**Article type:** systematic review with meta-analysis

Title: Nigella Sativa (Black Seed) Effects on Plasma Lipid Concentrations in Humans: a Systematic

Review and Meta-Analysis of Randomized Placebo-Controlled Trials

**Authors:** Amirhossein Sahebkar<sup>1,2</sup> – Guglielmo Beccuti<sup>3</sup> – Luis E. Simental-Mendía<sup>4</sup> – Valerio

Nobili<sup>5</sup> – Simona Bo<sup>3</sup>\*

**Affiliations:** 

AS: <sup>1</sup>Biotechnology Research Center, Mashhad University of Medical Sciences, Mashhad, Iran and

<sup>2</sup>Metabolic Research Centre, Royal Perth Hospital, School of Medicine and Pharmacology, University

of Western Australia, Perth, Australia

GB; SB: <sup>3</sup>Department of Medical Sciences, University of Turin, Turin, Italy

LES: <sup>4</sup>Biomedical Research Unit, Mexican Social Security Institute, Durango, Mexico

VN: <sup>5</sup>Hepato-Metabolic Disease Unit, Bambino Gesù Children's Hospital, IRCCS, Rome, Italy

**Corresponding Author:** Simona Bo, M.D.

Department of Medical Sciences; University of Turin; Corso Dogliotti 14, 10126, Turin, Italy

Telephone office: +39.011.6336036; Fax: +39.011.6335401; E-mail: simona.bo@unito.it

Key words: Nigella Sativa, Cholesterol, HDL-cholesterol, Triglycerides

Word count: text 3554

**Conflicts of interest:** none

#### **Abstract**

The effects of *Nigella Sativa* (NS) on plasma lipid concentrations are controversial. A systematic review and meta-analysis of randomized controlled trials (RCTs) was conducted to obtain a conclusive result in humans.

PubMed-Medline, SCOPUS, Web of Science, and Google Scholar databases were searched (up to August 2015) to identify RCTs investigating the impact of NS on total cholesterol, LDL-cholesterol (LDL-C), HDL-cholesterol (HDL-C), and triglycerides concentrations. A random-effects model and the generic inverse variance weighting method were used for quantitative data synthesis. Metaregression, sensitivity analysis, and publication bias assessments were performed using standard methods.

A total of 17 RCTs examining the effects of NS on plasma lipid concentrations were included. Meta-analysis suggested a significant association between NS supplementation and a reduction in total cholesterol (weighed-mean-difference [WMD]: -15.65 mg/dL, 95% CI: -24.67, -6.63, p =0.001), LDL-C (WMD: -14.10 mg/dL, 95% CI: -19.32, -8.88, p <0.001), and triglyceride levels (WMD: -20.64 mg/dL, 95% CI: -30.29, -11.00, p <0.001). No significant effect on HDL-C concentrations (WMD: 0.28 mg/dL, 95% CI: -1.96, 2.53, p =0.804) was found. A greater effect of NS seed oil versus seed powder was observed on serum total cholesterol and LDL-C levels, and an increase in HDL-C levels was found only after NS seed powder supplementation.

NS has a significant impact on plasma lipid concentrations, leading to lower total cholesterol, LDL-C, and TG levels while increased HDL-C is associated with NS powder only. Further RCTs are needed to explore the NS benefits on cardiovascular outcomes.

### 1. Introduction

Nigella Sativa (NS), popularly known as black seed or black cumin, is an annual plant of the Ranunculaceae family which grows widely in many Middle Eastern countries and Southwest Asia. The seeds of NS are claimed to play antibacterial [1, 2], anti-inflammatory [3], immune-potentiating [4], antioxidant [5, 6], hypoglycemic [7, 8], antihypertensive [9], anti-obesity [10], bronchodilatory [11], neuro- and cardio-protective [12, 13], and antidiarrhoic effects [14], Increasing evidence also supports hypolipidemic properties of NS [13, 15-18]. The NS seeds contain many bioactive constituents, such as antioxidant compounds (mainly represented by thymoquinone and dithymoquinone), flavonoids, sterols, and polyunsaturated fatty acids [15], and the lipid-lowering effect is likely mediated by a synergistic action of its different components. Different mechanisms have been postulated, such as activation of the peroxisome proliferator-activated receptor gamma (PPAR-gamma) [19], increasing uptake of low-density lipoprotein cholesterol (LDL-C) by upregulation of hepatic LDL receptors [20], de novo suppression of cholesterol synthesis [20], reduction in dietary cholesterol absorption, and prevention of lipid peroxidation [6], There is growing interest in finding safe natural alternatives to common drugs used to treat dyslipidemia, one of the most important risk factor for cardiovascular diseases, particularly in patients resistant to or intolerant of statins [21]. Therefore, the lipid-lowering action of NS has been evaluated both in different experimental models, varying from normal [22] to dyslipidemic [23, 24], dysmetabolic [25], and diabetic animals [26], and in clinical studies, performed in healthy [27], dyslipidemic [28], dysmetabolic [29], and diabetic subjects [30]. Supplementation with NS has been associated with decreased concentrations of serum total cholesterol [23, 24, 26-29, 31-45], triglycerides [2, 23, 24, 26, 28, 31, 32, 34-41, 43, 44, 46], and/or increased concentrations of highdensity lipoprotein cholesterol (HDL-C) [23, 25, 28, 32, 34-36, 39, 41-44]. With respect to clinical findings, although some randomized controlled trials (RCTs) have been

performed, the reported results have been controversial [9, 47-62]. Because of the variable duration

of studies, preparation of NS employed, study designs, and recruited populations, it is difficult to draw definitive conclusions on the hypolipidemic activity of this natural supplement. The aim of this study was therefore to perform a systematic review of RCTs and conduct a meta-analysis to evaluate the effect of NS supplementation on plasma lipid concentrations.

### 2. Methods

## 2.1 Search Strategy

This study was designed according to the guidelines of the 2009 preferred reporting items for systematic reviews and meta-analysis (PRISMA) statement [63]. PubMed-Medline, SCOPUS, Web of Science, and Google Scholar databases were searched using the following search terms in titles and abstracts: ("Nigella sativa" OR "black seed" OR "black cumin" OR thymoquinone) AND (placebo) AND (hyperlipidemia OR hyperlipidaemia OR hyperlipidaemic OR hyporlipidaemic OR hypolipidemic OR hypolipidaemic OR dyslipidaemia OR dyslipidaemia OR dyslipidaemia OR dyslipidaemic OR dyslipidaemic OR hypercholesterolemic OR hypercholesterolaemic OR hypercholesterolaemic OR "low-density lipoprotein" OR hypocholesterolaemic OR hypocholesterolaemic OR hypertriglyceridaemia OR hypertriglyceridaemia OR hypotriglyceridaemic OR hypotriglyceridaemic OR LDL OR LDL-C OR LDL-cholesterol OR HDL- OR HDL-C OR HDL-cholesterol). The wild-card term "\*" was used to increase the sensitivity of the search strategy. No language restriction was applied. The literature was searched from inception to August 18th, 2015.

# 2.2 Study Selection

The following criteria were used to identify eligible studies: (i) randomized placebo-controlled trials with either case-control or case-cross-over design, (ii) investigation of the effects of NS or thymoquinone on plasma/serum lipid concentrations, (iii) providing sufficient information on

baseline and end-trial plasma/serum lipid concentrations in both NS and control groups. Exclusion criteria were (i) experimental studies, (ii) observational studies, (iii) uncontrolled studies, and (iv) lack of sufficient information on baseline or end-trial lipid concentrations. In case of the latter item, authors of the article(s) were contacted and requested to provide necessary numerical data.

### 2.3 Data extraction

Eligible studies were reviewed, and the following data were abstracted: 1) first author's name; 2) year of publication; 3) country were the study was performed; 4) study design; 5) number of participants in the NS and control groups; 6) type and dose of NS supplement; 7) treatment duration; 9) age, gender, and body mass index (BMI) of study participants; and 9) data regarding baseline and follow-up plasma concentrations of total cholesterol, LDL-C, HDL-C, and triglycerides.

# 2.4 Quality assessment

A systematic assessment of bias in the included studies was performed using the Cochrane criteria [64]. The items used for the assessment of each study were as follows: adequacy of sequence generation, allocation concealment, blinding, addressing of dropouts (incomplete outcome data), selective outcome reporting, and other potential sources of bias. According to the recommendations of the Cochrane Handbook, a judgment of "yes" indicated low risk of bias, while "no" indicated high risk of bias. Labeling an item as "unclear" indicated an unclear or unknown risk of bias.

## 2.5 Quantitative Data Synthesis

Meta-analysis was conducted using Comprehensive Meta-Analysis (CMA) V2 software (Biostat, NJ) [65]. Net changes in measurements (change scores) were calculated as follows: measure at end of follow-up – measure at baseline. For single-arm cross-over trials, net change in plasma concentrations of lipid indices were calculated by subtracting the value after control intervention

from that reported after treatment. All values were collated to mg/dL. Standard deviations (SDs) of the mean difference were calculated using the following formula:  $SD = \text{square root } [(SD_{\text{pre-treatment}})^2 + (SD_{\text{post-treatment}})^2 - (2R \times SD_{\text{pre-treatment}} \times SD_{\text{post-treatment}})]$ , assuming a correlation coefficient (R) = 0.5. If the outcome measures were reported in median and range (or 95% confidence interval [CI]), mean and standard SD values were estimated using the method described by Wan et al [66]. When only the standard error of the mean (SEM) was reported, standard deviation (SD) was estimated using the following formula:  $SD = SEM \times sqrt(n)$ , where n is the number of subjects. In order to avoid unit-of-analysis error due to double-counting of subjects in the trials with more than 1 treatment arm, the control group was evenly (where possible) divided.

Net changes in measurements (change scores) were calculated for parallel and cross-over trials, as follows: (measure at the end of follow-up in the treatment group – measure at baseline in the treatment group) – (measure at the end of follow-up in the control group – measure at baseline in the control group). A random-effects model (using DerSimonian-Laird method) and the generic inverse variance method were used to compensate for the heterogeneity of studies in terms of study design, treatment duration, and the characteristics of populations being studied [67]. Effect sizes were expressed as weighted mean difference (WMD) and 95% confidence interval (CI). Inter-study heterogeneity was assessed using Cochran Q test and I<sup>2</sup> index. In order to evaluate the influence of each study on the overall effect size, sensitivity analysis was conducted using leave-one-out method, i.e. iteratively removing one study each time and repeating the analysis [68-70].

### 2.6 Meta-regression

A weighted random-effects meta-regression using unrestricted maximum likelihood model was performed to assess the association between the overall estimate of effect size and potential moderator variables including dose and duration of NS supplementation.

#### 2.7 Publication bias

Potential publication bias was explored using visual inspection of Begg's funnel plot asymmetry, Egger's weighted regression, and "fail safe N" tests. Duval & Tweedie "trim and fill" method was used to adjust the analysis for the effects of publication bias [71].

### 3. Results

# 3.1 Flow of studies

After multiple database searches, 52 published studies were identified, and the abstracts reviewed. Twenty-five did not meet the inclusion criteria and were excluded. Next, 22 full text articles were careful assessed and reviewed; of which 5 studies were excluded for not measuring plasma lipid concentrations (n=3), incomplete lipid data (n=1), and non-original article (n=1). Finally, 17 studies were eligible and included in the systematic review and meta-analysis. The study selection process is shown in **Figure 1**.

# 3.2 Characteristics of included studies

Data were pooled from 17 RCTs that included 1185 subjects, with 616 in the NS arm and 569 in the control arm. Included studies were published between 2008 and 2015. The clinical trials used different forms and doses of NS. Three studies investigated NS powder 1 g/day [49, 56, 57], two studies investigated NS powder 1.5 g/day [47, 50], one study investigated NS powder 1.6 g/day [58], three studies investigated NS powder 2 g/day [51, 61, 62], two studies investigated NS powder 2 spoons/day [52, 60], two studies investigated NS oil 5 ml/day [48, 55], one study investigated NS oil 100 mg/day [9], one study investigated NS oil 200 mg/day [9], one study investigated NS oil 1 g/day [53], and two studies investigated NS oil 3 g/day [54, 59]. The range of intervention periods was from 4 weeks [52, 62] up to 3 months [50, 54, 55, 58]. The most of included studies were parallel-group design [9, 47-57, 59-62], only one study was cross-over design [58]. Selected trials enrolled

subjects with metabolic syndrome [47, 56], overweight [51], obesity [50, 59], mild hypertension [9], hyperlipidemia [51, 52, 60-62], type 2 diabetes [53-55], menopausal women [56-58], and healthy subjects [48, 49] (**Table 1**).

## 3.3 Risk of bias assessment

Several of the included studies were characterized by lack of information about the allocation concealment and blinding of participants, personnel, and outcome assessors. Some trials did not provide sufficient information of sequence generation [9, 48, 50, 52, 53, 57, 60]. In addition, some trials had other biases related with the study design, such as open-label single arm [58] and single blind [52, 60]. However, most of evaluated studies showed low risk of bias according to incomplete outcome data and selective outcome reporting. Details of the quality of bias assessment are shown in **Table 2**.

# 3.4 Effect of NS supplementation on plasma lipid concentrations

Overall, the impact of NS supplementation on plasma concentrations of total cholesterol, LDL-C, HDL-C, and triglycerides was assessed in 15, 16, 16 and 18 treatment arms, respectively. NS supplementation was found to significantly reduce plasma concentrations of total cholesterol (WMD: -15.65 mg/dL, 95% CI: -24.67, -6.63, p = 0.001; **Figure 2**), LDL-C (WMD: -14.10 mg/dL, 95% CI: -19.32, -8.88, p < 0.001; **Figure 2**), and triglycerides (WMD: -20.64 mg/dL, 95% CI: -30.29, -11.00, p < 0.001; **Figure 2**), while no significant effect on HDL-C concentrations (WMD: 0.28 mg/dL, 95% CI: -1.96, 2.53, p = 0.804; **Figure 2**) was found. All these effects were robust in the sensitivity analysis (**Figure 3**), and the overall estimate of effect size was not significantly driven by a single study.

When the meta-analysis was stratified according to the type of NS supplement that was administered, a greater effect of NS seed oil versus seed powder was observed on serum total cholesterol and LDL-

cholesterol concentrations, while the effect of both supplement types on plasma triglycerides levels was comparable (**Figure 4**). With respect to plasma HDL-C concentrations, a significant elevation was found in the subgroups of studies with NS seed powder, but not NS seed oil (**Figure 4**).

## 3.5 Meta-regression

Meta-regression analysis was conducted to evaluate the association between changes in plasma lipid concentrations and potential confounders including dose and duration of supplementation with NS. No significant association was found between changes in lipid parameters and duration of supplementation with NS (**Figure 5**). With respect to dose, there were significant associations with changes in plasma total cholesterol (slope: 0.001; 95% CI: 0.0002, 0.015; p = 0.044) and HDL-C concentrations (slope: 0.004; 95% CI: 0.003, 0.005; p < 0.001), but not with LDL-C and triglyceride levels (**Figure 6**).

#### 3.6 Publication bias

Visual inspection of funnel plots suggested an asymmetry in the meta-analyses of NS's effects on plasma lipid concentrations. Using "trim and fill" method, 2, 3 and 3 potentially missing studies were imputed for the meta-analyses of triglycerides, LDL-C, and HDL-C (**Figure 7**). Corrected effect sizes (following imputation of potentially missing studies) and the results of Egger's linear regression, Begg's rank correlation, and "fail safe N" tests are summarized in **Table 3**.

### 4. Discussion

The results of the present meta-analysis of RCTs showed that supplementation with NS significantly reduces plasma concentrations of total cholesterol, LDL-C, and triglycerides. This effect was greater with seed oil, while seed powder was found to be associated with a significant elevation in HDL-C levels.

The effects of NS seem to be predominant on LDL-C. The ethanol extract of NS is an agonist of the PPAR-gamma gene [19], whose activation appears to be associated with the enhanced expression of CD36 [72], a cellular scavenger receptor for atherogenic LDL, and ATP-binding cassette transporter A1, a reverse cholesterol transporter involved in the cholesterol efflux from macrophages [73]. Thymoquinone, an active ingredient of NS oil, can up-regulate hepatic LDL receptors [20], inhibit 3-hydroxy-3-methylglutaryl-coenzyme A reductase gene [20], and down-regulate ApoB100 gene [74], thus leading to both increased clearance and reduced synthesis of LDL-C. NS could also stimulate bile acid excretion [75] and has a slight anorexic effect [76]. Moreover, many of phytochemicals present in the NS seeds may contribute to its hypocholesterolemic effects: β-sitosterol inhibits the intestinal absorption of dietary cholesterol [77, 78], anti-oxidants protect tissues from lipid peroxidation [6], and the rich content of unsaturated fatty acids may contribute to the reduction in and prevention of cholesterol oxidation [27].

Almost all previous experimental studies in animals [23, 24, 26, 31-44, 79-81], but one [25], showed a reduction in LDL-C after NS supplementation. Consistently, small clinical studies confirmed a significant LDL-C reduction after the consumption of NS in both healthy individuals [27] and dysmetabolic patients [28, 29, 45]. In our meta-analysis, seed oil showed a greater benefit on serum total cholesterol and LDL-C concentrations than seed powder. Owing to the different compositions and doses of NS preparations, along with the inter-study heterogeneities, it is difficult to justify this result with certainty. However, it has been demonstrated that, differently from the powder deriving from seed crushing, the preparation processes of the seed oil may lead to significant compositional changes in the active ingredients, with increased content of the thymoquinone, to which is attributed most of the biological activity of NS [82]. Controlled thermal processing of the seeds, responsible for thymoquinone accumulation at temperatures between 50 and 150°C, could explain the higher biological activity of the oil from heated seeds [82]. Furthermore, thymoquinone is soluble in fat, and the NS oil may be more effective than the aqueous extract form [59].

We found a highly significant effect of NS supplementation in reducing triglyceride concentrations, in line with many animal and clinical studies [2, 23, 24, 26, 28, 31, 32, 34-41, 43, 44, 46, 83-85], but not all [24, 25, 29, 33, 40, 45]. Possible mechanisms responsible for the triglyceride-lowering effect of NS may be due to its components. The high content of polyunsaturated fats may affect both the synthesis and catabolism of triglyceride-rich lipoproteins through increasing PPAR-gamma and lipoprotein lipase activities [59]. Triglyceride-rich lipoproteins exert pro-atherogenic effects, commonly attributed to their remnant lipoprotein particles [86, 87]. Remnants are involved in the mechanisms of endothelial dysfunction and atherosclerosis progression, and appear to be an independent risk factor for cardiovascular disease [86, 87]. Furthermore, the nigellamines A-(5) have been shown to reduce triglyceride levels *in vitro* similarly to clofibrate [88].

Finally, the impact of NS on HDL-C is more controversial: no overall significant effect was found in this meta-analysis, and data from literature are contrasting. Experimental and clinical studies have reported positive effects [23, 25, 28, 32, 34-36, 39, 41-44, 81, 83, 85], but others did not confirm these findings [2, 24, 26, 29, 31, 33, 34, 37, 38, 40, 45].

The divergent results may be due to differences in dosage and type of NS supplementation, dietary habits, physical activity level, duration of the intervention, ethnicity, laboratory and clinical characteristics of the patients studied. Indeed, an HDL-C increase was reported in studies where participants were assigned to an exercise program [51, 60], in experimental studies where NS determined a relevant decrement of triglycerides and insulin resistance, thus resulting in a lower clearance of HDL [25, 36, 54], or in studies administering seed powder [52]. A possible explanation for the superiority of seed powder on HDL-C may be the higher content of seeds of unsaturated fatty acids, mainly linoleic and oleic acids [16]. Regardless of the impact of NS supplementation on plasma HDL-C levels, it is noteworthy that the concept of HDL-C elevation for protection against atherosclerotic cardiovascular disease has been assailed by some recent large-scale trials [89-91]. In

fact, it is currently uncertain if HDL-C elevation could be considered as a treatment target to reduce the risk of cardiovascular outcomes [92].

It could be hypothesized that a longer period of NS supplementation might be necessary in order to impact on HDL-C values. Indeed, the RCTs included in this meta-analysis are short-lasting (maximum 12 weeks), while significant improvements were observed in clinical studies with 6 months supplementations [93].

A major point is that the value of a supplementation should be evaluated not only in term of surrogate endpoints, but also for its effects on morbidity and mortality. Many clinical studies have reported benefits of NS on fasting glucose, insulin resistance, and metabolic syndrome [30, 47, 50, 53-56], body weight and central obesity [10, 29, 50, 55, 59], arterial pressure [9, 94], and inflammation [50]. Lastly, additional cardio-protective effects have been demonstrated in experimental studies: NS inhibits the plaque formation [95] and arachidonic acid-induced platelet aggregation [96], improves endothelial function [97], reduces intima/media ratio [95], induces bradycardia [98], ameliorates cardiac hemodynamics [99], exerts cardio-protection against exogenous and endogenous toxic products [13]. Long-term RCTs are surely needed to evaluate the impact of NS on cardiovascular outcomes. These preliminary results, however, report a wide range of benefits other than the hypolipidemic effects, suggesting a potential use of NS as an adjunct to statin therapy, in consideration of both the significant residual cardiovascular risk in statin-treated individuals and the limitation of statins in achieving optimal LDL-C concentrations.

The safety of NS has been documented in clinical practice. Only a few adverse events have been reported in humans: two cases of contact dermatitis after topical use [100]. In addition to its pungent bitter taste, NS may cause mild, transient nausea and dyspepsia [47], exert a slight anorexic [76] and weight-loss effect [10, 29]. High doses of thymoquinone were found to induce hepatic oxidative stress in mice [101], but humans with solid tumors have tolerated a dose up to 2600 mg/day in a phase I clinical study, with no side effects [102]. NS seeds, particularly in the powdered form,

resulted in a potent hepato-protective effect in experimental models [6, 103], while a non-toxic increase in liver enzymes has been observed after NS oil administration in women [104]. In the included studies, there was no difference in the frequency of adverse effects between treated and placebo groups, and among all participants only one treated subject was reported to quit for nausea [47].

No significant association was found between changes in lipid parameters and duration of supplementation with NS in our meta-analysis. Ibrahim demonstrated that 1 month after cessation of supplementation, the lipid concentrations changed towards the pretreatment levels, thus suggesting the need of a life-long assumption [57]. On the other hand, in the available study with the longer follow-up (6 months), the effect of NS started after 4 days and continued for the entire follow-up [93].

A dose-dependent effect of NS was observed for changes in total cholesterol and HDL-C, but not for triglycerides and LDL-C. We showed in this meta-analysis that NS was more effective against triglyceride and LDL-C concentrations. It could be therefore hypothesized that any dosage was effective on these parameters, while higher doses of NS are needed to affect the other lipid variables, i.e. triglycerides and HDL-C.

RCTs with larger cohorts and longer follow-up are however needed to ascertain the most effective duration and dosage of NS supplementation, that at present are not defined.

# 4.1 Strengths and limitation

To the best of authors' knowledge, this is the first systematic review and meta-analysis of RTCs investigating the effect of NS on plasma lipid concentrations. However, a number of limitations should be mentioned. First of all, the heterogeneity of RCTs included in the meta-analysis have to be considered, as healthy individuals, patients with hypertension, type 2 diabetes, dyslipidemia, and metabolic syndrome were enrolled. Different preparations, doses, and durations of supplementation

were employed. Part of this inter-study heterogeneity was addressed by choosing a random-effects model for meta-analysis and performing subgroup and meta-regression analyses. Furthermore, some RCTs were not primarily designed to assess the effects of NS on lipid concentrations. Finally, the number of subjects studied in the present meta-analysis was relatively small, but the current pooled population size was sufficient to detect a significant lipid-lowering effect of NS. All observed effects in meta-analysis were robust in the sensitivity analysis, and the overall estimate of effect size was not significantly driven by a single study.

### 5. Conclusions

This present study provides for the first time a quantitative pooled estimate of the impact of NS supplementation on plasma lipid concentrations evaluated in RCTs, showing that NS significantly reduces plasma concentrations of total cholesterol, LDL-C, and triglycerides. These results are intriguing, considering the good safety profile and low cost of NS. Additional studies are required to define the optimal dosage and duration of this supplementation to obtain a favorable effect on lipid blood values. Finally, the value of adding NS supplements to conventional and novel LDL- [105-108] and triglyceride-lowering therapies [86, 109, 110] remains to be investigated in future studies.

# **Funding source**

No funding was received for this study.

### References

- [1] Bakathir HA, Abbas NA. Detection of the antibacterial effect of nigella sativa ground seedswith water. Afr J Tradit Complement Altern Med. 2011;8(2):159-64.
- [2] Shafiee-Nick R, Ghorbani A, Vafaee Bagheri F, Rakhshandeh H. Chronic administration of a combination of six herbs inhibits the progression of hyperglycemia and decreases serum lipids and aspartate amino transferase activity in diabetic rats. Adv Pharmacol Sci. 2012;2012:789796. doi: 10.1155/2012/789796.
- [3] Ghannadi A, Hajhashemi V, Jafarabadi H. An investigation of the analgesic and antiinflammatory effects of Nigella sativa seed polyphenols. J Med Food. 2005;8(4):488-93. doi: 10.1089/jmf.2005.8.488.
- [4] Swamy SMK, Tan BKH. Cytotoxic and immunopotentiating effects of ethanolic extract of Nigella sativa L. seeds. J Ethnopharmacol. 2000;70(1):1-7. doi: 10.1016/S0378-8741(98)00241-4.
- [5] Kanter M, Akpolat M, Aktas C. Protective effects of the volatile oil of Nigella sativa seeds on b-cell damage in streptozotocin-induced diabetic rats: A light and electron microscopic study. J Mol Histol. 2009;40(5-6):379-85. doi: 10.1007/s10735-009-9251-0.
- [6] Meral I, Yener Z, Kahraman T, Mert N. Effect of Nigella sativa on Glucose Concentration, Lipid Peroxidation, Anti-Oxidant Defence System and Liver Damage in Experimentally-Induced Diabetic Rabbits. J Vet Med A Physiol Pathol Clin Med. 2001;48(10):593-9. doi: 10.1046/j.1439-0442.2001.00393.x.
- [7] Salama RH. Hypoglycemic effect of lipoic acid, carnitine and Nigella sativa in diabetic rat model. Int J Health Sci. 2011;5(2):126-34.
- [8] Burits M, Bucar F. Antioxidant activity of Nigella sativa essential oil. Phytother Res. 2000;14(5):323-8. doi: 10.1002/1099-1573(200008)14:5<323::AID-PTR621>3.0.CO;2-Q.
- [9] Dehkordi FR, Kamkhah AF. Antihypertensive effect of Nigella sativa seed extract in patients with mild hypertension. Fundam Clin Pharmacol. 2008;22(4):447-52. Epub 2008/08/19. doi: 10.1111/j.1472-8206.2008.00607.x. PubMed PMID: 18705755.
- [10] Haque SF, Nasiruddin M, Najmi A. Indigenous herbal product Nigella sativa proved effective as an anti-obesity therapy in metabolic syndrome. Int J Medicobiol Res. 2011;1(3):173-6.
- [11] Gilani AH, Aziz N, Khurram IM, Chaudhary KS, Iqbal A. Bronchodilator, spasmolytic and calcium antagonist activities of Nigella sativa seeds (Kalonji): A traditional herbal product with multiple medicinal uses. J Pak Med Assoc. 2001;51(3):115-20.
- [12] Aboul Ezz HS, Khadrawy YA, Noor NA. The neuroprotective effect of curcumin and nigella sativa oil against oxidative stress in the pilocarpine model of epilepsy: A comparison with valproate. Neurochem Res. 2011;36(11):2195-204. doi: 10.1007/s11064-011-0544-9.
- [13] Shabana A, El-Menyar A, Asim M, Al-Azzeh H, Al Thani H. Cardiovascular benefits of black cumin (Nigella sativa). Cardiovasc Toxicol. 2013;13(1):9-21. doi: 10.1007/s12012-012-9181-z.
- [14] Duncker SC, Philippe D, Martin-Paschoud C, Moser M, Mercenier A, Nutten S. Nigella sativa (Black Cumin) seed extract alleviates symptoms of allergic diarrhea in mice, involving opioid receptors. PLoS ONE. 2012;7(6). doi: 10.1371/journal.pone.0039841.
- [15] Asgary S, Sahebkar A, Goli-Malekabadi N. Ameliorative effects of Nigella sativa on dyslipidemia. J Endocrinol Invest. 2015;38(10):1039-46. doi: 10.1007/s40618-015-0337-0.
- [16] Ahmad A, Husain A, Mujeeb M, Khan SA, Najmi AK, Siddique NA, et al. A review on therapeutic potential of Nigella sativa: A miracle herb. Asian Pac J Trop Biomed. 2013;3(5):337-52. doi: 10.1016/S2221-1691(13)60075-1.
- [17] Butt MS, Sultan MT. Nigella sativa: Reduces the risk of various maladies. Crit Rev Food Sci Nutr. 2010;50(7):654-65. doi: 10.1080/10408390902768797.
- [18] Heshmati J, Namazi N. Effects of black seed (Nigella sativa) on metabolic parameters in diabetes mellitus: A systematic review. Complement Ther Med. 2015;23(2):275-82. doi: 10.1016/j.ctim.2015.01.013.
- [19] Benhaddou-Andaloussi A, Martineau LC, Vallerand D, Haddad Y, Afshar A, Settaf A, et al. Multiple molecular targets underlie the antidiabetic effect of Nigella sativa seed extract in skeletal

- muscle, adipocyte and liver cells. Diabetes Obes Metab. 2010;12(2):148-57. doi: 10.1111/j.1463-1326.2009.01131.x.
- [20] Al-Naqeep G, Ismail M, Allaudin Z. Regulation of low-density lipoprotein receptor and 3-hydroxy-3- methylglutaryl coenzyme a reductase gene expression by thymoquinone-rich fraction and thymoquinone in HepG2 cells. J Nutrigenet Nutrigenomics. 2010;2(4-5):163-72. doi: 10.1159/000227264.
- [21] Reiner Z. Resistance and intolerance to statins. Nutr Metab Cardiovasc Dis. 2014;24(10):1057-66. doi: 10.1016/j.numecd.2014.05.009.
- [22] Zaoui A, Cherrah Y, Alaoui K, Mahassine N, Amarouch H, Hassar M. Effects of Nigella sativa fixed oil on blood homeostasis in rat. J Ethnopharmacol. 2002;79(1):23-6. doi: 10.1016/S0378-8741(01)00342-7.
- [23] Ahmad S, Beg ZH. Elucidation of mechanisms of actions of thymoquinone-enriched methanolic and volatile oil extracts from Nigella sativa against cardiovascular risk parameters in experimental hyperlipidemia. Lipids Health Dis. 2013;12(1). doi: 10.1186/1476-511X-12-86.
- [24] Ragheb A, Elbarbry F, Prasad K, Mohamed A, Ahmed MS, Shoker A. Attenuation of the development of hypercholesterolemic atherosclerosis by thymoquinone. Int J Angiol. 2008;17(4):186-92. doi: 10.1055/s-0031-1278307.
- [25] Parhizkar S, Latiff LA, Rahman SA, Dollah MA. Preventive effect of nigella sativa on metabolic syndrome in menopause induced rats. J Med Plant Res. 2011;5(8):1478-84.
- [26] Kaleem M, Kirmani D, Asif M, Ahmed Q, Bano B. Biochemical effects of Nigella sativa L seeds in diabetic rats. Indian J Exp Biol. 2006;44(9):745-8.
- [27] Bamosa AO, Ali BA, Sowayan SA. Effect of oral ingestion of Nigella sativa seeds on some blood parameters. Saudi Pharm J. 1997;5(2-3):126-9.
- [28] Bhatti IU, Ur Rehman F, Khan MA, Marwat SK. Effect of prophetic medicine kalonji [Nigella sativa I.] on lipid profile of human beings. An invivo approach. World Appl Sci J. 2009;6(8):1053-7.
- [29] Najmi A, Nasiruddin M, Khan R, Haque S. Effect of Nigella sativa oil on various clinical and biochemical parameters of insulin resistance syndrome. Int J Diabetes Dev Ctries. 2008;28(1):11-4. doi: 10.4103/0973-3930.41980.
- [30] Bamosa AO, Kaatabi H, Lebda FM, Al Elq AM, Al-Sultan A. Effect of Nigella Sativa seeds on the glycemic control of patients with type 2 diabetes mellitus. Indian J Physiol Pharmacol. 2010;54(4):344-54.
- [31] Sultan MT. Characterization of black cumin seed oil and exploring its role as a functional food. Faisalabad: University of Agriculture; 2009.
- [32] El-Dakhakhny M, Mady NI, Halim MA. Nigella sativa L. oil protects against induced hepatotoxicity and improves serum lipid profile in rats. Drug Res (Stuttg). 2000;50(9):832-6.
- [33] Asgary S, Ghannadi A, Dashti G, Helalat A, Sahebkar A, Najafi S. Nigella sativa L. improves lipid profile and prevents atherosclerosis: Evidence from an experimental study on hypercholesterolemic rabbits. J Funct Foods. 2013;5(1):228-34. doi: 10.1016/j.jff.2012.10.011.
- [34] Kocyigit Y, Atamer Y, Uysal E. The effect of dietary supplementation of Nigella sativa L. on serum lipid profile in rats. Saudi Med J. 2009;30(7):893-6.
- [35] Alobaidi AHA. Effect of nigella sativa and allium sativum coadminstered with simvastatin in dyslipidemia patients: A prospective, randomized, double-blind trial. Antiinflamm Antiallergy Agents Med Chem. 2014;13(1):68-74. doi: 10.2174/18715230113129990013.
- [36] Al-Rasheed N, Al-Rasheed N, Bassiouni Y, Faddah L, Mohamad AM. Potential protective effects of Nigella sativaand allium sativum against fructose-induced metabolic syndrome in rats. J Oleo Sci. 2014;63(8):839-48. doi: 10.5650/jos.ess14027.
- [37] Paul S, Paul KK, Palodhi S, Dutta S. Effect of atorvastatin and black seed (Nigella sativa) in experimentally induced hypercholesterolemia in rabbits. Pharmacologyonline. 2010;2:842-9.
- [38] Al-Nazawi MH, El-Bahr SM. Hypolipidemic and hypocholestrolemic effect of medicinal plant combination in the diet of rats: Black cumin seed (Nigella sativa) and turmeric (Curcumin). J Anim Vet Adv. 2012;11(12):2013-9. doi: 10.3923/javaa.2012.2013.2019.

- [39] Ebrahimzadeh Attari V, Pourghassem Gargari B, Rafraf M, Gorbani A, Tabibi H. Effect of ground black seed (Nigella sativa L.) on serum lipid profile, body weight and food intake in hyperlipidemic rabbits. J Zanjan Univ Med Sci Health Serv. 2010;18(70):31-43.
- [40] Ikram F, Hussain F. Antidiabetic efficacy of Nigella sativa Linn. in alloxan-induced diabetic rabbits. Int Med J Malaysia. 2014;13(1):13-8.
- [41] Parhizkar S, Latiff LA, Rahman SA, Hanachi P, Dollah MA. Metabolic impact of nigella sativa extracts on experimental menopause induced rats. J Appl Pharm Sci. 2011;1(9):38-42.
- [42] Suriyavathana Vedanarayanan M, Krishnan N. Ayurvedic Formulation of Liv-Pro-08 Reduces Nonalcoholic Fatty Liver Disease in Rats Fed with High-fat Diet. J Acupunct Meridian Stud. 2011;4(4):236-41. doi: 10.1016/j.jams.2011.09.014.
- [43] Nader MA, El-Agamy DS, Suddek GM. Protective effects of propolis and thymoquinone on development of atherosclerosis in cholesterol-fed rabbits. Arch Pharm Res. 2010;33(4):637-43. doi: 10.1007/s12272-010-0420-1.
- [44] Ahmad S, Beg ZH. Hypolipidemic and antioxidant activities of thymoquinone and limonene in atherogenic suspension fed rats. Food Chem. 2013;138(2–3):1116-24. doi: http://dx.doi.org/10.1016/j.foodchem.2012.11.109.
- [45] Najmi A, Nasiruddin M, Khan RA, Haque SF. Therapeutic effect of Nigella sativa in patients of poor glycemic control. Asian J Pharm Clin Res. 2012;5(SUPPL. 3):224-8.
- [46] Ghorbani A, Shafiee-Nick R, Rakhshandeh H, Borji A. Antihyperlipidemic Effect of a Polyherbal Mixture in Streptozotocin-Induced Diabetic Rats. J Lipids. 2013;2013:675759. doi: 10.1155/2013/675759. PubMed PMID: PMC3870091.
- [47] Amin F, Islam N, Anila N, Gilani AH. Clinical efficacy of the co-administration of Turmeric and Black seeds (Kalongi) in metabolic syndrome A double blind randomized controlled trial TAK-MetS trial. Complement Ther Med. 2015;23(2):165-74. doi: 10.1016/j.ctim.2015.01.008.
- [48] Amini M, Fallah Huseini H, Mohtashami R, Sadeqhi Z, Ghamarchehre MA. Hypolipidemic Effects of Nigella sativa L. Seeds Oil in Healthy Volunteers: a Randomized, Double-Blind, Placebo-Controlled Clinical Trial. J Med Plants. 2011;4(40):133-8.
- [49] Bin Sayeed MS, Asaduzzaman M, Morshed H, Hossain MM, Kadir MF, Rahman MR. The effect of Nigella sativa Linn. seed on memory, attention and cognition in healthy human volunteers. J Ethnopharmacol. 2013;148(3):780-6. doi: http://dx.doi.org/10.1016/j.jep.2013.05.004.
- [50] Datau EA, Wardhana, Surachmanto EE, Pandelaki K, Langi JA, Fias. Efficacy of Nigella sativa on serum free testosterone and metabolic disturbances in central obese male. Acta Med Indones. 2010;42(3):130-4. Epub 2010/08/21. PubMed PMID: 20724766.
- [51] Farzaneh E, Nia FR, Mehrtash M, Mirmoeini FS, Jalilvand M. The Effects of 8-week Nigella sativa Supplementation and Aerobic Training on Lipid Profile and VO2 max in Sedentary Overweight Females. Int J Prev Med. 2014;5(2):210-6. Epub 2014/03/15. PubMed PMID: 24627749; PubMed Central PMCID: PMCPMC3950745.
- [52] Fatima A, Shad MN, Asrar A, Murad S. Effects of Nigella Sativa on HDL-c & Body Weight. Pak J Med Health Sci. 2014;8(1):122-4.
- [53] Hadi S, Mirmiran P, Hosseinpour-Niazi S, Hedayati M, Azizi F. Effect of nigella sativa oil extract on lipid profiles in type 2 diabetic patients: A randomized, double blind, placebo-controlled clinical trial. Iran J Endocrinol Metab. 2015;16(6):411-8.
- [54] Heshmati J, Namazi N, Memarzadeh MR, Taghizadeh M, Kolahdooz F. Nigella sativa oil affects glucose metabolism and lipid concentrations in patients with type 2 diabetes: A randomized, double-blind, placebo-controlled trial. Food Res Int. 2015;70:87-93. doi: 10.1016/j.foodres.2015.01.030.
- [55] Hosseini MS, Mirkarimi SA, Amini M, Mohtashami R, Kianbakht S, Fallah Huseini H. Effects of Nigella sativa L. seed oil in type II diabetic patients: A randomized, double-blind, placebo Controlled clinical trial. J Med Plants. 2013;12(47):93-9.

- [56] Ibrahim RM, Hamdan NS, Ismail M, Saini SM, Rashid SNA, Latiff LA, et al. Protective effects of Nigella sativa on metabolic syndrome in menopausal women. Adv Pharm Bull. 2014;4(1):29-33. doi: 10.5681/apb.2014.005.
- [57] Ibrahim RM, Hamdan NS, Mahmud R, Imam MU, Saini SM, Rashid SNA, et al. A randomised controlled trial on hypolipidemic effects of Nigella Sativa seeds powder in menopausal women. J Transl Med. 2014;12(1). doi: 10.1186/1479-5876-12-82.
- [58] Latiff LA, Parhizkar S, Dollah MA, Tajuddin Syed Hassan S. Alternative supplement for enhancement of reproductive health and metabolic profile among perimenopausal women: A novel role of Nigella sativa. Iran J Basic Med Sci. 2014;17(12):980-5.
- [59] Mahdavi R, Namazi N, Alizadeh M, Farajnia S. Effects of Nigella sativa oil with a low-calorie diet on cardiometabolic risk factors in obese women: A randomized controlled clinical trial. Food Funct. 2015;6(6):2041-8. doi: 10.1039/c5fo00316d.
- [60] Moeen-ud-din H, Murad S, Fatima A. Placebo controlled study on comparison of effects of Nigella sativa and nicotinic acid along with low fat diet and physical exercise on LDL-cholesterol and HDL-cholesterol. Pak J Med Health Sci. 2014;8(2):306-9.
- [61] Qidwai W, Hamza HB, Qureshi R, Gilani A. Effectiveness, safety, and tolerability of powdered Nigella sativa (kalonji) seed in capsules on serum lipid levels, blood sugar, blood pressure, and body weight in adults: results of a randomized, double-blind controlled trial. J Altern Complement Med. 2009;15(6):639-44.
- [62] Sabzghabaee AM, Dianatkhah M, Sarrafzadegan N, Asgary S, Alireza G. Clinical evaluation of Nigella sativa seeds for the treatment of hyperlipidemia: a randomized, placebo controlled clinical trial. Med Arh. 2012;66(3):198-200. doi: 10.5455/medarh.2012.66.198-200.
- [63] Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. BMJ. 2009;339(7716):332-6. doi: 10.1136/bmj.b2535.
- [64] Cochrane Handbook for Systematic Reviews of Interventions. Chichester, UK: John Wiley & Sons, Ltd; 2008.
- [65] Borenstein M, Hedges L, Higgins J, Rothstein HR. Comprehensive Meta-analysis. Version 2 ed. Englewood, NJ: Biostat; 2005.
- [66] Wan X, Wang W, Liu J, Tong T. Estimating the sample mean and standard deviation from the sample size, median, range and/or interquartile range. BMC Med Res Methodol. 2014;14(1). doi: 10.1186/1471-2288-14-135.
- [67] Sutton AJ, Abrams KR, Jones DR, Sheldon TA, Song F. Methods for Meta-analysis in Medical Research. Chichester, UK: John Wiley & Sons; 2000.
- [68] Derosa G, Maffioli P, Sahebkar A. Plasma uric acid concentrations are reduced by fenofibrate: A systematic review and meta-analysis of randomized placebo-controlled trials. Pharmacol Res. 2015 Dec;102:63-70.
- [69] Sahebkar A. Are curcuminoids effective C-reactive protein-lowering agents in clinical practice? evidence from a meta-analysis. Phytother Res. 2014;28(5):633-42. doi: 10.1002/ptr.5045.
- [70] Ferretti G, Bacchetti T, Sahebkar A. Effect of statin therapy on paraoxonase-1 status: A systematic review and meta-analysis of 25 clinical trials. Prog Lipid Res. 2015;60:50-73. Epub 2015/09/30. doi: 10.1016/j.plipres.2015.08.003. PubMed PMID: 26416579.
- [71] Duval S, Tweedie R. Trim and fill: A simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. Biometrics. 2000;56(2):455-63.
- [72] Nagy L, Tontonoz P, Alvarez JGA, Chen H, Evans RM. Oxidized LDL regulates macrophage gene expression through ligand activation of PPARγ. Cell. 1998;93(2):229-40. doi: 10.1016/S0092-8674(00)81574-3.
- [73] Chawla A, Boisvert WA, Lee CH, Laffitte BA, Barak Y, Joseph SB, et al. A PPARy-LXR-ABCA1 pathway in macrophages is involved in cholesterol efflux and atherogenesis. Mol Cell. 2001;7(1):161-71. doi: 10.1016/S1097-2765(01)00164-2.

- [74] Al-Naqeeb G, Ismail M. Regulation of apolipoprotein A-1 and apolipoprotein B100 genes by thymoquinone rich fraction and thymoquinone in HEPG2 cells. J Food Lipids. 2009;16(2):245-58. doi: 10.1111/j.1745-4522.2009.01144.x.
- [75] El-Dakhakhany M, editor Some Pharmacological Properties of Some Constituents of Nigella Sativa L Seeds: The Carbonyl Fraction of Essential Oil. Proceedings of the 2nd International Conference on Islamic Medicine; 1982.
- [76] Le PM, Benhaddou-Andaloussi A, Elimadi A, Settaf A, Cherrah Y, Haddad PS. The petroleum ether extract of Nigella sativa exerts lipid-lowering and insulin-sensitizing actions in the rat. J Ethnopharmacol. 2004;94(2-3):251-9. doi: 10.1016/j.jep.2004.04.030.
- [77] Moghadasian MH, Frohlich JJ. Effects of dietary phytosterols on cholesterol metabolism and atherosclerosis: Clinical and experimental evidence. Am J Med. 1999;107(6):588-94. doi: 10.1016/S0002-9343(99)00285-5.
- [78] Atta MB. Some characteristics of nigella (Nigella sativa L.) seed cultivated in Egypt and its lipid profile. Food Chem. 2003;83(1):63-8. doi: http://dx.doi.org/10.1016/S0308-8146(03)00038-4.
- [79] Alimohamadi K, Taherpour K, Ghasemi HA, Fatahnia F. Comparative effects of using black seed (Nigella sativa), cumin seed (Cuminum cyminum), probiotic or prebiotic on growth performance, blood haematology and serum biochemistry of broiler chicks. J Anim Physiol Anim Nutr. 2014;98(3):538-46.
- [80] Khan SH, Anjum MA, Parveen A, Khawaja T, Ashraf NM. Effects of black cumin seed (Nigella sativa L.) on performance and immune system in newly evolved crossbred laying hens. Vet Q. 2013;33(1):15-21. doi: 10.1080/01652176.2013.782119.
- [81] Dahri AH, Chandiol AM, Rahoo AA, Memon RA. Effect of Nigella sativa (kalonji) on serum cholesterol of albino rats. J Ayub Med Coll Abbottabad. 2005;17(2):72-4.
- [82] Agbaria R, Gabarin A, Dahan A, Ben-Shabat S. Anticancer activity of Nigella sativa (black seed) and its relationship with the thermal processing and quinone composition of the seed. Drug Des Devel Ther. 2015;9:3119-24. doi: 10.2147/DDDT.S82938.
- [83] Siddiqui MN, Islam MT, Sayed MA, Hossain MA. Effect of dietary supplementation of acetone extracts of Nigella sativa L. Seeds on serum cholesterol and pathogenic intestinal bacterial count in broilers. J Anim Plant Sci. 2015;25(2):372-9.
- [84] Islam MT, Selim ASM, Sayed MA, Khatun MA, Siddiqui MNEA, Alam MS, et al. Nigella sativa L. supplemented diet decreases egg cholesterol content and suppresses harmful intestinal bacteria in laying hens. J Anim Feed Sci. 2011;20(4):587-98.
- [85] Boka J, Mahdavi AF, Samie AH, Jahanian R. Effect of different levels of black cumin (Nigella sativa L.) on performance, intestinal Escherichia coli colonization and jejunal morphology in laying hens. J Anim Physiol Anim Nutr. 2014;98(2):373-83.
- [86] Sahebkar A, Chew GT, Watts GF. Recent advances in pharmacotherapy for hypertriglyceridemia. Prog Lipid Res. 2014;56(1):47-66. doi: 10.1016/j.plipres.2014.07.002.
- [87] Hodis HN, Mack WJ. Triglyceride-rich lipoproteins and progression of atherosclerosis. Eur Heart J. 1998;19(SUPPL. A):A40-A4.
- [88] Morikawa T, Xu F, Ninomiya K, Matsuda H, Yoshikawa M. Nigellamines A3, A4, A5, and C, new dolabellane-type diterpene alkaloids, with lipid metabolism-promoting activities from the Egyptian medicinal food black cumin. Chem Pharm Bull. 2004;52(4):494-7. doi: 10.1248/cpb.52.494.
- [89] Boden WE, Probstfield JL, Anderson T, Chaitman BR, Desvignes-Nickens P, Koprowicz K, et al. Niacin in patients with low HDL cholesterol levels receiving intensive statin therapy. N Engl J Med. 2011;365(24):2255-67. doi: 10.1056/NEJMoa1107579.
- [90] Landray MJ, Haynes R, Hopewell JC, Parish S, Aung T, Tomson J, et al. Effects of extended-release niacin with laropiprant in high-risk patients. N Engl J Med. 2014;371(3):203-12. doi: 10.1056/NEJMoa1300955.
- [91] Tuteja S, Rader DJ. Dyslipidaemia: Cardiovascular prevention End of the road for niacin? Nat Rev Endocrinol. 2014;10(11):646-7. doi: 10.1038/nrendo.2014.159.

- [92] Siddiqi HK, Kiss D, Rader D. HDL-cholesterol and cardiovascular disease: Rethinking our approach. Curr Opin Cardiol. 2015;30(5):536-42. doi: 10.1097/HCO.000000000000011.
- [93] Tasawar Z, Siraj Z, Ahmad N, Lashari MH. The effects of nigella sativa (Kalonji) on lipid profile in patients with stable coronary artery disease in Multan, Pakistan. Pak J Nutr. 2011;10(2):162-7.
- [94] Fallah Huseini H, Amini M, Mohtashami R, Ghamarchehre ME, Sadeqhi Z, Kianbakht S, et al. Blood pressure lowering effect of Nigella sativa I. seed oil in healthy Volunteers: A randomized, double-blind, placebo-controlled clinical trial. Phytother Res. 2013;27(12):1849-53. doi: 10.1002/ptr.4944.
- [95] Al-Naqeep G, Al-Zubairi AS, Ismail M, Amom ZH, Esa NM. Antiatherogenic potential of nigella sativa seeds and oil in diet-induced hypercholesterolemia in rabbits. J Evid Based Complementary Altern Med. 2011;2011:213628. doi: 10.1093/ecam/neq071.
- [96] Enomoto S, Asano R, Iwahori Y, Narui T, Okada Y, Singab ANB, et al. Hematological studies on black cumin oil from the seeds of Nigella sativa L. Biol Pharm Bull. 2001;24(3):307-10. doi: 10.1248/bpb.24.307.
- [97] Idris-Khodja N, Schini-Kerth V. Thymoquinone improves aging-related endothelial dysfunction in the rat mesenteric artery. Naunyn Schmiedebergs Arch Pharmacol. 2012;385(7):749-58. doi: 10.1007/s00210-012-0749-8.
- [98] El Tahir KEH, Al-Ajmi MF, Al-Bekairi AM. Some cardiovascular effects of the dethymoquinonated Nigella sativa volatile oil and its major components  $\alpha$ -pinene and p-cymene in rats. Saudi Pharm J. 2003;11(3):104-10.
- [99] Al-Hariri MT, Yar T, Bamosa AO, El-Bahai MN. Effects of two-months Nigella sativa supplementation on cardiac hemodynamics and adrenergic responsiveness. J Pak Med Assoc. 2009;59(6):363-7.
- [100] Ali BH, Blunden G. Pharmacological and toxicological properties of Nigella sativa. Phytother Res. 2003;17(4):299-305. doi: 10.1002/ptr.1309.
- [101] Mansour MA, Ginawi OT, El-Hadiyah T, El-Khatib AS, Al-Shabanah OA, Al-Sawaf HA. Effects of volatile oil constituents of Nigella sativa on carbon tetrachloride-induced hepatotoxicity in mice: Evidence for antioxidant effects of thymoquinone. Res Commun Mol Pathol Pharmacol. 2001;110(3-4):239-51.
- [102] Al-Amri AM, Bamosa AO. Phase I safety and clinical activity study of thymoquinone in patients with advanced refractory malignant disease. Shiraz E Med J. 2009;10(3):107-11.
- [103] Coban S, Yildiz F, Terzi A, Al B, Aksoy N, Bitiren M, et al. The effects of Nigella sativa on bile duct ligation induced-liver injury in rats. Cell Biochem Funct. 2010;28(1):83-8. doi: 10.1002/cbf.1624.
- [104] Ibraheim ZZ. Effect of Nigella sativa seeds and total oil on some blood parameters in female volunteers. Saudi Pharm J. 2002;10(1-2):54-9.
- [105] Sahebkar A, Watts GF. New LDL-cholesterol lowering therapies: Pharmacology, clinical trials, and relevance to acute coronary syndromes. Clin Ther. 2013;35(8):1082-98. doi: 10.1016/j.clinthera.2013.06.019.
- [106] Banach M, Aronow WS, Serban C, Sahabkar A, Rysz J, Voroneanu L, et al. Lipids, blood pressure and kidney update 2014. Pharmacol Res. 2015;95-96:111-25. doi: 10.1016/j.phrs.2015.03.009.
- [107] Sahebkar A, Watts GF. New therapies targeting apoB metabolism for high-risk patients with inherited dyslipidaemias: What can the clinician expect? Cardiovasc Drugs Ther. 2013;27(6):559-67. doi: 10.1007/s10557-013-6479-4.
- [108] Sahebkar A, Watts GF. Managing recalcitrant hypercholesterolemia in patients on current best standard of care: Efficacy and safety of novel pharmacotherapies. Clin Lipidol. 2014;9(2):221-33. doi: 10.2217/clp.14.14.
- [109] Sahebkar A, Chew GT, Watts GF. New peroxisome proliferator-activated receptor agonists: potential treatments for atherogenic dyslipidemia and non-alcoholic fatty liver disease. Expert Opin Pharmacother. 2014;15(4):493-503. doi: 10.1517/14656566.2014.876992.

[110] Sahebkar A, Watts GF. Role of selective peroxisome proliferator-activated receptor modulators in managing cardiometabolic disease: Tale of a roller-coaster. Diabetes Obes Metab. 2014;16(9):780-92. doi: 10.1111/dom.12277.

**Table 1.** Demographic characteristics of the included studies.

Author	Study design	Target Population	Treatment duration	n	Study groups	Age, years	Female (n, %)	BMI, (kg/m <sup>2</sup> )	Systolic blood pressure (mmHg)	Diastolic blood pressure (mmHg)	Total cholesterol (mg/dl)	LDL cholesterol (mg/dl)	HDL cholesterol (mg/dl)	Triglycerides (mg/dl)
Amin et al. (2015)	Randomized, double-blind,	Men with metabolic	8 weeks											
(2013)	placebo- controlled	syndrome		62	NS powder 1.5 g/day	45.1±11.7	0 (0.0)	27.4±3.1	131.8±20.2	82.5±12.1	184.3±33.6	110.2±28.0	34.3±7.8	169.5±44.3
				63	Placebo	41.5±12.8	0 (0.0)	27.5±4.1	125.5±16.7	76.8±11.5	180.8±23.3	119.5±27.3	33.7±7.4	163.6±42.7
Bin Sayeed et al. (2013)	Randomized, double-blind,	Healthy male	9 weeks											
	placebo- controlled	volunteers		20	NS powder 1 g/day	55.8±0.57	0 (0.0)	24.77±0.34	ND	ND	148.9±8.4	113.7±8.4	25.1±12.0	116.0±3.1
				20	Placebo	55.9±0.65	0 (0.0)	24.55±0.18	ND	ND	151.2±5.3	112.1±13.8	29.0±12.0	114.3±2.2
Datau et al. (2010)	Randomized, double-blind.	Men with central	3 months											
( /	placebo- controlled	obesity		19	NS powder 1.5 g/day	ND	0 (0.0)	ND	130.53±13.11	80.53±13.93	ND	ND	35.60±5.21	202.05±134.31
				20	Placebo	ND	0 (0.0)	ND	123.50±12.68	80.0±7.96	ND	ND	39.14±7.65	115.05±81.06
Dehkordi et al. (2008)	Randomized, double-blind,	Men with	8 weeks											
ai. (2008)	placebo- controlled	hypertension		36	NS oil 100 mg/day	44.6±1.3	0 (0.0)	23.9±0.8	151.2±1.3	93.2±0.5	201.2±9.1	128.2±6.1	51.8±3.4	127.9±10.7
				39	NS oil 200 mg/day	43.7±1.3	0 (0.0)	24.1±0.8	149.5±1.3	95.1±0.8	200.7±8.2	127.7±6.8	52.7±3.1	118.0±9.7
				33	Placebo	43.1±1.4	0 (0.0)	24.5±0.7	148.2±1.2	94.5±0.8	189.7±8.7	123.5±5.4	47.2±2.9	121.2±10.3
Farzaneh et al. (2014)	Randomized, double-blind,	Overweight females with	8 weeks	8	NS powder 2 g/day	34.14±10.54	8 (100.0)	25.39±0.75	ND	ND	221.5±31.34	119.1±11.96	45.62±10.02	167.88±18.61
ai. (2014)	placebo- controlled	hypercholes- terolemia		8	Placebo	33.0±4.34	8 (100.0)	25.85±1.45	ND	ND	232.75±12.85	120.5±10.39	48.37±8.61	173.8±50.43
Fatima et al. (2014)	Single-blind,	Hyper- lipidemia	4 weeks											
(2014)	controlled	присши		30	NS powder 2 spoons/day	ND	ND	ND	ND	ND	ND	ND	31.7±3.11	ND
				30	Placebo	ND	ND	ND	ND	ND	ND	ND	35.87±2.22	ND

Heshmati et al. (2015)	Randomized, double-blind,	Type 2 diabetes	12 weeks											
(,	placebo- controlled			36	NS oil 3 g/day	45.3±6.5	54.3*	29.5±4.4	ND	ND	216.8±51.7	132.9±48.0	38.2±5.3	228.0±74.4
				36	Placebo	47.5±8.0	51.4*	28.6±4.3	ND	ND	228.3±49.0	140.6±44.8	38.5±5.5	244.4±59.8
Hosseini et al. (2013)	Randomized, double-blind,	Type 2 diabetes	3 months											
	placebo- controlled			35	NS oil 5 ml/day	48.74±7.33	21 (60.0)	30.81±3.55	ND	ND	250.0±30.0	171.8±27.0	47.7±9.2	182.5±62.3
				35	Placebo	50.72±5.69	19 (54.2)	30.92±3.67	ND	ND	246.9±31.1	168.0±7.1	46.8±9.0	179.9±52.4
Ibrahim et al. (2014) <sup>a</sup>	Randomized, placebo-	Menopausal women with	2 months											
	controlled	metabolic syndrome		18	NS powder 1 g/day	ND	18 (100.0)	ND	ND	ND	233.1±40.4	179.7±35.8	60.9±10.0	133.7±31.6
				17	Placebo	ND	17 (100.0)	ND	ND	ND	234.2±40.7	186.7±23.1	52.5±10.9	132.6±42.8
Ibrahim et al. (2014) <sup>b</sup>	Randomized, placebo-	Menopausal women	2 months											
	controlled			19	NS powder 1 g/day	53.22±2.16	19 (100.0)	27.18±4.34	129.33±15.44	77.13±9.16	235.5±40.2	179.8±34.4	50.7±8.5	155.9±31.0
				18	Placebo	53.71±3.57	18 (100.0)	27.75±4.38	138.40±18.90	83.93±15.73	234.3±39.4	177.5±18.6	51.0±9.7	152.3±46.9
Latiff et al. (2014)	Open label single arm,	Peri- menopausal	12 weeks			50.1±7.6								
	controlled	women		69	NS powder 1.6 g/day		69 (100.0)	26.31±4.89	122.32±15.25	79.07±7.76	225.1±47.2	165.9±204.216 6.3±198.0	60.3±14.7	125.8±74.4
				69	Placebo		69 (100.0)	26.03±4.66	121.42±14.89	78.42±8.48	225.8±46.0		58.8±17.4	124.9±70.0
Mahdavi et al. (2015)	Randomized, double-blind,	Obese women	8 weeks	43	NS oil 3 g/day	41.5±11.7	43 (100.0)	32.4±1.5	120.5±10.3	7.7±0.7	203.1±42.2	129.1±32.3	48.8±11.6	130.2±65.9
	placebo- controlled			41	Placebo	39.3±9.9	41 (100.0)	32.6±1.5	120.4±10.4	7.9±0.6	191.7±41.1	119.2±33.0	49.3±13.4	115.5±64.7
Moeen-Ud- Din et al.	Single-blind,	Hyper- lipidemia	6 weeks	27	NS powder 2 spoons/day	ND	ND	ND	ND	ND	ND	202.45±1.54	38.81±3.90	ND
(2014)	controlled	processia		30	Placebo	ND	ND	ND	ND	ND	ND	189.15±3.90	36.11±2.11	ND
Qidwai et al. (2009)	Randomized, double-blind,	Hypercholes -terolemia	6 weeks	39	NS powder 2 g/day	45.58±10.86	6 (10.0)	27.13±3.88	128.90±18.37	81.82±11.24	209.07±28.63	145.76±23.30	40.53±8.52	163.14±71.43
	placebo- controlled			34	Placebo	46.86±11.00	8 (14.0)	28.26±6.75	122.30±17.76	80.45±11.48	217.11±27.72	144.43±24.0	41.74±10.63	157.12±84.53

omized, Hypercholes	4 weeks	37	NS powder 2 g/day	40.38**	17 (45.9)	25.01**	ND	ND	235.24±28.29	144.58±19.06	51.48±15.45	173.91±69.35
olled		37	Placebo	38.4**	16 (43.2)	23.19**	ND	ND	233.39±26.24	138.36±19.05	48.03±8.70	173.88±47.92
omized, Type 2 bo- diabetes	8 weeks	23	NS oil 1 g/day	51.4±9.2	10 (43.5)	28.4±4.4	ND	ND	189.0±48.2	114.0±38.2	48.1±7.5	156.0±73.9
olled		20	Placebo	56.0±3.4	10 (50.0)	28.8±8.1	ND	ND	175.0±41.7	102.0±39.6	48.2±10.5	142±61.8
omized, Healthy bo- subjects with	8 weeks	35	NS oil 5 mL/day	42.3±13.8	17 (48.5)	ND	ND	ND	196.7±35.7	111.5±24.2	43.0±10.1	210.2±103.8
total 200 mg/dL total cholesterol 300 mg/dL		35	Placebo	36.3±13.6	18 (51.5)	ND	ND	ND	184.2±28.2	112.4±18.1	42.4±6.4	191.6±6.4
t	bo- omized, Type 2 bo- diabetes olled  omized, Healthy bo- subjects with 200 mg/dL< total cholesterol<	bo- bo- clied  Type 2 diabetes  bo- diabetes  bo- subjects with clied  200 mg/dL< total cholesterol<	bo- omized, Type 2 8 weeks 23 bo- diabetes olled 20 omized, Healthy 8 weeks 35 bo- subjects with olled 200 mg/dL 35 total cholesterol	bo- omized, Type 2 8 weeks 23 NS oil 1 g/day bo- diabetes omized, Healthy 8 weeks 35 NS oil 5 mL/day bo- subjects with olled 200 mg/dL< total cholesterol<	Second   S	bo- onlized, Type 2 8 weeks 23 NS oil 1 g/day 51.4±9.2 10 (43.5) bo- diabetes olled 20 Placebo 56.0±3.4 10 (50.0) omized, Healthy 8 weeks 35 NS oil 5 mL/day 42.3±13.8 17 (48.5) bo- subjects with olled 200 mg/dL         35 Placebo 36.3±13.6 18 (51.5) total cholesterol	bo- bolled -terolemia -terolemia 37 Placebo 38.4** 16 (43.2) 23.19**  omized, Type 2 8 weeks 23 NS oil 1 g/day 51.4±9.2 10 (43.5) 28.4±4.4  bo- diabetes olled 20 Placebo 56.0±3.4 10 (50.0) 28.8±8.1  omized, Healthy 8 weeks 35 NS oil 5 mL/day 42.3±13.8 17 (48.5) ND  omized, bo- subjects with olled 200 mg/dL	Second   S	December   First   December   D	bo- bolled -terolemia	-terolemia	bo- terolemia - te

Values are expressed as mean ± SD \*Percentage only Abbreviations: NS, *Nigella sativa*; ND, no data; BMI, body mass index.

**Table 2.** Risk of bias assessment of the included studies according to the Cochrane guidelines.

Study	Sequence generation	Allocation	Blinding of participants, personnel and outcome assessors	Incomplete outcome data	Selective outcome reporting	Other sources of bias
Amin et al. (2015)	L	L	L	L	L	L
Bin Sayeed et al. (2013)	L	L	L	L	L	L
Datau et al. (2010)	U	U	U	L	L	U
Dehkordi et al. (2008)	U	U	U	L	L	U
Farzaneh et al. (2014)	L	U	U	L	L	U
Fatima et al. (2014)	U	U	U	Н	U	U
Heshmati et al. (2015)	L	L	L	L	L	L
Hosseini et al. (2013)	L	U	U	L	U	U
Ibrahim et al. (2014) <sup>a</sup>	L	U	U	U	U	U
Ibrahim et al. (2014) <sup>b</sup>	U	U	U	L	L	U
Latiff et al. (2014)	Н	Н	Н	L	L	Н
Mahdavi et al. (2015)	L	L	L	L	L	L

Moeen-Ud-Din et al. (2014)	U	U	U	Н	U	U
Qidwai et al. (2009)	L	L	L	L	L	L
Sabzghabaee et al. (2012)	L	U	U	L	L	U
Hadi et al., (2015)	U	U	L	L	Н	L
Amini et al. (2013)	U	U	L	L	L	L

L, low risk of bias; H, high risk of bias; U, unclear risk of bias.

**Table 3.** Assessment of publication bias in the meta-analysis of NS effects on plasma concentrations of lipids.

	Correct	ed effect size <sup>a</sup>	Begg's rank corr	elation tes	st,	Egger's lin	Fail safe N test		
	WMD	95% CI	Kendall's <b>Tau</b> <sup>a</sup>	z-value	<i>p</i> -value	Intercept	95% CI	<i>p</i> -value	$n^b$
Total cholesterol	-	-	0.11	0.59	0.553	-0.32	-3.53, 2.89	0.834	432
LDL-C	-17.21	-22.66, -11.76	-0.06	0.32	0.753	-0.50	-2.54, 1.55	0.610	982
HDL-C	-0.63	-2.76, 1.50	-0.14	0.83	0.405	0.99	-1.68, 3.65	0.445	-
Triglycerides	-17.80	-27.10, -8.50	0.14	0.77	0.44	-2.04	-3.44, -0.65	0.007	347

<sup>&</sup>lt;sup>a</sup>With continuity correction; <sup>b</sup>Number of theoretically missing studies to bring the p-value to > 0.05.

#### FIGURE LEGENDS

- Figure 1. Flow chart of the number of studies identified and included into the meta-analysis.
- **Figure 2.** Forest plot displaying weighted mean difference and 95% confidence intervals for the impact of NS supplementation on plasma lipid concentrations.
- **Figure 3.** Results of leave-one-out sensitivity analysis for the impact of NS supplementation on plasma lipid concentrations.
- **Figure 4.** Forest plot displaying weighted mean difference and 95% confidence intervals for the impact of NS supplementation on plasma lipid concentrations in the subgroups of trials studying the effects of NS powder and black seed oil.
- **Figure 5.** Random-effects meta-regression plots of the association between mean changes in plasma concentrations of lipids and NS dose.
- **Figure 6.** Random-effects meta-regression plots of the association between mean changes in plasma concentrations of lipids and duration of NS supplementation.
- **Figure 7.** Funnel plot displaying publication bias in the studies reporting the impact of NS supplementation on plasma lipid concentrations.