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PHOTOCHEMICAL PRODUCTION OF ORGANIC MATTER TRIPLET STATES IN WATER SAMPLES FROM MOUNTAIN LAKES, LOCATED BELOW OR ABOVE THE TREE LINE

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Abstract

The production of triplet states (T*) of chromophoric dissolved organic matter (CDOM), reacting with the probe molecule 2,4,6-trimethylphenol (TMP) was measured upon irradiation of water samples, taken from lakes located in a mountain area (NW Italy) between 1450 and 2750 m above sea level. The lakes are located below or above the tree line and surrounded by different vegetation types (trees, alpine meadows or exposed rocks). The most photoactive samples belonged to lakes below the tree line and their fluorescence spectra and CDOM optical features suggested the presence of a relatively elevated amount of humic (allochthonous) material. The lowest (negligible) photoactivity was found for a lake surrounded by exposed rocks. Its CDOM showed an important autochthonous contribution (due to in-lake productivity) and considerably higher spectral slope compared to the other samples, suggesting low CDOM molecular weight and/or aromaticity. Among the samples, CDOM photoactivity (measured as the rate of TMP-reactive T* photoproduction) decreased with changing vegetation type in the order: trees, meadows, rocks. It

could be connected with decreasing contribution from catchment runoff and increasing contribution from autochthonous processes and possibly precipitation.

Keywords: lake water photochemistry; sensitised photolysis; inter-system crossing; 2,4,6-trimethylphenol; humic and fulvic acids; fluorescence matrix (EEM).

Introduction

Mountain lakes are peculiar environments that are highly impacted by solar radiation. A combination of altitude and low concentration of UV-absorbing substances favours the penetration of harmful UV radiation in the water column (Laurion et al., 2000). Despite the development of several adaptations by aquatic organisms (Sommaruga, 2010), alpine lakes are a potentially vulnerable environment to climate change because of the possible increase of UV doses connected to variations in organic matter inputs and, most notably, to decreases in ice cover (Neale et al., 2001; Leavitt et al., 2003; Rogora et al., 2003).

Chromophoric dissolved organic matter (CDOM) is the main radiation absorber in lake water in the 300-500 nm wavelength range (Bracchini et al., 2004). DOM/CDOM can be produced by aquatic organisms (autochthonous) or reach the lake by catchment runoff (allochthonous) (Gondar et al., 2008). In alpine environments, catchment runoff is expected to be a more important DOM source for lakes located below the tree line (Vinebrooke and Leavitt, 1998). A recent study has also shown that high-altitude alpine lakes are comparatively more impacted by atmospheric deposition, and that a significant fraction of organic matter including pollutant load would be of atmospheric origin (Mladenov et al., 2011).

An important feature of CDOM is its photoactivity, namely the ability to produce reactive species upon sunlight absorption. A key issue is the photoproduction of triplet states (T*), which can be reactive themselves and/or yield ${}^{1}O_{2}$ and ${}^{\bullet}OH$ (Canonica, 2007; Latch and McNeill, 2006). These reactive transients can induce transformation of dissolved pollutants or of naturally occurring DOM (Richard et al., 2007), and might also cause oxidative stress to living organisms (Souza et al., 2007). Such considerations account for the importance of measuring reactive species photoproduction upon irradiation of water from mountain lakes, and to our knowledge information on this subject is very scarce.

We have recently observed negligible CDOM photoactivity in rainwater (Albinet et al., 2010b). Considering that lakes located in different mountain environments across the tree line (dominated by forests, meadows or rocks) would be impacted to a different extent by catchment runoff, inwater productivity and atmospheric deposition, one might expect significant differences in CDOM photoactivity. However, to our knowledge such a hypothesis has never been tested. In this work, we measured the photochemical generation of TMP-reactive T* in water samples from lakes located below and above the tree line, surrounded by trees, meadows or exposed rocks. An attempt was made to relate CDOM photoactivity with water chemical composition and environmental features.

Experimental

Reagents and materials. H₃PO₄ (85%), NaNO₂ (>97.5%), 2,4-dinitrophenylhydrazine (98%) and 2,4,6-trimethylphenol (TMP, 99%) were purchased from Aldrich, NaNO₃ (>99.5%), NaHCO₃ (99%), Na₂CO₃ (99%), CCl₄ (Uvasol for spectroscopy) and methanol (LiChrosolv gradient grade) from VWR Int., Zero-grade air from SIAD (Bergamo, Italy). Water used was of Milli-Q quality.

Sampling and sample treatment. The study lakes are located in Torino province, NW Italy. Water samples were taken from the surface layer of the lakes under study with a glass bottle secured to a 10-m rope, to avoid entering the lake and perturbing the sediment. Samples were transported to the laboratory under refrigeration. Sampling dates and lake features are reported in Table 1. The samples were vacuum-filtered on Millipore MF membranes (cellulose acetate, pore diameter 0.45 μ m) and stored under refrigeration till further processing.

Determination of nitrate, nitrite, DOC and pH. Nitrate was determined by ion chromatography, nitrite by high performance liquid chromatography (HPLC) upon pre-column derivatisation with 2,4-dinitrophenylhydrazine (Kieber and Seaton, 1996). DOC was measured as Non-Purgeable Organic Carbon (NPOC) with a Shimadzu TOC-V_{CSH} Total Organic Carbon Analyzer. Sample pH was measured with a Metrohm 602 combined glass electrode, connected to a Metrohm 713 pH meter. Further details are reported as Supplementary Material (hereafter SM).

Fluorescence measurements. A Varian Cary Eclipse fluorescence spectrofluorimeter was used, adopting a 10 nm bandpass on both excitation and emission. Fluorescence excitation-emission matrix (EMM) was obtained at 5 nm intervals for excitation wavelengths from 250 to 460 nm and emission ones from 250 to 600 nm. Identification of fluorescent constituents in water samples was performed on the basis of literature data, using the main contours to identify fluorescence peaks.

Irradiation experiments. Lake water samples (20 mL) were spiked with 10 μ M TMP and placed into cylindrical Pyrex glass cells (diameter 4.0 cm, height 2.5 cm), tightly closed with a lateral screw cap. The cells were irradiated from the top under a set of five UVA lamps (TL K 40W/05, Philips, emission maximum 365 nm), with magnetic stirring. Two replicates were made of each irradiation experiment and the results were averaged. Blank runs were carried out, in the absence of irradiation, by wrapping the cells with aluminium foil and placing them under the lamp. The time evolution of TMP was also monitored upon irradiation in Milli-Q water.

The UV irradiance (295-400 nm) reaching the cells was 22 ± 1 W m⁻², measured with a CO.FO.ME.GRA. (Milan, Italy) power meter. The photon flux in solution was 1.6×10^{-5} Einstein L⁻¹ s⁻¹, actinometrically determined with the ferrioxalate method (Kuhn et al., 2004). Figure 1 shows the emission spectrum of the lamps, taken with an Ocean Optics SD2000 CCD spectrophotometer and normalised to the actinometry data, taking into account the Pyrex transmittance (Albinet et al., 2010a). The Figure also reports the absorption spectra of the filtered lake water samples (A₁(λ) = A(λ) b^{-1} , where A(λ) is the measured absorbance and *b* the optical path length), taken on a Varian CARY 300 UV-Vis spectrophotometer with a cylindrical quartz cuvette having b = 10 cm.

Irradiation time was up to 32 h, which approximately corresponds to the sunlight UV energy input of three fair-weather summer days at 45°N latitude (Maddigapu et al., 2011). TMP after irradiation was monitored by HPLC. The isocratic eluent was a 60:40 mixture of methanol: aqueous H_3PO_4 (pH 2.8) at a flow rate of 1.0 mL min⁻¹. The TMP retention time was 6.7 min, detection wavelength 210 nm.

Assessment of the rates of triplet state formation. Radiation absorption by organic compounds in surface water yields the excited singlet states, which by inter-system crossing can be transformed into the longer-lived triplet states, T* (Hoigné, 1990). T* would mainly react with O₂ to give ${}^{1}O_{2}$ or undergo various deactivation reactions. Such processes have a lumped rate constant $k' \approx 5 \cdot 10^{5} \text{ s}^{-1}$ (Canonica and Freiburghaus, 2001). Moreover, T* can also react with dissolved organic molecules.

TMP is a suitable probe to measure the formation rate of T*, R_{T*} (Halladja et al., 2007; Minella et al., 2011) due to its selectivity. TMP can also react with ¹O₂, but the ¹O₂ contribution to TMP transformation would be negligible compared to T* (see SM).

Assume R_{TMP} as the experimentally measured initial rate of TMP transformation in lake water, while R_0 is the rate found upon irradiation of TMP in Milli-Q water, due to different unaccounted for processes (*e.g.* direct photolysis, volatilisation). No TMP transformation was detected with lake

water in the dark, excluding important microbiological processes. Therefore, the difference R_{TMP} - R_{o} would represent the TMP transformation rate that is accounted for by T*. Considering the competition for T* between TMP (with second-order rate constant $k_{\text{TMP},\text{T*}} = 3.0 \cdot 10^9 \text{ M}^{-1} \text{ s}^{-1}$; Al Housari et al., 2010) and the other deactivation processes (with pseudo-first order rate constant $k' = 5 \cdot 10^5 \text{ s}^{-1}$, Canonica and Freiburghaus, 2001) and applying the steady-state approximation to T*, the initial rate of TMP-reactive triplet formation $R_{\text{T*}}$ can be obtained as follows (the complete kinetic treatment is reported as SM):

$$R_{T^{*}} = (R_{TMP} - R_{o}) \cdot \frac{k_{TMP,T^{*}} [TMP]_{o} + k'}{k_{TMP,T^{*}} [TMP]_{o}}$$
(1)

where $[TMP]_o = 10 \ \mu M$ is the initial TMP concentration. The TMP time trends were fitted with equations $[TMP]_t = [TMP]_o (1 - k t)$, obtaining $R_{TMP} = k \ [TMP]_o$. The linear time trend of TMP suggests zero-order kinetics, probably due to TMP being a major T* sink under the adopted conditions (Minella et al., 2011).

Numerical data treatment. Linear and non-linear fits (to calculate reaction rates and spectral slopes, respectively) were carried out with the FigP software package (BioSoft, UK).

Results and discussion

T* formation rate. Figure 2 reports the time trend of TMP, added to the lake water samples at 10 μ M initial concentration, upon UVA irradiation. The corresponding values of R_{TMP} and $R_{\text{T}*}$ are reported in Table 1. Two samples (from Lago Nero and Lago della Foppa, which are both located below the tree line) had markedly higher $R_{\text{T}*}$ values than the others. On the other hand, negligible $R_{\text{T}*}$ was observed in the case of Lago Rouen, which is mostly surrounded by exposed rocks. Such an environment is expected to decrease the importance of soil runoff as organic matter source (Laurion et al., 2000). Soil-derived organic matter usually includes a considerable fraction of humic substances (Vinebrooke and Leavitt, 1998) with significant photochemical activity (Halladja et al., 2007). Moreover, precipitation might be a significant contributor to lake water chemical composition above the tree line and with no meadows around the lake (Mladenov et al., 2011), and there is some evidence for rainwater CDOM to be less photoactive than that of surface water (Albinet et al., 2010b).

The polychromatic quantum yield of T* formation was assessed for the different lake water samples, as $\Phi_{T^*} = R_{T^*} P_a^{-1}$, where P_a is the photon flux absorbed by lake water. From the water absorption coefficient $A_1(\lambda)$ and the incident spectral photon flux density of the lamp, $p^{\circ}(\lambda)$ (Figure 1), one derives the absorbed photon flux:

$$P_a = \int_{\lambda} p^{\circ}(\lambda) \cdot [1 - 10^{-A_1(\lambda) \cdot b}] d\lambda$$
⁽²⁾

where b = 1.6 cm is the optical path length of the irradiated solutions. The lamp and lake water spectra overlap in the 300-500 nm wavelength range, where CDOM is expected to be the main radiation absorber (Bracchini et al., 2004). Φ_{Γ^*} values for the different samples are reported in Table 1, and Lago Verde and Lago della Foppa (both located below the tree line) showed the highest Φ_{Γ^*} .

Spectrophotometric characterisation of lake water CDOM. CDOM was characterised for DOC-specific absorbance (Rostan and Cellot, 1995; Westerhoff and Anning, 2000) and spectral slope (Vodacek et al., 1997). As DOC-specific absorbance we adopted the ratio $A_1(254\text{nm})$ DOC⁻¹ between the absorption coefficient at 254 nm (cm⁻¹) and the DOC value (g C L⁻¹) (Oliveira et al., 2006). The spectral slope *S* was determined by fitting the lake water $A_1(\lambda)$ data between 295 and 400 nm with the exponential equation $A_1(\lambda) = A_0 e^{-S \lambda}$ (Laurion et al., 2000), with A_0 and *S* as fit parameters. Most of the lake water $A_1(\lambda)$ spectra showed good exponential decays with wavelength (see Figure 1). The spectrum of Lago Rouen had a small peak between 300 and 335 nm (Figure 1), which is usually associated to material released from plankton (Vernet and Whitehead, 1996). The wavelengths associated to that peak were not considered in the exponential fit (Laurion et al., 2000), but this procedure had limited effect on *S*: peak elimination decreased the measured *S* value of Lago Rouen by less than 10%. A further spectrophotometric variable considered is the spectral index E_2/E_3 (ratio of the absorbance values at 250 and 365 nm).

The values of $A_1(254\text{nm}) \text{DOC}^{-1}$, E_2/E_3 and *S* for the samples are reported in Table 1. Note the elevated $A_1(254\text{nm}) \text{DOC}^{-1}$ of Lago Nero (> 20 cm L (g C)⁻¹), which might be associated to the presence of abundant Humic-like substances (possibly fulvic acids). In contrast, values lower than 10 observed for Lago Verde, Rosset and Rouen suggest significant occurrence of non-absorbing aliphatic material (Oliveira et al., 2006).

The *S* values are in the 12-19 μ m⁻¹ range, which is typical of mountain lakes (Laurion et al., 2000). Usually, *S* is negatively correlated to the molecular weight and/or aromaticity degree of CDOM, as is the index E₂/E₃ (Table 1) (Peuravuori and Pihlaja, 1997).

Interestingly, the Lago Rouen sample had the highest *S* and E_2/E_3 values, which is usually linked to low occurrence of humic substances (Oliveira et al., 2006). The latter mainly reach lake water from catchment runoff (Vinebrooke and Leavitt, 1998), but would be leached in limited amount from an environment dominated by exposed rocks (Laurion et al., 2000). Contribution of precipitation could also be important in such an environment (Mladenov et al., 2011).

It is interesting to compare the *S* values reported in Table 1 with those derived by exponential fit of rainwater spectra, collected in the same region of the lakes (Albinet et al., 2010b). The much scattered *S* values of rainwater range from 12 to over 100 μ m⁻¹, but they are on average considerably higher compared to lake water. This finding is reasonable, considering that organic compounds in the atmosphere have usually lower molecular weight than those in surface waters (Graber and Rudich, 2006). In fact, while photochemical reactions in the atmosphere could lead to formation of oligomers with molecular weight up to 900 Da (Grgic et al., 2010), humic substances in surface waters are over ten times larger (Retamal et al., 2007).

Fluorescence characterisation of lake water CDOM. Fluorescent constituents were identified on the basis of previously reported data (Coble, 1996; Baker, 2001). Figure 3 reports the EMM plots of Lago Nero, della Foppa, Nivolet and Rouen. Several types of fluorescence signals are observed that can be attributed to Tryptophan-like (autochthonous) and Humic-like (allochthonous) material. The other EEM plots (Lago Verde, Soprano, Sottano, Rosset) are reported in Figure 2-SM. The signals are characterised by Ex_{max} and Em_{max} , the respective wavelengths of maximum excitation and emission. The linear features are the first and second harmonic of the Rayleigh-Tyndall scatter (emission wavelength equal to or double than excitation wavelength), and the Raman scattering of water (Baker, 2002).

For Lago Nero we observed two main signals corresponding to an intense Humic-like peak A $(Ex_{max}/Em_{max} 250-275 \text{ nm}/400-480 \text{ nm})$ and to Humic-like peak C $(Ex_{max}/Em_{max} 305-330 \text{ nm}/410-450 \text{ nm})$. Lago Verde showed a major peak corresponding to Tryptophan-like compounds (peak T, $Ex_{max}/Em_{max} 270-280 \text{ nm}/330-360 \text{ nm})$ and, with lower intensity, Humic-like peak A at Ex_{max}/Em_{max} around 250/435 nm. In Lago Nivolet and Lago Rosset only the Tryptophan-like signal was observed. A similar signal was present in Lago della Foppa, together with a smaller fluorescence peak at $Ex_{max}/Em_{max} 250-254 \text{ nm}/420-460 \text{ nm}$ that can be attributed to Humic-like material ("peak A"; Coble, 1996).

Lago Soprano and Sottano, with similar EMM, have contours corresponding to Tryptophan-like compounds, peak A of Humic-like material (with very low intensity for Lago Sottano) and a further peak located at $Ex_{max}/Em_{max} = 310-320 \text{ nm}/370-400 \text{ nm}$, which was also found in Lago Rouen as its only important EEM feature. Such a peak (PD) has been described previously and assigned to material arising from phytoplankton decomposition (Zhang et al., 2009). It is most likely a signature of autochthonous organic matter.

All lake samples above the tree line have autochthonous organic material, which is consistent with an important role of in-lake productivity compared to catchment runoff. In contrast, Humiclike material was detected in all samples from lakes below the tree line.

Principal Component Analysis (PCA) of the data. After column autoscaling, the data matrix reported in Table 1 (underlined variables) was subjected to PCA to get insight into possible correlations. PCA was carried out with the free chemometric software V-Parvus 2008 (Forina et al., 2008). The first two principal components explained 67% of total variance, and PC3 explained a further 16%. Figure 4 reports the plot of PC1 *vs.* PC2. R_{T^*} and Φ_{T^*} , as well as inorganic carbon, showed good correlation with high positive loadings on PC₁ and low loadings on PC₂. PC₁ could thus be able to differentiate the photochemical activity of the samples. It is also interesting to see an anticorrelation (loadings pointing in opposite directions) between Φ_{T^*} and the variables *S* and E₂/E₃, which suggests that Φ_{T^*} would be higher in samples where CDOM has higher aromaticity/molecular weight (low *S* and E₂/E₃). This finding is consistent with the significant photochemical activity of humic substances (Halladja et al., 2007).

The plot of PC1 *vs.* PC2 does not show a clear differentiation between the lake water samples. However, the sample scores on PC1 suggest some agreement between photoactivity data and environmental features derived from the kind of vegetation surrounding the lakes. In fact, two of the samples from lakes below the tree line (Lago Nero and Lago della Foppa) have elevated positive scores on PC1. In contrast, Lago Rouen that is surrounded by rocks with practically no vegetation has a highly negative score on PC1, and all the other samples have relatively low absolute values.

It was unfortunately not possible to quantify the vegetation type. However, Spearman's rank correlation for ordinal data was applied with the following vegetation ranking: trees – meadows – rocks. The test gave statistically significant correlation between R_{T^*} and vegetation type (p = 0.039). By comparison, insignificant rank correlation with vegetation type was found for DOC, nitrate, nitrite or for the spectral features $A_1(254\text{nm}) \text{ DOC}^{-1}$, *S* and E_2/E_3 .

Conclusions

In the present study, the highest TMP-reactive T* photoproduction was observed in samples from lakes located below the tree line. The CDOM in the most photoactive samples had relatively high values of $A_1(254\text{nm}) \text{ DOC}^{-1}$ (Table 1) and significant levels of humic substances as shown by EEM (Figure 3). Samples from lakes above the tree line and surrounded by alpine meadows had usually lower values of T* photoproduction. Their CDOM spectral features were very variable, but EEM data suggested an important contribution from autochthonous CDOM sources. Water from Lago Rouen, located above the tree line and surrounded by rocks with no meadows showed negligible photoactivity and low $A_1(254\text{nm}) \text{ DOC}^{-1}$. Its CDOM also had the highest values of spectral slope *S* and E_2/E_3 index, suggesting low molecular weight and aromaticity. This implies low CDOM contribution from catchment runoff, while EEM suggested the presence of autochthonous material.

Overall, the generation rate of TMP-reactive T* showed better rank correlation than chemical composition with the vegetation type surrounding the lakes. This finding suggests that, at least in the studied samples, photochemistry could be more sensitive than water chemistry to the lake location with respect to the tree line. Further studies will be needed to check the validity of this assumption.

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Table 1. Features of the sampled lakes. IC: inorganic carbon $(H_2CO_3 + HCO_3^- + CO_3^{2-})$. Note: $(1.1\pm0.1)E-10$ is a compact notation for $(1.1\pm0.1)\cdot10^{-10}$, and so on. n/a: not applicable. The average depth of the lakes is in the range of 1-2 m. E_2/E_3 is the ratio between the absorbance values at 250 and 365 nm (Peuravuori and Pihlaja, 1997). The error bounds represent $\mu\pm\sigma$. The underlined variables were included in PCA.

Lake		Altitude, m Samp		Samplir	pling date Surface, km ²		Tree line	Surrounding environment			рН	P _a 10 ⁻⁷ Ein	
Lago Nero		20	70	1 Sep 2011		0.110	Below	Trees (fir), water plants, meadows			7.2	5.1	
Lago Verde		18	40	1 Sep 2011		0.012	Below	Trees (fir)			7.4	4.5	
Lago della Foppa		1450		1 Sep 2011		0.016	Below	Trees (deciduous)			7.5	0.96	
Lago Nivolet		2750		30 Aug 2011		0.010	Above	Meadows, some rocks			6.7	0.82	
Lago Rosset		27	00	30 Aug 2011		0.166	Above	Meadows			7.4	0.4	4
Lago Soprano		21	100 31 Aug		2011	0.018	Above	Meadows, some rocks			6.8	1.	6
Lago Sottano		22	10	31 Aug 2011		0.011	Above	Meadows, some rocks			6.8	0.80	
Lago Rouen		23	90	31 Aug 2011		0.018	Above	e Rocks, no meadows			7.5	0.28	
								1				1	
	R_{TMI}				$\underline{\Phi}_{\underline{\Gamma}^*}, 10^{-3}$	<u>DOC</u> ,	<u>IC</u> ,	<u>NO</u> 3 ⁼ , µМ	<u>NO</u> 2 ⁼ ,	<u>A₁(254nm)/</u>	<u>DOC</u> ,	<u>S</u> , μm ⁻¹	<u>E₂/E₃</u>
	10^{-11} M	I s ⁻¹				mg C L ⁻¹	mg C L^{-1}		μM	cm L/ g	C		
Nero	11±	:1 1.8		±0.1 3.5±0.2		1.80±0.06	28.4±6.8	< 0.8	0.36	29.8±1.0		15.5±0.1	5.28
Verde	3.2±0	0.1 0.4		±0.03 9.1±0.7		0.43±0.02	19.0±3.0	12.7	0.28	8.6±0.4		13.7±0.2	4.64
Foppa	12±2		1.9±0.4		2.0±0.4	0.53±0.01	23.6±4.4	6.01	< 0.02	19.0±0.4		15.6±0.1	5.80
Nivolet	3.6±0.1		0.49±0.01		6.0±0.2	0.62±0.01	14.9±2.1	1.68	0.04	13.0±0.2		12.1±0.2	5.05
Rosset	2.1±0.2		0.22±0.04		5.0±0.9	0.64±0.01	12.2±1.0	< 0.8	0.31	7.8±0.1		13.5±0.2	6.68
Soprano	4.1±0	2	0.57±0.05		3.6±0.3	0.84±0.04	1.08±0.01	25.4	1.07	18.7±0.9		15.0±0.1	5.37

Sottano	2.3±0.1	0.25±0.02	3.1±0.2	0.48±0.01	0.16±0.01	28.9	0.24	17.1±0.4	15.5±0.1	5.64
Rouen	0.23±0.01	Negligible	Negligible	0.53±0.01	0.56±0.01	8.52	0.69	9.1±0.2	18.9±0.4	11.3
Milli-Q	0.85±0.03	n/a	n/a	0.13±0.01	0	< 0.8	< 0.02	~ 0	n/a	n/a

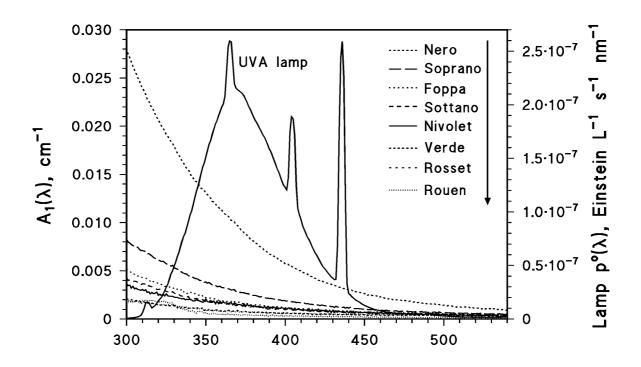


Figure 1. Emission spectrum (spectral photon flux density $p^{\circ}(\lambda)$) of the adopted UVA lamps (Philips TL K05). Absorption spectra of the studied lake water samples ($A_1(\lambda) = A(\lambda) b^{-1}$, b = 10 cm), listed from the most to the least intense.

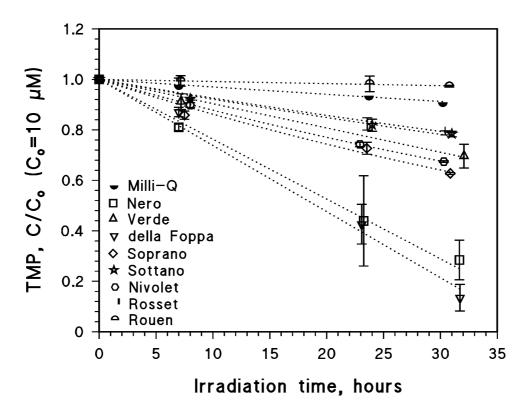


Figure 2. Time evolution of 10 μ M TMP, spiked to the lake water samples or to Milli-Q water, upon UVA irradiation. The error bars represent $\mu \pm \sigma$ of duplicate runs.

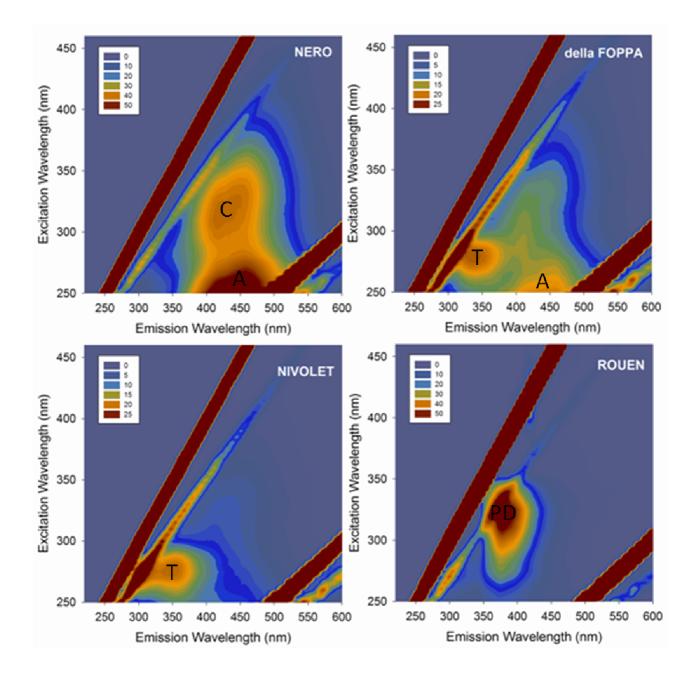


Figure 3. 3D fluorescence excitation-emission matrix (as contour plot) of Lago Nero, della Foppa, Nivolet and Rouen. Fluorescence intensity is scaled from 0 to 50 intensity units for Lago Nero and Rouen and from 0 to 25 for Lago della Foppa and Nivolet. A, C: humic-like peaks; T: tryptophan-like peak; PD: Phytoplankton Decomposition.

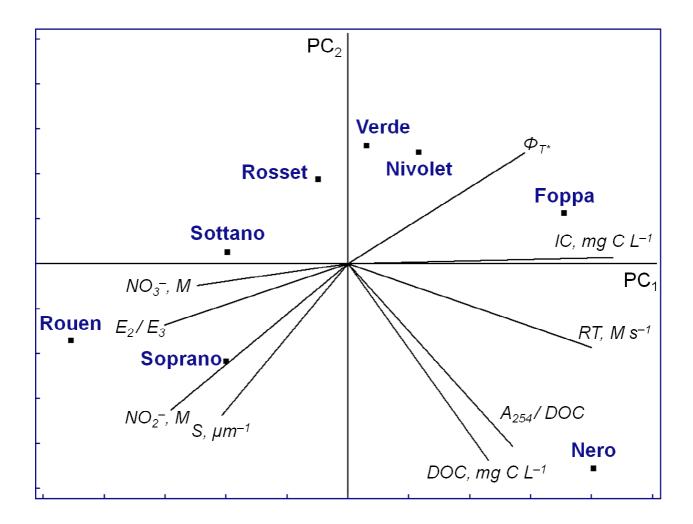


Figure 4. Principal Component Analysis (PC1 vs. PC2) of the numerical data reported in Table 1.