

# UNIVERSITÀ DEGLI STUDI DI TORINO

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5	Sex- and age-related variation in metal content of penguin feathers
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## 18 Abstract

The presence of xenobiotics, such as metals, in ecosystems is concerning due to their durability and 19 they pose a threat to the health and life of organisms. Moreover, mercury can biomagnify in many 20 marine food chains and, therefore, organisms at higher trophic levels can be adversely impacted. 21 Although feathers have been used extensively as a bio-monitoring tool, only a few studies have 22 addressed the effect of both age and sex on metal accumulation. In this study, the concentrations of 23 trace elements were determined in the feathers of all members of a captive colony of African 24 Penguins (Spheniscus demersus) housed in a zoological facility in Italy. Tests were performed by 25 Inductively Coupled Plasma - Mass Spectrometry (ICP-MS) to detect aluminum, arsenic, cadmium, 26 27 cobalt, chromium, copper, iron, manganese, nickel, lead, selenium, tin, vanadium, and zinc. Mercury was detected by a Direct Mercury Analyzer. Sexing was performed by a molecular 28 approach based on analyzing the chromo-helicase-DNA-binding1 (CHD1) gene, located on the sex 29 chromosomes. Sex- and age-related differences were studied in order to investigate the different 30 patterns of metal bioaccumulation between male and female individuals and between adults and 31 juveniles. Juvenile females had significantly higher arsenic levels than males, while selenium levels 32 increased significantly with age in both sexes. Penguins kept in controlled environments- given that 33 34 diet and habitat are under strict control- represent a unique opportunity to determine if and how metal bioaccumulation is related to sex and age. 35

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- 37 *Keywords:* African penguin, seabirds, *Spheniscus*, trace elements.

#### 38 **1. Introduction**

Over the last decade, there has been an increasing interest in the use of sentinel organisms for 39 pollution monitoring studies (Burger et al. 2008). Of these, birds have received particular attention, 40 41 because they are exposed to heavy metals, both through environmental exposure and diet (Roux and Marra 2007). While many host factors, such as size and age, have received considerable attention in 42 metal bioaccumulation studies in seabirds, gender has received relatively little attention (Burger et 43 al., 2003). Adults of larger species, which have longer lifespans and are at higher trophic position 44 were found to accumulate higher levels of mercury (Hg) in their feathers, due to biomagnification 45 phenomena through the food chain (Burger and Gochfeld, 2000; Carravieri et al., 2013). Moreover, 46 in long-living seabirds, adults have higher metal content in their feathers than chicks, as the time 47 48 interval available for accumulation is longer in adults (Burger and Gochfeld, 2000; Bond and Diamond, 2009b). However, there are sex-based differences in the excretion of contaminants and 49 sex-related differences in the fate and effects of chemicals in seabirds; sex can affect exposure and 50 susceptibility to contaminants and influences the ability to rid the body of these pollutants (Burger 51 et al., 2007). Most excretion methods are similar for both males and females, but females can also 52 excrete contaminants in their eggs and embryos (Burger et al. 2003). 53

Bird's feathers are useful non-invasive indicators of metal contamination, as a relatively high 54 proportion of the body burden of certain metals, such as mercury, is stored in the feathers, because 55 of their affinity to -SH groups of keratin. Moreover, several studies have shown that a high 56 correlation exists between levels of certain contaminants in the diet of birds and relative levels 57 detected in their feathers (e.g. mercury, Becker et al. 2002; Brasso et al. 2014). Another advantage 58 of using feathers in such analyses is that they can be easily collected and, if necessary, repeatedly 59 sampled without affecting the health and condition of the individuals being studied (Adout et al. 60 2007). However, in many studies using bird feathers as an indication of internal body burdens, the 61 sex of the different individuals is not determined (Burger and Gochfeld, 2000; Dauwe et al. 2002; 62 Markowski et al. 2013; Ansara-Ross et al. 2013). Accordingly, many bird species (including 63

penguins) are not obviously sexually dimorphic and sex determination is therefore impossible in the 64 field. In the very few studies considering sexual differences, a limited number of elements were 65 investigated. Dauwe et al. (2002) examined cadmium (Cd), lead (Pb), copper (Cu) and zinc (Zn) 66 levels in great and blue tits, considering both age and gender, and they found that only the Zn 67 concentration was significantly higher in males than in females. Lucia et al. (2010) investigated the 68 presence of aluminum (Al), arsenic (As), Cd, Cu, Pb, Hg, nickel (Ni), selenium (Se) and Zn in 69 feathers of aquatic birds. They found that Cd accumulation increased with age, while As was 70 influenced by sex, as female birds displayed higher concentrations in the liver and feathers 71 compared to male birds. Mansouri and Hoshyari (2012) investigated the levels of Ni in the feathers 72 of the Western Reef Heron (Egretta gularis) and Siberian Gull (Larus heuglini), but no significant 73 74 differences were found between gender and age groups in either species. Finally, very few studies have been designed to determine age- or gender-differences in metal content of penguin tissues. 75 These studies have mostly focused on Hg (e.g. Frias et al. 2012; Carravieri et al. 2013) and did not 76 consider other trace elements. Feather metal concentrations were generally found to be species-77 specific, and a high variance in metal levels between individuals for some species, but not for 78 others, were registered (Burger and Gochfeld, 2000). 79

The African or Jackass Penguin (Spheniscus demersus) is a colonial seabird endemic to South 80 81 Africa and Namibia. The current conservation status of this species is "endangered", according to the Red List of Threatened Species of the IUCN (International Union for Conservation of Nature), 82 and the wild African Penguin population has dramatically decreased in recent years to less than 83 75,000-80,000 mature individuals (Birdlife International, 2013). The African Penguin therefore 84 faces a high risk of extinction, and as a result, in-situ conservation programs are becoming 85 increasingly crucial. The body size of an African penguin is 45–68 cm in height and 3 kg in weight. 86 Sexual dimorphism is not evident, but males are generally larger than females, although 87 measurements tend to overlap. In the wild, individuals of the species tend to live from 10 to 27 88 89 years (Whittington et al. 2000) but captive birds live significantly longer. Sexual maturity is reached at 3-4 years in wild specimens, but captive penguins are observed to breed at 2-3 years. After obtaining the adult plumage, penguins molt annually, and the feather-shedding phase lasts for 12.7  $\pm$  1.4 days (Randall et al. 1986). As breeding and molting are energetically demanding activities in the annual cycle of the adult penguin, the timing of these events should coincide with periods of favorable environmental conditions, particularly with the availability of food.

Several African penguin colonies are housed in zoos and aquaria worldwide for *ex-situ* conservation 95 purposes. According to the International Species Information System (www.isis.org), 2394 96 Spheniscus demersus individuals live under human care (species holding data for Spheniscus as of 97 September 27, 2014). The use of captive birds to monitor pollutants offers several advantages over 98 99 the analysis of biotic and abiotic matrices (Falkowska et al. 2013). Of these, seabirds are considered 100 to be among the most reliable indicators of environmental changes; with specific reference to penguins, they are particularly relevant as they occupy a high position in many marine food chains 101 102 and accumulate metals and other toxic elements in their tissues at concentrations of several orders of magnitude above environmental levels (Barbieri et al. 2010). Moreover, penguins living in ex-103 situ colonies are confined to controlled areas and have a very special and homogeneous diet, usually 104 limited to one or two species of commercially-available fish. Therefore, they constitute a unique 105 opportunity to investigate sex- and age-related differences in trace elements using the feathers as a 106 107 noninvasive bio-monitoring tool. Our aim was to assess whether sex and/or age can affect the accumulation of essential (arsenic, chromium, copper, iron, manganese, nickel, selenium and zinc) 108 and non-essential (aluminum, cadmium, cobalt, lead, mercury, tin, vanadium) elements in a large 109 captive colony of African penguins. 110

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- 112 2. Methods and Materials
- 113 2.1 Ethical statement

This research conformed to the Ethical Guidelines for the Conduct of Research on Animals by Zoosand Aquariums (WAZA 2005), and was conducted with the approval of the Ethical Committee of

the Istituto Zooprofilattico Sperimentale del Piemonte Liguria e Valle d'Aosta (11168; 14 July

117 2014). During collection of feathers, we made every effort to minimize distress to the penguins.

## 118 *2.2 Penguins and collection of samples*

Feathers were collected in March 2014 from a captive colony of 46 penguins housed at the biopark Zoom Torino (Cumiana, Torino; 44°56' N, 7°25' E; www.zoomtorino.it). The colony was composed of 25 adult breeders and 21 sexually immature juveniles (< 2 years old). Penguins were maintained in an outdoor communal exhibit of 1500 m<sup>2</sup>, which included a pond of 120 m<sup>2</sup> (maximum depth: 3 m). All penguins were fed with herrings (*Clupea harengus*), purchased from an animal food retailer, which had been caught in the northeast Atlantic Ocean (FAO fishing area 27).

Samples of feathers up to a total weight of 0.3 - 0.5 g were cut from the back of each penguin. The cut was made with scissors between the *calamus* and the vane of the feather, in order not to cause any pain to the bird. Finally, a blood sample (1 mL) was extracted from a vein of the foot of each bird with a needle and a vacuum plastic tube (4.9 mL) with lithium heparin. After collection, all samples were stored for subsequent laboratory analyses.

130 *2.3 Molecular sexing* 

Blood samples were extracted using the Pure Link<sup>TM</sup> Genomic DNA Mini Kit (Invitrogen, Grand 131 Island, NY USA). PCR was carried out using the primers P8 and P2, previously described by 132 133 Griffiths (1998). Primer P2 was labelled with Hex fluorescent dye at the 5' end, to be used with capillary gel electrophoresis. PCR was carried out in a total volume of 25 µL containing 50-60 ng 134 of genomic DNA using HotStarTag Oiagen (1.5 U), Buffer containing Mg<sup>2+</sup> (1.5 mM), dNTPs (0.2 135 mM each) and the primers reported above (300 nM each). PCR was performed in a GeneAmp PCR 136 System 9700 thermal cycler (Applied Biosystems, Life Technologies Monza, Italy). An initial 137 activation step at 95 °C for 15 min was followed by 40 cycles of 95 °C for 30 sec, 48 °C for 30 sec, 138 and 65 °C for 1 min. A final run of 65 °C for 5 min completed the program. 139

Each amplification product, diluted at 1:100, was added to a mix containing Rox Size Standard (Life Technologies, Grand Island, NY USA) and formamide, and then subjected to capillary electrophoresis on an ABI 3130 Genetic Analyzer (Life Technologies, Grand Island, NY USA). The
size of the amplification products was determined by GeneMapper software analysis. Males were
characterized by just one peak at 364 bp, while females had two peaks at 364 and 380 bp.

145 2.4 Analytical methods

Surface lipids and contaminants were removed from the feathers in four cleaning steps: a Triton-x-146 100 (0.01%) bath (for at least 4 hours), followed by one rinse with deionized water, two successive 147 vigorous washes with methanol, and a final rinse with deionized water. Feathers were dried at 50°C 148 until a constant weight was obtained, and were then minced. Prior to analysis, samples were divided 149 into two sub-samples, one for mercury quantification with a Direct Mercury Analyzer (DMA-80 150 151 Analyzer from Milestone, Shelton, CT, USA) and the other for detecting all the other metals by 152 Inductively Coupled Plasma - Mass Spectrometry (ICP-MS Xseries II from Thermo Scientific, Bremen, Germany). The DMA-80 analyzer performed thermal decomposition, catalytic reduction, 153 amalgamation, desorption and atomic absorption spectroscopy without having to pre-treat the 154 samples, and with no waste generation. Between 0.05 g and 0.1 g of feathers, according to 155 availability, were directly weighed on graphite shuttles and processed with an output result, for 156 157 mercury content, in about 5 min (per sample).

Determining levels of Al, As, Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, Se, Sn, V and Zn was performed 158 159 after wet digestion with acids and oxidants (HNO<sub>3</sub> and H<sub>2</sub>O<sub>2</sub>) of the highest quality grade (Suprapure). In this case, between 0.05 and 0.27 g of feathers, according to availability, and 0.10 g 160 of pooled fish were subjected to microwave digestion (microwave oven ETHOS 1 from Milestone, 161 Shelton, CT, USA) with 7 mL of HNO<sub>3</sub> (70% v/v) and 1.5 mL of H<sub>2</sub>O<sub>2</sub> (30% v/v). Samples were 162 then brought to a final weight of 50 g with ultrapure water (Arium611VF system from Sartorius 163 Stedim Italy S.p.A., Antella - Bagno a Ripoli, FI, Italy). Multi-elemental determination was 164 performed with ICP-MS after daily optimization of instrumental parameters and using an external 165 standard calibration curve; Rodium and Germanium were used as internal standards. Analytical 166 167 performances were verified by processing Certified Reference Materials (Dogfish liver -DOLT-4

from the National Research Council of Canada, and Oyster Tissue-SRM 1566b from the National 168 Institute of Standard and Technology), along with blank reagents in each analytical session. The 169 recoveries for reference materials ranged from 85 to 120% for DOLT-4 and from 82 to 117% for 170 SRM 1566b. The limit of quantitation (LOQ) was 0.010 mg Kg<sup>-1</sup> for all elements, except for Hg 171 that was 0.034 mg Kg<sup>-1</sup>. The selenium:mercury molar ratio was calculated as follows: for each bird 172 class (adult males, adult females, juvenile males, juvenile females) the mercury and selenium 173 concentrations in ng  $g^{-1}$  (wet weight) were divided by their respective atomic weights (200.59 and 174 789.00) to obtain the molar concentrations (nmol  $g^{-1}$ ), which allowed us to calculate the mean 175 molar ratios. Note that this is a ratio of the mean values. 176

177 2.5 Statistical analysis

One half of the values of the respective limit of quantitation (LOQ) were substituted for those values below the limit of quantitation and used in statistical analysis. Data were tested for normality by using the Kolmogorov-Smirnov test. Since data distribution was normal, comparisons of the mean metal concentrations between age classes were carried out using a Student's *t-test*. All analyses were performed using the SPSS version 20.0 for Macintosh.

## 183 **3. Results and Discussion**

## 184 *3.1 Sex-related differences*

Metal concentrations in birds are highly species-specific (Burger and Gochfeld, 2000). Moreover, a 185 few gender-related differences have been found by Burger et al. (2007) in species with sexual 186 dimorphism in body size. In particular, females usually show higher levels of metals than males, 187 and this suggests that the mechanism of excretion into eggs and eggshells is not very effective, or 188 that uptake is greater (Burger et al., 2007). In our study, there were no significant sex-related 189 differences in concentrations of essential elements, with the exception of As (Figure 1a, Table 1). 190 Arsenic is a well-known environmental contaminant, potentially toxic even at trace levels but 191 essential for life at very low concentrations as ultra-trace element (Cox, 1995). The concentration 192 193 of arsenic was significantly higher in feathers of juvenile females than in those of juvenile males (t

= -2.97, df = 19, p < 0.005; Figure 1a), while in adults there were no significant sex-related 194 195 differences. Generally, arsenic uptake is via the diet in marine animals (Kubota et al. 2001) and arsenic does not biomagnified through food chains. Accordingly, as observed for other metals that 196 are not biomagnified, lower trophic marine animals show higher arsenic concentrations than higher 197 trophic marine animals (Rahaman et al. 2012). Studies on As levels in high-trophic-level marine 198 organisms are scarce, and researches are now focusing on the mechanism of As accumulation, 199 considering its possible role as an endocrine disruptor (Georgescu et al. 2011). Lucia et al. (2010) 200 found that female birds displayed higher As concentrations in the liver and feathers than male birds. 201 Similarly, Taggart et al. (2006) examined As, Zn, Se, Pb and Cu levels in the livers and bones of 202 203 five waterfowl species from SW Spain and found higher concentrations of As in female bones and -204 as in our case - higher concentrations in juveniles than in adults. As arsenic is an essential element, this difference could reflect a different physiological requirement at a particular growth stage. Egg 205 transfer is metal specific and although it is well known that mercury is effectively transferred from 206 females to eggs in seabirds (Robinson et al., 2012), limited information is available on the maternal 207 transfer of arsenicals to seabird eggs. Kubota and coauthors (2002) studied the maternal transfer of 208 arsenicals to eggs of the black-tailed gull. As composition in the eggs was similar to that in tissues 209 of the mother bird, and the percentage of As in eggs was about 11% of that of the mother. 210

211 Examining sex-related differences in response to chemicals is complicated in wild birds because of the differences in niches and forage (Burger et al. 2007), as well as the difficulties in determining 212 the sex of non-dimorphic species. Sex and age of vertebrate top predators are known to be involved 213 in the variation of tissue trace element concentrations (Kojadinovic et al. 2007a). Metal 214 accumulation in birds can be largely affected by contamination of the surrounding environment 215 (Markowski et al. 2013). Top-level piscivores, such as penguins accumulate much higher levels of 216 contaminants than birds that are lower on the food chain (Lodenius and Solonen 2013). Very few 217 studies consider sex differences in penguins and tend to only focus on Hg (e.g. Becker et al. 2002; 218 Frias et al. 2012). Despite being naturally occurring, Hg is a pervasive environmental contaminant 219

that negatively affects humans and wildlife. The diet and foraging ecology of penguins may play an 220 important role in explaining feather Hg levels in some penguin species, because ingestion of food is 221 the main route of Hg exposure in birds (Lodenius and Solonen 2013; Burger et al., 2014). In 222 particular, males of the Gentoo penguin (Pygoscelis papua) in South Georgia were shown to have 223 higher levels of Hg in their feathers than females (Becker et al. 2002). By contrast, Frias et al. 224 (2012) found that males and females of the Magellanic penguin (Spheniscus magellanicus) from the 225 Atlantic coast of Patagonia had similar median Hg levels in all age classes, although in adult males 226 the range was greater than in adult females. The Hg levels that we found in the feathers of African 227 penguins in this study did not differ significantly (p > 0.05) between males and females (Table 1) 228 and were in the range 1.30 to 2.80 mg Kg<sup>-1</sup> d.w. Our results support the hypothesis that sex-related 229 230 variation in metal content does not occur in closely related penguins of the genus Spheniscus. Differences in diet of wild penguins may explain the variation in Hg concentrations between similar 231 aged seabirds from different environments, while differing Hg concentrations between the sexes are 232 thought to be a result of metabolic differences between the sexes (Frias et al. 2012). Some authors 233 have suggested that male penguins could have more Hg than females because egg-laying may allow 234 excretion of Hg (Becker et al. 2002; Falkowska 2013). Penguins have a unique molting pattern 235 among birds; they renew all their feathers simultaneously, just before or just after the breeding 236 period, and Hg concentrations in feathers represent Hg exposure in the period of time elapsed since 237 the last molt. In fact, Hg excretion through the quill is a detoxification mechanism during 238 premolting (Becker et al. 2002), and once the boom ends its growth, the blood transport channel 239 atrophies leaving a permanent record until the next molt (Burger and Gochfeld 2002). There were 240 no significant differences in Hg content between males and female in our study, although gender 241 only seems to be of interpretive concern for species exhibiting different dietary habits between 242 sexes (Robinson et al. 2012), and diet is very homogeneous in captive penguins. 243

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The concentrations of metals in feathers reflect the body burden at the time that the feathers are grown (Furness et al. 1986). Unlike most other avian orders, penguins have a unique molt in the year, with fasting periods of 2-3 weeks on average (Adams and Brow 1990).

Age is an important factor to be considered in exposure to chemicals in wildlife. Young animals could have higher exposure because they are often restricted to nesting sites or brooding sites, while adults tend to roam far from the source of contamination (Lodenius and Solonen 2013).

Conversely, in our study, juveniles and adults had the same patterns of exposure and we did not find 252 age-related differences in the feathers of African penguins, except for selenium (t = 2.42, df = 44, p 253 < 0.05; Figure 1b). Se is a metalloid trace element that birds and other wildlife need in small 254 amounts to maintain good health (Ohlendorf and Heinz 2009). Se levels of 3.8 to 26 mg kg<sup>-1</sup> d.w. 255 (depending upon the species) in feathers results in mortality (Burger 1993), and 1.8 mg kg<sup>-1</sup> d.w. 256 results in sub-lethal adverse effects (Ohlendorf and Heinz 2009). Se levels in our samples (Table 1) 257 ranged from 2.142 (juvenile females) to 2.656 mg kg<sup>-1</sup> (adults males), and were similar or higher 258 than those detected by Metcheva et al. (2006) in the feathers of the Gentoo Penguin, but lower than 259 those reported by Jerez et al. (2011) in the Adélie penguin (Pygoscelis adeliae) and in the Chinstrap 260 penguin (Pygoscelis Antarctica). The increase in Se levels with age could be due to a chronic 261 exposure to trace elements such as Cd and Hg, since Se is also known to have a detoxifying effect 262 on these metals. Moreover, Ralston and Raymond (2010) suggested that the selenium:mercury 263 molar ratio is an important consideration for understanding the toxic effects of mercury. While there 264 is consensus that an excess of mercury compared to selenium is potentially hazardous, there is no 265 consensus about the levels of selenium deemed necessary to reduce mercury toxicity. Different 266 authors have suggested that Se:Hg molar ratios above 1 are probably protective (Ralston et al 2008; 267 Peterson et al. 2009). While this molar ratio has been examined in fish because of the potential 268 protective effects to humans consuming fish, recently Burger et al. (2014) suggested that it should 269 be protective of brain function in birds and other animals as well. In fact, the Se:Hg molar ratio has 270 been shown to vary significantly between tissues, with the highest ratios in brain and liver and the 271

lowest in feathers (Burger et al. 2014). In the present study, the Se:Hg molar ratio was above 1,
ranging from 2.51 (adult females) to 2.77 (adult males) and presumably protective for both genders
(Table 2).

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### 276 **4. Conclusions**

Sex-related effects should only be examined when males and females have been directly compared 277 under similar conditions. This unique type of experimental approach can be successfully carried out 278 with animals living in controlled environments, where the entire colony is fed with the same 279 identical food supply. Moreover, feathers are ideal for monitoring exposure and inferring effects 280 because they can be sampled non-invasively with minimal stress to the birds, especially relevant to 281 282 endangered species. In particular, birds with a synchronous molt are good candidates for bioindicators of metal contamination. Investigating sex- and age-related variations in environmental 283 contaminant concentrations in seabirds has produced contrasting results. In most studies on feathers, 284 no age-related variations were detected for several trace elements, while gender-related variations 285 were detected in species that exhibit different dietary habits between the sexes. 286

In the captive African penguins examined, Hg content in feathers seemed to reflect the bio-287 magnification phenomena through the penguin's fish diet but did not show any significant 288 289 relationship with sex and age. However, arsenic seems to be gender-related in juveniles, probably due to a different metabolic rate at that phase of growth. Finally, selenium concentration was shown 290 to increase with increasing age, according to its function of contrasting the toxicity of certain 291 292 metals. The concentrations of the other 13 trace elements considered appeared to be unaffected by sex or age, but the relationships between concentrations of many trace elements in internal tissues 293 and feathers concentrations remains to be determined. 294

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