

## UNIVERSITÀ DEGLI STUDI DI TORINO

*This is an author version of the contribution published on: Questa è la versione dell'autore dell'opera:* 

Beyond NPM-anaplastic lymphoma kinase driven lymphomagenesis: alternative drivers in anaplastic large cell lymphoma.

Tabbò F, Ponzoni M, Rabadan R, Bertoni F, Inghirami G; European T-cell LymphomaStudyGroup.CurrOpinHematol.2013Jul;20(4):374-81.doi:10.1097/MOH.0b013e3283623c07.Philadelphia Pa : Lippincott Williams And Wilkins

### The definitive version is available at:

La versione definitiva è disponibile alla URL: http://journals.lww.com/cohematology/pages/articleviewer.aspx?year=2013&issue=07000&article=00018&typ e=abstract

## Beyond NPM-ALK driven lymphomagenesis: alternative drivers in Anaplastic Large Cell Lymphoma

Fabrizio Tabbo'<sup>1\*</sup>, Maurilio Ponzoni<sup>2</sup>, Raul Rabadan<sup>3</sup>, Francesco Bertoni<sup>4,5</sup>, Giorgio Inghirami<sup>1+</sup> and the European T-cell Lymphoma Study Group.

<sup>1</sup>Department of Molecular Biotechnology and Health Science - Center for Experimental Research and Medical Studies (CeRMS), University of Torino, Torino, 10126 Italy; <sup>2</sup>Pathology & Lymphoid Malignancies Units, San Raffaele Scientific Institute, Milan, 20132 Italy; <sup>3</sup>Department of Biomedical Informatics, Center for Computational Biology and Bioinformatics, Columbia University, New York, NY 10027 USA; <sup>4</sup>Lymphoma and Genomics Research Program, IOR Institute of Oncology Research, 6500 Bellinzona, Switzerland. <sup>5</sup>Lymphoma Unit, IOSI Oncology Institute of Southern Switzerland, 6500 Bellinzona, Switzerland.

The European T-Cell Lymphoma Study Group: *Italy*: Sabrina Aliberti, Antonella Barreca, Luca Bessone, Ramona Crescenzo, Filomena Di Giacomo, Marcello Gaudiano, Giorgio Inghirami, Indira Landra, Elena Lasorsa, Rodolfo Machiorlatti, Elisabetta Mereu, Katia Messana, Domenico Novero, Elisa Pellegrino, Achille Pich, Roberto Piva, Irene Scarfo', Elisa Spaccarotella, Fabrizio Tabbo', Maria Todaro, Ivana Ubezzi, Susanna Urigu, Francesco Vittone (Azienda Ospedaliera Città della Salute e della Scienza di Torino, University of Torino); Francesco Abate, Elisa Ficarra, Andrea Acquaviva (Politecnico di Torino); Maurilio Ponzoni (San Raffaele Institute, Milano); Carmelo Stella (Istituto Clinico Humanitas, Milano), Claudio Agostinelli, Pier Paolo Piccaluga, Stefano Pileri (University of Bologna); Brunangelo Falini, Enrico Tiacci (University of Perugia) and *Switzerland*: Francesco Bertoni, Michela Boi (IOR-IOSI).

Corresponding author:

<sup>+</sup>Giorgio Inghirami <sup>1</sup>Department of Molecular Biotechnology and Health Science and CeRMS University of Torino Via Santena 7, Torino 10126, Italy E-mail: <u>giorgio.inghirami@unito.it</u> Tel +39-011-633-4623 Fax +39-011-633-6887

#### ABSTRACT

**PURPOSE OF THE REVIEW**: Anaplastic Large Cell Lymphomas (ALCLs) are rare entities whose tumorigenic events have only been found in well-defined subsets. The categorization of additional molecular fingerprints is needed to advance our knowledge and to deliver successful therapies.

**RECENT FINDINGS**: The discovery of Anaplastic Lymphoma Kinase (ALK) fusions has provided the basis for the characterization of distinct subsets among ALCL patients. Although the oncogenic addiction of ALK signaling is proven, the tumorigenic contribution of co-activating lesions is still missing. As ALK- and ALK+ share common signatures, it is plausible that analogous mechanisms of transformation may be operating in both subsets, as confirmed by the dis-regulated activation of c-MYC, and the loss of Blimp-1 and p53/p63 axis. Nonetheless, recurrent genetic alterations for ALK- ALCL or refractory leukemic ALK+ ALCL are lacking. Moreover, although conventional chemotherapies (anthracycline-based) are most successful, i.e. in ALK+ ALCL patients, the implementation of ALK inhibitors or of anti-CD30 based treatments provides innovatites solutions, particularly in pediatric ALK+ ALCL and in chemorefractory/relapsed patients.

**SUMMARY**: The complete portrayal of the landscape of genetic alterations in ALCL will dictate the development of innovative chemotherapeutic and targeted therapies that will fit most with the molecular and clinical profiling of individual patients.

#### Key words:

Anaplastic Lymphoma Kinase, signaling pathways, kinases' inhibitors, lymphoid differentiation, mouse models.

### **KEY POINTS**:

- Anaplastic Large Cell Lymphoma are an heterogeneous group of lymphoma some of which carry a restricted number of genetic defects mainly involving the Anaplastic Lymphoma Kinase (ALK+ ALCL) o less frequently display alternative translocations [t(2;x)(p23;x), t(6;7)(p25.3;q32.3) and inv(3)(q26q28) leading to TBL1XR1/TP63, etc.]. Nonetheless, specific lesions are still lacking for many ALCLs.
- The precise relationship and origin of ALCL remain unclear. Although ALK+ and ALK- share a set of genes and similar phenotypes, they are considered distinct groups with unique clinical features. However, it is unknown whether clinical differences and responses to conventional therapies may simply be related to different clinical stages and/or unique genomic lesions.
- Bioinformatics algorithms have identified several ALCL signaling classifiers demonstrating the preferential expression of a restricted number of pathways. The recognition of "common hubs", which can be targeted by selective inhibitors, represents a viable strategy for future therapeutic protocols.
- Taking advantage of *in vitro* and *in vivo* models, several groups have shown that the transforming properties of ALK fusions involve a plethora of alternative modules capable to regulate intrinsic (i.e. cytoskeleton, cell growth, etc.) and/or extrinsic (cell matrix invasion, tumor-host relationships, etc.) modalities.
- The identification of the driving lesion of ALCL will require the construction of international networks capable to synergize their activities and to construct large and clinically annotate tissue libraries. The collection of viable tissues will facilitate the generation of batteries of "Patient Derived Tumorgrafts".

#### **INTRODUCTION**

Peripheral T-cell lymphoma (PTCL) are a heterogeneous group of tumors derived from postthymic elements including leukemic/disseminated, nodal and extranodal diseases [1,2,3]. As orphan diseases (12 to 15% of all non-Hodgkin's lymphoma [NHL] in Western populations) [1,2,3,4], they include entities displaying a great variability in clinical, morphological, immunophenotypic, cytogenetic and molecular features. First described in 1985 [5], Anaplastic Large Cell Lymphoma (ALCL) of either adults (2-8% of NHL) or children (15-30% of NHL), nowadays correspond to specific subtypes of systemic peripheral T-cell lymphoma [1]. The presence of Anaplastic Lymphoma Kinase (ALK) gene fusions has provided the criteria for a new WHO classification, which contemplates a novel entity (i.e. ALCL ALK+) and proposes a provisional one, including ALCL patients, who lack ALK translocations (i.e. ALCL ALK-). Because of genetic, immunophenotypic, and clinical differences, cutaneous ALCLs (cALCL) are considered as a completely distinct subset a part from its systemic counterparts. Systemic ALCL share cytological, immunophenotypic and molecular features. However, ALK- ALCL patients have poorer performance status, more often B symptoms [6], and an overall survival (OS) rate of 36% versus 20% of PTCL-NOS patients. This suggests unique driving defects, with high oncogenic penetrance. In contrast, ALK+ ALCL have a more favorable clinical course [4], though ALK+ ALCL with an aggressive behavior could be encountered in the clinical practice [7]. It remains uncertain whether the molecular lesion(s) and/or other features determine the clinical course of ALCL patients. In fact, once patients are normalized by clinical parameters, ALK- and ALK+ ALCL display analogous prognosis (failure-free survival [FFS] and OS) [6]. Considering that ALK+ ALCL have a less complex karyotype [8,9,10,11], it plausible that ALK fusions are critical actors and that tumor progression is due to somatic mutations (minimal deletions, activating somatic mutations, etc.) disrupting the function of a limited set of genes. In contrast, the transformation of ALK- ALCL might require the consolidation/acquisition of many genetic defects that rapidly lead to systemic and more aggressive phenotype. This is in agreement suggested with their higher and heterogeneous karyotypes [8,9,10,11]. Nonetheless, the driving lesion(s) of ALK- ALCL are still to be identified, and co-drivers are lacking for both ALCL subgroups.

The lack of representative cell lines or animal models has definitively contributed in impairing our knowledge of mature T-cell lymphoma. Ultimately, this has jeopardized the design of successful therapies and the upgrade of clinical programs, particularly in patients with poor outcome (ALK- ALCL and PTCL-NOS). The recognition of the tumorigenic defects of PTCL is

expected to provide patient specific "molecular fingerprints" and thus more suitable tailored therapies.

#### **ONCOGENIC SIGNALING OF ALK FUSIONS**

Chromosomal translocations of the *ALK* gene are documented in many ALCL, although the percentage of ALK+ ALCL varies, as a result of the inclusion criteria of the ALK- ALCL. In absence of strong classifier(s) (like ALK), the distinction between ALK- ALCL, CD30 PTCL-NOS and some enteropathy associated T-cell lymphoma represents a diagnostic challenge.

*ALK* gene encodes a 210kDa tyrosine kinase receptor (CD247) belonging to the insulin growth factor receptor superfamily. Its expression is largely limited to the nervous system during embryogenesis and to focal areas of the adult brain [12]. Although the physiological role of ALK in mammals is unknown, it is involved in neuronal differentiation [13] and ALK activating mutations have been found in familial and sporadic neuroblastomas [14].

The breakpoints of ALK chimera invariably occur within the intron placed between the exons 19 and 20 (NM\_004304.3). Thus exons coding for the intracytoplasmic domain of ALK (exons 20-29) are then juxtaposed to different partners [15,16]. Seventy-80% of ALK+ ALCL harbor the t(2;5)(p23;q35) translocation (NPM–ALK chimera). The intracellular distribution of the fusions is due to the structure/function of ALK partners, enforcing either to nuclear/cytoplasmic, or cytoplasmic and in rare cases juxta-membranous localization. Virtually all partners (with the exclusion of MYH9-ALK) provide dimerization domains, leading to the homo/heterodimerization of the fusions and to constitutive activation of the kinase [16,17]. Conventional genomic approaches, and more recently next generation sequencing (NGS) have shown alternative ALK translocations in many types of human cancers, e.i. lung tumors [18,19].

ALCL display additional alterations involving many chromosomes [8,9,10,20], however frequent common/discrete secondary lesions are rare. Two translocations were reported in ALK-ALCL, involving the *DUSP22* gene, which is juxtaposed to the *FRA7H* fragile site, or to the gene coding for *IRF4* [21]. Boi et al. have recently shown that several ALCL display recurrent deletions affecting 17p13.3-p12 (25%) region, in which *TP53* gene is located, and 6q21 (19%) encompassing *PRDM1* and *ATG5* genes [11]. Finally, Vaismatzis et al. have recently described a set of genomic defects in DLBCL and PTCL/ALCL encoding fusion proteins homologous to  $\Delta$ Np63, a dominant-negative p63 isoform that inhibits the p53 pathway [22].

ALK chimera were originally proven to be oncogenic *in vivo* [23] and these data were largely confirmed in *in vitro* models [24] and then in genetically modified animals [25,26]. Understanding how ALK signals acts and defining the mechanisms responsible for its

deregulation is critical for dissecting the mechanisms, which mediate ALK cellular transformation and provide the basis for *rationale* therapeutic approaches. By a large array of methods, it is now know that ALK fusions and in particular the NPM-ALK chimera interact with a plethora of molecules and elicit many pathways. These include the RAS/Erk, PLC- $\gamma$ , PI3K, and Jak/Signal Transducers and Activators of Transcription (STAT), capable to control individually or in association, cell proliferation, survival, and cytoskeletal properties [19].

The activation of RAS/Erk pathway provides positive signals regulating cell growth and the inhibition of MEK (AZD6244 or shRNA) leads to cell cycle arrest, without significant changes in cell viability (Crescenzo R, personal communication). Similarly, NPM-ALK can down-modulate, via PI3K-AKT, the inhibitory action of FOXO3a, upregulating Cyclin D2 and down-regulating p27, and providing positive growth signals.

We and other groups have shown that the neoplastic phenotype of NPM-ALK is largely mediated by the STAT3. This enforces the transcription of a surplus of genes (coding and non coding), promoting cell growth and survival. In shRNA-based knockdown experiments, Piva et al. have demonstrated that several genes are directly regulated by STAT3, which display canonical STAT3 binding sites within their regulatory regions (Piva R, personal communication). Among them we mention CD30, granzyme, perforin, IL1RAP and IL2RA. From a diagnostic point of view, CD30, granzyme, perforin are known to be preferentially expressed by ALCL cells, and are commonly used in algorithms encompassing the differential diagnosis of different PTCL entities. Their transcription requires phosphorylated STAT3 complexes, which often include CEBPb and AP-1 transcription factors. Notably AP-1 members play an important role in ALK mediated transformation controlling tumor growth and positive host signals, via PDGF [27] (Fig. 1).

Zhang and coworkers have recently elucidated additional features of STAT3, demonstrating a STAT3-positive regulation of ICOS [28]. The same group had previously shown that PDL-1 expression is also regulated by STAT3 [29]. Collectively, these data demonstrate that ALCL cells engage ICOS to gain a growth advantage, and PDL-1 as novel mechanism of tumor escape, modulating the host responses. The overexpression of IL-21 [30] and deregulation of TNF/Fas/TNF [31] can also contribute to ALK tumorigenic phenotype, favoring the success of ALCL cells and overcoming host defenses. Finally, ALK signaling controls HiF1 $\alpha$ , a factor that impacts directly in the neo-angiogenesis and provides a positive growth advantage to the lymphoma cells [32,33] (Fig. 1).

Lastly, STAT3 itself can directly or via downstream mediators down-regulate the transcription of many genes. Approximately 60% of modulated STAT3 genes are repressed after shRNA KD.

Interestingly, Zhang et al have recently reported that STAT3, engaging the IL-2R $\gamma$  promoter, enhances the binding of DNA methyltransferases (DNMTs), leading ultimately to the transcriptional repression of IL-2R $\gamma$  gene. The knockdown of IL-2R $\gamma$  expression contributes to the neoplastic phenotype, as demonstrated by its forced/expression that leads to the loss of NPM-ALK protein expression, and then apoptosis. Ultimately, STAT3 down regulates T-cell associated molecules controlling T-cell identity of ALCL cells. In this context NPM-ALK provides signals capable to bypass TCR mediated activation [34,35] (Fig. 1).

STAT3 can similarly regulate the expression of several miRNA clusters (Spaccarotella E, personal communication) including the miRNA17-92 [36], known to have a role in human cancers. In ALCL, the miRNA17-92 overexpression overcomes in part the loss of STAT3 in an shRNA STAT3 inducible ALK+ ALCL model. More importantly, primary ALK+ ALCL display higher miRNA17-92 levels [36] compared to ALK- ALCL and cutaneous T-cell lymphoma and the usage of STAT3 inhibitors leading to the down-regulation of this cluster could represent an attractive strategy for the treatment of ALCL lymphoma (Lin C, personal communication).

The ability to successfully migrate and invade distant tissues contributes to the neoplastic phenotype, impairing clinical responses and long remissions. ALK signaling can efficiently module the cytoskeleton and promote invasion. The data reported by Ambrogio et al. [37] have recently been confirmed [38]. Dupuis-Coronas et al. have also shown that ALK, modulating the activity of PIKfyve, enhances the invasive capacities of NPM-ALK cells and their capacity to degrade the extracellular matrix [39]. Invasion of ALK+ ALCL cells is also modulated by the axis ALK-STAT3-Twist1 [40] (Fig. 1).

In conclusion, it is evident that the tumorigenic properties of ALK signaling are more complex that originally proposed, confirming that ALK is a powerful kinase capable to provide a complete and broad oncogenic addiction. These properties make ALK an excellent therapeutic target.

# ALKR AND ALKS ANAPLASTIC LARGE CELL LYMPHOMAS: TWO SIDES OF THE SAME COIN?

The debate on distinct entities among PTCLs remains open. Novel hypotheses are emerging on the origin and relationship of different PTCL entities. The concept that ALK- ALCL should be lumped within PTCL-NOS has been recently sponsored. Alternatively, a scenario in which all ALCL are incorporated in a single group, irrespectively of the ALK expression has been contemplated. This level of uncertainty is corroborated by the fact that, once ALCL patients are stratified by stage, IPI etc. either groups display similar characteristics. In this landscape, CD30+

PTCL represent a puzzling/confounding group [41]. Their precise definition is critical and sometime questionable. Immunophenotypically, they express weak/partial CD30, and in same cases CD15 [42,43]. Cytologically display a certain monomorphism and they often have a functional TCR signaling (NFATc positive etc.) [44,45]. Clinically, CD30+ PTCL share a more aggressive clinical course, justifying their distinction and a closer relationship to PTCL-NOS. We strongly believe that these uncertainties will be solved only when distinct molecular defects will be discovered in different PTCLs.

Another similar confusing topic regards the ALCL origin, and their putative normal counterpart elements. Several hypotheses have been proposed, taking in account their expression profile and unique immune-phenotype. The expression of perforin, T1A1 and granzyme has been interpreted as a specific fingerprint, supporting the idea that ALCL may derive from cytotoxic Tlymphocytes. Alternatively, we speculate that the phenotype of ALCL may rather be the result of the deregulated expression of unique pathways and/or specific defects, which impose unique/fixed profiles. It is known that transcription factors can play a critical role in T-cell differentiation and once constitutively activated can undermine physiological programs and rerouted their development. Based on this assumption, we could speculate that the constitutive activation of STAT3 might be responsible for the cytotoxic phenotype of ALK+ ALCL cells, even in cells that were committed to different lineages and/or function. This leaves the open question, why ALK- ALCL display a cytotoxic phenotype? To solve this question, we have analyzed a large cohort of ALCL samples and found that a subset of ALK- ALCL clearly shares a STAT3 expression profile and detectable nuclear pSTAT3. Moreover, both ALK+ and ALK-ALCL reveal signatures, linked to the activation of c-MYC, NOTCH-1, or NFkB, and RAS/ERK, suggesting the existence of upstream activators. Interestingly, it now evident that several ALCL co-share overlapping signatures suggesting multiple activating defects or alternatively the presence of unique lesions capable, like ALK fusions, to efficiently and concomitantly fire multiple pathways. Search of ALCL pathogenetic lesions is under evaluation and it is predicted that new information will be available soon (Fig. 2).

# CAN WE USE PRECISION MEDICINE DATA TO IMPROVE THERAPUTIC COMPLIANCE?

The definition of the molecular fingerprints of neoplasms is now possible through the implementation of the impressive technologies. The NGS platforms are currently entering the clinical arena and it is plausible that, once interconnected and clinical based networks of laboratories, many patients will have individualized molecular identikits. Nonetheless, caveats

on the tumorigenic contribution of individual lesions and their functional role in the maintenance of the neoplastic phenotypes remain untouched. This are a critical issues, which should be added to the overwhelming capacity of tumor cells to adapt rapidly to the environment and to stress imposed by drugs and host changes. Thus, the search the "magic bullet" may fail. Instead, the association of multiple "smart" compounds could provide higher response rates and overcome resistance. Since the cost for a novel drug is around 1billion and requires approximately 12-15 years, we need to overcome impairing inefficiencies. It is agreed that many improvements in discovery programs need to be rapidly put in place, meliorating company inefficiencies (structural and operation), selection of viable targets, defining good therapeutic biopredictors, innovative technologies, more efficient and reliable screening tests and faster and less expensive clinical tracks in molecularly defined and/or naïve patients.

While pharmaceutical companies are reshaping their pipelines, a small number of drugs is successfully introduced into clinics. This ineffective result is seemingly related to update preclinical models, heavily relaying on *in vitro* models and "xenografts mouse platforms". Indeed, the most frequently used cell lines poorly represent human tumors [46]. This has encouraged many institutions and drug industry to acquire large library of cell lines, which can be interrogated with HTP platforms (NSG, phosphomapping etc.). The hope is to define better criteria and relationships between the genome and responses to therapies. The hope is of predicting more reliable clinical responses and dissecting responders and refractory patients. But, cell lines lack the host and its regulatory networks, have undergone ferocious in vitro selections and do not represent tumor heterogeneity.

To solve some of these issues, implants of fresh primary neoplasms are frequently introduced in severally-immunocompromised mice [47]. The generation of individualized cancer models represents an unprecedented opportunity to test battery of drugs for each individual and provide personalized oncology programs. However, these strategies need to be linked to defined genetic defects. Only combining HTP and innovative models, we can deliver a list of targetable lesions, which once validated in animals, should provide reasonable expectations. Since the successful growth of tumorgraft implants may require long period of time, new technologies interrogating the functional network in cancer and the efficacy of chemical libraries *in vitro* may provide alternative routes for the execution of pre-clinical trials *in vivo*. Our group has recently embarked in such a program and generated a battery of ALCL "Patients Derived Tumorgrafts" (PDT) [48]. These retain the immunophenotypic, genomic features of their corresponding primary tumors and display responses to conventional and innovative protocols that closely mimic those seen in donor patients. Their molecular characterization has demonstrated the presence of unique

genomic defects and allowed to discover new pathogenetic translocations and activating somatic mutations. The definition of a molecular identikit in PDTs will provide not only patients' fingerprints but also models to test the efficacy of selected drugs targeting hypothetical tumorigenic defects in each patient in vivo.

#### CONCLUSION

Little is yet known of mechanisms leading to T-cell lymphomagenesis. Nonetheless, the systematic usage of high throughput platforms has recently demonstrated that recurrent defects may be present in specific subsets of PTCLs. Although ALKb ALCL and ALK- ALCL display heterogeneous complex karyotypes, they share common expression signa- tures and dysregulated signalling pathways. The use of NGS approaches will be instrumental for the more complete discovery of mechanisms driving the pathogenesis of ALCL. New molecular lesions, even in small subgroups of patients, will provide objective diagnostic criteria and the bases for 'intelligent' therapies, to be first validated in the most informative preclinical models (i.e. PDT and so on)

#### ACKNOWLEGMENTS

GI is supported by the Italian Association for Cancer Research (AIRC) Special Program in Clinical Molecular Oncology, Milan (5x1000 No. 10007); Regione Piemonte (ONCOPROT, CIPE 25/2005); ImmOnc (Innovative approaches to boost the immune responses, Programma Operativo Regionale, Piattaforme Innovative BIO F.E.S.R. 2007/13, Asse 1 'Ricerca e innovazione' della LR 34/2004) and the Oncology Program of Compagnia di San Paolo, Torino. RR is supported by Partnership for Cure, NIH 1 P50 MH094267-01, NIH 1 U54 CA121852-05, NIH 1R01CA164152-01. FB sponsored by the Oncosuisse KLS-02403-02-2009 (Bern, Switzerland); Anna Lisa Stiftung (Ascona, Switzerland); Nelia and Amadeo Barletta Foundation (Lausanne, Switzerland). We thank Drs Vigliani C, Fioravanti A, and Mossino M for their technical support.

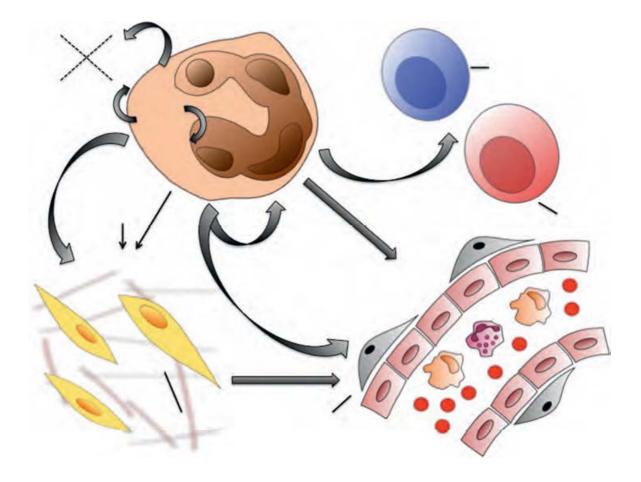
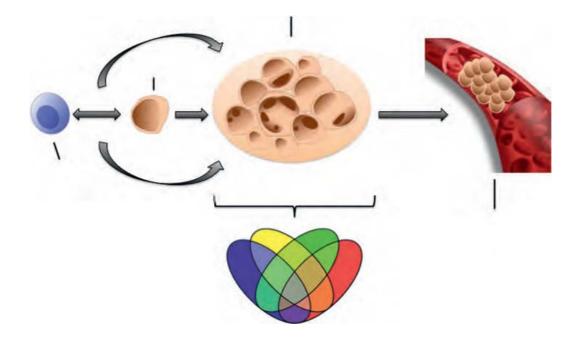


FIGURE 1. Anaplastic large cell lymphoma cells dysregulate and control the host environment. The oncogenic drivers of ALCL reroute intrinsic pathways leading to a self-autonomous cell growth as well as overcome host confinements, modulating immuno-responses of regulatory and effector T-cells. This is accomplished through alternative mechanisms including antigencamouflaging and lymphokines production obliterating immune surveillance (i.e. TNF/FasR, PD-L1/PD-L1R). Through cytoskeleton changes and production of pro-invasive mechanisms, tumour cells have the ability to migrate locally and disseminate to distant organs. Production of pro-angiogenetic factors stimulates vessel formation providing the necessary growth support for tumour survival. The JAK – STAT signaling pathway represents a master culprit modulating gene expression transcription of critical players. ALCL, anaplastic large cell lymphoma; CDC42, cell division control protein 42 homolog; FasR, FAS receptor; HIF1a, hypoxia-inducible factor 1 alpha; IL, interleukin; JAK, Janus kinase; MMPs, matrix metalloproteinases; PDGF, plateletderived growth factor; PD-L1, programmed cell death 1 ligand 1; PIKfyve, finger-containing phosphoinositide kinase; pSTAT3, phosphorylated signal transducer and activator of transcription 3; TNF, tumour necrosis factor; TWIST1, twist-related protein 1; VEGF, vascular endothelial growth factor.



**FIGURE 2**. Tumourigenic model for anaplastic large cell lymphoma transformation. ALCL may derive from a common stem cell precursor or alternatively from partially committed T element(s). Through the acquisition of powerful oncogenetic drivers, ALCL cells acquire unique phenotypes and display restricted signaling pathways. The progressive acquisition of selected genetic defects (loss of TP53, TP63, BLIMP1 and constitutive activation of c-MYC) is eventually responsible for tumour progression and more aggressive clinical behaviours. ALCL, anaplastic large cell lymphoma; ALK, anaplastic lymphoma kinase; IRF4, interferon regulatory factor 4; PRDM1, PR domain zinc finger protein 1; STAT3, signal transducer and activator of transcription; TKR, tyrosine kinase receptor; TP, tumour protein.

#### REFERENCES

- 1. Swerdlow SH, Campo E, Haris NL, et al. WHO classification of Tumors of Haemotolopoietic and Lymphoid tissues. 4th Edition. Edited by Swerdlow SH, Campo E., Haris NL, et al. Lyon: International Agency for Research on Cancer; 2008. pp. 312-317
- 2. Inghirami G, Pileri SA. Anaplastic large-cell lymphoma. Semin Diagn Pathol 2011; 28: 190-201.
- 3. Rodriguez-Abreu D, Filho VB, Zucca E. Peripheral T-cell lymphomas, unspecified (or not otherwise specified): a review. Hematol Oncol 2008; 26: 8-20.

- 4. Vose J, Armitage J, Weisenburger D. International peripheral T-cell and natural killer/T-cell lymphoma study: pathology findings and clinical outcomes. J Clin Oncol 2008; 26: 4124-4130.
- 5. Stein H, Mason DY, Gerdes J *et al.* The expression of the Hodgkin's disease associated antigen Ki-1 in reactive and neoplastic lymphoid tissue: evidence that Reed-Sternberg cells and histiocytic malignancies are derived from activated lymphoid cells. Blood 1985; 66: 848-858.
- 6. Savage KJ, Harris NL, Vose JM, *et al.* ALK- anaplastic large-cell lymphoma is clinically and immunophenotypically different from both ALK+ ALCL and peripheral T-cell lymphoma, not otherwise specified: report from the International Peripheral T-Cell Lymphoma Project. Blood 2008; 111: 5496-5504.
- 7. Grewal JS, Smith LB, Winegarden JD 3rd, *et al.* Highly aggressive ALK-positive anaplastic large cell lymphoma with a leukemic phase and multi-organ involvement: a report of three cases and a review of the literature. Ann Hematol 2007; 86: 499-508.
- 8. Zettl A, Rüdiger T, Konrad MA, *et al.* Genomic profiling of peripheral T-cell lymphoma, unspecified, and anaplastic large T-cell lymphoma delineates novel recurrent chromosomal alterations. Am J Pathol 2004; 164: 1837-1848.
- 9. Salaverria I, Beà S, Lopez-Guillermo A, *et al.* Genomic profiling reveals different genetic aberrations in systemic ALK-positive and ALK-negative anaplastic large cell lymphomas. Br J Haematol 2008; 140: 516-526.
- 10. Boi M, Stathis A, Zucca E, *et al.* Genetic alterations in systemic nodal and extranodal noncutaneous lymphomas derived from mature T cells and natural killer cells. Cancer science 2012; 103: 1397-1404.
- 11. Boi M, Rinaldi A, Piva R *et al.* BLIMP1 Is Commonly Inactivated In Anaplastic Large T-Cell Lymphomas (ALCL). Blood (ASH Annual Meeting Abstracts) 2011; 118: 1131-1132.
- 12. Iwahara T, Fujimoto J, Wen D *et al.* Molecular characterization of ALK, a receptor tyrosine kinase expressed specifically in the nervous system. Oncogene 1997; 14: 439-449.
- 13. Souttou B, Carvalho NB, Raulais D, Vigny M. Activation of anaplastic lymphoma kinase receptor tyrosine kinase induces neuronal differentiation through the mitogen-activated protein kinase pathway. J Biol Chem 2001; 276: 9526-9531.
- 14. Mossé YP, Laudenslager M, Longo L, *et al.* Identification of ALK as a major familial neuroblastoma predisposition gene. Nature 2008; 455: 930-935.
- 15. Morris SW, Kirstein MN, Valentine MB, *et al.* Fusion of a kinase gene, ALK, to a nucleolar protein gene, NPM, in non-Hodgkin's lymphoma. Science 1994; 263: 1281-1284.
- 16. Barreca A, Lasorsa E, Riera L, *et al.* Anaplastic lymphoma kinase in human cancer. J Mol Endocrinol 2011; 47: R11-23.
- 17. Chiarle R, Voena C, Ambrogio C, *et al.* The anaplastic lymphoma kinase in the pathogenesis of cancer. Nat Rev Cancer 2008; 8: 11-23.
- 18. Soda M, Takada S, Takeuchi K, *et al.* A mouse model for EML4-ALK-positive lung cancer. Proc Natl Acad Sci U S A 2008; 105: 19893-19897.
- 19. Tabbò F, Barreca A, Piva R, Inghirami G. ALK Signaling and Target Therapy in Anaplastic Large Cell Lymphoma. Front Oncol 2012; 2: 41.
- 20. Ott G, Katzenberger T, Siebert R, DeCoteau JF *et al.* Chromosomal abnormalities in nodal and extranodal CD30+ anaplastic large cell lymphomas: infrequent detection of the t(2;5) in extranodal lymphomas. Genes Chromosomes Cancer 1998; 22:114-121.
- 21. Feldman, AL, Dogan A, Smith DI *et al.* Discovery of recurrent t(6;7)(p25.3;q32.3) translocations in ALK-negative anaplastic large cell lymphomas by massively parallel genomic sequencing. Blood; 2011 117: 915-919.
- 22.\* Vasmatzis G, Johnson SH, Knudson RA *et al.* Genome-wide analysis reveals recurrent structural abnormalities of TP63 and other p53-related genes in peripheral T-cell lymphomas. Blood 2012; 120:2280-2289.

- 23. Kuefer MU, Look AT, Pulford K *et al.* Retrovirus-mediated gene transfer of NPM-ALK causes lymphoid malignancy in mice. Blood 1997; 90:2901-2910.
- 24. Bai RY, Dieter P, Peschel C, *et al* Nucleophosmin-anaplastic lymphoma kinase of large-cell anaplastic lymphoma is a constitutively active tyrosine kinase that utilizes phospholipase C-gamma to mediate its mitogenicity. Mol Cell Biol 1998; 18:6951-6961.
- 25. Chiarle R, Gong JZ, Guasparri I *et al.* NPM-ALK transgenic mice spontaneously develop T-cell lymphomas and plasma cell tumors. Blood 2003; 101:1919-1927.
- 26. Turner SD, Alexander DR. What have we learnt from mouse models of NPM-ALK-induced lymphomagenesis? Leukemia 2005: 19;1128-1134.
- 27.\* Laimer D, Dolznig H, Kollmann K *et al.* PDGFR blockade is a rational and effective therapy for NPM-ALK-driven lymphomas. Nat Med 2012; 18:1699-1704.
- Taking advantage of engineered mouse modeling the authors highlight the pathogenetic role of AP-1 transcription factors and their downstream molecules (PDGF) in ALK-driven lymphomagenesis.
- 28. Zhang Q, Wang H, Kantekure K *et al.* Oncogenic tyrosine kinase NPM-ALK induces expression of the growth-promoting receptor ICOS. Blood 2011; 118:3062-3071.
- 29. Marzec M,Zhang Q, Goradia A *et al.* Oncogenic kinase NPM/ALK induces through STAT3 expression of immunosuppressive protein CD274 (PD-L1, B7-H1). Proc Natl Acad Sci U S A 2008; 105:20852-20857.
- 30. Dien Bard, J, Gelebart P, Anand M *et al.* IL-21 contributes to JAK3/STAT3 activation and promotes cell growth in ALK-positive anaplastic large cell lymphoma. Am J Pathol 2009; 175:825-834.
- 31. Wu F, Wang P, Zhang J *et al.* Studies of phosphoproteomic changes induced by nucleophosmin-anaplastic lymphoma kinase (ALK) highlight deregulation of tumor necrosis factor (TNF)/Fas/TNF-related apoptosis-induced ligand signaling pathway in ALK-positive anaplastic large cell lymphoma. Mol Cell Proteomics 2010; 9:1616-1632.
- 32. Marzec M, Liu X, Wong W *et al.* Oncogenic kinase NPM/ALK induces expression of HIF1alpha mRNA. Oncogene 2010; 30:1372-1378.
- 33. Dejean E, Renalier MH, Foisseau M *et al.* Hypoxia-microRNA-16 downregulation induces VEGF expression in anaplastic lymphoma kinase (ALK)-positive anaplastic large-cell lymphomas. Leukemia 2011; 25:1882-1890.
- 34. Bonzheim I, Geissinger E, Roth S *et al.* Anaplastic large cell lymphomas lack the expression of T-cell receptor molecules or molecules of proximal T-cell receptor signaling. Blood 2004; 104:3358-3360.
- 35. Ambrogio C, Martinengo C, Voena C *et al.* NPM-ALK oncogenic tyrosine kinase controls T-cell identity by transcriptional regulation and epigenetic silencing in lymphoma cells. Cancer Res 2009; 69:8611-8619.
- 36. Merkel O, Hamacher F, Laimer D *et al.* Identification of differential and functionally active miRNAs in both anaplastic lymphoma kinase (ALK)+ and ALK- anaplastic large-cell lymphoma. Proc Natl Acad Sci U S A 2010; 107:16228-16233.
- 37. Ambrogio C, Voena C, Manazza AD *et al.* The anaplastic lymphoma kinase controls cell shape and growth of anaplastic large cell lymphoma through Cdc42 activation. Cancer Res 2008; 68:8899-8907.
- 38. Colomba A, Giuriato S, Dejean E *et al*. Inhibition of Rac controls NPM-ALK-dependent lymphoma development and dissemination. Blood Cancer 2011; 1(6):e2.
- 39. Dupuis-Coronas S, Lagarrigue F, Ramel D *et al*. The nucleophosmin-anaplastic lymphoma kinase oncogene interacts, activates, and uses the kinase PIKfyve to increase invasiveness. J Biol Chem 2011; 286:32105-32114.
- 40. Zhang J, Wang P, Wu F *et al.* Aberrant expression of the transcriptional factor Twist1 promotes invasiveness in ALK-positive anaplastic large cell lymphoma. Cell Signal 2012 24:852-858.

- 41. Fornari A, Piva R, Chiarle R, *et al.* Anaplastic large cell lymphoma: one or more entities among T-cell lymphoma? Hematol Oncol 2009; 27:161-170.
- 42. Barry TS, Jaffe ES, Sorbara L, *et al.* Peripheral T-cell lymphomas expressing CD30 and CD15. Am J Surg Pathol 2003; 27:1513-1522.
- 43. Gorczyca W, Tsang P, Liu Z *et al.* CD30-positive T-cell lymphomas co-expressing CD15: an immunohistochemical analysis. Int J Oncol 2003; 22:319-324.
- 44. Piva R, Agnelli L, Pellegrino E *et al.* Gene expression profiling uncovers molecular classifiers for the recognition of anaplastic large-cell lymphoma within peripheral T-cell neoplasms. J Clin Oncol 2010; 28:1583-1590.
- 45.\* Agnelli L, Mereu E, Pellegrino E *et al.* Identification of a 3-gene model as a powerful diagnostic tool for the recognition of ALK-negative anaplastic large-cell lymphoma. Blood 2012; 120:1274-1281.
- The authors investigate, through a gene expression platform, the feasibility of a molecular stratification among T-NHL, highlighting diagnostic predictors capable to improve the correct identification of questionable cases, including CD30+ PTCL.
- 46. Garber K. From human to mouse and back: 'tumorgraft' models surge in popularity. J Natl Cancer Inst 2009; 101:6-8.
- 47. Shultz LD, Ishikawa F, Greiner DL. Humanized mice in translational biomedical research. Nat Rev Immunol 2007; 7:118-130.
- 48. Tabbò F, Barreca A, Machiorlatti R *et al.* Humanized NOD/Scid/IL2g-/- tumorgrafts recapitulate primary Anaplastic Large Cell Lymphoma. AACR Meeting Abstracts 2013.