregulated (191). Co-inhibition of PI3K overcomes trastuzumab resistance, placing PI3K as an essential co-target in future HER2 therapies (117, 326). This therapeutic strategy has been under evaluation in preclinical models and potentially provides clinical benefit (198). In addition to oncogenic mutations of PI3K, activating splicing of ECD of HER2 was also discovered in tumors resistant to trastuzumab. Masking of the antibody-binding site within the ECD leads to a silent response of HER2 to trastuzumab without impairing its intracellular signaling as shown by its capacity of maintaining hyperactivation of PI3K/Akt (6, 244), which in contrast is abolished by lapatinib (243). This indicates that the truncated HER2 with loss of trastuzumab recognition is potentially persistently active, which is partially confirmed in a recent report (30). Recent studies also revealed a nuclear fraction of the truncated form of HER2 that may also contribute to resistance (315). Another form of HER2 splicing is the truncation within its ECD, resulting from the deletion of exon 16. Tumors carrying this truncation showed increased growth, metastasis (4), and dimerization activity of HER2, resulting in antagonizing trastuzumab-induced complex disruption (177). Interestingly, this splicing variant was sensitive to trastuzumab in a mouse xenograft model. Therefore, it was suggested that this splicing variant may predict the likelihood of acquired trastuzumab resistance (32). Somatic HER2 mutations have been occasionally found in some tumors but are rarely associated with increased HER2 expression levels. It is unclear how or if these mutations contribute to cancer development and drug resistance.

With the support of more sensitive NGS technologies, a few activating mutations were identified in human breast cancers (9, 26, 66, 195, 255, 276). The roles of such emerging mutations are still being characterized, and the current data support the notion that some point mutations are possibly

responsible for either primary or acquired resistance to HER2-targeted therapies (5, 21). In addition to the gain-of-function mutations of PI3KCA, increased activity of Src kinase in response to HER2-targeting has been shown to activate downstream PI3K/Akt bypassing HER2 in resistant breast cancer cells (212). Similar to PI3KCA, co-inhibition of Src kinase overcomes lapatinib-elicited resistance in vivo (230). Enhanced Src activation is also triggered by ECD masking with trastuzumab, and co-targeting both HER2 and Src has demonstrated synergistic effect in animal models (334). These observations provide a scientific rationale for Src kinase as a core component of targeted therapies in breast cancer (3, 268). Furthermore, induced overexpression of other oncogenes, mainly receptor tyrosine kinases, also contributes to resistant phenotypes compensating HER2 deactivation in resistant tumors. For example, IGF-IR overexpression can override trastuzumab-induced tumor regression through rescue of the cell-cycle program (154), which could be reversed upon blockade of IGF-IR (113). Consistent with this observation, enhanced oligomerization between HER2 and IGF-IR was observed in trastuzumab-resistant breast cancer cells (107, 149). Another key regulator of trastuzumab resistance is c-Met (257). c-Met is also reported to be overexpressed in a subset of invasive breast cancers and correlates with poor outcome (119, 321). Interestingly, c-Met was found to form active protein complexes with HER2 in NSCLC, implying a synergistic role in driving downstream PI3K/Akt and ERK signaling (FIGURE 4).

Some members of the mucin protein family, such as mucin-1 and mucin-4, were also found upregulated and masked the trastuzumab-binding site on HER2 in resistant breast cancer cells (76, 192). It was also reported that dimerization of HER2 and HER3, rather than EGFR, plays an essential role in mediating downstream PI3K/Akt signaling (139). This dimerization pattern is not readily disrupted by trastuzumab (1), and increased HER3 expression upon TKI treatment may effectively enhance HER2/HER3 interaction (252), thus contributing to a reduced response to HER2 targeting. Following this rationale, disruption of dimerization with pertuzumab (Perjeta, Roche), another FDA-approved humanized blocking antibody that specifically targets dimerization functionality of HER2, dramatically restored sensitivity to trastuzumab treatment (13, 27, 143). This synergistic efficacy was also confirmed in a clinical study (12).

C. Resistance to Mutant BRAF^{V600}. Targeting Therapy in Metastatic Melanoma

Approximately 50% of metastatic melanomas harbor an oncogenic mutation on BRAF which acts as a major driver to fuel out-of-control proliferation of tumor cells through MAPK signaling. Among the human melanomas with a

gain-of-function mutation on BRAF the substitution on V600 is: V600E (~70%), V600K (10~15%), and V600R (3~7%). V600E/K mutations biochemically mimic the phosphorylation-dependent active conformation of wild-type BRAF, thus resulting in constitutive activation as a monomer in a RAS-independent manner. Although ARAF and CRAF share high sequence similarity with BRAF in the kinase domain, mutations of ARAF or CRAF are very rare. BRAF mutations are not only restricted to melanoma, but also frequently detected in thyroid cancer (30~70%), ovarian cancer (~30%), and colon cancer (~10%) (51). Therefore, targeting mutant BRAF is a promising therapeutic strategy.

Two FDA and EMA approved small molecular BRAF inhibitors (BRAFi), vemurafenib/Zelboraf and dabrafenib/Tafinlar, specifically inhibit the BRAF kinase activity and lead to prolonged overall survival of melanoma patients (272). However, the majority of patients stop responding to BRAFi within 6–9 mo (36), indicating an acquired resistance to BRAFi. BRAFi-induced resistance can be mediated by several different mechanisms (FIGURE 4) including re-dimerization of the kinase-dead form of mutant BRAF with endogenous CRAF (96, 221), unfavorable disruption of the ERK-dependent negative feedback signaling loop (148), acquiring activating mutations of RAS and MEK (23, 67, 298, 300), upregulation of other pro-oncogenic kinases, such as COT (115), RTKs [platelet-derived growth factor receptor (PDGFR), EGFR, and fibroblast growth factor receptor (FGFR)] (193, 319) or RTK ligands like HGF (279). All of these mechanisms ultimately trigger the reactivation of MAPK and PI3K/Akt signaling that support melanoma cell survival. In fact, despite those spontaneously occurring mutations of RAS, MEK and possibly other undiscovered oncogenes, targeting mutant BRAF simply enhances physiological RAS/RAF/MAPK signaling in three ways. First, BRAFi induces self-activation of RAF signaling through an existing downstream feedback loop. This effect is essentially achieved through ERK-dependent transcriptomic programming of physiological inhibitors of RAF kinases (148, 222) or direct phospho-inhibition of physical binding between RAS and RAFs (60, 233). As a consequence, transient downregulation of ERK may also lead to proliferative activation of another pathway, mTOR/PI3K/Akt, through signaling cross-talk mediated by ERK (38, 214). Prolonged exposure of cancer cells to BRAFi eventually reactivates ERK. This indicates a fundamental role of ERK reactivation in BRAFi resistance. Indeed, ERK activation is a universal phenotype for both BRAF- and MEKi-induced resistance (95, 184). Simultaneous blocking of mutant BRAF and MEK improves anti-tumor activity of BRAFi (77, 152). On the basis of three positive phase 3 trials, the combination of BRAF (vemurafenib, dabrafenib) and MEK inhibitors (cobimetinib, trametinib) is a new standard for BRAF mutant melanoma. Second, BRAFi transactivates RAF signaling, mimicking functional assembly of a wild-type signalosome.

Functional RAF signaling requires dimerization of RAF family members such as BRAF and CRAF to drive downstream MAPK activation. This is triggered by upstream RAS signaling in a CRAF-dependent manner. In melanoma cells with mutant BRAF, unlike the dimerization-dependent activation of wild-type RAF, BRAF^{V600E/K} performs its kinase function as a monomer. BRAFi treatment only blocks its kinase activity but disguises the kinase-dead form as a “wild-type” BRAF that still has the capacity to dimerize with CRAF. As a consequence, the entire MAPK pathway is reactivated in this setting. Third, BRAFi can induce RTK signaling through a feedback loop. Feedback-induced over-expression includes PDGFR, HER3, and RAS, which potentially promotes Akt phosphorylation through classical RTK/PI3K/Akt signaling (280). Investigating BRAFi resistance has unravelled a complex intracellular signaling cross-talk whose deregulation leads to unresponsiveness to targeted therapies.

D. Inhibition-Enhanced Activation: Feedback and Cross-talk Loops of PI3K/Akt/mTOR Axis at a Glance

The physiological roles of PI3K/Akt in gate-keeping cell proliferation and anti-apoptosis place this pathway as one of the most important defensive signaling cascades. This vital function is often malignantly hijacked due to an increased demand of metabolism in pathological contexts. In almost all cancer types, PI3K/Akt pathway is deregulated and essential for cancer cell proliferation. Tumor relapse from drug resistance also strongly relies on Akt-dominated anti-apoptosis. Thus targeting this signaling axis could potentially induce cancer cell death. Many different small-molecule inhibitors and biological substances specifically targeting each component along this pathway are currently in clinical studies. Although in vivo studies in cancer-modeled mice show a profound benefit in attenuating tumor growth and abrogating therapy-induced drug resistance through mono-targeting PI3K, Akt, and mTOR, or by dual-inhibition of PI3K/mTOR, inhibition of PI3K/Akt/mTOR also elicits resistance through feedback loops similar to other targeted therapies.

Several mechanisms are involved in inducing resistance. The first important mechanism is the interference of a pathway with its own physiological self-regulating feedback loop (FIGURE 5). Nutrient-stimulated Akt activation through insulin and insulin growth factor-like receptors (IR and IGF-IR, respectively) is the major driver of downstream signaling in a physiological context. Along this signaling axis, the p85 regulatory subunit of PI3K is recruited to IR/IGF-IR through the insulin receptor substrate (IRS), an adaptor protein that anchors the assembly of the active PI3K kinase complex. Upon activation, Akt triggers the activation of ribosomal protein S6 kinase (S6K) and 4EBP-1 in a mTOR-dependent manner (135–137, 262).

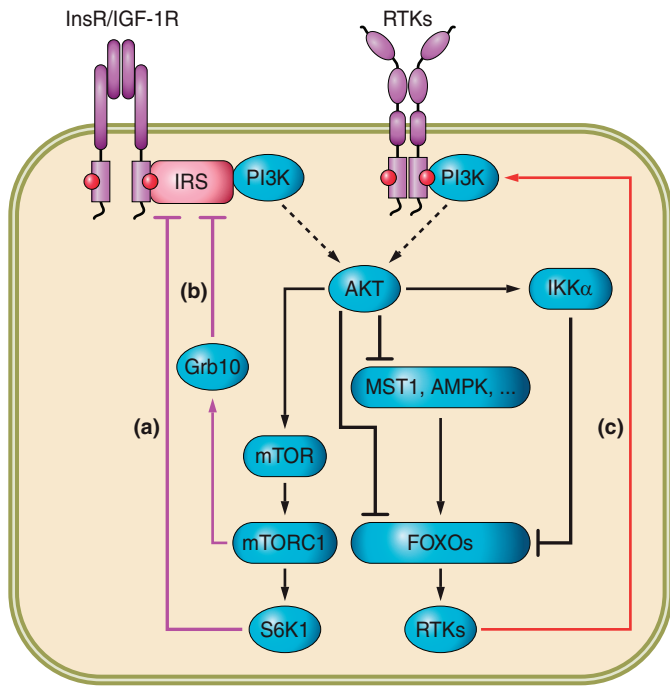


FIGURE 5. Physiological negative-feedback regulation and bypassing activating signaling loops of PI3K/Akt/mTOR pathway. PI3K/Akt/mTOR activation is initiated through PI3K by binding to RTKs. This process can be direct or mediated by a linker protein such as IRS. Under physiological conditions, insulin signaling is a strong driver of PI3K activation, which subsequently leads to ribosomal protein S6 kinase [S6K1] activation. To avoid unwanted PI3K activity, S6K1 is able to phospho-inhibit IRS that promotes the dissociation of PI3K from InsR or IGF-1R to reduce PI3K activity (a). Activated mTORC1 can also phosphorylate Grb10, an adaptor protein that interferes with IRS binding to InsR/IGF-1R, an alternative route to control PI3K activity (b). Several RTKs are transcriptionally regulated by FOXO proteins whose activities are negatively regulated by Akt through direct phosphorylation. Indirectly, Akt is able to activate IKK signaling that inhibits FOXO, or Akt inhibits MST1 and AMPK which activate FOXOs. Upon inhibition of PI3K/Akt, FOXO proteins rapidly translocate into nucleus and form active transcriptional complex that drives mRNA expression of RTKs (c).

Physiological homeostasis of metabolism is maintained through negative feedback signaling of S6K-driven phosphoinactivation of IRS, which avoids overtime activation of the PI3K/Akt/mTOR pathway (176). In addition, another parallel route was also discovered: mTORC1 phosphoactivates growth factor receptor-bound protein 10 (Grb10) (103), an adaptor protein that negatively regulates the activities of IR/IGF-IR and IRS (329). As a result, cooperation of these two major negative feedback loops provides a balanced Akt activity for proper cell proliferation. In parallel, activation of Akt sequesters transcription factor FOXO proteins in the cytoplasm through direct phosphorylation. FOXO proteins are considered negative regulators of cell proliferation through transcriptional inhibition of cell-cycle promoters like cyclin D (247) and through induction of several cell-cycle blockers, such as p21 (53) and p27 (57, 170). On the other hand, FOXO proteins are also involved in other cell functions such as driving RTK expression directly through transcriptional regulation. This is interesting

because, for example, FOXO 1, 3, and 4 have been found to upregulate platelet-derived growth factor receptor (PDGFR) in neuroblastoma, a childhood malignancy (171). Additionally, other RTKs like insulin and insulin-like receptors (InsR and IGF-IR) (164), HER2 and HER3 (34, 82, 253) are emerging as transcriptional targets of FOXO family members that closely associate with the PI3K/Akt/mTOR activities in different cancer types. Moreover, several studies also pointed out the importance of the transcriptional interregulations between FOXO proteins (72). Due to the oncogenic demand for accelerated metabolism, RTKs including IR/IGF-IR signaling are often hyperactivated, thus continuously providing energy equivalents to fuel Akt activation (218). In addition to Akt, the I κ B kinase (IKK), a downstream substrate of Akt, was also shown to phosphoinhibit FOXO activities (106), further preventing an unwanted activation of RTKs and cell apoptosis in stress conditions.

Interestingly, some proapoptotic kinases such as MST, the mammalian Hippo ortholog, and stress-activated AMP-activated protein kinase (AMPK), which are both inhibited by Akt, can directly phosphoactivate FOXO family members (62, 91, 141, 294, 331). This remarkable increase of PI3K/Akt activity in cancer cells quarantines their survival phenotype while antagonizing genomic instability-triggered apoptosis through (at least partly) FOXO transcription factors. In this regard, blockade of PI3K/Akt/mTOR pathway has been considered as one of the most promising strategies to suppress cancer cell proliferation. This concept has indeed been proven in a number of clinical studies with monospecific or dual-specific compounds targeting the PI3K/Akt/mTOR pathway. Everolimus, Temsirolimus (both mTOR inhibitors), and Idelalisib (a PI3K δ inhibitor) have been approved for clinical use. However, based on the emerging image of the signaling landscape of autoregulation and compensatory reactivation (240), it is assumed that inhibition of PI3K/Akt/mTOR will eventually activate upstream RTK signalosomes. This multi-faced reactivation of RTK-mediated signaling reupregulates a variety of downstream oncogenes, which then leads to resistance to targeted inhibition therapies (125, 150, 188, 249).

IV. TUMOR HETEROGENEITY PROMOTES RESISTANCE

Tumor heterogeneity, namely, the intertumor and intratumor diversities, has been realized for a long time in clinic (99, 273) and is confirmed with improved sequencing technologies (14). Such diversities promoted through stemness of cancer cell and clonal evolution, and reflected by cell morphology, gene expression profile, metabolic, proliferative and migratory patterns have been observed in many types of cancer (254). Genetically, extrinsic factors such as exposure to radiation and cytotoxic reagents, as well as intrinsic genomic instability induce somatic driver muta-

tions (22, 58, 85) to allow accelerated cell proliferation and actively resist to environmental stress. Furthermore, such genetic heterogeneity is often accompanied by epigenetically induced deregulation of gene expression and pathway activation to elevate genetic plasticity that is in favor of cancer cell survival (282, 327).

Evolutionary reprogramming of tumor genome frequently correlates with decreased sensitivity in response to chemotherapy (56). The selective clonal expansion supports tumor cells to better adapt to stress conditions. In addition to the preexisting somatic driver mutations in certain tumor-initiating cells that actively accelerate their cell cycle, deactivation of single oncogenic signaling axis by targeted therapies ultimately triggers Darwinian selection to guide continuous cancer cell proliferation. This passive process leads to rapid growth of those cancer cells that are insensitive to the therapies. In fact, emerging studies have shown that the selectively expanded cancer cells are the source of tumor malignancy and account for tumor relapse in cancer patients (166, 236). The relapsed tumors often become more heterogeneous (197) and decline the response to cytotoxic reagents. A better studied model is the core node driving cell survival, the PI3K/PTEN/Akt pathway. In melanoma patients with disease progression posttreatment, the resistant tumors exhibited branched evolution marked by high frequency of oncogenic driver mutations along PI3K/PTEN/Akt and MAPK pathways (124, 260), indicating that the melanoma genomic heterogeneity is the key factor responsible for reduced efficacy of BRAFi treatment. Similarly, the PI3K mutation (H1047R) can activate a multipotent genetic program and cell plasticity at the early stage of breast tumor initiation and establish future intratumoural heterogeneity (128, 295). Independent clinical trial studies have also demonstrated that rapid clonal expansion of the cells harboring oncogenic mutations correlates with worse responsiveness (160, 197) and accelerated malignancy (245, 291). Taken together, tumor heterogeneity is an essential factor responsible for drug resistance through clonal evolution and expansion (185). As a consequence, current targeted chemotherapies fail to provide considerable survival benefit to the cancer patients due to the lack of systematic targeting of these genetically evolved, selectively expanded clones that escape from the targeted therapies. Therefore, functional targeting of individual oncogenic pathways is unlikely an effective therapeutic strategy; rather, it is more important to investigate the details of genetic divergence in each tumor, to design personalized, targeted combination therapies to avoid the selective enrichment of the insensitive clones. This has been demonstrated by recent efforts on strategic combination therapies that have shown promising potentials as effective therapies (144, 187). In addition, antibody-mediated specific cytotoxic targeting that directly and rapidly kills the cancer cells may effectively suppress clonal expansion and overcome drug resistance (65, 165). These ongoing studies strongly imply that heterogeneity

may be used as a biomarker in clinic for determining personalized therapy (84), and targeting tumor heterogeneity resulted from deregulation of DNA-repair machinery (123, 237) and the gateway of gene activation (93, 263) in cancer cells emerges as a fundamental strategy to kill cancer cells.

V. IMPACT OF DEFECTIVE IMMUNOSURVEILLANCE IN RESISTANCE

It has been known for a long time that tumors are surrounded by diverse types of immune cells that impact tumor progression. Data from clinical studies have revealed that infiltration of certain types of immune cells, such as T helper 2 (Th2), regulatory T cells (Treg), T helper 17 (Th17), as well as macrophages and neutrophils can be associated with a poor prognosis (79, 92, 219, 228, 241), indicating a tumor-promoting role of a deregulated immune response. Tumor-infiltrating lymphocytes (TILs) are directed towards the tumor vicinity through individual chemokine signaling (318). Trafficking of TILs to this region often leads to a biological remodeling of the tumor microenvironment mediated by the local enrichment of secreted factors, which consequently signal to cancer cells to promote or suppress the anti-tumor functions of the TILs (45, 87). In addition to the cancer cell-TIL interaction, TIL-TIL interaction can also contribute to deregulated immune response such as infiltrating CD4⁺ T-cell-mediated macrophage differentiation from M1 to M2 stage (55). At the same time, there is evidence indicating that the infiltration of tumors by CD8⁺ effector cells can improve the prognosis of certain cancer entities. However, upon survival of intrinsic or extrinsic stress, including host immune defense, cancer cells actively disregard the host immunosurveillance and gain tolerance (61). Therefore, extensive studies have been focusing on the characterization of the unique roles of individual type of lymphocytes towards to their clinical value as biomarkers.

This neglect of the immune defense can be governed by T-cell anergy (190) and direct suppression of host immune cell functionality by cancer cells (54, 297). Selected cancer cells that survive environmental stress such as chemo- or radiotherapy are capable of undergoing genetic and/or epigenetic modifications to evade immune detection, an organized process called cancer immunoediting (248). As a consequence, this gain-of-capacity generally increases the malignancy of cancer cells. Under physiological conditions, the immune activity is attenuated through inhibitory signaling networks, among which the members of B7 protein family play a predominant role. B7 proteins, including B7-1, B7-2, B7-H1 (also called programmed cell death ligand 1, PD-L1), and B7-DC (also called programmed death ligand 2, PD-L2), are expressed on the surface of infiltrating lymphocytes with differential preference in distinct cell lineages (90). Negative feed-forward signals direct the inhibition of TCR-mediated proliferation to prevent tissue damage from

unfavorable immune responses through intercellular pairing with their targeted receptors, such as cytotoxic T-lymphocyte-associated protein 4 (CTLA-4, for B7-1 and B7-2) and programmed death 1 protein (PD-1) expressed on T cells, macrophages, dendritic cells (DCs), B cells, and NK cells (39). Under pathological conditions, disruption of this homeostatic regulatory machinery leads to T-cell apoptosis, thereby helping cancer cells to evade an immune attack (208). Advances from recent studies discovered significant basal level of PD-L1/2 on the surface of cancer cells (179, 211). The overexpression of PD-L1/2 empowers cancer cells to subvert the action of the immune system (116, 293). It has been observed to achieve this by blocking the signaling interaction PD-1/PD-L1/2 and B7-1/2/CTLA-4 between cancer cells and host immune cells potentially inducing apoptosis of resistant cancer cells (293, 322). This indicates another resistance mechanism stemming from an unfavorable deactivation of the immune response and possibly triggered by targeted therapies (FIGURE 6). Interestingly, PD-L1 expression is tightly associated with and possibly controlled by PI3K/Akt activation (318). This is consistent with the observation of downregulated PD-L1 expression by specific inhibition of Akt with pharmacological inhibitor MK-2206 in triple-negative breast cancer (179). Although the underlying molecular mechanisms are not yet fully understood, this signaling route seems to be a universal event as it was also reported in prostate cancer (46), mutant BRAF-harboring melanoma (7, 114), pancreatic cancer (336), and human

glioma (210). Furthermore, overexpression of PD-L1 in tumors has been found to correlate with poor clinical prognosis, and its expression status could be useful to predict drug resistance (168, 169).

VI. CURRENT STRATEGIES TO OVERCOME RESISTANCE

Developmental homeostasis requires meticulous and tightly regulated switching of signaling activation. This is commonly achieved through functional interactions between different pathways. A representative module is the interplay between PI3K/Akt, MAPK and the Hippo pathways in which integral interactions of both help maintain proper cell proliferation and organ size. Despite the self-regulated feedback loops (FIGURE 5), inter-inhibitory mechanisms between PI3K/Akt and MAPK signaling were also widely acknowledged (172, 275). Activated Akt can attenuate MAPK signaling by phospho-inhibition of Raf (180, 338), or indirectly through the mTORC1/S6K/IRS1/RAS/MAPK signaling feedback loop. Active ERK can also mediate the inhibition of IRS1 through phospho-activation of raptor-dependent mTORC1/S6K signaling (29) or inhibition of TSC1/2 (159). Such inverse inter-regulation has been confirmed *in vivo* (28). Moreover, with its suppressive function, Hippo signaling is involved in the regulation of PTEN, thus preventing over-activation of PI3K/Akt through p53

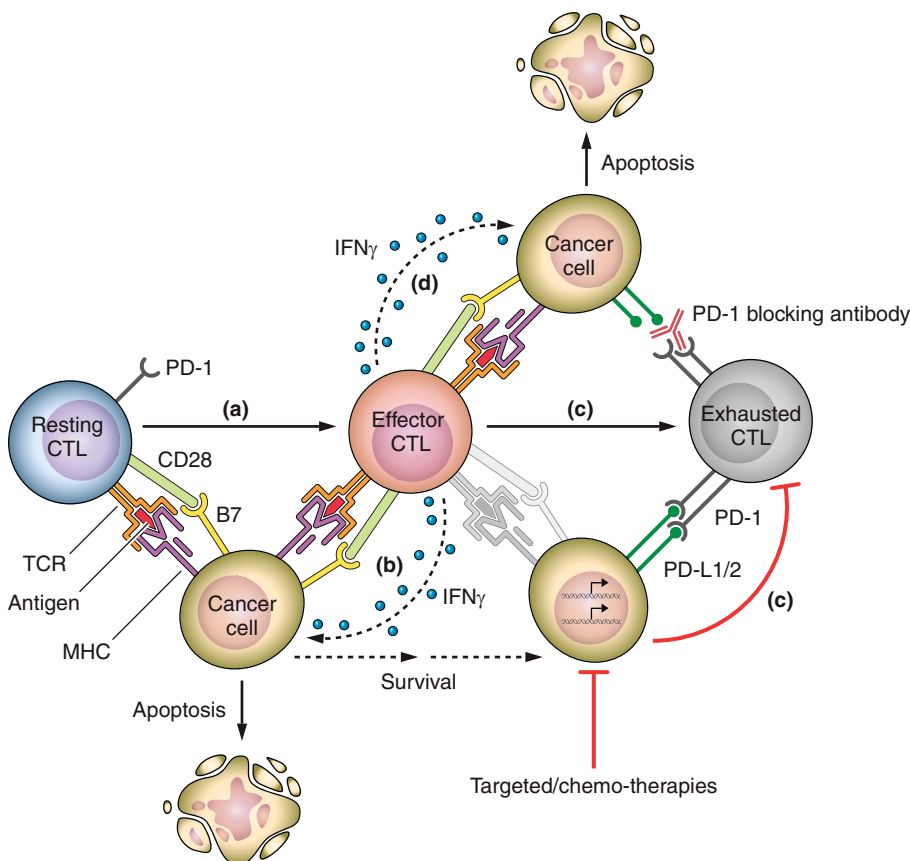


FIGURE 6. Mechanisms of cancer cell-mediated CTL activation and inhibition. Upon activation of CD8⁺ T cells mediated by binding to a cancer cell through TCR/Ag/MHC and CD28/B7 formation, resting CD8⁺ T cells are activated to become effector T cells (a) that release pro-apoptotic molecules like IFN γ that mediate apoptotic death of the vast majority of cancer cells (b). However, cancer cell survivors from T-cell attack may evolve to increase the expression level of B7 family members like PD-L1/2, which binds to PD-1 on the surface of CD8⁺ T cells. This interaction masks the formation of functional complex TCR/Ag/MHC and CD28/B7 and consequently induces T-cell exhaustion to protect cancer cells from immunosurveillance (c). Clinical targeted therapies often upregulate PD-L1/2 expression on the cancer cell surface. When this inhibitory interaction is disrupted with PD-1 blocking antibody, T-cell exhaustion is inhibited, and the killing effect is restored due to reaccumulation of functional cytotoxic T cells (d).

(274, 330, 335). Given the importance of shared signaling nodes downstream of a number of signaling cascades, it is not surprising that a compensatory activation will be triggered upon deactivation of any pathway. In fact, this is frequently reflected by the outcome of drug resistance in clinical monotherapies of cancer. Therefore, combination therapies including bispecific inhibition that simultaneously block compensatory activation of self or parallel signaling axes come to the center stage, and several clinical trials have shown clinical benefits to this over monotherapies. As mentioned above, two phase III studies of coinhibition of mutant BRAF and MEK with dabrafenib and trametinib in melanoma patients increased the overall survival compared with dabrafenib monotherapy (151, 234) (Combi-d trial: ClinicalTrials.gov Identifier NCT01584648; Combi-v trial: ClinicalTrials.gov Identifier NCT01597908). Combination between chemotherapy, trastuzumab, and pertuzumab has also become the standard of care in Her2-positive breast cancer patients (288), and another combined chemotherapy also exhibits significant advances in treating triple negative breast cancer patients (118). Other combinatorial strategies, such as cytotoxic reagents paired with several negative immunologic regulators, have been explored and seem to be promising (101, 138, 158, 229). Certainly, it will be interesting to investigate the therapeutic outcome when targeted kinase inhibition is combined with immunotherapy (105). A number of trials are currently running.

VII. DISCUSSION AND PERSPECTIVE

A. Targeting Oncogenic Mutations

Physiological signaling supports cell proliferation, differentiation, migration, and acute responses to overcome epigenetic and genetic stress. Functional interplay between pro-survival and pro-apoptotic signaling ensures homeostatic development. When the metabolic stability in a cell is interfered with and eventually disrupted, the cell fate becomes uncontrollable, often resulting in an accelerated cell cycle and a high degree of genomic instability that triggers further oncogenic mutations. Therefore, the development-oriented signaling cross-talk is a key factor to be considered for targeted inhibition. The first generation of small RTK inhibitors shares similar mechanistic actions and are generally reversible, such as FDA-approved gefitinib and erlotinib for NSCLC therapy. They competitively bind to the catalytic domain of EGFR to inhibit the phosphorylation of key tyrosine residues in a reversible manner. Although these types of inhibitors are effective, the majority of the patients stop responding to the therapy and inevitably acquire resistance in a rather short period of time. Such resistance can broadly be categorized into two types: “self-activating” and “nonself-activating.” Self-activating resistance results from the reactivation of on-target or on-pathway ele-

ments, while nonself-activating resistance is mediated through alternative targets or pathways. On-target reactivation is often associated with acquired or clonally expanded mutations that desensitize the cancer cells to inhibitors, for instance, T790M mutations of EGFR in gefitinib-resistant NSCLC (206). Similarly, a substantial number of on-target mutations directly linked to resistance have been discovered, such as KIT (97) and ALK (242); on-pathway resistance can be mediated by either activating mutations or genomic amplifications on different components along the same pathway, which is represented by the MEK mutation upon BRAF^{V600E} inhibition (67, 298) and PI3KCA gene amplification in resistance to trastuzumab (15). The second-generation inhibitors (200) aiming to overcome this drawback specialize in irreversible binding to the adjacent sites of the kinase pocket that form stable covalent bond. For example, the second-generation EGFR inhibitor afatinib potently circumvents EGFR^{T790M}-induced resistance to gefitinib (68). Nevertheless, it is also important to point out that such irreversible inhibition may target structurally similar members of the same family, including wild-type kinases. The persistent inactivation of physiological required enzymes with this class of irreversible inhibitors may cause relevant toxicity (121, 323). With the help of next-generation sequencing technology, the spatial and temporal resolution of specific clonal sequences of the tumor is dramatically improved and drives the discovery of genetic alterations that are druggable. Selective targeting of non-self oncogenic targets may substantially reduce toxicity, increase specificity, and avoid emergence of resistance.

B. Targeting Gene Amplifications

As described above, oncogenic amplification without activating mutation is another factor driving drug resistance in cancer (FIGURES 2 AND 4). Amplified genes can be on both the “self-pathway” and on “non-self pathways.” Mechanistically, gene amplification can “mistarget” the physiological feedback loops in the signaling network. To overcome this type of resistance, two major approaches are being explored in clinical studies. The first approach is a “combinatory blockade,” namely, simultaneous targeting of two or three proteins as a cocktail therapy. Clinical data have already shown significant benefit through enhanced anti-tumor potency and delayed drug resistance. Cotargeting several cancer markers is not only applicable to the activation of non-self pathways but also to the activation of the downstream components on the same pathway (37, 104, 152). The other approach is to optimize the drug doses and schedules. It was shown that the dose of imatinib in clinical phase II and III studies can influence on-site occurrence of resistance (18, 19), so the dosing schedule could also potentially be a critical factor to attenuate vemurafenib-triggered resistance in melanoma therapy (50).

C. Targeting Cancer-Directed Immunosuppression

Recent advances in understanding the exhaustion of immune cells in the cancer environment significantly evoke the importance of immunotherapy. With the discovery of cancer cell-directed blockade of immunological checkpoints, in particular by CTLA-4 and PD-1 signaling, the therapeutic strategies have rapidly shifted to immunotherapy. As a consequence, we have seen a large number of trials with anti-CTLA-4 and anti-PD-1/PD-L1 antibodies, which reactivate the anti-tumor responses of the human immune system, resulting in durable and long-lasting responses in some cancer patients. However, similar to the on-targeted therapies, such checkpoints are intrinsically important for homeostatic immune reaction. Thus prolonged activation of a T-cell response potentiates toxicities in many different tissues/organs (306, 307); in some cases this can be severely detrimental to the patient (102). So, although immunotherapy targeting the checkpoints CTLA-4 and PD-1/PD-L1 has convincingly demonstrated a therapeutic benefit in a variety of malignant cancers, further studies on optimizing dose and schedule are required for clinic safety. Currently it is foreseen that, in combination with TKIs, interference with the checkpoint regulation in cancer may not only suppress the activities of the hyperactivated oncoproteins, but also induce anti-tumor memory of the immune system, achieving the synergistic effect from two directions. Although promising, relevant safety issues became evident in a few clinical trials that have been ongoing to evaluate a combination between immunotherapy and chemotherapy or targeted therapy (231). In fact, more observations have emerged that in certain circumstances the resistance to targeted therapies essentially associates with suppressed anti-tumor activity of T cells and results from impaired immune checkpoint controls (169). A number of immunological regulators are dysregulated despite adequate CTLA-4 and PD-1/PD-L1 function. Clearly, future discoveries of novel targetable checkpoint regulators hold the promise to strengthen the immunologic anti-tumor efficacy.

D. Targeting “Histologic Transformation”

It is also suggested that “histologic transformation” may cause acquired resistance. This category mainly includes epithelial-mesenchymal transition (EMT) (267, 299) and phenotypic changes (such as from NSCLC to SCLC) (251). EMT is also a key event during embryonic development at an early stage. Such a transition is crucial for cell migration and differentiation and exhibits high plasticity to facilitate cell fate and organ formation. EMT has been shown to support cancer cell anti-apoptosis and metastasis; therefore, it is also hypothesized to contribute to resistance. Nevertheless, due to its highly dynamic and plastic nature, it is still unclear whether and how EMT mechanistically promotes acquired resistance. In addition, while EMT is dem-

onstrated in established cell lines and animal models, it is much more difficult to be identified in human tumors. Although some of these EMT-drivers do not seem to be important during postnatal development but are upregulated in metastatic cancers (for example, Twist; Ref. 316), it remains to be determined whether they are suitable targets for therapy.

Taken together, a better understanding of the mechanisms of cancer drug resistance is ultimately the driving force to develop novel therapeutic tools in the future.

ACKNOWLEDGMENTS

Address for reprint requests and other correspondence: G. Xue, Dept. of Mechanisms of Cancer, Friedrich Miescher Institute for Biomedical Research, Maulbeerstrasse 66, 4056 Basel, Switzerland (e-mail: gongda.xue@fmi.ch) or Laboratory of Medical Oncology, Dept. of Biomedicine, University Hospital Basel, Hebelstrasse 20, 4031 Basel, Switzerland.

GRANTS

The research projects in the authors’ laboratories are supported by Novartis Research Foundation, Swiss National Science Foundation Grant 31–130838 (to B. A. Hemmings and G. Xue), Swiss Cancer Res Foundation Grants KFS-3170-02-2013 and KFS-3501-08-2014 (to A. Wicki), Associazione Italiana per la Ricerca sul Cancro (A.I.R.C. 5xmille Ref. 12237) P.I. A.F. (to M. Mandalà), Fondazione Cassa di Risparmio di Pistoia & Pescia (ID 154/2014) (to D. Massi), AIRC IG2010-10104DT & IG2013-14201DT (to D. Taverna), and National Natural Science Foundation of China Grant 381570056 (to H. Tang).

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

REFERENCES

1. Agus DB, Akita RW, Fox WD, Lewis GD, Higgins B, Pisacane PI, Lofgren JA, Tindell C, Evans DP, Maiese K, Scher HI, Sliwkowski MX. Targeting ligand-activated ErbB2 signaling inhibits breast and prostate tumor growth. *Cancer Cell* 2: 127–137, 2002.
2. Ahmad AS, Ormiston-Smith N, Sasieni PD. Trends in the lifetime risk of developing cancer in Great Britain: comparison of risk for those born from 1930 to 1960. *Br J Cancer* 112: 943–947, 2015.
3. Alajati A, Guccini I, Pinton S, Garcia-Escudero R, Bernasocchi T, Sarti M, Montani E, Rinaldi A, Montemurro F, Catapano C, Bertoni F, Alimonti A. Interaction of CDCP1 with HER2 enhances HER2-driven tumorigenesis and promotes trastuzumab resistance in breast cancer. *Cell Reports* 11: 564–576, 2015.
4. Alajati A, Sausgruber N, Aceto N, Duss S, Sarret S, Voshol H, Bonenfant D, Bentires-Alj M. Mammary tumor formation and metastasis evoked by a HER2 splice variant. *Cancer Res* 73: 5320–5327, 2013.

5. Ali SM, Alpaugh RK, Downing SR, Stephens PJ, Yu JQ, Wu H, Buell JK, Miller VA, Lipson D, Palmer GA, Ross JS, Cristofanilli M. Response of an ERBB2-mutated inflammatory breast carcinoma to human epidermal growth factor receptor 2-targeted therapy. *J Clin Oncol* 32: e88–91, 2014.
6. Anido J, Scaltriti M, Bech Serra JJ, Santiago Josefát B, Todo FR, Baselga J, Arribas J. Biosynthesis of tumorigenic HER2 C-terminal fragments by alternative initiation of translation. *EMBO J* 25: 3234–3244, 2006.
7. Atefi M, Avramis E, Lassen A, Wong DJ, Robert L, Foulad D, Cerniglia M, Titz B, Chodon T, Graeber TG, Comin-Anduix B, Ribas A. Effects of MAPK and PI3K pathways on PD-L1 expression in melanoma. *Clin Cancer Res* 20: 3446–3457, 2014.
8. Bader AG, Kang S, Zhao L, Vogt PK. Oncogenic PI3K deregulates transcription and translation. *Nature Rev Cancer* 5: 921–929, 2005.
9. Banerji S, Cibulskis K, Rangel-Escareno C, Brown KK, Carter SL, Frederick AM, Lawrence MS, Sivachenko AY, Sougnez C, Zou L, Cortes ML, Fernandez-Lopez JC, Peng S, Ardlie KG, Auclair D, Bautista-Pina V, Duke F, Francis J, Jung J, Maffuz-Aziz A, Onofrio RC, Parkin M, Pho NH, Quintanar-Jurado V, Ramos AH, Rebollar-Vega R, Rodriguez-Cuevas S, Romero-Cordoba SL, Schumacher SE, Stransky N, Thompson KM, Uribe-Figueroa L, Baselga J, Beroukhir R, Polyak K, Sgroi DC, Richardson AL, Jimenez-Sanchez G, Lander ES, Gabriel SB, Garraway LA, Golub TR, Melendez-Zajgla J, Tokar A, Getz G, Hidalgo-Miranda A, Meyerson M. Sequence analysis of mutations and translocations across breast cancer subtypes. *Nature* 486: 405–409, 2012.
10. Bardelli A, Siena S. Molecular mechanisms of resistance to cetuximab and panitumumab in colorectal cancer. *J Clin Oncol* 28: 1254–1261, 2010.
11. Baselga J, Bradbury I, Eidtmann H, Di Cosimo S, de Azambuja E, Aura C, Gomez H, Dinh P, Fauria K, Van Dooren V, Aktan G, Goldhirsch A, Chang TW, Horvath Z, Coccia-Portugal M, Domont J, Tseng LM, Kunz G, Sohn JH, Semiglazov V, Lerzo G, Palacova M, Probachai V, Pusztai L, Untch M, Gelber RD, Piccart-Gebhart M, Neo AST. Lapatinib with trastuzumab for HER2-positive early breast cancer (Neo-ALTTO): a randomised, open-label, multicentre, phase 3 trial. *Lancet* 379: 633–640, 2012.
12. Baselga J, Gelmon KA, Verma S, Wardley A, Conte P, Miles D, Bianchi G, Cortes J, McNally VA, Ross GA, Fumoleau P, Gianni L. Phase II trial of pertuzumab and trastuzumab in patients with human epidermal growth factor receptor 2-positive metastatic breast cancer that progressed during prior trastuzumab therapy. *J Clin Oncol* 28: 1138–1144, 2010.
13. Baselga J, Swain SM. Novel anticancer targets: revisiting ERBB2 and discovering ERBB3. *Nature Rev Cancer* 9: 463–475, 2009.
14. Bedard PL, Hansen AR, Ratain MJ, Siu LL. Tumour heterogeneity in the clinic. *Nature* 501: 355–364, 2013.
15. Berns K, Horlings HM, Hennessy BT, Madiredjo M, Hijmans EM, Beelen K, Linn SC, Gonzalez-Angulo AM, Stemke-Hale K, Hauptmann M, Beijersbergen RL, Mills GB, van de Vijver MJ, Bernards R. A functional genetic approach identifies the PI3K pathway as a major determinant of trastuzumab resistance in breast cancer. *Cancer Cell* 12: 395–402, 2007.
16. Beroukhir R, Mermel CH, Porter D, Wei G, Raychaudhuri S, Donovan J, Barretina J, Boehm JS, Dobson J, Urashima M, Mc Henry KT, Pinchback RM, Ligon AH, Cho YJ, Haery L, Greulich H, Reich M, Winckler W, Lawrence MS, Weir BA, Tanaka KE, Chiang DY, Bass AJ, Loo A, Hoffman C, Prensner J, Liefeld T, Gao Q, Yecies D, Signoretti S, Maher E, Kaye FJ, Sasaki H, Tepper JE, Fletcher JA, Tabernero J, Baselga J, Tsao MS, Demichelis F, Rubin MA, Janne PA, Daly MJ, Nucera C, Levine RL, Ebert BL, Gabriel S, Rustgi AK, Antonescu CR, Ladanyi M, Letai A, Garraway LA, Loda M, Beer DG, True LD, Okamoto A, Pomeroy SL, Singer S, Golub TR, Lander ES, Getz G, Sellers WR, Meyerson M. The landscape of somatic copy-number alteration across human cancers. *Nature* 463: 899–905, 2010.
17. Blackwell KL, Burstein HJ, Storniolo AM, Rugo H, Sledge G, Koehler M, Ellis C, Casey M, Vukelja S, Bischoff J, Baselga J, O'Shaughnessy J. Randomized study of Lapatinib alone or in combination with trastuzumab in women with ErbB2-positive, trastuzumab-refractory metastatic breast cancer. *J Clin Oncol* 28: 1124–1130, 2010.
18. Blanke CD, Demetri GD, von Mehren M, Heinrich MC, Eisenberg B, Fletcher JA, Corless CL, Fletcher CD, Roberts PJ, Heinz D, Wehre E, Nikolova Z, Joensuu H. Long-term results from a randomized phase II trial of standard- versus higher-dose imatinib mesylate for patients with unresectable or metastatic gastrointestinal stromal tumors expressing KIT. *J Clin Oncol* 26: 620–625, 2008.
19. Blanke CD, Rankin C, Demetri GD, Ryan CW, von Mehren M, Benjamin RS, Raymond AK, Bramwell VH, Baker LH, Maki RG, Tanaka M, Hecht JR, Heinrich MC, Fletcher CD, Crowley JJ, Borden EC. Phase III randomized, intergroup trial assessing imatinib mesylate at two dose levels in patients with unresectable or metastatic gastrointestinal stromal tumors expressing the kit receptor tyrosine kinase: S0033. *J Clin Oncol* 26: 626–632, 2008.
20. Bollag G, Tsai J, Zhang J, Zhang C, Ibrahim P, Nolop K, Hirth P. Vemurafenib: the first drug approved for BRAF-mutant cancer. *Nature Rev Drug Discovery* 11: 873–886, 2012.
21. Bose R, Kavuri SM, Searleman AC, Shen W, Shen D, Koboldt DC, Monsey J, Goel N, Aronson AB, Li S, Ma CX, Ding L, Mardis ER, Ellis MJ. Activating HER2 mutations in HER2 gene amplification negative breast cancer. *Cancer Discovery* 3: 224–237, 2013.
22. Burrell RA, McGranahan N, Bartek J, Swanton C. The causes and consequences of genetic heterogeneity in cancer evolution. *Nature* 501: 338–345, 2013.
23. Callahan MK, Rampal R, Harding JJ, Klimek VM, Chung YR, Merghoub T, Wolchok JD, Solit DB, Rosen N, Abdel-Wahab O, Levine RL, Chapman PB. Progression of RAS-mutant leukemia during RAF inhibitor treatment. *N Engl J Med* 367: 2316–2321, 2012.
24. Cameron D, Casey M, Press M, Lindquist D, Pienkowski T, Romieu CG, Chan S, Jagiello-Gruszfeld A, Kaufman B, Crown J, Chan A, Campone M, Viens P, Davidson N, Gorbounova V, Raats JJ, Skarlos D, Newstat B, Roychowdhury D, Paoletti P, Oliva C, Rubin S, Stein S, Geyer CE. A phase III randomized comparison of lapatinib plus capecitabine versus capecitabine alone in women with advanced breast cancer that has progressed on trastuzumab: updated efficacy and biomarker analyses. *Breast Cancer Res Treatment* 112: 533–543, 2008.
25. Camidge DR, Doebele RC. Treating ALK-positive lung cancer—early successes and future challenges. *Nature Rev Clin Oncol* 9: 268–277, 2012.
26. Cancer Genome Atlas. Comprehensive molecular portraits of human breast tumours. *Nature* 490: 61–70, 2012.
27. Capelan M, Pugliano L, De Azambuja E, Bozovic I, Saini KS, Sotiriou C, Loi S, Piccart-Gebhart MJ. Pertuzumab: new hope for patients with HER2-positive breast cancer. *Ann Oncol* 24: 273–282, 2013.
28. Carracedo A, Ma L, Teruya-Feldstein J, Rojo F, Salmena L, Alimonti A, Egia A, Sasaki AT, Thomas G, Kozma SC, Papa A, Nardella C, Cantley LC, Baselga J, Pandolfi PP. Inhibition of mTORC1 leads to MAPK pathway activation through a PI3K-dependent feedback loop in human cancer. *J Clin Invest* 118: 3065–3074, 2008.
29. Carriere A, Romeo Y, Acosta-Jaquez HA, Moreau J, Bonnell E, Thibault P, Fingar DC, Roux PP. ERK1/2 phosphorylate Raptor to promote Ras-dependent activation of mTOR complex 1 (mTORC1). *J Biol Chem* 286: 567–577, 2011.
30. Carvajal-Hausdorf DE, Schalper KA, Pusztai L, Psyrri A, Kologeras KT, Kotoula V, Fountzilas G, Rimm DL. Measurement of domain-specific HER2 (ERBB2) expression may classify benefit from trastuzumab in breast cancer. *J Natl Cancer Inst* 107: 2015.
31. Casalini P, Iorio MV, Galmozzi E, Menard S. Role of HER receptors family in development and differentiation. *J Cell Physiol* 200: 343–350, 2004.
32. Castagnoli L, Iezzi M, Ghedini GC, Ciravolo V, Marzano G, Lamolinara A, Zappasodi R, Gasparini P, Campiglio M, Amici A, Chiodoni C, Palladini A, Lollini PL, Triulzi T, Menard S, Nanni P, Tagliabue E, Pupa SM. Activated d16HER2 homodimers and SRC kinase mediate optimal efficacy for trastuzumab. *Cancer Res* 74: 6248–6259, 2014.
33. Catling AD, Reuter CW, Cox ME, Parsons SJ, Weber MJ. Partial purification of a mitogen-activated protein kinase kinase activator from bovine brain. Identification as B-Raf or a B-Raf-associated activity. *J Biol Chem* 269: 30014–30021, 1994.
34. Chandarlapaty S, Sawai A, Scaltriti M, Rodrik-Outmezguine V, Grbovic-Huezo O, Serra V, Majumder PK, Baselga J, Rosen N. AKT inhibition relieves feedback suppression of receptor tyrosine kinase expression and activity. *Cancer Cell* 19: 58–71, 2011.
35. Chang L, Karin M. Mammalian MAP kinase signalling cascades. *Nature* 410: 37–40, 2001.
36. Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J, Dummer R, Garbe C, Testori A, Maio M, Hogg D, Lorigan P, Lebbe C, Jouary T, Schadendorf D, Ribas A, O'Day SJ, Sosman JA, Kirkwood JM, Eggermont AM, Dreno B, Nolop K, Li J, Nelson B, Hou J, Lee RJ, Flaherty KT, McArthur GA, Group BS. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med* 364: 2507–2516, 2011.

37. Chapman PB, Solit DB, Rosen N. Combination of RAF and MEK inhibition for the treatment of BRAF-mutated melanoma: feedback is not encouraged. *Cancer Cell* 26: 603–604, 2014.
38. Chen B, Tardell C, Higgins B, Packman K, Boylan JF, Niu H. BRAFV600E negatively regulates the AKT pathway in melanoma cell lines. *PLoS One* 7: e42598, 2012.
39. Chen DS, Irving BA, Hodi FS. Molecular pathways: next-generation immunotherapy–inhibiting programmed death-ligand 1 and programmed death-1. *Clin Cancer Res* 18: 6580–6587, 2012.
40. Chen FL, Xia W, Spector NL. Acquired resistance to small molecule ErbB2 tyrosine kinase inhibitors. *Clin Cancer Res* 14: 6730–6734, 2008.
41. Chen WS, Xu PZ, Gottlob K, Chen ML, Sokol K, Shivanova T, Roninson I, Weng W, Suzuki R, Tobe K, Kadowaki T, Hay N. Growth retardation and increased apoptosis in mice with homozygous disruption of the Akt1 gene. *Genes Dev* 15: 2203–2208, 2001.
42. Cho H, Mu J, Kim JK, Thorvaldsen JL, Chu Q, Crenshaw EB 3rd, Kaestner KH, Bartolomei MS, Shulman GI, Birnbaum MJ. Insulin resistance and a diabetes mellitus-like syndrome in mice lacking the protein kinase Akt2 (PKB beta). *Science* 292: 1728–1731, 2001.
43. Cho H, Thorvaldsen JL, Chu Q, Feng F, Birnbaum MJ. Akt1/PKBalpha is required for normal growth but dispensable for maintenance of glucose homeostasis in mice. *J Biol Chem* 276: 38349–38352, 2001.
44. Cooper JA. Transforming mutations in protein-tyrosine kinase genes. *BioEssays* 4: 9–15, 1986.
45. Coussens LM, Pollard JW. Leukocytes in mammary development and cancer. *Cold Spring Harbor Perspect Biol* 3: 2011.
46. Crane CA, Panner A, Murray JC, Wilson SP, Xu H, Chen L, Simko JP, Waldman FM, Pieper RO, Parsa AT. PI(3) kinase is associated with a mechanism of immunoresistance in breast and prostate cancer. *Oncogene* 28: 306–312, 2009.
47. Cuello M, Ettenberg SA, Clark AS, Keane MM, Posner RH, Nau MM, Dennis PA, Lipkowitz S. Down-regulation of the erbB-2 receptor by trastuzumab (herceptin) enhances tumor necrosis factor-related apoptosis-inducing ligand-mediated apoptosis in breast and ovarian cancer cell lines that overexpress erbB-2. *Cancer Res* 61: 4892–4900, 2001.
48. D'Uva G, Aharonov A, Lauriola M, Kain D, Yahalom-Ronen Y, Carvalho S, Weisinger K, Bassat E, Rajchman D, Yifa O, Lysenko M, Konfino T, Hegesh J, Brenner O, Neeman M, Yarden Y, Leor J, Sarig R, Harvey RP, Tzahor E. ERBB2 triggers mammalian heart regeneration by promoting cardiomyocyte dedifferentiation and proliferation. *Nature Cell Biol* 17: 627–638, 2015.
49. D'Uva G, Tzahor E. The key roles of ERBB2 in cardiac regeneration. *Cell Cycle* 14: 2383–2384, 2015.
50. Das Thakur M, Salangsang F, Landman AS, Sellers WR, Pryer NK, Levesque MP, Dummer R, McMahon M, Stuart DD. Modelling vemurafenib resistance in melanoma reveals a strategy to forestall drug resistance. *Nature* 494: 251–255, 2013.
51. Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Clegg S, Teague J, Woffendin H, Garnett MJ, Bottomley W, Davis N, Dicks E, Ewing R, Floyd Y, Gray K, Hall S, Hawes R, Hughes J, Kosmidou V, Menzies A, Mould C, Parker A, Stevens C, Watt S, Hooper S, Wilson R, Jayatilake H, Gusterson BA, Cooper C, Shipley J, Hargrave D, Pritchard-Jones K, Maitland N, Chenevix-Trench G, Riggins GJ, Bigner DD, Palmieri G, Cossu A, Flanagan A, Nicholson A, Ho JW, Leung SY, Yuen ST, Weber BL, Seigler HF, Darrow TL, Paterson H, Marais R, Marshall CJ, Wooster R, Stratton MR, Futreal PA. Mutations of the BRAF gene in human cancer. *Nature* 417: 949–954, 2002.
52. De Azambuja E, Holmes AP, Piccart-Gebhart M, Holmes E, Di Cosimo S, Swaby RF, Untch M, Jackisch C, Lang I, Smith I, Boyle F, Xu B, Barrios CH, Perez EA, Azim HA Jr, Kim SB, Kuemmel S, Huang CS, Vuylsteke P, Hsieh RK, Gorbunova V, Eniu A, Dreosti L, Tavarikiladze N, Gelber RD, Eidmann H, Baselga J. Lapatinib with trastuzumab for HER2-positive early breast cancer (NeoALTTO): survival outcomes of a randomised, open-label, multicentre, phase 3 trial and their association with pathological complete response. *Lancet Oncol* 15: 1137–1146, 2014.
53. De Keizer PL, Packer LM, Szybowska AA, Riedl-Polderman PE, van den Broek NJ, de Bruin A, Dansen TB, Marais R, Brenkman AB, Burgering BM. Activation of forkhead box O transcription factors by oncogenic BRAF promotes p21^{cip1}-dependent senescence. *Cancer Res* 70: 8526–8536, 2010.
54. De Visser KE, Eichten A, Coussens LM. Paradoxical roles of the immune system during cancer development. *Nature Rev Cancer* 6: 24–37, 2006.
55. DeNardo DG, Barreto JB, Andreu P, Vasquez L, Tawfik D, Kolchak N, Coussens LM. CD4(+) T cells regulate pulmonary metastasis of mammary carcinomas by enhancing protumor properties of macrophages. *Cancer Cell* 16: 91–102, 2009.
56. Dexter DL, Leith JT. Tumor heterogeneity and drug resistance. *J Clin Oncol* 4: 244–257, 1986.
57. Dijkers PF, Medema RH, Pals C, Banerji L, Thomas NS, Lam EW, Burgering BM, Raaijmakers JA, Lammers JW, Koenderman L, Coffey PJ. Forkhead transcription factor FKHR-L1 modulates cytokine-dependent transcriptional regulation of p27(KIP1). *Mol Cell Biol* 20: 9138–9148, 2000.
58. Ding L, Ley TJ, Larson DE, Miller CA, Koboldt DC, Welch JS, Ritchey JK, Young MA, Lamprecht T, McLellan MD, McMichael JF, Wallis JW, Lu C, Shen D, Harris CC, Dooling DJ, Fulton RS, Fulton LL, Chen K, Schmidt H, Kalicki-Veizer J, Magrini VJ, Cook L, McGrath SD, Vickery TL, Wendl MC, Heath S, Watson MA, Link DC, Tomasson MH, Shannon WD, Payton JE, Kulkarni S, Westervelt P, Walter MJ, Graubert TA, Mardis ER, Wilson RK, DiPersio JF. Clonal evolution in relapsed acute myeloid leukaemia revealed by whole-genome sequencing. *Nature* 481: 506–510, 2012.
59. Doebele RC, Pilling AB, Aisner DL, Kutateladze TG, Le AT, Weickhardt AJ, Kondo KL, Linderman DJ, Heasley LE, Franklin WA, Varella-Garcia M, Camidge DR. Mechanisms of resistance to crizotinib in patients with ALK gene rearranged non-small cell lung cancer. *Clin Cancer Res* 18: 1472–1482, 2012.
60. Dougherty MK, Muller J, Ritt DA, Zhou M, Zhou XZ, Copeland TD, Conrads TP, Veenstra TD, Lu KP, Morrison DK. Regulation of Raf-1 by direct feedback phosphorylation. *Mol Cell* 17: 215–224, 2005.
61. Drake CG, Jaffee E, Pardoll DM. Mechanisms of immune evasion by tumors. *Adv Immunol* 90: 51–81, 2006.
62. Du X, Shi H, Li J, Dong Y, Liang J, Ye J, Kong S, Zhang S, Zhong T, Yuan Z, Xu T, Zhuang Y, Zheng B, Geng JG, Tao W. Mst1/Mst2 regulate development and function of regulatory T cells through modulation of Foxo1/Foxo3 stability in autoimmune disease. *J Immunol* 192: 1525–1535, 2014.
63. Dummer B, Tschopp O, Hynx D, Yang ZZ, Dirnhofer S, Hemmings BA. Life with a single isoform of Akt: mice lacking Akt2 and Akt3 are viable but display impaired glucose homeostasis and growth deficiencies. *Mol Cell Biol* 26: 8042–8051, 2006.
64. Easton RM, Cho H, Roovers K, Shineman DW, Mizrahi M, Forman MS, Lee VM, Szabolcs M, de Jong R, Oltersdorf T, Ludwig T, Efstratiadis A, Birnbaum MJ. Role for Akt3/protein kinase Bgamma in attainment of normal brain size. *Mol Cell Biol* 25: 1869–1878, 2005.
65. Eisenstein M. Medicine: eyes on the target. *Nature* 527: S110–112, 2015.
66. Ellis MJ, Ding L, Shen D, Luo J, Suman VJ, Wallis JW, Van Tine BA, Hoog J, Goiffon RJ, Goldstein TC, Ng S, Lin L, Crowder R, Snider J, Ballman K, Weber J, Chen K, Koboldt DC, Kandoth C, Schierding WS, McMichael JF, Miller CA, Lu C, Harris CC, McLellan MD, Wendl MC, DeSchryver K, Allred DC, Esserman L, Unzeitig G, Margenthaler J, Babiera GV, Marcom PK, Guenther JM, Leitch M, Hunt K, Olson J, Tao Y, Maher CA, Fulton LL, Fulton RS, Harrison M, Oberkell B, Du F, Demeter R, Vickery TL, Elhammali A, Piwnica-Worms H, McDonald S, Watson M, Dooling DJ, Ota D, Chang LW, Bose R, Ley TJ, Piwnica-Worms D, Stuart JM, Wilson RK, Mardis ER. Whole-genome analysis informs breast cancer response to aromatase inhibition. *Nature* 486: 353–360, 2012.
67. Emery CM, Vijayendran KG, Zipser MC, Sawyer AM, Niu L, Kim JJ, Hatton C, Chopra R, Oberholzer PA, Karpova MB, MacConaill LE, Zhang J, Gray NS, Sellers WR, Dummer R, Garraway LA. MEK1 mutations confer resistance to MEK and B-RAF inhibition. *Proc Natl Acad Sci USA* 106: 20411–20416, 2009.
68. Engelman JA, Zejnullahu K, Gale CM, Lifshits E, Gonzales AJ, Shimamura T, Zhao F, Vincent PW, Naumov GN, Bradner JE, Althaus IW, Gandhi L, Shapiro GI, Nelson JM, Heymach JV, Meyerson M, Wong KK, Janne PA. PF00299804, an irreversible pan-ERBB inhibitor, is effective in lung cancer models with EGFR and ERBB2 mutations that are resistant to gefitinib. *Cancer Res* 67: 11924–11932, 2007.
69. Engelman JA, Zejnullahu K, Mitsudomi T, Song Y, Hyland C, Park JO, Lindeman N, Gale CM, Zhao X, Christensen J, Kosaka T, Holmes AJ, Rogers AM, Cappuzzo F, Mok T, Lee C, Johnson BE, Cantley LC, Janne PA. MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. *Science* 316: 1039–1043, 2007.

70. Erhardt P, Troppmair J, Rapp UR, Cooper GM. Differential regulation of Raf-1 and B-Raf and Ras-dependent activation of mitogen-activated protein kinase by cyclic AMP in PC12 cells. *Mol Cell Biol* 15: 5524–5530, 1995.
71. Erickson SL, O'Shea KS, Ghaboosi N, Loverro L, Frantz G, Bauer M, Lu LH, Moore MW. ErbB3 is required for normal cerebellar and cardiac development: a comparison with ErbB2-and heregulin-deficient mice. *Development* 124: 4999–5011, 1997.
72. Essaghir A, Dif N, Marbehan CY, Coffey PJ, Demoulin JB. The transcription of FOXO genes is stimulated by FOXO3 and repressed by growth factors. *J Biol Chem* 284: 10334–10342, 2009.
73. Esteva FJ, Valero V, Booser D, Guerra LT, Murray JL, Pusztai L, Cristofanilli M, Arun B, Esmali B, Fritsche HA, Sneige N, Smith TL, Hortobagyi GN. Phase II study of weekly docetaxel and trastuzumab for patients with HER-2-overexpressing metastatic breast cancer. *J Clin Oncol* 20: 1800–1808, 2002.
74. Eychene A, Dusanter-Fourt I, Barnier JV, Papin C, Charon M, Gisselbrecht S, Calothy G. Expression and activation of B-Raf kinase isoforms in human and murine leukemia cell lines. *Oncogene* 10: 1159–1165, 1995.
75. Fayard E, Xue G, Parcellier A, Bozovic L, Hemmings BA. Protein kinase B (PKB/Akt), a key mediator of the PI3K signaling pathway. *Curr Top Microbiol Immunol* 346: 31–56, 2010.
76. Fessler SP, Wotkowicz MT, Mahanta SK, Bamdad C. MUC1* is a determinant of trastuzumab (Herceptin) resistance in breast cancer cells. *Breast Cancer Res Treatment* 118: 113–124, 2009.
77. Flaherty KT, Infante JR, Daud A, Gonzalez R, Kefford RF, Sosman J, Hamid O, Schuchter L, Cebon J, Ibrahim N, Kudchadkar R, Burris HA, 3rd Falchook G, Algazi A, Lewis K, Long GV, Puzanov I, Lebowitz P, Singh A, Little S, Sun P, Allred A, Ouellet D, Kim KB, Patel K, Weber J. Combined BRAF and MEK inhibition in melanoma with BRAF V600 mutations. *N Engl J Med* 367: 1694–1703, 2012.
78. Friboulet L, Li N, Katayama R, Lee CC, Gainor JF, Crystal AS, Michellys PY, Awad MM, Yanagitani N, Kim S, Pferdekamper AC, Li J, Kasibhatla S, Sun F, Sun X, Hua S, McNamara P, Mahmood S, Lockerman EL, Fujita N, Nishio M, Harris JL, Shaw AT, Engelman JA. The ALK inhibitor ceritinib overcomes crizotinib resistance in non-small cell lung cancer. *Cancer Discovery* 4: 662–673, 2014.
79. Fridman WH, Pages F, Sautes-Fridman C, Galon J. The immune contexture in human tumours: impact on clinical outcome. *Nature Rev Cancer* 12: 298–306, 2012.
80. Fruman DA, Rommel C. PI3K and cancer: lessons, challenges and opportunities. *Nature Rev Drug discovery* 13: 140–156, 2014.
81. Garofalo RS, Orena SJ, Rafidi K, Torchia AJ, Stock JL, Hildebrandt AL, Coskran T, Black SC, Brees DJ, Wicks JR, McNeish JD, Coleman KG. Severe diabetes, age-dependent loss of adipose tissue, and mild growth deficiency in mice lacking Akt2/PKB beta. *J Clin Invest* 112: 197–208, 2003.
82. Garrett JT, Olivares MG, Rinehart C, Granja-Ingram ND, Sanchez V, Chakrabarty A, Dave B, Cook RS, Pao W, McKinley E, Manning HC, Chang J, Arteaga CL. Transcriptional and posttranslational up-regulation of HER3 (ErbB3) compensates for inhibition of the HER2 tyrosine kinase. *Proc Natl Acad Sci USA* 108: 5021–5026, 2011.
83. Gassmann M, Casagrande F, Orioli D, Simon H, Lai C, Klein R, Lemke G. Aberrant neural and cardiac development in mice lacking the ErbB4 neuregulin receptor. *Nature* 378: 390–394, 1995.
84. Gebhart G, Lamberts LE, Wimana Z, Garcia C, Emonts P, Ameye L, Stroobants S, Huizing M, Aftimos P, Tol J, Oyen WJ, Vugts DJ, Hoekstra OS, Schroder CP, Menkevan der Houven van Oordt CW, Guiot T, Brouwers AH, Awada A, de Vries EG, Flamen P. Molecular imaging as a tool to investigate heterogeneity of advanced HER2-positive breast cancer and to predict patient outcome under trastuzumab emtansine (T-DM1): the ZEPHIR trial. *Ann Oncol* 27: 619–624, 2016.
85. Gerlinger M, Rowan AJ, Horswell S, Larkin J, Endesfelder D, Gronroos E, Martinez P, Matthews N, Stewart A, Tarpey P, Varela I, Phillimore B, Begum S, McDonald NQ, Butler A, Jones D, Raine K, Latimer C, Santos CR, Nohadani M, Eklund AC, Spencer-Dene B, Clark G, Pickering L, Stamp G, Gore M, Szallasi Z, Downward J, Futreal PA, Swanton C. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N Engl J Med* 366: 883–892, 2012.
86. Geyer CE, Forster J, Lindquist D, Chan S, Romieu CG, Pienkowski T, Jagiello-Gruszfeld A, Crown J, Chan A, Kaufman B, Skarlos D, Campone M, Davidson N, Berger M, Oliva C, Rubin SD, Stein S, Cameron D. Lapatinib plus capecitabine for HER2-positive advanced breast cancer. *N Engl J Med* 355: 2733–2743, 2006.
87. Ghajar CM. On leukocytes in mammary development and cancer. *Cold Spring Harbor Perspect Biol* 4: 2012.
88. Golding JP, Trainor P, Krumlauf R, Gassmann M. Defects in pathfinding by cranial neural crest cells in mice lacking the neuregulin receptor ErbB4. *Nature Cell Biol* 2: 103–109, 2000.
89. Gottlob K, Majewski N, Kennedy S, Kandel E, Robey RB, Hay N. Inhibition of early apoptotic events by Akt/PKB is dependent on the first committed step of glycolysis and mitochondrial hexokinase. *Genes Dev* 15: 1406–1418, 2001.
90. Greenwald RJ, Freeman GJ, Sharpe AH. The B7 family revisited. *Annu Rev Immunol* 23: 515–548, 2005.
91. Greer EL, Dowlatshahi D, Banko MR, Villen J, Hoang K, Blanchard D, Gygi SP, Brunet A. An AMPK-FOXO pathway mediates longevity induced by a novel method of dietary restriction in *C. elegans*. *Curr Biol* 17: 1646–1656, 2007.
92. Gregory AD, Houghton AM. Tumor-associated neutrophils: new targets for cancer therapy. *Cancer Res* 71: 2411–2416, 2011.
93. Hamamoto R, Nakamura Y. Dysregulation of protein methyltransferases in human cancer: an emerging target class for anticancer therapy. *Cancer Sci*. In press.
94. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 144: 646–674, 2011.
95. Hatzivassiliou G, Liu B, O'Brien C, Spoeerke JM, Hoeflich KP, Haverty PM, Soriano R, Forrest WF, Heldens S, Chen H, Toy K, Ha C, Zhou W, Song K, Friedman LS, Amler LC, Hampton GM, Moffat J, Belvin M, Lackner MR. ERK inhibition overcomes acquired resistance to MEK inhibitors. *Mol Cancer Ther* 11: 1143–1154, 2012.
96. Heidorn SJ, Milagre C, Whittaker S, Noury A, Niculescu-Duvas I, Dhomen N, Husain J, Reis-Filho JS, Springer CJ, Pritchard C, Marais R. Kinase-dead BRAF and oncogenic RAS cooperate to drive tumor progression through CRAF. *Cell* 140: 209–221, 2010.
97. Heinrich MC, Maki RG, Corless CL, Antonescu CR, Harlow A, Griffith D, Town A, McKinley A, Ou WB, Fletcher JA, Fletcher CD, Huang X, Cohen DP, Baum CM, Demetri GD. Primary and secondary kinase genotypes correlate with the biological and clinical activity of sunitinib in imatinib-resistant gastrointestinal stromal tumor. *J Clin Oncol* 26: 5352–5359, 2008.
98. Hemmings BA, Restuccia DF. PI3K-PKB/Akt pathway. *Cold Spring Harbor Perspect Biol* 4: a011189, 2012.
99. Heppner GH. Tumor heterogeneity. *Cancer Res* 44: 2259–2265, 1984.
100. Hill MM, Clark SF, Tucker DF, Birnbaum MJ, James DE, Macaulay SL. A role for protein kinase Bbeta/Akt2 in insulin-stimulated GLUT4 translocation in adipocytes. *Mol Cell Biol* 19: 7771–7781, 1999.
101. Hodi FS, Lee S, McDermott DF, Rao UN, Butterfield LH, Tarhini AA, Leming P, Puzanov I, Shin D, Kirkwood JM. Ipilimumab plus sargramostim vs ipilimumab alone for treatment of metastatic melanoma: a randomized clinical trial. *JAMA* 312: 1744–1753, 2014.
102. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, Akerley W, van den Eertwegh AJ, Lutzky J, Lorigan P, Vaubel JM, Linette GP, Hogg D, Ottensmeier CH, Lebbe C, Peschel C, Quirt I, Clark JL, Wolchok JD, Weber JS, Tian J, Yellin MJ, Nichol GM, Hoos A, Urba WJ. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 363: 711–723, 2010.
103. Hsu PP, Kang SA, Rameseder J, Zhang Y, Ottina KA, Lim D, Peterson TR, Choi Y, Gray NS, Yaffe MB, Marto JA, Sabatini DM. The mTOR-regulated phosphoproteome reveals a mechanism of mTORC1-mediated inhibition of growth factor signaling. *Science* 332: 1317–1322, 2011.
104. Hu-Lieskovan S, Mok S, Homet Moreno B, Tsoi J, Robert L, Goedert L, Pinheiro EM, Koya RC, Graeber TG, Comin-Anduix B, Ribas A. Improved antitumor activity of immunotherapy with BRAF and MEK inhibitors in BRAF(V600E) melanoma. *Science Transl Med* 7: 279ra241, 2015.
105. Hu-Lieskovan S, Robert L, Homet Moreno B, Ribas A. Combining targeted therapy with immunotherapy in BRAF-mutant melanoma: promise and challenges. *J Clin Oncol* 32: 2248–2254, 2014.

106. Hu MC, Lee DF, Xia W, Golfman LS, Ou-Yang F, Yang JY, Zou Y, Bao S, Hanada N, Saso H, Kobayashi R, Hung MC. IkkappaB kinase promotes tumorigenesis through inhibition of forkhead FOXO3a. *Cell* 117: 225–237, 2004.
107. Huang X, Gao L, Wang S, McManaman JL, Thor AD, Yang X, Esteve FJ, Liu B. Heterotrimerization of the growth factor receptors erbB2, erbB3, and insulin-like growth factor- α receptor in breast cancer cells resistant to herceptin. *Cancer Res* 70: 1204–1214, 2010.
108. Hubbard SR, Miller WT. Receptor tyrosine kinases: mechanisms of activation and signaling. *Curr Opin Cell Biol* 19: 117–123, 2007.
109. Hynes NE, Lane HA. ERBB receptors and cancer: the complexity of targeted inhibitors. *Nature Rev Cancer* 5: 341–354, 2005.
110. Iwahara T, Fujimoto J, Wen D, Cupples R, Bucay N, Arakawa T, Mori S, Ratzkin B, Yamamoto T. Molecular characterization of ALK, a receptor tyrosine kinase expressed specifically in the nervous system. *Oncogene* 14: 439–449, 1997.
111. Izumi Y, Xu L, di Tomaso E, Fukumura D, Jain RK. Tumour biology: herceptin acts as an anti-angiogenic cocktail. *Nature* 416: 279–280, 2002.
112. Jaiswal RK, Weissinger E, Kolch W, Landreth GE. Nerve growth factor-mediated activation of the mitogen-activated protein (MAP) kinase cascade involves a signaling complex containing B-Raf and HSP90. *J Biol Chem* 271: 23626–23629, 1996.
113. Jerome L, Alami N, Belanger S, Page V, Yu Q, Paterson J, Shiry L, Pegram M, Leyland-Jones B. Recombinant human insulin-like growth factor binding protein 3 inhibits growth of human epidermal growth factor receptor-2-overexpressing breast tumors and potentiates herceptin activity in vivo. *Cancer Res* 66: 7245–7252, 2006.
114. Jiang X, Zhou J, Giobbie-Hurder A, Wargo J, Hodi FS. The activation of MAPK in melanoma cells resistant to BRAF inhibition promotes PD-L1 expression that is reversible by MEK and PI3K inhibition. *Clin Cancer Res* 19: 598–609, 2013.
115. Johannessen CM, Boehm JS, Kim SY, Thomas SR, Wardwell L, Johnson LA, Emery CM, Stransky N, Cogdill AP, Barretina J, Caponigro G, Hieronymus H, Murray RR, Salehi-Ashtiani K, Hill DE, Vidal M, Zhao JJ, Yang X, Alkan O, Kim S, Harris JL, Wilson CJ, Myer VE, Finan PM, Root DE, Roberts TM, Golub T, Flaherty KT, Dummer R, Weber BL, Sellers WR, Schlegel R, Wargo JA, Hahn WC, Garraway LA. COT drives resistance to RAF inhibition through MAP kinase pathway reactivation. *Nature* 468: 968–972, 2010.
116. Jonsson G, Busch C, Knappskog S, Geisler J, Miletic H, Ringner M, Lillehaug JR, Borg A, Lonning PE. Gene expression profiling-based identification of molecular subtypes in stage IV melanomas with different clinical outcome. *Clin Cancer Res* 16: 3356–3367, 2010.
117. Junttila TT, Akita RW, Parsons K, Fields C, Lewis Phillips GD, Friedman LS, Sampath D, Sliwkowski MX. Ligand-independent HER2/HER3/PI3K complex is disrupted by trastuzumab and is effectively inhibited by the PI3K inhibitor GDC-0941. *Cancer Cell* 15: 429–440, 2009.
118. Katakami N, Atagi S, Goto K, Hida T, Horai T, Inoue A, Ichinose Y, Kobayashi K, Takeda K, Kiura K, Nishio K, Seki Y, Ebisawa R, Shahidi M, Yamamoto N. LUX-Lung 4: a phase II trial of afatinib in patients with advanced non-small-cell lung cancer who progressed during prior treatment with erlotinib, gefitinib, or both. *J Clin Oncol* 31: 3335–3341, 2013.
119. Kang JY, Dolled-Filhart M, Ocal IT, Singh B, Lin CY, Dickson RB, Rimm DL, Camp RL. Tissue microarray analysis of hepatocyte growth factor/Met pathway components reveals a role for Met, matrix metalloproteinase, and hepatocyte growth factor activator inhibitor 1 in the progression of node-negative breast cancer. *Cancer Res* 63: 1101–1105, 2003.
120. Karnoub AE, Weinberg RA. Ras oncogenes: split personalities. *Nature Rev Mol Cell Biol* 9: 517–531, 2008.
121. Katakami N, Atagi S, Goto K, Hida T, Horai T, Inoue A, Ichinose Y, Kobayashi K, Takeda K, Kiura K, Nishio K, Seki Y, Ebisawa R, Shahidi M, Yamamoto N. LUX-Lung 4: a phase II trial of afatinib in patients with advanced non-small-cell lung cancer who progressed during prior treatment with erlotinib, gefitinib, or both. *J Clin Oncol* 31: 3335–3341, 2013.
122. Katayama R, Shaw AT, Khan TM, Mino-Kenudson M, Solomon BJ, Halmos B, Jessop NA, Wain JC, Yeo AT, Benes C, Drew L, Saeh JC, Crosby K, Sequist LV, Iafrate AJ, Engelman JA. Mechanisms of acquired crizotinib resistance in ALK-rearranged lung cancers. *Sci Transl Med* 4: 120ra117, 2012.
123. Kelley MR, Logsdon D, Fishel ML. Targeting DNA repair pathways for cancer treatment: what's new? *Future Oncol* 10: 1215–1237, 2014.
124. Kemper K, Krijgsman O, Cornelissen-Steijger P, Shahrabi A, Weeber F, Song JY, Kuilman T, Vis DJ, Wessels LF, Voest EE, Schumacher TN, Blank CU, Adams DJ, Haanen JB, Peepers DS. Intra- and inter-tumor heterogeneity in a vemurafenib-resistant melanoma patient and derived xenografts. *EMBO Mol Med* 7: 1104–1118, 2015.
125. Kloet DE, Burgering BM. The PKB/FOXO switch in aging and cancer. *Biochim Biophys Acta* 1813: 1926–1937, 2011.
126. Kobayashi S, Boggon TJ, Dayaram T, Janne PA, Kocher O, Meyerson M, Johnson BE, Eck MJ, Tenen DG, Halmos B. EGFR mutation and resistance of non-small-cell lung cancer to gefitinib. *N Engl J Med* 352: 786–792, 2005.
127. Kolch W. Coordinating ERK/MAPK signalling through scaffolds and inhibitors. *Nature Rev Mol Cell Biol* 6: 827–837, 2005.
128. Koren S, Reavie L, Couto JP, De Silva D, Stadler MB, Roloff T, Britschgi A, Eichlsberger T, Kohler H, Aina O, Cardiff RD, Bentires-Alj M. PIK3CA(H1047R) induces multipotency and multi-lineage mammary tumours. *Nature* 525: 114–118, 2015.
129. Kornblum HI, Hussain R, Wiesen J, Miettinen P, Zurcher SD, Chow K, Derynck R, Werb Z. Abnormal astrocyte development and neuronal death in mice lacking the epidermal growth factor receptor. *J Neurosci Res* 53: 697–717, 1998.
130. Krause DS, Van Etten RA. Tyrosine kinases as targets for cancer therapy. *N Engl J Med* 353: 172–187, 2005.
131. Kris MG, Natale RB, Herbst RS, Lynch TJ Jr, Prager D, Belani CP, Schiller JH, Kelly K, Spiridonidis H, Sandler A, Albain KS, Cella D, Wolf MK, Averbuch SD, Ochs JJ, Kay AC. Efficacy of gefitinib, an inhibitor of the epidermal growth factor receptor tyrosine kinase, in symptomatic patients with non-small cell lung cancer: a randomized trial. *JAMA* 290: 2149–2158, 2003.
132. Kupriyanova TA, Kandror KV. Akt-2 binds to Glut4-containing vesicles and phosphorylates their component proteins in response to insulin. *J Biol Chem* 274: 1458–1464, 1999.
133. Kwak EL, Bang YJ, Camidge DR, Shaw AT, Solomon B, Maki RG, Ou SH, Dezube BJ, Janne PA, Costa DB, Varella-Garcia M, Kim WH, Lynch TJ, Fidias P, Stubbs H, Engelmann JA, Sequist LV, Tan W, Gandhi L, Mino-Kenudson M, Wei GC, Shreeve SM, Ratain MJ, Settleman J, Christensen JG, Haber DA, Wilner K, Salgia R, Shapiro GI, Clark JW, Iafrate AJ. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 363: 1693–1703, 2010.
134. Kwak EL, Sordella R, Bell DW, Godin-Heymann N, Okimoto RA, Brannigan BW, Harris PL, Driscoll DR, Fidias P, Lynch TJ, Rabindran SK, McGinnis JP, Wissner A, Sharma SV, Isselbacher KJ, Settleman J, Haber DA. Irreversible inhibitors of the EGF receptor may circumvent acquired resistance to gefitinib. *Proc Natl Acad Sci USA* 102: 7665–7670, 2005.
135. Laplante M, Sabatini DM. mTOR signaling. *Cold Spring Harbor Perspect Biol* 4: a011593, 2012.
136. Laplante M, Sabatini DM. mTOR signaling at a glance. *J Cell Sci* 122: 3589–3594, 2009.
137. Laplante M, Sabatini DM. mTOR signaling in growth control and disease. *Cell* 149: 274–293, 2012.
138. Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao CD, Schadendorf D, Dummer R, Smylie M, Rutkowski P, Ferrucci PF, Hill A, Wagstaff J, Carlini MS, Haanen JB, Maio M, Marquez-Rodas I, McArthur GA, Ascierto PA, Long GV, Callahan MK, Postow MA, Grossmann K, Sznol M, Dreno B, Bastholt L, Yang A, Rolin LM, Horak C, Hodi FS, Wolchok JD. Combined nivolumab and ipilimumab or monotherapy in untreated melanoma. *N Engl J Med* 373: 23–34, 2015.
139. Lee-Hoeflich ST, Crocker L, Yao E, Pham T, Munroe X, Hoeflich KP, Sliwkowski MX, Stern HM. A central role for HER3 in HER2-amplified breast cancer: implications for targeted therapy. *Cancer Res* 68: 5878–5887, 2008.
140. Lee KF, Simon H, Chen H, Bates B, Hung MC, Hauser C. Requirement for neuregulin receptor erbB2 in neural and cardiac development. *Nature* 378: 394–398, 1995.
141. Lehtinen MK, Yuan Z, Boag PR, Yang Y, Villen J, Becker EB, DiBacco S, de la Iglesia N, Gygi S, Blackwell TK, Bonni A. A conserved MST-FOXO signaling pathway mediates oxidative-stress responses and extends life span. *Cell* 125: 987–1001, 2006.
142. Lemmon MA. Membrane recognition by phospholipid-binding domains. *Nature Rev Mol Cell Biol* 9: 99–111, 2008.

143. Li B, Meng Y, Zheng L, Zhang X, Tong Q, Tan W, Hu S, Li H, Chen Y, Song J, Zhang G, Zhao L, Zhang D, Hou S, Qian W, Guo Y. Bispecific antibody to ErbB2 overcomes trastuzumab resistance through comprehensive blockade of ErbB2 heterodimerization. *Cancer Res* 73: 6471–6483, 2013.
144. Li JY, Perry SR, Muniz-Medina V, Wang X, Wetzel LK, Rebelatto MC, Hinrichs MJ, Bezabeh BZ, Fleming RL, Dimasi N, Feng H, Toader D, Yuan AQ, Xu L, Lin J, Gao C, Wu H, Dixit R, Osbourn JK, Coats SR. A biparatopic HER2-targeting antibody-drug conjugate induces tumor regression in primary models refractory to or ineligible for HER2-targeted therapy. *Cancer Cell* 29: 117–129, 2016.
145. Li X, Maretzky T, Weskamp G, Monette S, Qing X, Issuree PD, Crawford HC, McIlwain DR, Mak TW, Salmon JE, Blobel CP. *rRhoms 1 and 2* are essential upstream regulators of ADAM17-dependent EGFR signaling. *Proc Natl Acad Sci USA* 112: 6080–6085, 2015.
146. Lin W, Sanchez HB, Deerinck T, Morris JK, Ellisman M, Lee KF. Aberrant development of motor axons and neuromuscular synapses in erbB2-deficient mice. *Proc Natl Acad Sci USA* 97: 1299–1304, 2000.
147. Lipson D, Capelletti M, Yelensky R, Otto G, Parker A, Jarosz M, Curran JA, Balasubramanian S, Bloom T, Brennan KW, Donahue A, Downing SR, Frampton GM, Garcia L, Juhn F, Mitchell KC, White E, White J, Zwirko Z, Peretz T, Nechushtan H, Soussan-Gutman L, Kim J, Sasaki H, Kim HR, Park SI, Ercan D, Sheehan CE, Ross JS, Cronin MT, Janne PA, Stephens PJ. Identification of new ALK and RET gene fusions from colorectal and lung cancer biopsies. *Nature Med* 18: 382–384, 2012.
148. Lito P, Pratilas CA, Joseph EW, Tadi M, Halilovic E, Zubrowski M, Huang A, Wong WL, Callahan MK, Merghoub T, Wolchok JD, de Stanchina E, Chandraratna S, Poulikakos PI, Fagin JA, Rosen N. Relief of profound feedback inhibition of mitogenic signaling by RAF inhibitors attenuates their activity in BRAFV600E melanomas. *Cancer Cell* 22: 668–682, 2012.
149. Liu B, Fan Z, Edgerton SM, Yang X, Lind SE, Thor AD. Potent anti-proliferative effects of metformin on trastuzumab-resistant breast cancer cells via inhibition of erbB2/IGF-1 receptor interactions. *Cell Cycle* 10: 2959–2966, 2011.
150. Logue JS, Morrison DK. Complexity in the signaling network: insights from the use of targeted inhibitors in cancer therapy. *Genes Dev* 26: 641–650, 2012.
151. Long GV, Stroyakovskiy D, Gogas H, Levchenko E, de Braud F, Larkin J, Garbe C, Jouary T, Hauschild A, Grob JJ, Chiarion-Sileni V, Lebbe C, Mandala M, Millward M, Arance A, Bondarenko I, Haanen JB, Hansson J, Utikal J, Ferraresi V, Kovalenko N, Mohr P, Probachai V, Schadendorf D, Nathan P, Robert C, Ribas A, DeMarini DJ, Irani JG, Swann S, Legos JJ, Jin F, Mookerjee B, Flaherty K. Dabrafenib and trametinib versus dabrafenib and placebo for Val600 BRAF-mutant melanoma: a multicentre, double-blind, phase 3 randomised controlled trial. *Lancet* 386: 444–451, 2015.
152. Long GV, Stroyakovskiy D, Gogas H, Levchenko E, de Braud F, Larkin J, Garbe C, Jouary T, Hauschild A, Grob JJ, Chiarion-Sileni V, Lebbe C, Mandala M, Millward M, Arance A, Bondarenko I, Haanen JB, Hansson J, Utikal J, Ferraresi V, Kovalenko N, Mohr P, Probachai V, Schadendorf D, Nathan P, Robert C, Ribas A, DeMarini DJ, Irani JG, Casey M, Ouellet D, Martin AM, Le N, Patel K, Flaherty K. Combined BRAF and MEK inhibition versus BRAF inhibition alone in melanoma. *N Engl J Med* 371: 1877–1888, 2014.
153. Lovly CM, McDonald NT, Chen H, Ortiz-Cuaran S, Heukamp LC, Yan Y, Florin A, Ozretic L, Lim D, Wang L, Chen Z, Chen X, Lu P, Paik PK, Shen R, Jin H, Buettner R, Ansen S, Perner S, Brockmann M, Bos M, Wolf J, Gardizi M, Wright GM, Solomon B, Russell PA, Rogers TM, Suehara Y, Red-Brewer M, Tieu R, de Stanchina E, Wang Q, Zhao Z, Johnson DH, Horn L, Wong KK, Thomas RK, Ladanyi M, Pao W. Rationale for co-targeting IGF-1R and ALK in ALK fusion-positive lung cancer. *Nature Med* 20: 1027–1034, 2014.
154. Lu Y, Zi X, Zhao Y, Mascarenhas D, Pollak M. Insulin-like growth factor-1 receptor signaling and resistance to trastuzumab (Herceptin). *J Natl Cancer Inst* 93: 1852–1857, 2001.
155. Luetette NC, Qiu TH, Fenton SE, Troyer KL, Riedel RF, Chang A, Lee DC. Targeted inactivation of the EGF and amphiregulin genes reveals distinct roles for EGF receptor ligands in mouse mammary gland development. *Development* 126: 2739–2750, 1999.
156. Luetette NC, Qiu TH, Peiffer RL, Oliver P, Smithies O, Lee DC. TGF alpha deficiency results in hair follicle and eye abnormalities in targeted and waved-1 mice. *Cell* 73: 263–278, 1993.
157. Lynch TJ, Bell DW, Sordella R, Gurubhagavata S, Okimoto RA, Brannigan BW, Harris PL, Haserlat SM, Supko JG, Haluska FG, Louis DN, Christiani DC, Settleman J, Haber DA. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med* 350: 2129–2139, 2004.
158. Lynch TJ, Bondarenko I, Luft A, Serwatowski P, Barlesi F, Chacko R, Sebastian M, Neal J, Lu H, Cuillerot JM, Reck M. Ipilimumab in combination with paclitaxel and carboplatin as first-line treatment in stage IIIB/IV non-small-cell lung cancer: results from a randomized, double-blind, multicenter phase II study. *J Clin Oncol* 30: 2046–2054, 2012.
159. Ma L, Chen Z, Erdjument-Bromage H, Tempst P, Pandolfi PP. Phosphorylation and functional inactivation of TSC2 by Erk implications for tuberous sclerosis and cancer pathogenesis. *Cell* 121: 179–193, 2005.
160. Majewski JJ, Nuciforo P, Mittemperger L, Bosma AJ, Eidtmann H, Holmes E, Sotiriou C, Fumagalli D, Jimenez J, Aura C, Prudkin L, Diaz-Delgado MC, de la Pena L, Loi S, Ellis C, Schultz N, de Azambuja E, Harbeck N, Piccart-Gebhart M, Bernards R, Baselga J. PIK3CA mutations are associated with decreased benefit to neoadjuvant human epidermal growth factor receptor 2-targeted therapies in breast cancer. *J Clin Oncol* 33: 1334–1339, 2015.
161. Mancini M, Gaborit N, Lindzen M, Meir Salame T, Dall’Ora M, Sevilla-Sharon M, Abdul-Hai A, Downward J, Yarden Y. Combining three antibodies nullifies feedback-mediated resistance to erlotinib in lung cancer. *Science Signaling* 8: ra53, 2015.
162. Manning BD. Balancing Akt with S6K: implications for both metabolic diseases and tumorigenesis. *J Cell Biol* 167: 399–403, 2004.
163. Marais R, Light Y, Paterson HF, Mason CS, Marshall CJ. Differential regulation of Raf-1, A-Raf, and B-Raf by oncogenic ras and tyrosine kinases. *J Biol Chem* 272: 4378–4383, 1997.
164. Marr MT 2nd, D’Alessio JA, Puig O, Tjian R. IRES-mediated functional coupling of transcription and translation amplifies insulin receptor feedback. *Genes Dev* 21: 175–183, 2007.
165. Martinez MT, Perez-Fidalgo JA, Martin-Martorell P, Cejalvo JM, Pons V, Bermejo B, Martin M, Albanell J, Lluch A. Treatment of HER2 positive advanced breast cancer with T-DMI: a review of the literature. *Crit Rev Oncol Hematol* 97: 96–106, 2016.
166. Marusyk A, Tabassum DP, Altmann PM, Almendro V, Michor F, Polyak K. Non-cell-autonomous driving of tumour growth supports sub-clonal heterogeneity. *Nature* 514: 54–58, 2014.
167. Mason CS, Springer CJ, Cooper RG, Superti-Furga G, Marshall CJ, Marais R. Serine and tyrosine phosphorylations cooperate in Raf-1, but not B-Raf activation. *EMBO J* 18: 2137–2148, 1999.
168. Massi D, Brusa D, Merelli B, Ciano M, Audrito V, Serra S, Buonincontri R, Baroni G, Nassini R, Minocci D, Cattaneo L, Tamborini E, Carobbio A, Rulli E, Deaglio S, Mandala M. PD-L1 marks a subset of melanomas with a shorter overall survival and distinct genetic and morphological characteristics. *Ann Oncol* 25: 2433–2442, 2014.
169. Massi D, Brusa D, Merelli B, Falcone C, Xue G, Carobbio A, Nassini R, Baroni G, Tamborini E, Cattaneo L, Audrito V, Deaglio S, Mandala M. The status of PD-L1 and tumor-infiltrating immune cells predict resistance and poor prognosis in BRAFi-treated melanoma patients harboring mutant BRAFV600. *Ann Oncol* 26: 1980–1987, 2015.
170. Medema RH, Kops GJ, Bos JL, Burgering BM. AFX-like Forkhead transcription factors mediate cell-cycle regulation by Ras and PKB through p27kip1. *Nature* 404: 782–787, 2000.
171. Mei Y, Wang Z, Zhang L, Zhang Y, Li X, Liu H, Ye J, You H. Regulation of neuroblastoma differentiation by forkhead transcription factors FOXO1/3/4 through the receptor tyrosine kinase PDGFRA. *Proc Natl Acad Sci USA* 109: 4898–4903, 2012.
172. Mendoza MC, Er EE, Blenis J. The Ras-ERK and PI3K-mTOR pathways: cross-talk and compensation. *Trends Biochem Sci* 36: 320–328, 2011.
173. Mercer K, Giblett S, Oakden A, Brown J, Marais R, Pritchard C. A-Raf and Raf-1 work together to influence transient ERK phosphorylation and G1/S cell cycle progression. *Oncogene* 24: 5207–5217, 2005.
174. Meyer D, Birchmeier C. Multiple essential functions of neuregulin in development. *Nature* 378: 386–390, 1995.

175. Miettinen PJ, Berger JE, Meneses J, Phung Y, Pedersen RA, Werb Z, Derynck R. Epithelial immaturity and multiorgan failure in mice lacking epidermal growth factor receptor. *Nature* 376: 337–341, 1995.
176. Mieulet V, Lamb RF. Tuberous sclerosis complex: linking cancer to metabolism. *Trends Mol Med* 16: 329–335, 2010.
177. Mitra D, Brumlik MJ, Okamgba SU, Zhu Y, Duplessis TT, Parvani JG, Lesko SM, Brogi E, Jones FE. An oncogenic isoform of HER2 associated with locally disseminated breast cancer and trastuzumab resistance. *Mol Cancer Ther* 8: 2152–2162, 2009.
178. Mitsudomi T, Kosaka T, Endoh H, Horio Y, Hida T, Mori S, Hatooka S, Shinoda M, Takahashi T, Yatabe Y. Mutations of the epidermal growth factor receptor gene predict prolonged survival after gefitinib treatment in patients with non-small-cell lung cancer with postoperative recurrence. *J Clin Oncol* 23: 2513–2520, 2005.
179. Mittendorf EA, Philips AV, Meric-Bernstam F, Qiao N, Wu Y, Harrington S, Su X, Wang Y, Gonzalez-Angulo AM, Akcakanat A, Chawla A, Curran M, Hwu P, Sharma P, Litton JK, Mollndrem JJ, Alatrash G. PD-L1 expression in triple-negative breast cancer. *Cancer Immunol Res* 2: 361–370, 2014.
180. Moelling K, Schad K, Bosse M, Zimmermann S, Schweneker M. Regulation of Raf-Akt Cross-talk. *J Biol Chem* 277: 31099–31106, 2002.
181. Mok TS, Wu YL, Thongprasert S, Yang CH, Chu DT, Saijo N, Sunpawaravong P, Han B, Margono B, Ichinose Y, Nishiwaki Y, Ohe Y, Yang JJ, Chewaskulyong B, Jiang H, Duffield EL, Watkins CL, Armour AA, Fukuoka M. Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. *N Engl J Med* 361: 947–957, 2009.
182. Molina MA, Codony-Servat J, Albanell J, Rojo F, Arribas J, Baselga J. Trastuzumab (herceptin), a humanized anti-Her2 receptor monoclonal antibody, inhibits basal and activated Her2 ectodomain cleavage in breast cancer cells. *Cancer Res* 61: 4744–4749, 2001.
183. Moodie SA, Paris MJ, Kolch W, Wolfman A. Association of MEK1 with p21ras. GMP-PNP is dependent on B-Raf. *Mol Cell Biol* 14: 7153–7162, 1994.
184. Morris EJ, Jha S, Restaino CR, Dayananth P, Zhu H, Cooper A, Carr D, Deng Y, Jin W, Black S, Long B, Liu J, Dinunzio E, Windsor W, Zhang R, Zhao S, Angagaw MH, Pinheiro EM, Desai J, Xiao L, Shipps G, Hruza A, Wang J, Kelly J, Paliwal S, Gao X, Babu BS, Zhu L, Daublain P, Zhang L, Lutterbach BA, Pelletier MR, Philippar U, Silphaiwanh P, Witter D, Kirschmeier P, Bishop WR, Hicklin D, Gilliland DG, Jayaraman L, Zawel L, Fawell S, Samatar AA. Discovery of a novel ERK inhibitor with activity in models of acquired resistance to BRAF and MEK inhibitors. *Cancer Discovery* 3: 742–750, 2013.
185. Morrissy AS, Garzia L, Shih DJ, Zuyderduyn S, Huang X, Skowron P, Remke M, Cavalli FM, Ramaswamy V, Lindsay PE, Jelveh S, Donovan LK, Wang X, Luu B, Zayne K, Li Y, Mayh C, Thiessen N, Mercier E, Mungall KL, Ma Y, Tse K, Zeng T, Shumansky K, Rtoaj, Shah S, Farooq H, Kijima N, Holgado BL, Lee JJ, Matan-Lithwick S, Liu J, Mack SC, Manno A, Michealraj KA, Nor C, Peacock J, Qin L, Reimand J, Rolider A, Thompson YY, Wu X, Pugh T, Ally A, Bilenky M, Butterfield YS, Carlsen R, Cheng Y, Chuah E, Corbett RD, Dhalla N, He A, Lee D, Li H, Long W, Mayo M, Plettner P, Qian JQ, Schein JE, Tam A, Wong T, Birol I, Zhao Y, Faria CC, Pimentel J, Nunes S, Shalaby T, Grotzer M, Pollack IF, Hamilton RL, Li XN, Bendel AE, Fults DW, Walter AW, Kumabe T, Tominaga T, Collins VP, Cho YJ, Hoffman C, Lyden D, Wisoff JH, Garvin JH, Stearns DS, Massimi L, Schuller U, Sterba J, Zitterbart K, Puget S, Ayrault O, Dunn SE, Tirapelli DP, Carlotti CG, Wheeler H, Hallahan AR, Ingram W, MacDonald TJ, Olson JJ, Van Meir EG, Lee JY, Wang KC, Kim SK, Cho BK, Pietsch T, Fleischhack G, Tippelt S, Ra YS, Bailey S, Lindsey JC, Clifford SC, Eberhart CG, Cooper MK, Packer RJ, Massimino M, Garre ML, Bartels U, Tabori U, Hawkins CE, Dirks P, Bouffet E, Rutka JT, Wechsler-Reya RJ, Weiss WA, Collier LS, Dupuy AJ, Korshunov A, Jones DT, Kool M, Northcott PA, Pfister SM, Largaespada DA, Mungall AJ, Moore RA, Jabado N, Bader GD, Jones SJ, Malkin D, Marra MA, Taylor MD. Divergent clonal selection dominates medulloblastoma at recurrence. *Nature*. In press.
186. Mullard A. 2014 FDA drug approvals. *Nature Rev Drug Discovery* 14: 77–81, 2015.
187. Muller P, Kreuzaler M, Khan T, Thommen DS, Martin K, Glatz K, Savic S, Harbeck N, Nitz U, Gluz O, von Bergwelt-Baildon M, Kreipe H, Reddy S, Christgen M, Zippelius A. Trastuzumab emtansine (T-DM1) renders HER2+ breast cancer highly susceptible to CTLA-4/PD-1 blockade. *Science Transl Med* 7: 315ra188, 2015.
188. Muranen T, Selfors LM, Worster DT, Iwanicki MP, Song L, Morales FC, Gao S, Mills GB, Brugge JS. Inhibition of PI3K/mTOR leads to adaptive resistance in matrix-attached cancer cells. *Cancer Cell* 21: 227–239, 2012.
189. Muss HB, Thor AD, Berry DA, Kute T, Liu ET, Koerner F, Cirrincione CT, Budman DR, Wood WC, Barcos M. c-erbB-2 expression and response to adjuvant therapy in women with node-positive early breast cancer. *N Engl J Med* 330: 1260–1266, 1994.
190. Nagaraj S, Gabrilovich DI. Tumor escape mechanism governed by myeloid-derived suppressor cells. *Cancer Res* 68: 2561–2563, 2008.
191. Nagata Y, Lan KH, Zhou X, Tan M, Esteva FJ, Sahin AA, Klos KS, Li P, Monia BP, Nguyen NT, Hortobagyi GN, Hung MC, Yu D. PTEN activation contributes to tumor inhibition by trastuzumab, and loss of PTEN predicts trastuzumab resistance in patients. *Cancer Cell* 6: 117–127, 2004.
192. Nagy P, Friedlander E, Tanner M, Kapanen AI, Carraway KL, Isola J, Jovin TM. Decreased accessibility and lack of activation of ErbB2 in JIMT-1, a herceptin-resistant, MUC4-expressing breast cancer cell line. *Cancer Res* 65: 473–482, 2005.
193. Nazarian R, Shi H, Wang Q, Kong X, Koya RC, Lee H, Chen Z, Lee MK, Attar N, Sazegar H, Chodon T, Nelson SF, McArthur G, Sosman JA, Ribas A, Lo RS. Melanomas acquire resistance to B-RAF(V600E) inhibition by RTK or N-RAS upregulation. *Nature* 468: 973–977, 2010.
194. Negro A, Brar BK, Lee KF. Essential roles of Her2/erbB2 in cardiac development and function. *Recent Prog Hormone Res* 59: 1–12, 2004.
195. Ng CK, Schultheis AM, Bidard FC, Weigelt B, Reis-Filho JS. Breast cancer genomics from microarrays to massively parallel sequencing: paradigms and new insights. *J Natl Cancer Inst* 107: djv015, 2015.
196. Nomura M, Shigematsu H, Li L, Suzuki M, Takahashi T, Estess P, Siegelman M, Feng Z, Kato H, Marchetti A, Shay JW, Spitz MR, Wistuba II, Minna JD, Gazdar AF. Polymorphisms, mutations, and amplification of the EGFR gene in non-small cell lung cancers. *PLoS Med* 4: e125, 2007.
197. Normanno N, Rachiglio AM, Lambiasi M, Martinelli E, Fenizia F, Esposito C, Roma C, Troiani T, Rizzi D, Tatangelo F, Botti G, Maiello E, Colucci G, Ciardiello F, CG Investigators. Heterogeneity of KRAS, NRAS, BRAF and PIK3CA mutations in metastatic colorectal cancer and potential effects on therapy in the CAPRI GOIM trial. *Ann Oncol* 26: 1710–1714, 2015.
198. O'Brien NA, McDonald K, Tong L, von Euw E, Kalous O, Conklin D, Hurvitz SA, di Tomaso E, Schnell C, Linnartz R, Finn RS, Hirawat S, Slamon DJ. Targeting PI3K/mTOR overcomes resistance to HER2-targeted therapy independent of feedback activation of AKT. *Clin Cancer Res* 20: 3507–3520, 2014.
199. O'Donovan KJ, Ma K, Guo H, Wang C, Sun F, Han SB, Kim H, Wong JK, Charron J, Zou H, Son YJ, He Z, Zhong J. B-RAF kinase drives developmental axon growth and promotes axon regeneration in the injured mature CNS. *J Exp Med* 211: 801–814, 2014.
200. Ou SH. Second-generation irreversible epidermal growth factor receptor (EGFR) tyrosine kinase inhibitors (TKIs): a better mousetrap? A review of the clinical evidence. *Crit Rev Oncol Hematol* 83: 407–421, 2012.
201. Paez JG, Janne PA, Lee JC, Tracy S, Greulich H, Gabriel S, Herman P, Kaye FJ, Lindeman N, Boggon TJ, Naoki K, Sasaki H, Fujii Y, Eck MJ, Sellers WR, Johnson BE, Meyerson M. EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science* 304: 1497–1500, 2004.
202. Paik S, Bryant J, Park C, Fisher B, Tan-Chiu E, Hyams D, Fisher ER, Lippman ME, Wickerham DL, Wolmark N. erbB-2 and response to doxorubicin in patients with axillary lymph node-positive, hormone receptor-negative breast cancer. *J Natl Cancer Inst* 90: 1361–1370, 1998.
203. Palazuelos J, Crawford HC, Klingener M, Sun B, Karelis J, Raines EW, Aguirre A. TACE/ADAM17 is essential for oligodendrocyte development and CNS myelination. *J Neurosci* 34: 11884–11896, 2014.
204. Pao W, Chmielecki J. Rational, biologically based treatment of EGFR-mutant non-small-cell lung cancer. *Nature Rev Cancer* 10: 760–774, 2010.
205. Pao W, Miller V, Zakowski M, Doherty J, Politi K, Sarkaria I, Singh B, Heelan R, Rusch V, Fulton L, Mardis E, Kupfer D, Wilson R, Kris M, Varmus H. EGF receptor gene mutations are common in lung cancers from “never smokers” and are associated with sensitivity of tumors to gefitinib and erlotinib. *Proc Natl Acad Sci USA* 101: 13306–13311, 2004.
206. Pao W, Miller VA, Politi KA, Riely GJ, Somwar R, Zakowski MF, Kris MG, Varmus H. Acquired resistance of lung adenocarcinomas to gefitinib or erlotinib is associated with a second mutation in the EGFR kinase domain. *PLoS Med* 2: e73, 2005.

207. Papin C, Denouel-Galy A, Laugier D, Calothy G, Eychene A. Modulation of kinase activity and oncogenic properties by alternative splicing reveals a novel regulatory mechanism for B-Raf. *J Biol Chem* 273: 24939–24947, 1998.
208. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nature Rev Cancer* 12: 252–264, 2012.
209. Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. *CA Cancer J Clin* 55: 74–108, 2005.
210. Parsa AT, Waldron JS, Panner A, Crane CA, Parney IF, Barry JJ, Cachola KE, Murray JC, Tihan T, Jensen MC, Mischel PS, Stokoe D, Pieper RO. Loss of tumor suppressor PTEN function increases B7–H1 expression and immunoresistance in glioma. *Nature Med* 13: 84–88, 2007.
211. Patel SP, Kurzrock R. PD-L1 expression as a predictive biomarker in cancer immunotherapy. *Mol Cancer Ther* 14: 847–856, 2015.
212. Peiro G, Ortiz-Martinez F, Gallardo A, Perez-Balaguer A, Sanchez-Paya J, Ponce JJ, Tibau A, Lopez-Vilaro L, Escuin D, Adrover E, Barnadas A, Lerma E. Src, a potential target for overcoming trastuzumab resistance in HER2-positive breast carcinoma. *Br J Cancer* 111: 689–695, 2014.
213. Peng XD, Xu PZ, Chen ML, Hahn-Windgassen A, Skeen J, Jacobs J, Sundararajan D, Chen WS, Crawford SE, Coleman KG, Hay N. Dwarfism, impaired skin development, skeletal muscle atrophy, delayed bone development, and impeded adipogenesis in mice lacking Akt1 and Akt2. *Genes Dev* 17: 1352–1365, 2003.
214. Perna D, Karreth FA, Rust AG, Perez-Mancera PA, Rashid M, Iorio F, Alifrangis C, Arends MJ, Bosenberg MW, Bollag G, Tuveson DA, Adams DJ. BRAF inhibitor resistance mediated by the AKT pathway in an oncogenic BRAF mouse melanoma model. *Proc Natl Acad Sci USA* 112: E536–545, 2015.
215. Perry MC, Dufour CR, Eichner LJ, Tsang DW, Deblois G, Muller WJ, Giguere V. ERBB2 deficiency alters an E2F-1-dependent adaptive stress response and leads to cardiac dysfunction. *Mol Cell Biol* 34: 4232–4243, 2014.
216. Pirker R, Pereira JR, Szczesna A, von Pawel J, Krzakowski M, Ramlau R, Vynnychenko I, Park K, Yu CT, Ganul V, Roh JK, Bajetta E, O'Byrne K, de Marinis F, Eberhardt W, Goddemeier T, Emig M, Gatzemeier U, Team FS. Cetuximab plus chemotherapy in patients with advanced non-small-cell lung cancer (FLEX): an open-label randomised phase III trial. *Lancet* 373: 1525–1531, 2009.
217. Polizzotti BD, Ganapathy B, Walsh S, Choudhury S, Ammanamanchi N, Bennett DG, dos Remedios CG, Haubner BJ, Penninger JM, Kuhn B. Neuregulin stimulation of cardiomyocyte regeneration in mice and human myocardium reveals a therapeutic window. *Science Transl Med* 7: 281ra245, 2015.
218. Pollak M. Insulin and insulin-like growth factor signalling in neoplasia. *Nature Rev Cancer* 8: 915–928, 2008.
219. Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. *Nature Rev Cancer* 4: 71–78, 2004.
220. Pollmann M, Parwaresch R, Adam-Klages S, Kruse ML, Buck F, Heidebrecht HJ. Human EML4, a novel member of the EMAP family, is essential for microtubule formation. *Exp Cell Res* 312: 3241–3251, 2006.
221. Poulikakos PI, Zhang C, Bollag G, Shokat KM, Rosen N. RAF inhibitors transactivate RAF dimers and ERK signalling in cells with wild-type BRAF. *Nature* 464: 427–430, 2010.
222. Pratilas CA, Taylor BS, Ye Q, Viale A, Sander C, Solit DB, Rosen N. (V600E)BRAF is associated with disabled feedback inhibition of RAF-MEK signaling and elevated transcriptional output of the pathway. *Proc Natl Acad Sci USA* 106: 4519–4524, 2009.
223. Press MF, Bernstein L, Thomas PA, Meisner LF, Zhou JY, Ma Y, Hung G, Robinson RA, Harris C, El-Naggar A, Slamon DJ, Phillips RN, Ross JS, Wolman SR, Flom KJ. HER-2/neu gene amplification characterized by fluorescence in situ hybridization: poor prognosis in node-negative breast carcinomas. *J Clin Oncol* 15: 2894–2904, 1997.
224. Press MF, Slamon DJ, Flom KJ, Park J, Zhou JY, Bernstein L. Evaluation of HER-2/neu gene amplification and overexpression: comparison of frequently used assay methods in a molecularly characterized cohort of breast cancer specimens. *J Clin Oncol* 20: 3095–3105, 2002.
225. Pritchard CA, Bolin L, Slattery R, Murray R, McMahon M. Post-natal lethality and neurological and gastrointestinal defects in mice with targeted disruption of the A-Raf protein kinase gene. *Curr Biol* 6: 614–617, 1996.
226. Pritchard CA, Samuels ML, Bosch E, McMahon M. Conditionally oncogenic forms of the A-Raf and B-Raf protein kinases display different biological and biochemical properties in NIH 3T3 cells. *Mol Cell Biol* 15: 6430–6442, 1995.
227. Pulford K, Lamant L, Morris SW, Butler LH, Wood KM, Stroud D, Delsol G, Mason DY. Detection of anaplastic lymphoma kinase (ALK) and nucleolar protein nucleophosmin (NPM)-ALK proteins in normal and neoplastic cells with the monoclonal antibody ALK1. *Blood* 89: 1394–1404, 1997.
228. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell* 141: 39–51, 2010.
229. Reck M, Bondarenko I, Luft A, Serwatowski P, Barlesi F, Chacko R, Sebastian M, Lu H, Cuillerot JM, Lynch TJ. Ipilimumab in combination with paclitaxel and carboplatin as first-line therapy in extensive-disease-small-cell lung cancer: results from a randomized, double-blind, multicenter phase 2 trial. *Ann Oncol* 24: 75–83, 2013.
230. Rexer BN, Ham AJ, Rinehart C, Hill S, Granja-Ingram Nde M, Gonzalez-Angulo AM, Mills GB, Dave B, Chang JC, Liebler DC, Arteaga CL. Phosphoproteomic mass spectrometry profiling links Src family kinases to escape from HER2 tyrosine kinase inhibition. *Oncogene* 30: 4163–4174, 2011.
231. Ribas A, Hodi FS, Callahan M, Kotto C, Wolchok J. Hepatotoxicity with combination of vemurafenib and ipilimumab. *N Engl J Med* 368: 1365–1366, 2013.
232. Riethmacher D, Sonnenberg-Riethmacher E, Brinkmann V, Yamaai T, Lewin GR, Birchmeier C. Severe neuropathies in mice with targeted mutations in the ErbB3 receptor. *Nature* 389: 725–730, 1997.
233. Ritt DA, Monson DM, Specht SI, Morrison DK. Impact of feedback phosphorylation and Raf heterodimerization on normal and mutant B-Raf signaling. *Mol Cell Biol* 30: 806–819, 2010.
234. Robert C, Karaszewska B, Schachter J, Rutkowski P, Mackiewicz A, Stroiakovski D, Lichinitser M, Dummer R, Grange F, Mortier L, Chiarion-Sileni V, Drucis K, Krajsova I, Hauschild A, Lorigan P, Wolter P, Long GV, Flaherty K, Nathan P, Ribas A, Martin AM, Sun P, Crist W, Legos J, Rubin SD, Little SM, Schadendorf D. Improved overall survival in melanoma with combined dabrafenib and trametinib. *N Engl J Med* 372: 30–39, 2015.
235. Robinson DR, Wu YM, Lin SF. The protein tyrosine kinase family of the human genome. *Oncogene* 19: 5548–5557, 2000.
236. Roesch A, Fukunaga-Kalabis M, Schmidt EC, Zabierowski SE, Brafford PA, Vultur A, Basu D, Gimotty P, Vogt T, Herlyn M. A temporarily distinct subpopulation of slow-cycling melanoma cells is required for continuous tumor growth. *Cell* 141: 583–594, 2010.
237. Roos WP, Thomas AD, Kaina B. DNA damage and the balance between survival and death in cancer biology. *Nature Rev Cancer* 16: 20–33, 2016.
238. Rosell R, Molina MA, Costa C, Simonetti S, Gimenez-Capitan A, Bertran-Alamillo J, Mayo C, Moran T, Mendez P, Cardenal F, Isla D, Provencio M, Cobo M, Insa A, Garcia-Campelo R, Reguart N, Majem M, Viteri S, Carcereny E, Porta R, Massuti B, Queralt C, de Aguirre I, Sanchez JM, Sanchez-Ronco M, Mate JL, Ariza A, Benloch S, Sanchez JJ, Bivona TG, Sawyers CL, Taron M. Pretreatment EGFR T790M mutation and BRCA1 mRNA expression in erlotinib-treated advanced non-small-cell lung cancer patients with EGFR mutations. *Clin Cancer Res* 17: 1160–1168, 2011.
239. Rosell R, Moran T, Queralt C, Porta R, Cardenal F, Camps C, Majem M, Lopez-Vivanco G, Isla D, Provencio M, Insa A, Massuti B, Gonzalez-Larriba JL, Paz-Ares L, Bover I, Garcia-Campelo R, Moreno MA, Catot S, Rolfó C, Reguart N, Palmero R, Sanchez JM, Bastus R, Mayo C, Bertran-Alamillo J, Molina MA, Sanchez JJ, Taron M, Spanish Lung Cancer Group. Screening for epidermal growth factor receptor mutations in lung cancer. *N Engl J Med* 361: 958–967, 2009.
240. Rozengurt E, Soares HP, Sinnet-Smith J. Suppression of feedback loops mediated by PI3K/mTOR induces multiple overactivation of compensatory pathways: an unintended consequence leading to drug resistance. *Mol Cancer Ther* 13: 2477–2488, 2014.
241. Ruffell B, DeNardo DG, Affara NI, Coussens LM. Lymphocytes in cancer development: polarization towards pro-tumor immunity. *Cytokine Growth Factor Rev* 21: 3–10, 2010.
242. Sasaki T, Koivunen J, Ogino A, Yanagita M, Nikiforow S, Zheng W, Lathan C, Marcoux JP, Du J, Okuda K, Capelletti M, Shimamura T, Ercan D, Stumpfova M, Xiao Y, Weremowicz S, Butaney M, Heon S, Wilner K, Christensen JG, Eck MJ, Wong KK,

- Lindeman N, Gray NS, Rodig SJ, Janne PA. A novel ALK secondary mutation and EGFR signaling cause resistance to ALK kinase inhibitors. *Cancer Res* 71: 6051–6060, 2011.
243. Scaltriti M, Chandarlapaty S, Prudkin L, Aura C, Jimenez J, Angelini PD, Sanchez G, Guzman M, Parra JL, Ellis C, Gagnon R, Koehler M, Gomez H, Geyer C, Cameron D, Arribas J, Rosen N, Baselga J. Clinical benefit of lapatinib-based therapy in patients with human epidermal growth factor receptor 2-positive breast tumors coexpressing the truncated p95HER2 receptor. *Clin Cancer Res* 16: 2688–2695, 2010.
244. Scaltriti M, Rojo F, Ocana A, Anido J, Guzman M, Cortes J, Di Cosimo S, Matias-Guiu X, Ramon y Cajal S, Arribas J, Baselga J. Expression of p95HER2, a truncated form of the HER2 receptor, and response to anti-HER2 therapies in breast cancer. *J Natl Cancer Inst* 99: 628–638, 2007.
245. Scheffler M, Bos M, Gardizi M, Konig K, Michels S, Fassunke J, Heydt C, Kunstlinger H, Ihle M, Ueckerthof F, Albus K, Serke M, Gerigk U, Schulte W, Topelt K, Nogova L, Zander T, Engel-Riedel W, Stoelben E, Ko YD, Randerath W, Kaminsky B, Panse J, Becker C, Hellmich M, Merkelbach-Bruse S, Heukamp LC, Buttner R, Wolf J. PIK3CA mutations in non-small cell lung cancer (NSCLC): genetic heterogeneity, prognostic impact and incidence of prior malignancies. *Oncotarget* 6: 1315–1326, 2015.
246. Schlessinger J. Receptor tyrosine kinases: legacy of the first two decades. *Cold Spring Harbor Perspect Biol* 6: 2014.
247. Schmidt M, Fernandez de Mattos S, van der Horst A, Klompmaaker R, Kops GJ, Lam EW, Burgering BM, Medema RH. Cell cycle inhibition by FoxO forkhead transcription factors involves downregulation of cyclin D. *Mol Cell Biol* 22: 7842–7852, 2002.
248. Schreiber RD, Old LJ, Smyth MJ. Cancer immunoeediting: integrating immunity's roles in cancer suppression and promotion. *Science* 331: 1565–1570, 2011.
249. Schwartz S, Wongvipat J, Trigwell CB, Hancox U, Carver BS, Rodrik-Outmezguine V, Will M, Yellen P, de Stanchina E, Baselga J, Scher HI, Barry ST, Sawyers CL, Chandarlapaty S, Rosen N. Feedback suppression of PI3Kalpha signaling in PTEN-mutated tumors is relieved by selective inhibition of PI3Kbeta. *Cancer Cell* 27: 109–122, 2015.
250. Sebolt-Leopold JS, Herrera R. Targeting the mitogen-activated protein kinase cascade to treat cancer. *Nature Rev Cancer* 4: 937–947, 2004.
251. Sequist LV, Waltman BA, Dias-Santagata D, Digumarthy S, Turke AB, Fidias P, Bergethon K, Shaw AT, Gettinger S, Cospoer AK, Akhavanfard S, Heist RS, Temel J, Christensen JG, Wain JC, Lynch TJ, Vernovsky K, Mark EJ, Lanuti M, Iafrate AJ, Mino-Kenudson M, Engelman JA. Genotypic and histological evolution of lung cancers acquiring resistance to EGFR inhibitors. *Sci Transl Med* 3: 75ra26, 2011.
252. Sergina NV, Rausch M, Wang D, Blair J, Hann B, Shokat KM, Moasser MM. Escape from HER-family tyrosine kinase inhibitor therapy by the kinase-inactive HER3. *Nature* 445: 437–441, 2007.
253. Serra V, Scaltriti M, Prudkin L, Eichhorn PJ, Ibrahim YH, Chandarlapaty S, Markman B, Rodriguez O, Guzman M, Rodriguez S, Gili M, Russillo M, Parra JL, Singh S, Arribas J, Rosen N, Baselga J. PI3K inhibition results in enhanced HER signaling and acquired ERK dependency in HER2-overexpressing breast cancer. *Oncogene* 30: 2547–2557, 2011.
254. Shackleton M, Quintana E, Fearon ER, Morrison SJ. Heterogeneity in cancer: cancer stem cells versus clonal evolution. *Cell* 138: 822–829, 2009.
255. Shah SP, Roth A, Goya R, Oloumi A, Ha G, Zhao Y, Turashvili G, Ding J, Tse K, Haffari G, Bashashati A, Prentice LM, Khattra J, Burleigh A, Yap D, Bernard V, McPherson A, Shumansky K, Crisan A, Giuliany R, Heravi-Moussavi A, Rosner J, Lai D, Birol I, Varhol R, Tam A, Dhalla N, Zeng T, Ma K, Chan SK, Griffith M, Moradian A, Cheng SW, Morin GB, Watson P, Gelmon K, Chia S, Chin SF, Curtis C, Rueda OM, Pharoah PD, Damaraju S, Mackey J, Hoon K, Harkins T, Tadigotla V, Sigaroudinia M, Gascard P, Tlsty T, Costello JF, Meyer IM, Eaves CJ, Wasserman WW, Jones S, Huntsman D, Hirst M, Caldas C, Marra MA, Aparicio S. The clonal and mutational evolution spectrum of primary triple-negative breast cancers. *Nature* 486: 395–399, 2012.
256. Sharma SV, Bell DW, Settleman J, Haber DA. Epidermal growth factor receptor mutations in lung cancer. *Nature Rev Cancer* 7: 169–181, 2007.
257. Shattuck DL, Miller JK, Carraway KL 3rd, Sweeney C. Met receptor contributes to trastuzumab resistance of Her2-overexpressing breast cancer cells. *Cancer Res* 68: 1471–1477, 2008.
258. Shaw AT, Hsu PP, Awad MM, Engelman JA. Tyrosine kinase gene rearrangements in epithelial malignancies. *Nature Rev Cancer* 13: 772–787, 2013.
259. Shaw AT, Kim DW, Mehra R, Tan DS, Felip E, Chow LQ, Camidge DR, Vansteenkiste J, Sharma S, De Pas T, Riely GJ, Solomon BJ, Wolf J, Thomas M, Schuler M, Liu G, Santoro A, Lau YY, Goldwasser M, Boral AL, Engelman JA. Ceritinib in ALK-rearranged non-small-cell lung cancer. *N Engl J Med* 370: 1189–1197, 2014.
260. Shi H, Hugo W, Kong X, Hong A, Koya RC, Moriceau G, Chodon T, Guo R, Johnson DB, Dahlman KB, Kelley MC, Kefford RF, Chmielowski B, Glaspy JA, Sosman JA, van Baren N, Long GV, Ribas A, Lo RS. Acquired resistance and clonal evolution in melanoma during BRAF inhibitor therapy. *Cancer Discovery* 4: 80–93, 2014.
261. Shih JY, Gow CH, Yang PC. EGFR mutation conferring primary resistance to gefitinib in non-small-cell lung cancer. *N Engl J Med* 353: 207–208, 2005.
262. Shimobayashi M, Hall MN. Making new contacts: the mTOR network in metabolism and signalling crosstalk. *Nature Rev Mol Cell Biol* 15: 155–162, 2014.
263. Shinjo K, Kondo Y. Targeting cancer epigenetics: Linking basic biology to clinical medicine. *Adv Drug Delivery Rev* 95: 56–64, 2015.
264. Sibilina M, Steinbach JP, Stingl L, Aguzzi A, Wagner EF. A strain-independent postnatal neurodegeneration in mice lacking the EGF receptor. *EMBO J* 17: 719–731, 1998.
265. Sibilina M, Wagner EF. Strain-dependent epithelial defects in mice lacking the EGF receptor. *Science* 269: 234–238, 1995.
266. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2015. *CA Cancer J Clin* 65: 5–29, 2015.
267. Singh A, Settleman J. EMT, cancer stem cells and drug resistance: an emerging axis of evil in the war on cancer. *Oncogene* 29: 4741–4751, 2010.
268. Singh JC, Jhaveri K, Esteva FJ. HER2-positive advanced breast cancer: optimizing patient outcomes and opportunities for drug development. *Br J Cancer* 111: 1888–1898, 2014.
269. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science* 235: 177–182, 1987.
270. Slamon DJ, Godolphin W, Jones LA, Holt JA, Wong SG, Keith DE, Levin WJ, Stuart SG, Udove J, Ullrich A. Studies of the HER-2/neu proto-oncogene in human breast and ovarian cancer. *Science* 244: 707–712, 1989.
271. Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y, Ishikawa S, Fujiwara S, Watanabe H, Kurashina K, Hatanaka H, Bando M, Ohno S, Ishikawa Y, Aburatani H, Niki T, Sobara Y, Sugiyama Y, Mano H. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature* 448: 561–566, 2007.
272. Sosman JA, Kim KB, Schuchter L, Gonzalez R, Pavlick AC, Weber JS, McArthur GA, Hutson TE, Moschos SJ, Flaherty KT, Hersey P, Kefford R, Lawrence D, Puzanov I, Lewis KD, Amaravadi RK, Chmielowski B, Lawrence HJ, Shyr Y, Ye F, Li J, Nolop KB, Lee RJ, Joe AK, Ribas A. Survival in BRAF V600-mutant advanced melanoma treated with vemurafenib. *N Engl J Med* 366: 707–714, 2012.
273. Sprenull EN, Dexter DL. Human tumor cell heterogeneity and metastasis. *J Clin Oncol* 1: 496–509, 1983.
274. Stambolic V, MacPherson D, Sas D, Lin Y, Snow B, Jang Y, Benchimol S, Mak TW. Regulation of PTEN transcription by p53. *Mol Cell* 8: 317–325, 2001.
275. Steelman LS, Chappell WH, Abrams SL, Kempf RC, Long J, Laidler P, Mijatovic S, Maksimovic-Ivanic D, Stivala F, Mazzarino MC, Donia M, Fagone P, Malaponte G, Nicoletti F, Libra M, Milella M, Tafuri A, Bonati A, Basecke J, Cocco L, Evangelisti C, Martelli AM, Montalto G, Cervello M, McCubrey JA. Roles of the Raf/MEK/ERK and PI3K/PTEN/Akt/mTOR pathways in controlling growth and sensitivity to therapy-implications for cancer and aging. *Aging* 3: 192–222, 2011.
276. Stephens PJ, Tarpey PS, Davies H, Van Loo P, Greenman C, Wedge DC, Nik-Zainal S, Martin S, Varela I, Bignell GR, Yates LR, Papaemmanuil E, Beare D, Butler A, Cheverton A, Gumble J, Hinton J, Jia M, Jayakumar A, Jones D, Latimer C, Lau KW, McLaren S, McBride DJ, Menzies A, Mudie L, Raine K, Rad R, Chapman MS, Teague J, Easton D, Langerod A, Oslo Breast Cancer C, Lee MT, Shen CY, Tee BT, Huimin BW, Broeks A, Vargas AC, Turashvili G, Martens J, Fatima A, Miron P, Chin SF, Thomas G, Boyault S, Mariani O, Lakhani SR, van de Vijver M, van't Veer L, Foekens J, Desmedt C, Sotiriou C, Tutt A, Caldas C, Reis-Filho JS, Aparicio SA, Salomon AV, Borresen-Dale AL, Richardson AL, Campbell PJ, Futreal PA, Stratton MR. The landscape of cancer genes and mutational processes in breast cancer. *Nature* 486: 400–404, 2012.
277. Storm SM, Cleveland JL, Rapp UR. Expression of raf family proto-oncogenes in normal mouse tissues. *Oncogene* 5: 345–351, 1990.

278. Stransky N, Cerami E, Schalm S, Kim JL, Lengauer C. The landscape of kinase fusions in cancer. *Nature Commun* 5: 4846, 2014.
279. Straussman R, Morikawa T, Shee K, Barzily-Rokni M, Qian ZR, Du J, Davis A, Mongare MM, Gould J, Frederick DT, Cooper ZA, Chapman PB, Solit DB, Ribas A, Lo RS, Flaherty KT, Ogino S, Wargo JA, Golub TR. Tumour micro-environment elicits innate resistance to RAF inhibitors through HGF secretion. *Nature* 487: 500–504, 2012.
280. Su F, Bradley WD, Wang Q, Yang H, Xu L, Higgins B, Kolinsky K, Packman K, Kim MJ, Trunzer K, Lee RJ, Schostack K, Carter J, Albert T, Germer S, Rosinski J, Martin M, Simcox ME, Lestini B, Heimbrook D, Bollag G. Resistance to selective BRAF inhibition can be mediated by modest upstream pathway activation. *Cancer Res* 72: 969–978, 2012.
281. Su KY, Chen HY, Li KC, Kuo ML, Yang JC, Chan WK, Ho BC, Chang GC, Shih JY, Yu SL, Yang PC. Pretreatment epidermal growth factor receptor (EGFR) T790M mutation predicts shorter EGFR tyrosine kinase inhibitor response duration in patients with non-small-cell lung cancer. *J Clin Oncol* 30: 433–440, 2012.
282. Swanton C. Intratumor heterogeneity: evolution through space and time. *Cancer Res* 72: 4875–4882, 2012.
283. Tebbutt N, Pedersen MW, Johns TG. Targeting the ERBB family in cancer: couples therapy. *Nature Rev Cancer* 13: 663–673, 2013.
284. Terai H, Soejima K, Yasuda H, Nakayama S, Hamamoto J, Arai D, Ishioka K, Ohgino K, Ikemura S, Sato T, Yoda S, Satomi R, Naoki K, Betsuyaku T. Activation of the FGF2-FGFR1 autocrine pathway: a novel mechanism of acquired resistance to gefitinib in NSCLC. *Mol Cancer Res* 11: 759–767, 2013.
285. Thor AD, Berry DA, Budman DR, Muss HB, Kute T, Henderson IC, Barcos M, Cirrincione C, Edgerton S, Allred C, Norton L, Liu ET. erbB-2, p53, and efficacy of adjuvant therapy in lymph node-positive breast cancer. *J Natl Cancer Inst* 90: 1346–1360, 1998.
286. Threadgill DW, Dlugosz AA, Hansen LA, Tennenbaum T, Lichti U, Yee D, LaMantia C, Mourton T, Herrup K, Harris RC. Targeted disruption of mouse EGF receptor: effect of genetic background on mutant phenotype. *Science* 269: 230–234, 1995.
287. Tidcombe H, Jackson-Fisher A, Mathers K, Stern DF, Gassmann M, Golding JP. Neural and mammary gland defects in ErbB4 knockout mice genetically rescued from embryonic lethality. *Proc Natl Acad Sci USA* 100: 8281–8286, 2003.
288. Tolaney SM, Barry WT, Dang CT, Yardley DA, Moy B, Marcom PK, Albain KS, Rugo HS, Ellis M, Shapira I, Wolff AC, Carey LA, Overmoyer BA, Partridge AH, Guo H, Hudis CA, Krop IE, Burstein HJ, Winer EP. Adjuvant paclitaxel and trastuzumab for node-negative, HER2-positive breast cancer. *N Engl J Med* 372: 134–141, 2015.
289. Torkamani A, Verkhivker G, Schork NJ. Cancer driver mutations in protein kinase genes. *Cancer Lett* 281: 117–127, 2009.
290. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *CA Cancer J Clin* 65: 87–108, 2015.
291. Trejo CL, Green S, Marsh V, Collisson EA, Iezza G, Phillips WA, McMahon M. Mutationally activated PIK3CA(H1047R) cooperates with BRAF(V600E) to promote lung cancer progression. *Cancer Res* 73: 6448–6461, 2013.
292. Tschopp O, Yang ZZ, Brodbeck D, Dummler BA, Hemmings-Mieszczak M, Watanabe T, Michaelis T, Frahm J, Hemmings BA. Essential role of protein kinase B gamma (PKB gamma/Akt3) in postnatal brain development but not in glucose homeostasis. *Development* 132: 2943–2954, 2005.
293. Twyman-Saint Victor C, Rech AJ, Maity A, Rengan R, Pauken KE, Stelekati E, Benci JL, Xu B, Dada H, Odorizzi PM, Herati RS, Mansfield KD, Patsch D, Amaravadi RK, Schuchter LM, Ishwaran H, Mick R, Pryma DA, Xu X, Feldman MD, Gangadhar TC, Hahn SM, Wherry EJ, Vonderheide RH, Minn AJ. Radiation and dual checkpoint blockade activate non-redundant immune mechanisms in cancer. *Nature* 520: 373–377, 2015.
294. Valis K, Prochazka L, Boura E, Chladova J, Obsil T, Rohlena J, Truksa J, Dong LF, Ralph SJ, Neuzil J. Hippo/Mst1 stimulates transcription of the proapoptotic mediator NOXA in a FoxO1-dependent manner. *Cancer Res* 71: 946–954, 2011.
295. Van Keymeulen A, Lee MY, Ousset M, Brohee S, Rorive S, Girardi RR, Wuidart A, Bouvencourt G, Dubois C, Salmon I, Sotiriou C, Phillips WA, Blanpain C. Reactivation of multipotency by oncogenic PIK3CA induces breast tumour heterogeneity. *Nature* 525: 119–123, 2015.
296. Vanhaesebroeck B, Stephens L, Hawkins P. PI3K signalling: the path to discovery and understanding. *Nature Rev Mol Cell Biol* 13: 195–203, 2012.
297. Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ. Natural innate and adaptive immunity to cancer. *Annu Rev Immunol* 29: 235–271, 2011.
298. Villanueva J, Infante JR, Krepler C, Reyes-Urabe P, Samanta M, Chen HY, Li B, Swoboda RK, Wilson M, Vultur A, Fukunaba-Kalabis M, Wubbenhorst B, Chen TY, Liu Q, Sproesser K, DeMarini DJ, Gilmer TM, Martin AM, Marmorstein R, Schultz DC, Speicher DW, Karakousis GC, Xu W, Amaravadi RK, Xu X, Schuchter LM, Herlyn M, Nathanson KL. Concurrent MEK2 mutation and BRAF amplification confer resistance to BRAF and MEK inhibitors in melanoma. *Cell Rep* 4: 1090–1099, 2013.
299. Voulgari A, Pintzas A. Epithelial-mesenchymal transition in cancer metastasis: mechanisms, markers and strategies to overcome drug resistance in the clinic. *Biochim Biophys Acta* 1796: 75–90, 2009.
300. Wagle N, Emery C, Berger MF, Davis MJ, Sawyer A, Pochanard P, Kehoe SM, Johannessen CM, Macconail LE, Hahn WC, Meyerson M, Garraway LA. Dissecting therapeutic resistance to RAF inhibition in melanoma by tumor genomic profiling. *J Clin Oncol* 29: 3085–3096, 2011.
301. Wagner MJ, Stacey MM, Liu BA, Pawson T. Molecular mechanisms of SH2- and PTB-domain-containing proteins in receptor tyrosine kinase signaling. *Cold Spring Harbor Perspect Biol* 5: a008987, 2013.
302. Wang YC, Morrison G, Gillihan R, Guo J, Ward RM, Fu X, Botero MF, Healy NA, Hilsenbeck SG, Phillips GL, Charness GC, Rimawi MF, Osborne CK, Schiff R. Different mechanisms for resistance to trastuzumab versus lapatinib in HER2-positive breast cancers—role of estrogen receptor and HER2 reactivation. *Breast Cancer Res* 13: R121, 2011.
303. Ward CW, Lawrence MC, Streltsov VA, Adams TE, McKern NM. The insulin and EGF receptor structures: new insights into ligand-induced receptor activation. *Trends Biochem Sci* 32: 129–137, 2007.
304. Ware KE, Hinz TK, Kleczko E, Singleton KR, Marek LA, Helfrich BA, Cummings CT, Graham DK, Astling D, Tan AC, Heasley LE. A mechanism of resistance to gefitinib mediated by cellular reprogramming and the acquisition of an FGF2-FGFR1 autocrine growth loop. *Oncogenesis* 2: e39, 2013.
305. Webb TR, Slavish J, George RE, Look AT, Xue L, Jiang Q, Cui X, Rentrop WB, Morris SW. Anaplastic lymphoma kinase: role in cancer pathogenesis and small-molecule inhibitor development for therapy. *Expert Rev Anticancer Ther* 9: 331–356, 2009.
306. Weber JS, Dummer R, de Pril V, Lebke C, Hodi FS, Investigators MDX. Patterns of onset and resolution of immune-related adverse events of special interest with ipilimumab: detailed safety analysis from a phase 3 trial in patients with advanced melanoma. *Cancer* 119: 1675–1682, 2013.
307. Weber JS, Kahler KC, Hauschild A. Management of immune-related adverse events and kinetics of response with ipilimumab. *J Clin Oncol* 30: 2691–2697, 2012.
308. Wetterskog D, Shiu KK, Chong I, Meijer T, Mackay A, Lambros M, Cunningham D, Reis-Filho JS, Lord CJ, Ashworth A. Identification of novel determinants of resistance to lapatinib in ERBB2-amplified cancers. *Oncogene* 33: 966–976, 2014.
309. Wheeler DL, Huang S, Kruser TJ, Nechrebecki MM, Armstrong EA, Benavente S, Gondi V, Hsu KT, Harari PM. Mechanisms of acquired resistance to cetuximab: role of HER (ErbB) family members. *Oncogene* 27: 3944–3956, 2008.
310. Wilson FH, Johannessen CM, Piccioni F, Tamayo P, Kim JW, Van Allen EM, Corsello SM, Capelletti M, Calles A, Butaney M, Sharifnia T, Gabriel SB, Mesirov JP, Hahn WC, Engelman JA, Meyerson M, Root DE, Janne PA, Garraway LA. A functional landscape of resistance to ALK inhibition in lung cancer. *Cancer Cell* 27: 397–408, 2015.
311. Wixler V, Smola U, Schuler M, Rapp U. Differential regulation of Raf isozymes by growth versus differentiation inducing factors in PC12 pheochromocytoma cells. *FEBS Lett* 385: 131–137, 1996.
312. Wojnowski L, Stancato LF, Larner AC, Rapp UR, Zimmer A. Overlapping and specific functions of Braf and Craf-1 proto-oncogenes during mouse embryogenesis. *Mech Dev* 91: 97–104, 2000.
313. Wojnowski L, Stancato LF, Zimmer AM, Hahn H, Beck TW, Larner AC, Rapp UR, Zimmer A. Craf-1 protein kinase is essential for mouse development. *Mech Dev* 76: 141–149, 1998.

314. Wojnowski L, Zimmer AM, Beck TW, Hahn H, Bernal R, Rapp UR, Zimmer A. Endothelial apoptosis in Braf-deficient mice. *Nature Genet* 16: 293–297, 1997.
315. Xia W, Liu Z, Zong R, Liu L, Zhao S, Bacus SS, Mao Y, He J, Wulfkuehle JD, Petricoin EF, 3rd Osada T, Yang XY, Hartman ZC, Clay TM, Blackwell KL, Lysterly HK, Spector NL. Truncated ErbB2 expressed in tumor cell nuclei contributes to acquired therapeutic resistance to ErbB2 kinase inhibitors. *Mol Cancer Ther* 10: 1367–1374, 2011.
316. Xue G, Hemmings BA. Phosphorylation of basic helix-loop-helix transcription factor Twist in development and disease. *Biochem Soc Trans* 40: 90–93, 2012.
317. Xue G, Hemmings BA. PKB/Akt-dependent regulation of cell motility. *J Natl Cancer Inst* 105: 393–404, 2013.
318. Xue G, Zippelius A, Wicki A, Mandala M, Tang F, Massi D, Hemmings BA. Integrated Akt/PKB Signaling in immunomodulation and its potential role in cancer immunotherapy. *J Natl Cancer Inst* 107: 2015.
319. Yadav V, Zhang X, Liu J, Estrem S, Li S, Gong XQ, Buchanan S, Henry JR, Starling JJ, Peng SB. Reactivation of mitogen-activated protein kinase (MAPK) pathway by FGF receptor 3 (FGFR3)/Ras mediates resistance to vemurafenib in human B-RAF V600E mutant melanoma. *J Biol Chem* 287: 28087–28098, 2012.
320. Yamada T, Takeuchi S, Nakade J, Kita K, Nakagawa T, Nanjo S, Nakamura T, Matsumoto K, Soda M, Mano H, Uenaka T, Yano S. Paracrine receptor activation by microenvironment triggers bypass survival signals and ALK inhibitor resistance in EML4-ALK lung cancer cells. *Clin Cancer Res* 18: 3592–3602, 2012.
321. Yamashita J, Ogawa M, Yamashita S, Nomura K, Kuramoto M, Saishoji T, Shin S. Immunoreactive hepatocyte growth factor is a strong and independent predictor of recurrence and survival in human breast cancer. *Cancer Res* 54: 1630–1633, 1994.
322. Yang H, Bueso-Ramos C, DiNardo C, Estecio MR, Davanlou M, Geng QR, Fang Z, Nguyen M, Pierce S, Wei Y, Parmar S, Cortes J, Kantarjian H, Garcia-Manero G. Expression of PD-L1, PD-L2, PD-1 and CTLA4 in myelodysplastic syndromes is enhanced by treatment with hypomethylating agents. *Leukemia* 28: 1280–1288, 2014.
323. Yang JC, Hirsh V, Schuler M, Yamamoto N, O'Byrne KJ, Mok TS, Zazulina V, Shahidi M, Lungershausen J, Massey D, Palmer M, Sequist LV. Symptom control and quality of life in LUX-Lung 3: a phase III study of afatinib or cisplatin/pemetrexed in patients with advanced lung adenocarcinoma with EGFR mutations. *J Clin Oncol* 31: 3342–3350, 2013.
324. Yang ZZ, Tschopp O, Di-Poi N, Bruder E, Baudry A, Dummler B, Wahli W, Hemmings BA. Dosage-dependent effects of Akt1/protein kinase B α (PKB α) and Akt3/PKB γ on thymus, skin, and cardiovascular and nervous system development in mice. *Mol Cell Biol* 25: 10407–10418, 2005.
325. Yang ZZ, Tschopp O, Hemmings-Mieszczyk M, Feng J, Brodbeck D, Perentes E, Hemmings BA. Protein kinase B α /Akt1 regulates placental development and fetal growth. *J Biol Chem* 278: 32124–32131, 2003.
326. Yao E, Zhou W, Lee-Hoeflich ST, Truong T, Haverty PM, Eastham-Anderson J, Lewin-Koh N, Gunter B, Belvin M, Murray LJ, Friedman LS, Sliwkowski MX, Hoeflich KP. Suppression of HER2/HER3-mediated growth of breast cancer cells with combinations of GDC-0941 PI3K inhibitor, trastuzumab, and pertuzumab. *Clin Cancer Res* 15: 4147–4156, 2009.
327. Yap TA, Gerlinger M, Futreal PA, Pusztai L, Swanton C. Intratumor heterogeneity: seeing the wood for the trees. *Science Transl Med* 4: 127ps110, 2012.
328. Yu HA, Arcila ME, Hellmann MD, Kris MG, Ladanyi M, Riely GJ. Poor response to erlotinib in patients with tumors containing baseline EGFR T790M mutations found by routine clinical molecular testing. *Ann Oncol* 25: 423–428, 2014.
329. Yu Y, Yoon SO, Pouligiannis G, Yang Q, Ma XM, Villen J, Kubica N, Hoffman GR, Cantley LC, Gygi SP, Blenis J. Phosphoproteomic analysis identifies Grb10 as an mTORC1 substrate that negatively regulates insulin signaling. *Science* 332: 1322–1326, 2011.
330. Yuan F, Xie Q, Wu J, Bai Y, Mao B, Dong Y, Bi W, Ji G, Tao W, Wang Y, Yuan Z. MST1 promotes apoptosis through regulating Sirt1-dependent p53 deacetylation. *J Biol Chem* 286: 6940–6945, 2011.
331. Yuan Z, Lehtinen MK, Merlo P, Villen J, Gygi S, Bonni A. Regulation of neuronal cell death by MST1-FOXO1 signaling. *J Biol Chem* 284: 11285–11292, 2009.
332. Yun CH, Mengwasser KE, Toms AV, Woo MS, Greulich H, Wong KK, Meyerson M, Eck MJ. The T790M mutation in EGFR kinase causes drug resistance by increasing the affinity for ATP. *Proc Natl Acad Sci USA* 105: 2070–2075, 2008.
333. Yuryev A, Wennogle LP. The RAF family: an expanding network of post-translational controls and protein-protein interactions. *Cell Res* 8: 81–98, 1998.
334. Zhang S, Huang WC, Li P, Guo H, Poh SB, Brady SW, Xiong Y, Tseng LM, Li SH, Ding Z, Sahin AA, Esteva FJ, Hortobagyi GN, Yu D. Combating trastuzumab resistance by targeting SRC, a common node downstream of multiple resistance pathways. *Nature Med* 17: 461–469, 2011.
335. Zhang W, Cohen SM. The Hippo pathway acts via p53 and microRNAs to control proliferation and proapoptotic gene expression during tissue growth. *Biol Open* 2: 822–828, 2013.
336. Zhang Y, Zhang J, Xu K, Xiao Z, Sun J, Xu J, Wang J, Tang Q. PTEN/PI3K/mTOR/B7–H1 signaling pathway regulates cell progression and immuno-resistance in pancreatic cancer. *Hepato-gastroenterology* 60: 1766–1772, 2013.
337. Zhang Z, Lee JC, Lin L, Olivas V, Au V, LaFramboise T, Abdel-Rahman M, Wang X, Levine AD, Rho JK, Choi YJ, Choi CM, Kim SW, Jang SJ, Park YS, Kim WS, Lee DH, Lee JS, Miller VA, Arcila M, Ladanyi M, Moonsamy P, Sawyers C, Boggon TJ, Ma PC, Costa C, Taron M, Rosell R, Halmos B, Bivona TG. Activation of the AXL kinase causes resistance to EGFR-targeted therapy in lung cancer. *Nature Genet* 44: 852–860, 2012.
338. Zimmermann S, Moelling K. Phosphorylation and regulation of Raf by Akt (protein kinase B). *Science* 286: 1741–1744, 1999.