

UNIVERSITÀ DEGLI STUDI DI TORINO

This is an author version of the contribution published on:

G. CARNEVALE; T.W. PIETSCH †Caruso, a new genus of anglerfishes from the Eocene of Monte Bolca, Italy, with a comparative osteology and phylogeny of the family Lophiidae (Teleostei: Lophiiformes) JOURNAL OF SYSTEMATIC PALAEONTOLOGY (2012) 10

2	<i>†Caruso</i> , a new genus of anglerfishes from the Eocene of Monte
3	Bolca, Italy, with a comparative osteology and phylogeny
4	of the family Lophiidae (Teleostei: Lophiiformes)
5	
6	Giorgio Carnevale ^{a,*} and Theodore W. Pietsch ^b
7	
8	^a Dipartimento di Scienze della Terra, Università degli Studi di Torino, Via Valperga Caluso,
9	35 I-10125 Torino, Italy; ^b School of Aquatic and Fishery Sciences, College of the
10	Environment, University of Washington, Campus Box 355020, Seattle, Washington 98195-
11	5020, USA
12	
13	
14	Abstract
15	<i>†Caruso</i> , a new genus of lophiid anglerfishes, erected to contain <i>†Lophius brachysomus</i>
16	Agassiz from the Eocene of Monte Bolca, Italy, is described and compared osteologically
17	with all known representatives of the family, both extinct and extant. Together with <i>†Sharfia</i> ,
18	known from a single specimen also from Monte Bolca, † <i>Caruso</i> is the oldest member of the
19	teleost family Lophiidae known to date, based on articulated skeletal remains. It possesses
20	several autapomorphic features as well as a unique combination of character states that
21	clearly separate it from all other known lophiiform fishes. Evidence is provided to show that
22	<i>†Sharfia</i> is sister group of all other known lophiid taxa, that <i>†Caruso</i> is most closely related
23	phylogenetically to the extant genus Sladenia, and that these two genera together form the

^{*}Corresponding author. E-mail: giorgio.carnevale@unito.it

24 sister group of all remaining members of the family. *†Caruso brachysomus* is the third valid extinct taxon of the Lophidae known from articulated skeletal remains. Candidate 25 developmental mechanisms that may have contributed to the origin of the lophiiform body 26 27 plan are also suggested. The hypertrophic development of the jaws, suspensorium, hyoid apparatus and branchial arches typical of lophiiforms may be correlated, possibly under the 28 influence of discrete sets of genes. The synapomorphic elongation of the pectoral-fin radials 29 might be related to a heterochronic shift of the onset of the expression of the gene sonic 30 *hedgehog*. Finally, the highly modified spinous dorsal fin may have arisen by 31 32 submodularization and co-option of the first dorsal-fin module, whereas its forward shift over the neurocranium may represent an anterior shift of the stripes of competency for dorsal-fin 33 formation along the dorsal midline. 34 35

Keywords: Teleostei; Lophiiformes; Lophiidae; comparative osteology; phylogenetic
analysis; Eocene; Monte Bolca

38

39

40 Introduction

Anglerfishes of the family Lophiidae are the sole members of the Lophioidei, one of the five 41 suborders of the order Lophiiformes (see Pietsch & Grobecker 1987; Pietsch & Orr 2007), a 42 43 morphologically derived group of marine teleost fishes that includes some of the most bizarre and anatomically peculiar representatives of the animal kingdom. The Lophiidae consists of 44 four extant genera with fewer than 30 described species (e.g., Regan 1903; Caruso & Bullis 45 1976; Caruso 1981; 1983) that inhabit tropical, subtropical and temperate waters of the 46 Atlantic, Indian and Pacific oceans. These fishes, also known as monkfishes or goosefishes, 47 are benthic on different substrates at depths ranging from the shoreline to greater than 1500 48

meters. They are characterized by having a dorsoventrally compressed head, a wide 49 cavernous mouth and a luring apparatus, the latter primarily consisting of a highly modified 50 anterior spine of the dorsal fin, commonly called the illicium. The angling behavior of 51 lophiids was reported for the first time by Aristotle in his Historia animalium and 52 subsequently by many naturalists in the eighteenth and nineteenth centuries (e.g., Parsons 53 1750; Hanow 1768; Montin 1779; Geoffroy St. Hilaire 1807; 1824; 1827; Bailly 1824). 54 Lophiid anglerfishes are opportunistic feeders with a common feeding strategy; they are lie-55 and-wait predators settling on soft-bottom substrates and luring their prey by wriggling the 56 57 illicium (e.g., Wilson 1937; Gudger 1945). Some species of this family, mostly belonging to the genus Lophius, support important commercial fisheries, and for this reason several 58 aspects of their life history and reproductive biology are extremely well documented (see, 59 60 e.g., Fariña et al. 2008). However, despite the economic relevance and abundance of these fishes, their osteological structure remains poorly known and almost totally restricted to 61 species of the genus Lophius (Brühl 1856; Morrow 1882; Supino 1908; Regan 1912; Gregory 62 63 1933; Montcharmont 1950; Eaton et al. 1954; Monod 1960; Field 1966; Oliva et al. 1968; Le Danois 1974). 64

The family Lophiidae is scarcely represented in the fossil record, mostly by 65 disarticulated fragmentary bones (Lawley 1876; De Stefano 1910; Leriche 1910; 1926; Ray 66 et al. 1968; Landini 1977; Purdy et al. 2001; Schultz 2006), otoliths (see, e.g., Nolf 1985) and 67 teeth (Leriche 1906; 1908; Hasegawa et al. 1988). Articulated skeletal remains are extremely 68 rare. Eocene material is restricted to a few specimens from the localities of Monte Bolca, 69 Italy (e.g., Agassiz 1833-1844; Zigno 1874) and Gornyi Luch, North Caucasus, Russia 70 (Bannikov 2004); whereas Neogene fossils are known from Algeria (Arambourg 1927), 71 Azerbaijan (Sychevskaya & Prokofiev 2010), and Italy (Sorbini 1988). 72

73 In this study we describe a new genus of the Lophiidae, including its anatomy, based on material from the Eocene of Monte Bolca, previously referred to the genus Lophius (Volta 74 1796; Agassiz 1833-1844; see synonymy below). Previous to this study, the family Lophiidae 75 76 had never been carefully investigated osteologically and phylogenetically. We therefore set out to examine the comparative osteology of the Lophiidae phylogenetically. This work is 77 another in a series of papers focused on the fossil record of lophilform fishes (Carnevale & 78 Pietsch 2006; 2009a; 2009b; 2010; in press; Carnevale et al. 2008; Pietsch & Carnevale, in 79 press). 80

81

82 Materials and methods

Specimens were examined under binocular microscopes equipped with camera lucida 83 84 drawing arms. Standard length (SL) is used throughout. Methods for taking counts and measurements mostly follow Caruso (1981), whereas osteological terminology follows 85 86 Montcharmont (1950) and Pietsch (1981). Extant comparative material was cleared and double stained with alizarin red S and alcian blue following the trypsin digestion technique of 87 Potthoff (1984). Fossil specimens were mechanically prepared whenever possible using 88 89 mounting entomological needles. All extinct taxa are marked with daggers ([†]) preceding their names. The methodologies employed in the phylogenetic analysis are presented below. 90 Material examined is housed in the following institutions: Borysiak Paleontological 91 92 Institute, Russian Academy of Sciences, Moscow (PIN); Burke Museum of Natural History and Culture, University of Washington, Seattle (UW); Commonwealth Scientific and 93 Industrial Research Organization, Hobart, Tasmania (CSIRO); Museo Civico di Storia 94 Naturale, Verona (MCSNV); Muséum National d'Histoire Naturelle, Paris (MNHN); 95 Museum of Comparative Zoology, Harvard University, Cambridge (MCZ); National 96

97	Museum of Victoria, Melbourne (NMV); Natural History Museum, London (NHM);
98	Queensland Museum, Brisbane (QM); Tulane University, New Orleans (TU).
99	Comparative lophiiform material is listed below, all but fossils were cleared and double
100	stained:
101	Antennariidae: Antennarius striatus, UW 20768, 57 mm SL.
102	Brachionichthyidae: Brachionichthys australis, UW 116842, 38.7 mm SL; UW 116843,
103	55 mm SL; Brachionichthys hirsutus, NMV A.19954, 80 mm SL; †Histionotophorus
104	bassani, MCSNV I.G.23163, 50.5 mm SL; NHM 19060, 16.4 mm SL; †Orrichthys
105	longimanus, MCSNV T.160/161, 63 mm SL; Sympterichthys unipennis, UW 116844, 31.6
106	mm SL; UW 116845, 34 mm SL.
107	Chaunacidae: Chaunax pictus, UW 20770, 90 mm SL.
108	Lophichthyidae: Lophichthys boschmai, UW 20773, 47 mm SL.
109	Lophiidae: † <i>Eosladenia caucasica</i> , PIN 4425-72, 83 mm SL; <i>Lophiodes caulinaris</i> ,
110	MCZ 51260, 33.5 mm SL; TU 72942, 152 mm SL; Lophiodes monodi, MCZ 40928, 92 mm
111	SL; Lophiodes spilurus, TU 78474, 71 mm SL; Lophiomus setigerus, TU 81104, 166 mm SL;
112	Lophius americanus, MCZ 51259, 121 mm SL; †Sharfia mirabilis, MNHN Bol 38-39, 39.9
113	mm SL; Sladenia remiger, CSIRO H.2559-02, 133 mm SL.
114	Tetrabrachiidae: Tetrabrachium ocellatum, QM I.27988, 49 mm SL; QM I.30596, 56
115	mm SL.
116	Anatomical abbrevations: alpmx , alveolar process of the premaxilla; ar , articular;
117	aspmx , ascending process of the premaxilla; cl , cleithrum; co , coracoid; cs , cleithral spine;
118	ctb, ceratobranchial; d, dentary; epi, epioccipital; epb, epibranchial; f, frontal; iop,
119	interopercle; h, hyomandibula; hyb, hypobranchial; ih, interhyal; le, lateral ethmoid; mtp,
120	metapterygoid; mx , maxilla; op , opercle; pa , parietal; pal , palatine; pas , parasphenoid; pcl ,
121	postcleithrum; phb , pharyngobranchial; pmx , premaxilla; pop , preopercle; pte , pterygoid;

pto, pterotic; ptt, posttemporal; q, quadrate; rad, pectoral-fin radial; sca, scapula; scl,
supracleithrum; soc, supraoccital; sop, subopercle; sos, supraocular spine of the frontal; spo,
sphenotic; sym, symplectic; tpc, transverse process of the caudal centrum; v, vertebra; vo,
vomer.

127 Systematic palaeontology

128	Subdivision Teleostei sensu Patterson & Rosen, 1977
129	Order Lophiiformes Garman, 1899
130	Suborder Lophioidei Regan, 1912
131	Family Lophiidae Rafinesque, 1810
132	Genus † <i>Caruso</i> nov.
133	(Figs. 1-8)

134

135 Diagnosis. A member of the Lophiidae with body moderately depressed, rounded, and relatively globose, and a compressed and moderately elongate tail; frontals heavily 136 137 constricted in the interorbital region (distance between the lateral margins of the anteromedial extensions of the frontals is about 18% of that measured between the outer margins of the 138 pterotics); dorsolateral ridge of the frontals smooth; elongate frontal fontanel; supraocular 139 spines prominent, representing the anterolateral corners of the postorbital sector of the 140 neurocranium; dorsal aspect of the parasphenoid narrow; symphysial process of the dentary 141 absent; retroarticular process of the articular well developed; maxillary process of the palatine 142 simple, with rounded profile; pterygoids (endo- + ecto-) greatly enlarged, ovoid in outline; 143 subopercle with fimbriate posteroventral margin; 19 vertebrae; haemal spines of the 144 abdominal vertebrae well developed and anteroposteriorly expanded; neural and haemal 145 spines of the penultimate vertebra widely expanded; three post-cephalic dorsal-fin spines; 146

nine dorsal-fin rays; anterior pterygiophore of the soft dorsal fin shortened and
anteroposteriorly expanded, bearing two rays, one in supernumerary association;
posteriormost dorsal-fin pterygiophore bearing two rays; six anal-fin rays; anteriormost and
posteriormost pterygiophores bearing two rays (one supernumerary on the anterior
pterygiophore); distal end of the posteriormost anal-fin pterygiophore expanded, notably
globose with rounded profile; cleithral spine short; pectoral-fin radials greatly elongate, the
ventralmost measuring about 23% SL; pectoral fin paddle-shaped; 14-16 pectoral-fin rays.
Etymology. We are pleased to name this genus in honor of John H. Caruso of Tulane
University for his years of service to the world ichthyological community and for his many
published contributions to anglerfish systematics.
Included species. Monotypic.
<i>†Caruso brachysomus</i> (Agassiz, 1835)
1796 Lophius piscatorius Volta: pl. 42, fig. 3 (misidentification).
1818 Lophius piscatorius var. Gonelli Risso; De Blainville: 340, 342 (in part;
misidentification based on Lophius piscatorius Volta).
1835 Lophius brachysomus Agassiz (name only): 292 (in part).
1844 Lophius brachysomus Agassiz; Agassiz: 114, vol. 5, pl. 1, figs 1-2.
1874 Lophius brachysomus Agassiz; Zigno: 105 (in part).
Non 1876 Lophius brachyostomus Agassiz; Lawley: 77, pl. 5, figs 2a-c.
1901 Lophius brachysomus Agassiz; Woodward: 591 (in part).
1905 Lophius brachysomus Agassiz; Eastman: 31 (in part).

- 172 1922 Lophius brachysomus Agassiz; D'Erasmo: 140-141 (in part).
- 173 1927 'Lophius' brachysomus Agassiz; Arambourg: 216.
- 174 1967 Lophius brachysomus Agassiz; Andrews, Gardiner, Miles & Patterson: 660.
- 175 Non 1972 Lophius brachysomus Agassiz; Sorbini; pl. 18 (misidentification).
- 176 1980 Lophius brachysomus Agassiz; Blot: 353-354 (in part).
- 177 1983 Lophius brachysomus Agassiz; Sorbini; pl. 9.
- 178 Non 1991 Lophius brachysomus Agassiz: Frickhinger: 694 (misidentification).
- 179 Non 1996 *Lophius brachysomus* Agassiz: Long: 157, fig. 2 (misidentification).

180

181 **Diagnosis**. As given for the genus.

182

- **Holotype**. MNHN Bol42/43, a relatively complete skeleton in part and counterpart, 125.7
- 184 mm SL; late early Eocene, Ypresian; Monte Bolca, Pesciara cave site (Fig. 1).

185

- 186 Additional material. MCSNV T.978, a complete well-preserved skeleton, 136.5 mm SL,
- 187 from the type locality (Fig. 2A); this specimen is part of the Baja's fossil collection of the
- 188 Museo Civico di Storia Naturale, Verona, and was figured in a photographic catalogue
- published by Lorenzo Sorbini (1983; pl. 9). MCSNV B.13, a nearly complete skeleton, 92.1

190 mm SL, from the type locality (Fig. 3); not previously recognized.

191

Horizon and locality. Late early Eocene, Ypresian, *Alveolina dainelli* Zone; Monte Bolca,
Pesciara cave site.

194

Biotope and palaeoecology. The finely laminated micritic limestone of the Pesciara cave site
has provided a huge amount of fossils, including thousands of exceptionally well-preserved,

fully articulated fish skeletons. The fish assemblage consists of more than 200 species of 197 sharks, batoids, pycnodontiforms and teleosts, representing the earliest evidence of an 198 acanthomorph dominated ichthyofauna, which also includes several representatives of the 199 200 fish groups found on coral reefs today (Patterson 1993; Bellwood 1996; Landini & Sorbini 1996). The fish assemblage from Monte Bolca therefore marks the starting point in the 201 known evolution of many reef fish groups and provides a reliable evidence of the 202 morphological and compositional stability of tropical and subtropical marine ichthyofaunas 203 throughout the Cenozoic (Bellwood & Wainwright 2002). 204

205 To date there is no consensus about the interpretation of the physiography and palaeoecological setting of the depositional environment during the sedimentation of the 206 micritic limestone. Based on a wide comprehensive palaeocological analysis of the fish 207 208 assemblage, Landini & Sorbini (1996) concluded that sedimentation of the fish-bearing limestone occurred at a short distance from the coast, many dozens of meters in depth in 209 close proximity to coral reefs, seagrass beds and open ocean, under the seasonal influence of 210 well-developed river systems. The fish carcasses possibly accumulated in a silled depression 211 that occasionally favored restricted circulation and bottom anoxia. Sedimentological, 212 palaeontological and taphonomic evidences, as well as the ecological spectrum of the fossil 213 assemblage, concur to indicate that the Pesciara cave laminated micritic limestone represents 214 215 an obrutionary stagnation deposit (Seilacher et al. 1985).

216

Remarks. *†Lophius brachysomus* was erected by Louis Agassiz (1835) to allocate the two
lophiid anglerfish specimens from Monte Bolca, previously documented in the *Ittiolitologia Veronese*, the voluminous monograph by Abbot Giovanni Serafino Volta (1796), and referred
to the extant species *Lophius piscatorius* and *Loricaria plecostomus* respectively. The two
lophiid specimens, both in part and counterpart, were part of the Gazola collection of fossil

fishes from Monte Bolca that was confiscated by the armies of Napoleon, and deposited in 222 the Muséum National d'Histoire Naturelle in Paris (see Pietsch and Carnevale, in press). 223 According to Agassiz (1844), the specimens represent different ontogenetic stages of a single 224 species; the smaller one was formerly assigned to the catfish species Loricaria plecostomus 225 by Volta (1796), while the larger one was considered as the lithified counterpart of the 226 Atlantic-Mediterranean anglerfish species Lophius piscatorius. Several authors (see 227 synonymy) followed the taxonomic interpretation of Agassiz up to the second half of the 228 1920s when Camille Arambourg (1927) questioned the generic identity of these fossils and, 229 230 mainly based on their number of vertebrae, considered them to be related to the extant genus Lophiomus. Subsequently, Andrews et al. (1967) emphasized the necessity of a new detailed 231 systematic study of these fossils in order to confirm their position. More recently, Blot (1980) 232 233 cursorily discussed the generic identity of these specimens suggesting that a new generic name would be necessary to include them. A recent examination of the specimens in April 234 2010, however, has revealed the existence of two different taxa among the type material 235 housed in the MNHN in Paris. The smaller specimen (MNHN Bol38/39) that was formerly 236 assigned to Loricaria plecostomus by Volta (1796) has been described as a new genus and 237 species (Pietsch & Carnevale in press). The morphology of the larger specimen is described 238 herein together with that of additional specimens residing among the collections of the 239 MCSNV. 240

241

242 Anatomical descriptions

Despite the small number of specimens (see Material section above), the available material of *†Caruso brachysomus* is relatively complete and reasonably well exposed from the matrix;
therefore, although certain details are not evident, it has been possible to properly realize a
morphological analysis of this taxon (see Fig. 2B).

247 Measurements are summarized in Table 1.

The body is moderately depressed with a large, rounded and relatively globose head, and a compressed and moderately elongate tail.

250 There is no evidence of dermal spinules so that the skin appears to be naked.

251

Neurocranium. The neurocranium (Fig. 4) appears to be greatly depressed dorso-ventrally 252 and elongate, with its maximum width measured between the lateral extensions of the 253 pterotics evidently less than its length (cranial width about 65% of the length). There is an 254 255 evident regionalization of the neurocranium with a sharp differentiation into preorbital (rostral), orbital and postorbital sectors clearly related to a strong constriction at the level of 256 the midlength of the frontals. The orbital-postorbital portion of the neurocranium is broadly 257 258 expanded laterally with respect to the preorbital portion, which is narrow and linear; the 259 distance between the lateral margins of the anteromedial extensions of the frontals equals about 18% of that measured between the outer margins of the pterotics. The dorsocranial 260 261 architecture is recognizable, at least in part, in all the examined specimens; as a consequence, there is no evidence of the basicranial morphology. Because of the partial flattening of the 262 bones due to the fossilization process, the presence of the spines that characterize the dorsal 263 aspect of the neurocrania of extant lophiids cannot be observed. On the other hand, the 264 original presence of prominent ridges that developed throughout the dorsal surface of the 265 266 skull roof can be easily interpreted; in extant lophiids dorsocranial spines recurrently emerge in specific points of these ridges (see Caruso 1985), thereby suggesting that their presence in 267 the fossils may be predicted, at least in certain cases (parietal, epioccipital, sphenotic). 268 Another general morphological feature of the neurocranium is the presence of a moderately 269 deep medial depression, developed throughout the neurocranial length; such a long concavity, 270 which corresponds to the illicial trough of Waterman (1948), is more developed rostrally, in 271

order to accommodate the ascending processes of the premaxillae. The posterior margin of
the neurocranium is moderately concave on each side of the vertebral column, forming a
large surface for the insertion of the epaxial muscles.

The frontals are the largest bones of the skull roof. These bones are greatly expanded 275 laterally in their posterior half, becoming abruptly narrow anteriorly; such a marked 276 difference in the lateral expansion in the two halves of the frontals occurs at the level of the 277 supraocular spine (sensu Montcharmont 1950), which, as a consequence, forms the massive 278 anterior corner of the orbital-postorbital sector of the neurocranium. The bony tissue of the 279 frontals is cancellous in juveniles (MCSNV B.13), becoming dense and generally hard in 280 adult specimens (MNHN Bol42/43; MCSNV T.9787). The two contralateral frontals are 281 separated for most of their length by a large fontanel that apparently reduces in size during 282 283 ontogeny. The lateral profile of the frontals is concave in their laterally expanded posterior portion, becoming more linear anteriorly, sometimes with reduced knobs and/or crenulations. 284 Each frontal bears a dorsolateral longitudinal ridge, which represents the anterior part of a 285 developed ridge system that originates posteriorly at the boundary between the first vertebra 286 and the epioccipital. This smooth ridge can be divided into two portions, a supraorbital one 287 that culminates with the prominent anterolaterally directed supraocular spine, and a 288 preorbital-rostral portion that extends anteriorly to the lateral ethmoids. Each frontal 289 290 articulates medially (before and behind the fontanel) with its opposite member, anterior and 291 anterolaterally with the lateral ethmoid, which it partially overlies, posterolaterally with the sphenotic, posteriorly with the parietal and posteromedially with the supraoccipital. 292

The lateral ethmoids are large, crescent-shaped bones, with a gently rounded lateral profile. The medial sector of each is notably depressed, and the posterior portion lies ventral to the anteriormost extension of the respective frontal. Each lateral ethmoid articulates

296 medially with the vomer, posteriorly with the frontals, and ventromedially with the297 parasphenoid.

The vomer is a flattened and anteriorly broadened bone, which makes contact posteriorly with the parasphenoid and posterolaterally with the lateral ethmoids. Like in other lophiids, the vomer appears to be characterized by having an anterior concave edge.

There is no evidence of an ossified mesethmoid. The reduced space between the two contralateral lateral ethmoids was probably occupied by the ethmoid cartilage in origin.

The parasphenoid is only partially exposed in the lectotype and MCSNV T.978. This median bone appears to be narrow, not expanded laterally.

The parietals are polygonal in shape. These bones lie lateral to the supraoccipital and articulate anteriorly with the frontals, laterally with the sphenotic, posterolaterally with the pterotic and posttemporal, and posteromedially with the epioccipital. A prominent ridge runs longitudinally through this bone; three additional ridges merge radially into the main longitudinal ridge; a spine probably was present at the intersection of all of these ridges. The supraoccipital is a large median ovoid bone with a concave dorsal surface. This bone articulates anteriorly with the frontals, laterally with the parietals, and posteriorly and

312 posterolaterally with the epioccipitals.

The sphenotic is a robust bone that forms a rounded flange culminating with a thick blunt spine, which extends outward in lateral direction, considerably beyond the width of the supraocular spine of the frontal. The dorsal surface of this bone bears a longitudinal ridge; a spine was possibly present on this ridge originally. Each sphenotic articulates anteriorly with the frontals, laterally with the parietal and posteriorly with the pterotic.

The pterotic is thickened and flange-like laterally and more massive medially; its lateral flange terminates with a blunt rounded spine. A prominent pterotic ridge radiates medially from that spine. The distance measured between the lateral ends of the pterotic spines

coincides with the maximum width of the neurocranium. Each pterotic makes contact
anteriorly with the sphenotic, laterally with the parietal, posteromedially with the epioccipital,
and posteriorly with the posttemporal.

324 The epioccipitals are large and irregular bones that form the posteromedial border of the skull roof. There is a conspicuous longitudinal ridge, nearly sigmoid in shape, which is 325 continuous with that of the parietal; a spine was possibly present along this ridge. Each 326 epioccipital articulates medially through interdigitation with its opposite member in the 327 midline of the neurocranium, anteromedially with the supraoccipital, anteriorly with the 328 329 parietal, anterolaterally with the sphenotic and pterotic, and laterally with the posttemporal. As in other lophilforms the posttemporals are sutured to the neurocranium at its 330 dorsolateral corner. A ridge, which terminates posteriorly with a blunt and thick process, 331 332 radiates medially on the dorsolateral surface of this bone. Each posttemporal articulates 333 medially and anteromedially with the epioccipital, and anteriorly with the pterotic.

334

Jaws, suspensorium and opercular series. As in other lophiids, the gape of the mouth is
wide; the mandible length reaches more than 32% SL.

Each premaxilla (Figs. 5-6) has a long and distally spatulate alveolar process, an expanded and obliquely oriented articular process, a shallow postmaxillary process with a gently rounded dorsal profile, and a thick and labiolingually compressed autogenous ascending process. Two or three series of large conical and usually depressible teeth are visible along the anterior two thirds of the alveolar process. Fixed conical and recurved teeth insert along the alveolar surface of the distal portion of the premaxilla.

The maxillae (Fig. 5) consist of a wide posterior portion and an expanded anterior head; this bone is characterized by having a smooth external surface and, during the juvenile phase, by cancellous texture.

The mandibles (Fig. 5) are very large, characterized by having a cancellous texture in 346 juveniles. The dentaries are strongly ossified and curved bones that bear a number of strong 347 depressible conical teeth arranged in two or three rows. These bones are deeply forked 348 posteriorly to accommodate the anterior pointed extensions of the articulars. The articular has 349 a well developed retroarticular process (sensu Montcharmont 1950) that originates just 350 posterior to the glenoid fossa (quadrate-articular joint). There is no evidence of spines along 351 352 the external surface of the articular. The retroarticular is a small bone located posteromedially on the mandible. 353

354 The hyomandibulae (Fig. 5) are large and stout bones divided into three portions: the broad articular head, main vertical shaft, and anterior flange. The articular head has two 355 condyles that articulates with the lateral otic region of the neurocranium, and the opercular 356 357 process that articulates with the articulating condyle of the opercle. In the basal sector of the 358 articular head is a stout anteriorly directed spine. The hyomandibular shaft tapers distally and articulates anteriorly with the posterior margin of the metapterygoid, anteroventrally with the 359 dorsal and posterior margin of the symplectic and the dorsal margin of the quadrate, and 360 posteriorly with the anterior margin of the preopercle. The anterior flange is a thin bony 361 lamina that articulates ventrally with the process emerging from the posterodorsal corner of 362 the metapterygoid. 363

The quadrates (Fig. 5) are stout and high bones, nearly triangular in outline, with a developed articular facet for the articular at their anteroventral corner. Each quadrate articulates at the anterodorsal corner with the pterygoids, and dorsally with the symplectic, hyomandibula and metapterygoid. Quadrate spines are not preserved.

The symplectics (Fig. 5) are long rod-like bones somewhat stouter posteriorly than anteriorly. Each symplectic articulates dorsally with hyomandibula, ventrally with the quadrate, and anteriorly with the metapterygoid.

The metapterygoids (Fig. 5) are relatively large and thin plates with an irregular outer margin. A posterodorsally directed process emerges from the posterodorsal corner of these bones. Each metapterygoid articulates dorsally and posteriorly with the hyomandibula, posteriorly with the symplectic, and ventrally with the quadrate.

The pterygoids (endo- + ecto-) (Fig. 5) are extremely large and ovoid in outline, connected with the palatine anteriorly and the quadrate posteriorly. The posterior sector of these bones is thick and slightly sculptured, becoming extremely thin, nearly translucent, anteriorly. A thick longitudinal ridge, which possibly represents the ontogenetic suture between the ectopterygoid and endopterygoid (see Morrow 1882; Matsuura & Yoneda 1987), extends from the posterior margin to the anterior tip of each of these bones.

The palatines (Fig. 5) are relatively large stout bones with an expanded maxillary process (*sensu* Montcharmont 1950) and a spatulate distal portion. Some strong depressible and slightly recurved teeth insert along the ventromedial margin of these bones. Each palatine articulates anteriorly with the maxilla, anteromedially with the lateral ethmoid, and posteriorly with the pterygoid.

The preopercles (Fig. 5) are long, subcylindrical and gently curved. Functionally, these bones are part of the suspensorium. Each preopercle articulates anterodorsally with the hyomandibula and anteroventrally with the quadrate.

The interopercles (Fig. 5) are large and flattened bones, subtriangular in outline, with an anteroventrally directed apex. Interopercular spines are not visible, possibly due to inadequate preservation.

The opercles (Fig. 5, 7) are deeply indented posteriorly making them notably bifurcate; the dorsal limb is filamentous and posteriorly directed, whereas the ventrolaterally directed lower limb is stout and characterized by having a strong thickening along the anterior margin.

Each subopercle (Fig. 5, 7) is a broad and morphologically complex bone that bears a stout and pointed process emerging from the dorsal margin, ventral and posterior flattened elongate fimbriations, and a spiny anterior process; the subopercle articulates with the opercle through the process that arise centrally from its dorsal margin.

399

Visceral arches. Of the hyoid apparatus, the elongate acinaciform branchiostegal rays can be
observed in all the examined specimens. The interhyals (Fig. 5) are cylindrical and slightly
curved.

403 The gill arches are only partially preserved. The bones are usually disarticulated and displaced from their original position. The first pharyngobranchial seems to be absent. The 404 second pharyngobranchial is preserved in MCSNV B.13 (Fig. 8); this bone is moderately 405 406 wide and bears a thick process emerging from its dorsal surface, and several strong and recurved teeth. What appears to be a large right third pharyngobranchial (Fig. 5) is preserved 407 in MCSNV T.978. The epibranchials are thin and cylindrical (Fig. 5). The ceratobranchials (? 408 409 I to IV) are long, stout and cyclindrical, gradually tapering and curved distally. The fifth ceratobrachial (Fig. 5) is massive and subtriangular in outline; the dorsal surface of this bone 410 has a linear longitudinal ridge; strong recurved teeth appear to be restricted to the medial and 411 lateral margins of the fifth ceratobranchial. Hypobranchials and basibranchials are not 412 preserved in any of the available specimens. 413

414

Vertebral column. The vertebral column (Fig. 2B) is compact with the abdominal portion arcuate in a kyphotic curve, with the concave side toward the ventrum. There are 19 vertebrae (including the last hemicentrum fused to the hypural complex). As in other lophilforms, the first vertebra is closely connected to the neurocranium (see, e.g., Rosen & Patterson 1969). The vertebral centra are massive and squared except for the second that is nearly rectangular,

higher than long. The neural spines of the vertebrae three through eight incline posteriorly 420 and are laterally flattened and anteroposteriorly expanded. More posteriorly, the neural 421 spines are elongate and cylindrical but become progressively shorter and more inclined in the 422 423 caudal region. Neural prezygapophyses are well-developed on all vertebrae. The first haemal spine occurs on the sixth vertebra. The first two haemal spines are poorly developed and 424 remarkably inclined posteriorly, bent over the anteroventral sector of the succeeding 425 vertebrae. Haemal arches and spines of the vertebrae eight through twelve are notably broad 426 and laterally compressed, becoming considerably shorter more posteriorly. 427

428

Median fins and support. The caudal skeleton (Fig. 2B) is consistent with that of other 429 lophilforms. The neural and haemal arches and spines of the penultimate vertebra are strongly 430 431 expanded anteroposteriorly and spatulate distally. The hypural complex consists of the fusion of the ural centra with the first preural centrum, hypurals and parhypural (see Rosen & 432 Patterson 1969). The hypural plate is triangular with a slight notch along the posterior margin 433 434 that continues on the plate as a shallow median furrow. A single well-developed and laterally compressed epural is also present. The caudal fin is slightly rounded and contains eight rays, 435 of which the central six are bifurcated. 436

The cephalic portion of the spinous dorsal fin (including the illicial apparatus) is not 437 preserved in any of the examined specimens. Like in *†Eosladenia*, Lophiomus and Lophius, 438 439 the post-cephalic portion of the spinous dorsal fin (Fig. 2B) consists of three spines progressively decreasing in length posteriorly. The first spine is located over the fourth 440 vertebral centrum, the second spine apparently inserts at the level of the fifth vertebral 441 centrum, and the third post-cephalic spine is located above the seventh vertebral centrum. 442 The soft-rayed dorsal fin (Fig. 2B) inserts at the level of the tenth vertebra and contains 443 nine distally bifurcated rays supported by seven pterygiophores. The first ray is in 444

supernumerary association on the first dorsal-fin pterygiophore, and the posteriormost dorsal-445 fin pterygiophore supports two rays. The rays gradually increase in length from the first to the 446 fourth, and succeeding elements are of progressively decreasing size. The first pterygiophore 447 448 of the soft dorsal fin is massive, short and anteroposteriorly enlarged, whereas the succeeding elements are characterized by having dorsally rounded heads and elongate narrow vertical 449 shafts interdigitating in the underlying interneural spaces. The posterior two dorsal-fin 450 pterygiophores insert in the space between the neural spines of the vertebrae 12th and 13th. 451 The head of the posteriormost pterygiophore has a hypertrophied rounded posterior flange. 452 453 The anal fin is not adequately preserved in the available material. It appears to be composed of six rays supported by four pterygiophores. The pterygiophores are 454 subrectangular, stout and closely associated to the ventral margin of the overlying vertebral 455 456 centra. The first anal-fin ray is in supernumerary association on the first anal-fin ptervgiophore, and two rays are supported by the posterior pterygiophore. 457

458

459 Paired fins and support. The paddle-shaped pectoral fin (Figs. 1-3) comprises 14 to 16 simple rays. The fin is supported by two greatly elongate (22.8 - 23.3 % SL) and thick radials 460 (Fig. 2B, 5). The dorsal radial is cylindrical, about two thirds the length of the ventral one. 461 The ventral pectoral-fin radial is considerably expanded distally to form an articulatory 462 surface for the pectoral-fin rays; the distal flat and dorsally convex expansion of the ventral 463 464 radial shows some radial striae that possibly represent the traces of the ontogenetic fusion of additional radial elements. Scapulae and coracoids are poorly preserved in the examined 465 material. The posttemporal, the dorsalmost element of the pectoral girdle, is firmly connected 466 to the posterolateral corner of the neurocranium. The supracleithrum is approximately ovoid 467 and laminar, closely associated with the lateral surface of the dorsal part of the ventral arm of 468 the cleithrum. The cleithrum is the largest bone of the pectoral girdle. The horizontal and 469

anteromedially directed arm is always nearly totally hidden by other cranial bones or partially
covered by the sediment. The vertical arm is broad, laterally flattened and dorsomedially
oriented; a short pointed and stout cleithral (=humeral) spine emerges in the basal portion of
the vertical arm of the claithrum along its posterior margin. The lateral surface of this bone is
slightly ornamented by small pits in the junction area between the vertical and horizontal
arms. A single rod-like postcleithrum appears to be present.

The pelvic fins are not exposed in the examined material. A single displaced
basipterygium can be observed in MCSNV T.978 (Fig. 2B); this bone is triradiate with a long
cylindrical anterior limb and an expanded articular posterior part. The medial process is
flattened with a straight profile and possibly served as a contact surface for its respective
counterpart.

481

482 **Phylogenetic relationships**

The present analysis is based on an examination of seven lophioid genera (*†Caruso*, 483 *†Eosladenia, Lophiodes, Lophiomus, Lophius, †Sharfia, and Sladenia), representing all* 484 known members of the family Lophiidae (see Caruso 1985), plus two outgroup antennarioid 485 genera Antennarius and Brachonichthys (Pietsch 2009:175-179, fig. 188). The purpose of 486 this section is to place the extinct genera *Caruso*, *Eosladenia*, and *Sharfia*, and the four 487 extant lophiid genera in a phylogenetic framework of the family. A data matrix of nine taxa 488 489 and 38 characters was constructed (Appendix 1). All characters were treated as unordered and unweighted. All characters except number 35 (see below) were binary. Characters states that 490 could not be determined from the fossils because of inadequate preservation are coded as 491 unknown, and indicated in the data matrix by a question mark. The matrix was analyzed with 492 PAUP (Swofford 2002), using the branch and bound algorithm, with accelerated 493 494 transformation (ACCTRAN) to optimize characters. To evaluate branch support, a heuristic

bootstrap analysis of 1000 replicates was conducted, with simple addition sequence and TBR
(tree bisection-reconnection) branch-swapping options. Bremer decay values (Bremer 1988)
were calculated using TREEROT (Sorenson 1999).

498

Character descriptions. The descriptions of the phylogenetically relevant characters for the 499 analyzed taxa are arranged below by discrete anatomical complexes. A description of each 500 character is followed by a summary of the recognized character state of each character. 501 Consistency and retention indices (CI and RI) were produced as a whole and for each 502 503 character individually. Both CI and RI are given after each character description. The condition of the different features in the genera included in the phylogenetic analysis is 504 briefly discussed. The numbered characters listed below correspond to the character numbers 505 506 in the matrix. Some of the characters were previously included in the analysis generated by 507 Caruso (1985). The intrageneric relationships hypothesized by Caruso (1985) were primarily based on external morphological features and spine distribution across the head, 508 509 suspensorium, jaws and opercular apparatus; because of inadequate preservation, these characters are in many cases not observable in the fossil material and for this reason are not 510 511 included in our analysis.

512

513 Cranium

1. *Mesethmoid: present (0); absent (1)* (CI 1.00, RI 1.00). Most lophiiforms, including both outgroups, possess an ossified mesethmoid lying between the lateral ethmoids and the anterior extensions of the frontals and above the parasphenoid (e.g., Regan & Trewavas 1932; Pietsch 1974; 1981). Regan (1912) and Le Danois (1974) described and figured an ossified mesethmoid in lophiids. The presence of the mesethmoid in extant and fossil lophiids has not been confirmed by our observations, which are consistent with those of Morrow (1882),

Supino (1908), and Montcharmont (1950). According to Supino (1908), a greatly reduced 520 and thin cartilage located between the lateral ethmoids testifies to the presence of a 521 mesethmoid in lophiid fishes. Based on functional considerations, Montcharmont (1950) 522 523 suggested that the possible homolog of the mesethmoid should be searched among the tissues associated with the autogenous ascending processes of the premaxillae. The analysis of 524 cleared and double stained specimens and dried skeletons of extant lophiids (Fig. 9), as well 525 526 as the morphological study of fossil taxa have not revealed the presence of any trace of a mesethmoid either bony or cartilaginous. 527

528 2. Interorbital width: greater than 30% of that measured between the outer margins of the pterotics (0); considerably less than 30% of that measured between the outer margins of 529 the pterotics (1) (CI 1.00, RI 1.00). The cranial width measured at the level of the medial 530 531 extensions of the frontals anterior to the supraocular spine is unusually narrow, notably less than 30% of that measured between the outer margins of the pterotics in *†Caruso* (Fig. 4) and 532 Sladenia (Fig. 9A), whereas it is always greater than 30% of that measured between the outer 533 margins of the pterotics in both outgroups and other lophiids (Fig. 9; Bannikov 2004; Pietsch 534 & Carnevale submitted). 535

3. Dorsolateral ridge of the frontals: smooth and uninterrupted (0); rugose, bearing
 short conical spines or low rounded knobs (1) (CI 0.50, RI 0.00). This character was
 originally described by Caruso (1985). The ornamented rugosities of the dorsolateral ridge of
 the frontal bones are unique to *Lophiomus* and *Lophius*, in which they increase during
 ontogeny. In both outgroups and other lophiids, including the fossil genera †*Caruso*,
 †*Eosladenia*, and †*Sharfia*, the dorsolateral ridge of the frontals is smooth (Fig. 9; Bannikov
 2004; Pietsch & Carnevale submitted).

4. *Parasphenoid width: narrow (0); laterally expanded (1)* (CI 1.00, RI 1.00). The
parasphenoid of most lophiiform fishes is elongate, narrow and well ossified; both outgroups

and the lophiid genera *†Caruso*, *†Eosladenia*, *Lophiodes*, *†Sharfia* and *Sladenia* display the
typical lophiiform condition (Fig. 9A-B; Bannikov 2004; Pietsch & Carnevale submitted).
The parasphenoid is characterized by having a remarkably wide lateral expansion in the
extant genera *Lophiomus* and *Lophius* (Figs 9C-D).

549

550 Jaws

55. *Ascending process of the premaxilla: fused to the alveolar process (0); autogenous* 552 *(1)* (CI 1.00, RI 0.00). The premaxilla of the vast majority of lophiiforms, including the 553 outgroups, is characterized by having a narrow ascending process fused to an elongate 554 alveolar process. All the lophiid genera exhibit an autogenous and highly mobile ascending 555 process of the premaxilla (Fig. 10; Bannikov 2004; Carnevale & Pietsch submitted).

6. *External surface of the maxilla: smooth (0); ornamented (1)* (CI 1.00, RI 0.00). This character was described by Caruso (1985). A smooth outer surface of the maxilla is typical of the outgroups and all the lophiid genera (including the fossils), except for *Lophiomus* in which it is conspicuously ornamented with short conical spines (Fig. 10C).

Symphysial spine: present (0); absent (1) (CI 0.33, RI 0.33). The dentaries of 7. 560 Antennarius and the extant lophiid genera Lophiomus, Lophius, and Sladenia bear a 561 prominent and often stout symphysial spine (Figs 11A, C-D). This spine is absent in 562 Brachionichthys (Carnevale & Pietsch 2010) and in the lophiid genera †Caruso, †Eosladenia, 563 564 Lophiodes, and *†Sharfia* (Figs. 5, 11B; Bannikov 2004; Pietsch & Carnevale submitted). External surface of the dentary: smooth (0); ornamented (1) (CI 1.00, RI 0.00). This 8. 565 character was described by Caruso (1985). The dentary of Lophiomus is unique in having a 566 dense covering of prominent rounded spines along its lateral surface (Fig. 11C). The 567 outgroups and the other lophiid genera are characterized by having dentaries with smooth 568 569 external surfaces.

9. Retroarticular process of the articular: extremely short and high (0); well-developed
and shallow (1) (CI 0.50, RI 0.50). The lower jaw terminates posteriorly more or less at the
articulation with the quadrate in both outgroups and *Sladenia* (Fig. 11A), but extends
posteriorly well past the glenoid fossa in all other lophiids (Figs 5, 11B-D; Bannikov 2004;
Pietsch & Carnevale submitted).

575

576 Suspensorium

577 10. Maxillary process of the palatine: simple, with rounded profile (0); bifurcate (1)

578 (CI 1.00, RI 1.00). The maxillary process of the palatine is simple, often expanded with

579 rounded profile, in *Antennarius*, *Brachionichthys*, †*Caruso*, †*Eosladenia*, *Lophiodes*,

†Sharfia and *Sladenia* (Figs 5, 11A-B; Bannikov 2004; Pietsch & Carnevale submitted). In *Lophiomus* and *Lophius* this process bears a couple of divergent acute spines dorsally that
make its dorsal aspect remarkably bifurcate (Figs 11C-D).

11. Arrangement of ectopterygoid and endopterygoid: separate from each other (0); 583 fused into a single element (1) (CI 1.00, RI 1.00). In the vast majority of lophiiforms, 584 including Antennarius (Pietsch 1981) and Brachionichthys (Carnevale & Pietsch 2010), the 585 ectopterygoid and endopterygoid are distinctly separate elements. In all lophiids a large 586 triangular, subrectangular or ovoid bony plate is always present in the region of the 587 suspensorium usually occupied by the ectopterygoid and endopterygoid (Figs 5, 11; 588 589 Bannikov 2004; Pietsch & Carnevale submitted); in all the lophiid genera, both extant and fossils, a longitudinal ridge emerges on the medial surface of this element. The anatomical 590 identity of this bone is rather problematic. According to Morrow (1882), the single pterygoid 591 element constitutes the product of the fusion between ectopterygoid and endopterygoid. 592 Supino (1908) hypothesized that the ectopterygoid is absent in lophiids and that the single 593 bone actually represents the endopterygoid. The structure of this bone was not clear in the 594

description and illustration provided by Gregory (1933). Montcharmont (1950) figured a 595 single element (figs 11-14) formed by the fusion of the ectopterygoid and endopterygoid but 596 surprisingly described two apparently separate bones. Le Danois (1974) proposed that the 597 598 large bony plate is the homolog of the endopterygoid of other teleosts and that a small ectopterygoid is nearly fused to the proximal portion of the palatine. Such a controversial 599 anatomical issue has been apparently resolved by Matsuura & Yoneda (1987) who, based on 600 a complete ontogenetic series of Lophius gastrophysus, documented the existence of two 601 bones recognizable as separate elements in specimens with notochord length of 20 mm. In 602 603 summary, we tentatively consider the pterygoid plate of lophiids to be the result of an ontogenetic fusion of the ectopterygoid and endopterygoid. 604

605

606 **Opercular series**

12. Opercular shape: subtriangular (0); strongly bifurcate (1) (CI 1.00, RI 1.00). The 607 possession of a fully ossified polygonal opercle is plesiomorphic for acanthomorphs and 608 609 more generally for teleost fishes. The opercle is subtriangular in outline in the outgroups Antennarius and Brachionichthys, and the Eocene lophiid *†Sharfia* (Pietsch & Carnevale 610 submitted). Such a condition has also been observed in lophichthyids, tetrabrachiids, 611 ogcocephaloids and chaunacoids (see Pietsch 1981). In all other members of the Lophiidae 612 the opercles are characterized by having a deep indentation along the posterior margin, which 613 614 makes them strongly bifurcated (Figs 5, 7, 12; Bannikov 2004; Pietsch & Carnevale submitted). A bifurcated opercle also occurs in most ceratioids (see, e.g., Regan & Trewavas 615 1932; Pietsch 2009), possibly representing the result of an independent derivation. 616 13. Posteroventral margin of the subopercle: simple (0); fimbriate (1) (CI 0.50, RI 617 0.67). The subopercle of most lophilform fishes is a morphologically heterogeneous bone 618 with entire margins (e.g., Regan & Trewavas 1932; Pietsch 1974; 1981). In antennarioids 619

(including Antennarius and Brachionichthys), ceratioids, chaunacoids, and ogcocephaloids, 620 the subopercle is relatively elongate, often broad, crescent-shaped and ovoid. The 621 posteroventral margin of the subopercle is therefore linear and entire in the taxa of the 622 623 lophiiform suborders Antennarioidei, Ceratioidei, Chaunacoidei and Ogcocephaloidei, as well as in the lophioid genera *†Sharfia* and *Sladenia* (Fig. 12A; Pietsch & Carnevale 624 submitted). The posteroventral margin of the subopercle of the other lophioid genera is 625 extremely fimbriated (Figs 5, 7, 12B-D; Bannikov 2004; Pietsch & Carnevale submitted), 626 even in very small larvae (see Matsuura & Yoneda 1987). 627

14. Anterodorsal process of the subopercle: short, reduced to a spine (0); prominent, articulating through connective tissue with the anteroventral margin of the opercle (1) (CI 1.00, RI 1.00). The subopercles of a number of lophiiforms (including both the outgroups) bear a median spiny process on the anterodorsal margin. A stout prominent ascending process arising from the anterodorsal margin of the subopercle, making contact with the anterior margin of the opercle is unique to the Lophiidae (Figs 5, 7, 12; Bannikov 2004; Pietsch & Carnevale submitted).

635

636 Gill arches

15. Distal end of the first epibranchial: simple (0); bifurcate (1) (CI 1.00, RI 1.00). The
distal tip of the first epibranchial has a single articular head in *Antennarius* and *Brachionichthys* (Pietsch 1981; Carnevale & Pietsch 2010), but two in the lophiid genera *Lophiodes*, *Lophiomus*, *Lophius*, and *Sladenia* (Fig. 13). The fossil genera †*Caruso*, **Eosladenia*, and **Sharfia* are coded as unknown.

16. *Third hypobranchial: present (0); absent (1)* (CI 1.00, RI 0.00). *Antennarius* has
three hypobranchials, as do many other lophiiforms (see, e.g., Pietsch 1974; 1981). The third
hypobranchial is absent in *Brachionichthys* (see Carnevale & Pietsch 2010), *Lophiodes*,

Lophiomus, Lophius, and *Sladenia* (Fig. 13). This character is unknown in †*Caruso*, *†Eosladenia*, and *†Sharfia*.

647 17. Second hypobranchial: present (0); absent (1) (CI 1.00, RI 0.00). The second
648 hypobranchial is present in the outgroups Antennarius and Brachionichthys (Pietsch 1981;
649 Carnevale & Pietsch 2010), and in the lophiids Lophiodes, Lophiomus, and Sladenia (Figs
650 13A-C), but absent in Lophius (Fig. 13D). The fossil genera are coded as unknown.

18. Arrangement of teeth on the fifth ceratobranchial: dense cluster (0); restricted to discrete rows along the lateral and medial margins (1) (CI 1.00, RI 1.00). In all lophiiform taxa except the lophiids, the teeth are arranged in a dense cluster on the inner surface of the fifth ceratobranchial (e.g., Pietsch 1981). The lophiid genera, including the fossils, are unique in having the fifth ceratobranchial teeth arranged in one or two rows along the outer margins of the bony plate (Figs 5, 13; Field 1966).

657

658 Axial skeleton and caudal fin

19. Number of vertebrae: 18-22 (0); 26-31 (1) (CI 1.00, RI 0.00). This character was 659 described by Caruso (1985). The outgroups and the lophiid genera *†Caruso*, *†Eosladenia*, 660 Lophiodes, Lophiomus, †Sharfia, and Sladenia usually have 22 or fewer vertebrae. Lophius 661 has 26 to 31 vertebrae. The apomorphic high vertebral number of Lophius has been 662 considered (Arambourg 1927) as further evidence of the so-called Jordan's rule (Jordan 663 664 1892), which states that number of vertebrae in fishes increase with latitude. The vertebral number in fishes, however, is subject to the influence of many different (often interacting) 665 factors (see, e.g., McDowall 2008), which makes it very difficult to evaluate the plausibility 666 of the possible effects of the Jordan's rule in anglerfishes of the genus Lophius. 667 20. Haemal spines of abdominal vertebrae: well developed and anteroposteriorly 668 expanded (0); reduced in size (1) (CI 1.00, RI 1.00). The abdominal haemal spines are widely 669

expanded and well developed in *Antennarius*, *Brachionichthys* (Pietsch 1981; Carnevale &
Pietsch 2010), and the lophiid genera †*Caruso*, †*Sharfia*, and *Sladenia* (Figs 2B, 14A; Pietsch
& Carnevale submitted). The abdominal vertebrae of the other lophiid genera (†*Eosladenia*, *Lophiodes*, *Lophiomus*, *Lophius*) possess reduced haemal spines (Figs 14B-D; Bannikov
2004).

21. Neural and haemal spines of the penultimate vertebra: well developed and 675 anteroposteriorly expanded (0); narrow, reduced in size (1) (CI 1.00, RI 1.00). In the vast 676 majority of lophilforms, including the outgroups Antennarius and Brachionichthys (Pietsch 677 678 1981; Carnevale & Pietsch 2010), and the lophiids *†Caruso*, *†Sharfia*, and *Sladenia*, the penultimate vertebra bears considerably enlarged and anteroposteriorly expanded neural and 679 haemal spines (Figs 2B, 14A; Pietsch & Carnevale submitted). These spines are narrow and 680 681 anteroposteriorly shortened in *†Eosladenia*, Lophiodes, Lophiomus, and Lophius (Figs 14B-D; Bannikov 2004). 682

Caudal centrum: rounded in cross section (0); depressed with lateral transverse
processes (1) (CI 1.00, RI 1.00). This character was described and discussed by Caruso
(1985). The caudal centrum of *Antennarius*, *Brachionichthys*, †*Caruso*, *Lophiodes*, †*Sharfia*,
and *Sladenia* (Figs 2B, 14A-B; Pietsch 1981; Carnevale & Pietsch 2010; Pietsch & Carnevale
submitted) is nearly circular in cross section. In †*Eosladenia*, *Lophiomus*, and *Lophius*, the
caudal centrum is in some ways depressed and bears well-developed lateral transverse
processes (Figs 14C-D; Bannikov 2004).

690

691 Median fins

692 23. Number of caudal-fin rays: nine (0); eight (1) (CI 1.00, RI 1.00). The caudal fin of
693 antennarioids, chaunacoids, ogcocephaloids and most ceratioids contains nine rays. All
694 members of the Lophiidae exhibit a caudal fin with eight rays.

24. *Cephalic dorsal-fin spines: three (0); two (1)* (CI 1.00, RI 0.00). This character was
described by Caruso (1985). The two outgroups and the extant lophiid genera *Lophiodes*, *Lophiomus*, and *Lophius* are characterized by having three cephalic dorsal-fin spines. The
third cephalic dorsal-fin spine is absent in *Sladenia*. This character could not be scored for
any of the fossil taxa.

25. Interdigitation between the anterior soft dorsal-fin pterygiophores and the *underlying neural spines: absent (0); present (1)* (CI 1.00, RI 1.00). The anterior soft dorsalfin pterygiophores of *Antennarius, Brachionichthys*, †*Caruso*, †*Sharfia* and *Sladenia* lie
above the corresponding neural spines (Fig. 2B, 14A; Pietsch 1981; Carnevale & Pietsch
2010; Pietsch & Carnevale submitted), but interdigitate with the neural spines in

705 *†Eosladenia, Lophiodes, Lophiomus, and Lophius (Figs 14B-D; Bannikov 2004).*

70626. Interdigitation between the anal-fin pterygiophores and the overlying haemal

spines: present (0); absent (1) (CI 0.50, RI 0.67). The anal-fin pterygiophores of Antennarius,

708 Brachionichthys, Lophius, and †Sharfia are closely associated with the corresponding haemal

spines and insert in the interhaemal spaces (Fig. 14D; Pietsch 1981; Carnevale & Pietsch

710 2010; Pietsch & Carnevale submitted). The anal-fin pterygiophores of the lophiid genera

⁷¹¹ *†Caruso*, *†Eosladenia*, *Lophiodes*, *Lophiomus*, and *Sladenia* lie well below the corresponding

haemal spines (Figs. 2B, 14A-C; Bannikov 2004).

713 27. Proximal shaft of the soft dorsal-fin pterygiophores: slender and elongate (0);

anteroposteriorly expanded (1) (CI 1.00, RI 1.00). The soft dorsal-fin pterygiophores of the

outgroups Antennarius and Brachionichthys (Pietsch 1981; Carnevale & Pietsch 2010), and

the lophiids *†Caruso*, *Lophiodes*, *†Sharfia* and *Sladenia* consist of a slender, elongate

proximal shaft and an expanded articular distal portion (Figs 2B, 14A-B; Pietsch & Carnevale

submitted). The proximal shaft of these pterygiophores is considerably expanded in

719 Lophiomus and Lophius (Figs 14C-D). †*Eosladenia* is coded as unknown.

720 28. *Rays articulating with the anteriormost dorsal-fin pterygiophore: two, one*

supernumerary (0); one (1) (CI 0.33, RI 0.00). The anterior dorsal-fin pterygiophore supports
a supernumerary ray in *Antennarius*, †*Caruso*, *Lophiomus*, †*Sharfia*, and *Sladenia* (Fig. 2B,

=

14A, C; Pietsch 1981; Pietsch & Carnevale submitted). The supernumerary ray on the first

dorsal-fin pterygiophore is absent in *Brachionichthys*, *Lophiodes*, and *Lophius* (Figs 14B, D;

725 Carnevale & Pietsch 2010). This character is unknown in *†Eosladenia*.

726 29. Rays articulating with the posteriormost dorsal-fin pterygiophore: one (0); two (1)

727 (CI 0.50, RI 0.50). The last dorsal-fin pterygiophore of *Antennarius*, *Brachionichthys*, and

Lophius support a single ray (Fig. 14D; Pietsch 1981; Carnevale & Pietsch 2010). In the

129 lophiid genera *†Caruso*, *Lophiodes*, *Lophiomus*, *†Sharfia*, and *Sladenia* two rays are

associated with the posteriormost dorsal-fin pterygiophore (Figs 2B, 14A-C; Pietsch &

731 Carnevale submitted). *†Eosladenia* is coded as unknown.

30. *Rays articulating with the anteriormost anal-fin pterygiophore: two, one*

supernumerary (0); one (1) (CI 1.00, RI 0.00). The anterior anal-fin pterygiophore of the

outgroups and of the lophiids *†Caruso*, *Lophiodes*, *Lophiomus*, and *Sladenia* supports two

rays, one of which is in supernumerary association (Figs 2B, 14A-C; Pietsch 1981; Carnevale

736 & Pietsch 2010). *Lophius* exhibits a single ray on the anteriormost anal-fin pterygiophore

737 (Fig. 14D). This character is coded as unknown in *†Eosladenia* and *†Sharfia*.

738 31. Rays articulating with the posteriormost dorsal-fin pterygiophore: one (0); two (1)

(CI 0.50, RI 0.50). The posteriormost anal-fin pterygiophore supports a single ray in

740 Antennarius, Brachionichthys, and Lophius (Fig. 14D; Pietsch 1981; Carnevale & Pietsch

2010), but two rays are associated with this element in *†Caruso*, *Lophiodes*, *Lophiomus*, and

742 *Sladenia* (Figs 2B, 14A-C). *†Eosladenia* and *†Sharfia* are coded as unknown.

32. Posteriorly directed expansion of the distal end of the posteriormost dorsal-fin

744 *pterygiophore: absent (0); present (1)* (CI 1.00, RI 1.00). The distal portion of the

posteriormost dorsal-fin pterygiophore is similar to those of the preceding elements in both
the outgroups, and in the lophiids *†Eosladenia*, *Lophiodes*, *Lophiomus*, *Lophius*, and *†Sharfia* (Figs 14B-D; Pietsch 1981; Bannikov 2004; Carnevale & Pietsch 2010; Pietsch &
Carnevale submitted). In *†Caruso* and *Sladenia* the distal sector of the posteriormost dorsalfin pterygiophore bears a broad posteriorly directed expansion with rounded profile (Figs 2B,
14A).

Number of dorsal-fin rays: nine to nineteen (0); eight (1) (CI 0.33, RI 0.33). This
character was described by Caruso (1985) and partially incorporated into his phylogenetic
study. The soft dorsal fin contains nine to nineteen rays in the outgroups *Antennarius* and *Brachionichthys*, and the lophiid genera *†Eosladenia, Lophius*, and *†Sharfia*. In the
remaining lophiid genera (*†Caruso, Lophiodes, Lophiomus*, and *Sladenia*) the number of soft
dorsal-fin rays is reduced to eight.

757

758 **Pectoral fin and girdle**

34. *Postcleithrum: rod-like (0); filamentous (1)* (CI 1.00, RI 0.00). The postcleithrum is
a stout rod-like bone in antennarioids (e.g., Pietsch 1981), chaunacoids, ogcocephaloids, and
many ceratioids (Regan & Trewavas 1932; Pietsch 1974; Bertelsen & Krefft 1988), and the
lophioids (Figs 15A-C), except *Lophius*. The latter genus has a long filamentous

763 postcleithrum (Fig. 15D).

35. *Cleithral spine (= humeral spine): absent (0); weakly developed (1); greatly*

reduced (2) (CI 1.00, RI 1.00). This character was partially described by Caruso (1985). A

cleithral spine is absent in the outgroups *Antennarius* and *Brachionichthys*. Lacking sufficient

material for skeletal preparations, Caruso (1985) erroneously stated that this spine is absent

also in *Sladenia*. *Sladenia*, as well as the extinct lophiids †*Caruso* and †*Sharfia*, possess a

short, weakly developed cleithral spine (Figs 5, 15A; Pietsch & Carnevale submitted). The

cleithral spine is extremely well developed in *†Eosladenia*, *Lophiodes*, *Lophiomus*, and *Lophius* (Figs 15B-D; Bannikov 2004).

36. Extent of ventralmost pectoral-fin radial: relatively short, measuring less than 20% 772 773 SL (0); long, greater than 20% SL (1) (CI 0.33, RI 0.33). The ventralmost pectoral-fin radial is relatively short in Antennarius, *†Eosladenia*, Lophiomus, Lophius, and *†Sharfia*, but 774 remarkably elongate in *Brachionichthys* and the lophiids *†Caruso*, *Lophiodes*, and *Sladenia*. 775 37. Shape of pectoral fin: paddle-like (0); fan-shaped (1) (CI 1.00, RI 1.00). Distinctly 776 paddle-like pectoral fins are characteristic of the outgroups Antennarius and Brachionichthys 777 778 (e.g., Pietsch & Grobecker 1987), and the lophiid genera *†Caruso*, *†Eosladenia*, Lophiodes, *†Sharfia*, and *Sladenia* (Figs 1-3, 5; Caruo & Bullis 1976; Caruso 1981; Bannikov 2004; 779 Pietsch & Carnevale submitted). Lophiomus and Lophius possess large fan-shaped pectoral 780 781 fins (e.g., Caruso 1983).

782

783 Integument

38. *Skin: covered with dermal spinules (0); naked (1)* (CI 1.00, RI 1.00). The skin of the
outgroups *Antennarius* and *Brachionichthys* is characterized by having a dense covering of
close-set dermal spinules. All known lophiid genera exhibit a smooth naked skin.

787

788 **Results**

The phylogenetic analysis produced a single tree (Fig. 16), with a total length of 53, a

consistency index of 0.7358, and a retention index of 0.7407 (Fig. 00). Monophyly of the

extant genera of the Lophiidae was reconfirmed as recognized by Caruso (1985), as well as

that of the family as a whole by the inclusion of the extinct genera *†Sharfia*, *†Caruso*, and

793 *†Eosladenia* (with bootstrap support of 99.9% and a Bremer value of 9). Eight characters

vithout homoplasy (unique and unreversed within the Lophiidae) support lophiid

795 monophyly: mesethmoid absent (character 1, state 1); ascending process of premaxilla 796 autogenous (5, 1); ectopterygoid and endopterygoid fused to form a single element (11, 1); anterodorsal process of subopercle prominent, articulating through connective tissue with 797 798 anteroventral margin of opercle (14, 1); teeth on fifth ceratobranchial restricted to discrete rows along lateral and medial margins (18, 1); caudal-fin rays eight (23, 1); cleithral spine 799 present (35, 1 and 2); and skin naked (38, 1). Monophyly of lophiid genera to the exclusion of 800 *Sharfia* was supported (with bootstrap support of 71.9% and a Bremer value of 2) by only a 801 single unique and unreversed character: opercle strongly bifurcate (12, 1). A sister-group 802 803 relationship between *†Caruso* and *Sladenia* and a sister group relationship of this clade relative to all remaining lophiids was supported (with bootstrap support of 62.8% and a 804 Bremer value of 1) by two characters without homoplasy: interorbital width narrow, 805 806 considerably less than distance between outer margins of pterotics (2, 1); and posteriorly 807 directed expansion of distal end of posteriormost dorsal-fin pterygiophore present (32, 1). Monophyly of a clade containing *†Eosladenia* and the remaining extant lophiid genera was 808 809 supported (with bootstrap support of 77.5% and a Bremer value of 2) by three characters: haemal spines of abdominal vertebrae reduced (20, 1); neural and haemal spines of 810 penultimate vertebra narrow and reduced (21, 1); and interdigitation between anterior soft 811 dorsal-fin pterygiophores and underlying neural spines present (25, 1). A clade containing 812 *†Eosladenia, Lophiomus* and *Lophius*, to the exclusion of *Lophiodes*, was supported by only 813 814 a single character: caudal centrum depressed, bearing lateral transverse processes (22, 1). Finally, a sister-group relationship between Lophiomus and Lophius was confirmed following 815 Caruso (1985), supported by four unique and unreversed characters: dorsolateral ridge of 816 817 frontals rugose, bearing short conical spines or low rounded knobs (3, 1); parasphenoid laterally expanded (4, 1); maxillary process of palatine bifurcate (10, 1); and proximal shaft 818 819 of soft dorsal-fin pterygiophores anteroposteriorly expanded (27, 1).

820

821 Discussion

822 *†Caruso brachysomus* is the third valid extinct taxon of the family Lophiidae known from articulated skeletal remains. All of these fossils are characterized by having a modern body 823 824 architecture, suggesting that the lophiid body plan was already established in the early Eocene. Based on an analysis of the fossil record and phylogenetic considerations, Patterson 825 & Rosen (1989), and subsequently Carnevale & Pietsch (2006), concluded that all the 826 lophiiform lineages (antennarioids, ceratioids, chaunacoids, lophioids, and ogcocephaloids) 827 were already in existence in the early Eocene. Unfortunately, the fossil record does not 828 provide precise data about the minimum age for the origin of the Lophiiformes in general and 829 the lophilform subgroups in particular, as well as the order of events in the phylogeny of the 830 831 order. Two different hypotheses resulted from recent molecular-clock analyses of divergence times: according to Alfaro et al. (2009) and Santini et al. (2009), the origin of the lophiiform 832 833 body plan should be searched for in the lower part of the Paleogene, thereby suggesting that the divergence of the lophiiform lineages occurred within a relatively short time interval; on 834 the contrary, a Cretaceous origin, in an interval between 130 and 100 Myr ago, has been 835 836 proposed by Miya et al. (2010).

The dramatic anatomical diversity of lophilform subgroups, including the large number 837 of morphological peculiarities make it very difficult to interpret the origin of these fishes, as 838 839 well as to hypothesize the possible appearance of a primitive lophilform. Apart for some synapomorphic features related to specific characters of the skeleton (cranial and caudal; 840 Pietsch 1981; Pietsch & Grobecker 1987) or to reproductive biology (Rasquin 1958; Pietsch 841 1981), the lophiiform body plan may be defined by four relevant aspects that were 842 documented and cursorily discussed by Gregory & Conrad (1936): (1) the enormous mouth 843 and throat, and consequent macrophagous habits; (2) the gill opening restricted to a tube-like 844

opening located close to the pectoral-fin base; (3) the elongation of pectoral-fin radials that
results in their pediculate appearance; and (4) the cephalic spinous dorsal fin with the anterior
element modified to serve as a luring apparatus. All but the tube-like gill opening refer to
skeletal features, which can be also observed in the fossils.

The huge increase in size of the mouth and throat is related to the hypertrophic 849 development of the jaws, suspensorium, hyoid apparatus, and branchial arches. As in other 850 vertebrates, the development of these elements of the head in the fish embryo is characterized 851 by having a hierarchy of cell movements and interactions between neural crest and 852 853 mesodermal mesenchyme, and surrounding epithelia (see, e.g., Le Douarin et al. 1994). Several genes that function in the development of this region of the head have been 854 identified. Recent studies (e.g., Neuhauss et al. 1996; Piotrowski et al. 1996; Schilling et al. 855 856 1996) have emphasized the existence of discrete sets of genes that influence the correlated diversification of both jaws and branchial arches (including the hyoid apparatus and 857 suspensorium). 858

The elongation of the pectoral fins and girdles of lophilforms and their resemblance to 859 tetrapod limbs was evidenced by Cuvier (1829) who called these fishes "pediculate" (little 860 feet), a term subsequently used to identify the whole group (see, e.g., Valenciennes 1837; 861 Günther 1861; Gill 1883, 1909). The lophiiform pectoral girdle, or pseudobrachium (Monod 862 1960) is primarily characterized by having enlarged pectoral-fin radials, which in certain 863 864 cases are employed in a tetrapod fashion to walk over the substrate (see Pietsch & Grobecker 1987; Edwards 1989). The elongation of the pectoral-fin radials exhibits considerable 865 variation within lophiiforms, with the maximum lengths observed in certain lophioids (see 866 above) and antennarioids (Carnevale & Pietsch 2010). In all vertebrates the embryonic 867 development of paired appendages passes through three main phases, namely positioning, 868 initiation, and outgrowth. During the first phase paired appendages arise from bud initials that 869

originate in the lateral plate mesoderm at positions considered to be specified by *Hox* gene 870 expression in somatic mesoderm (see Burke et al. 1995). The initiation phase of the pectoral 871 fins (=forelimbs) follows the positioning phase. The transcription factor gene Tbx5 is 872 expressed in the pectoral-fin bud (Tamura et al. 1999), during both the initiation and 873 outgrowth phases, mediated by interactions with Wnt2b and Fgf10 (Takeuchi et al. 2003). 874 Fibroblast growth factors (FGFs) are involved in both the initiation and the following 875 outgrowth phases; the FGFs are expressed in the apical ectodermal ridge of the fin bud 876 promoting fin outgrowth. The anteroposterior pattern is controlled by an equivalent of the 877 878 zone of polarizing activity of the amniote limb, located at the posterior margin of the fin bud. The gene *Sonic hedgehog* (*Shh*) is a major upstream factor in development expressed in this 879 sector of the fin bud. Neumann et al. (1999) demonstrated that Shh is required to establish 880 881 some aspects of the anteroposterior polarity, for normal development of the fin bud, and formation of the fin endoskeleton. In a recent paper, Sakamoto et al. (2009) found that a 882 temporal shift of *Shh* activity alters the size of the endoskeletal elements in paired fins. 883 884 According to those authors, a heterochronic shift of the onset of *Shh* expression influences the proliferation of cells that contribute to the formation of the endoskeletal disk, with 885 implications in the final size of the pectoral-fin radials. We may therefore hypothesize that 886 the synapomorphic elongation of the pectoral-fin radials in lophiiforms could be related to a 887 heterochronic shift of the onset of *Shh* expression. However, it is also interesting to note that 888 889 the enlargement of jaws, suspensorium, hyoid apparatus, branchial arches and pectoral-fin endoskeleton may be linked together and that sets of genes that influence both cranial and 890 appendicular skeletal morphology may also contribute to correlated fin and cranial 891 enlargement (Neuhauss et al. 1996; Piotrowski et al. 1996; Schilling et al. 1996). 892 The highly modified spinous dorsal fin of lophilforms certainly represents one of the 893 most spectacular morphological characteristics of these fishes. The anterior elements of this 894

fin are shifted anteriorly over the neurocranium and are modified to serve as a luring 895 apparatus, involving a vast series of associated modifications of the general architecture of 896 the neurocranium and of the musculature and innervations associated with the dorsal fin (see 897 898 Bertelsen 1951; Bradbury 1967; Rosen & Patterson 1969). The anteriormost spine, or illicium, usually bears an esca at its tip, which exhibits considerable diversity and is 899 extremely useful in alpha-level taxonomy; in female ceratioid anglerfishes the esca is unique 900 901 in having bioluminescent bacteria and pheromone-producing secretory glands used to attract a conspecific male (e.g., Munk 1992; Pietsch 2009), while in ogcocephalids (and perhaps in 902 903 some antennariids) it apparently produces a chemical attractant used to lure buried benthic preys (Pietsch & Grobecker 1987; Bradbury 1988; Nagareda & Shenker 2009). The anterior 904 migration of the anterior (=cephalic) dorsal-fin spines occurs during the larval stage (e.g., 905 906 Matsuura & Yoneda 1986), resulting from the forward extension of the cartilaginous basal pterygiophore inside the subepidermal space (Matsuura & Yoneda 1987). The forward 907 migration of the dorsal-fin spines continues until the first two spines are well anterior to the 908 909 eyes. The pterygiophores of the spinous dorsal fin develop from a single condensation of tissue that separates into independent pterygiophores during development (e.g., Everly 2002). 910 In basal lophiiforms the spinous dorsal fin separates into two discrete units, comprising the 911 cephalic and post-cephalic spines respectively (see Everly 2002). The spinous dorsal fin is a 912 major innovation of acanthomorph fishes. Mabee et al. (2002) hypothesized that the spinous 913 dorsal fin essentially is an anterior duplication of the soft-rayed fin typical of all the 914 actinopterygians. In this context, the evolution of a discrete, anatomically regionalized and 915 spatially separated structure is an example of duplication and divergence (sensu Raff 1996), 916 enhanced by modular organization (Gilbert 2010). Such an hypothesis fits well with the 917 configuration of the dorsal fins observed in lophilforms. However, lophilforms exhibit a 918 really complex dorsal-fin structure; as documented above, the spinous dorsal fin becomes 919

separated into discrete independent units during ontogeny, thereby resulting in a 920 submodularization of the anterior dorsal fin. The cephalic unit of the submodularized dorsal 921 fin includes the luring apparatus and associated muscles and nerves. The possibility of a 922 923 further modularization of the first dorsal-fin module (sensu Mabee et al. 2002) in lophiiforms may be confirmed by the recurrent suppression of the post-cephalic unit of the spinous dorsal 924 fin in antennarioids, ceratioids, chaunacoids, ogcocephaloids, and some lophioids (see, e.g., 925 Pietsch 1981; 2009). The structural complexity of the luring apparatus of lophiiforms, with its 926 vast array of highly derived characters and functions, may be envisaged as a remarkable case 927 928 of co-option (sensu Raff 1996), involving both morphological and functional transformation of a pre-existing submodularized first dorsal-fin module. The forward shift of the cephalic 929 unit of the submodularized spinous dorsal fin also represents a peculiarity of lophiiform 930 931 fishes. A few other teleost groups are characterized by having an extensive anterior migration of the dorsal fin (see Nelson 2006; Bannikov & Carnevale 2011). The condition typical of 932 lophilforms, however, is extreme considering the broad rearrangements of cranial architecture 933 934 observed in these fishes. Median fin development apparently involves the same genetic programs that operate in paired appendages, and expression of Hoxd and Tbx18 genes (which 935 specify paired limb position) defines the position of median fins (Freitas et al. 2006) along 936 continuous stripes of competency on the midline of the body (Yonei-Tamura et al. 2008). 937 Therefore, we may hypothesize that in lophilforms, the forward migration of the dorsal fin 938 939 over the neurocranium could be produced by an anterior shift of the stripes of competency for dorsal-fin formation along the dorsal midline. 940

941

942 Acknowledgements

We wish to thank Walter Landini, Università di Pisa, Pisa, for suggestions and a critical
review of an early draft of the text. Presubmission reviews of the manuscript were also

provided by James W. Orr and Duane E. Stevenson, both of NOAA Fisheries, Seattle. We are 945 particularly obliged to Gaël Clement and Monette Veran, Muséum National d'Histoire 946 Naturelle, Paris, and Roberto Zorzin and Anna Vaccari, Museo Civico di Storia Naturale di 947 Verona, for access to fossil material under their care and for hospitality during our visits to 948 Paris and Verona in April and February 2010 respectively. Loans of additional fossil material 949 were kindly provided by Zerina Johanson, Natural History Museum, London and Alexandre 950 F. Bannikov, Borysiak Paleontological Institute, Russian Academy of Sciences, Moscow. 951 Thanks are also due to the following individuals for providing access to extant comparative 952 953 material, and permission to clear and stain specimens for osteological study: Katherine P. Maslenikov, University of Washington, Seattle; Karsten Hartel, Museum of Comparative 954 Zoology, Harvard University, Cambridge; John H. Caruso, Tulane University, New Orleans; 955 956 Dianne J. Bray, Museum of Victoria, Melbourne; Jeffrey Johnson, Queensland Museum, 957 South Brisbane. Many thanks go to Anna Vaccari, Museo Civico di Storia Naturale di Verona, and Philippe Loubry, Muséum National d'Histoire Naturelle, Paris for photographs. 958 The work was supported in part by U.S. National Science Foundation Grant DEB-0314637, 959 T. W. Pietsch, principal investigator. Visit to Paris of the senior author was supported by the 960 SYNTHESYS grants of the Muséum National d'Histoire Naturelle, Paris. 961

962

963

964 **References**

965 Agassiz, L. 1833-1844. *Recherches sur les poissons fossiles*. Petitpierre, Neuchâtel, 1420 pp.

- 966 Agassiz, L. 1835. Kritische Revision der in der Ittiologia Veronese abgebildeten fossile
 967 Fische. *Neues Jahrbuch für Mineralogie, Geognosie, Geologie und Petrefakenkunde*,
- 967 Fische. Neues Jahrbuch für Mineralogie, Geognosie, Geologie und Petrefakenkunde,
 968 Jahrgang 1835, 290-316.

969	Alfaro, M. E., Santini, F., Brock, C., Alamillo, H., Dornburg, A., Rabovsky, D. L.,
970	Carnevale, G. & Harmon, L. J. 2009. Nine exceptional radiations plus high turnover
971	explain species diversity in jawed vertebrates. Proceedings of the National Academy
972	of Sciences of the United States of America, 106, 133410-13414.
973	Andrews, S. M., Gardiner, B. G., Miles, R. S. & Patterson, C. 1967. Chapter 26 – Pisces.
974	Pp. 637-683 in W. B. Harland, C. H. Holland, H. R House, N. F. Hughes, A. B.
975	Reynolds, M. J. S. Rudwick, J. Satterthwaite, L. B. H. Tarlo, & E. C. Willey (eds) The
976	Fossil Record. Geological Society of London, London.
977	Arambourg, C. 1927. Les poissons fossiles d'Oran. Matériaux pour la Carte géologique de
978	l'Algérie, I ^{ère} Série, Paléontologie, 6 , 1-218.
979	Bailly, E. M. 1824. Déscription des feuillets pêcheurs de la Baudroie. Annales des Sciences
980	<i>Naturelles</i> , 2 , 523-533.
981	Bannikov, A. F. 2004. The first discovery of an anglerfish (Teleostei, Lophiidae) in the
982	Eocene of the Northern Caucasus. Paleontological Journal, 38, 67-72.
983	Bannikov, A. F. & Carnevale, G. 2011. Enigmatic spiny-rayed fish from the Eocene of
984	Monte Bolca, Italy. Geological Journal,
985	Bellwood, D. R. 1996. The Eocene fishes from Monte Bolca: the earliest coral reef fish
986	assemblage. Coral Reefs, 15, 11-19.
987	Bellwood, D. R. & Wainwright, P. C. 2002. The history and biogeography of fishes on coral
988	reefs. Pp. 5-32 in P. F. Sale (ed) Coral Reef Fishes: Dynamics and Diversity in a
989	Complex Ecosystem. Academic Press, San Diego.
990	Bertelsen, E. 1951. The ceratioid fishes. Ontogeny, taxonomy, distribution and biology.
991	Dana Report, 39 , 1-276.
992	Bertelsen, E. & Krefft, G. 1988. The ceratioid family Himantolophidae (Pisces,
993	Lophiiformes). Steenstrupia, 14, 9-89.

994	Blainville, H. D. de 1818. Des ichthyolites du Monte Bolca, ou Vestena Nuova dans le
995	Veronais. Nouveau Dictionaire d'Histoire Naturelle Appliqué aux Arts, à
996	l'Agriculture, à l'Economie Rurale et Domestique, à la Medicine, etc., 27, 344-361.
997	Blot, J. 1980. La faune ichthyologique des gisements du Monte Bolca (Province de Vérone,
998	Italie). Catalogue systématique présentant l'état actuel des recherches concernant cette
999	faune. Bulletin du Muséum National d'Histoire Naturelle, Paris, C, 4, 339-396.
1000	Bradbury, M. G. 1967. The genera of batfishes (family Ogcocephalidae). Copeia, 1967,
1001	399-422.
1002	Bradbury, M. G. 1988. Rare fishes of the deep-sea genus Halieutopsis: A review with
1003	descriptions of four new species (Lophiiformes: Ogcocephalidae). Fieldiana, Zoology,
1004	44, 1-22.
1005	Bremer, K. 1988. The limitis of amino acid sequence data in angiosperm phylogenetic
1006	reconstruction. Evolution, 42, 795-803.
1007	Brühl, C. B. 1856. Osteologisches aus dem Pariser Pflanzergarten. Wien.
1008	Burke, A. C., Nelson, C. E., Morgan, B. A. & Tabin, C. 1995. Hox genes and the evolution
1009	of vertebrate axial morphology. Development, 121, 333-346.
1010	Carnevale, G. & Pietsch, T. W. 2006. Filling the gap: a fossil frogfish, genus Antennarius
1011	(Teleostei, Lophiiformes, Antennariidae), from the Miocene of Algeria. Journal of
1012	<i>Zoology</i> , 270 , 448-457.
1013	Carnevale, G. & Pietsch, T. W. 2009a. The deep-sea anglerfish genus Acentrophryne
1014	(Teleostei, Ceratioidei, Linophryne) in the Miocene of California. Journal of
1015	Vertebrate Paleontology, 29, 372-378.
1016	Carnevale, G. & Pietsch, T. W. 2009b. An Eocene frogfish from Monte Bolca, Italy: The
1017	earliest knonwn skeletal record for the family. Palaeontology, 52, 745-752.

1018	Carnevale, G. & Pietsch, T. W. 2010. Eocene handfishes from Monte Bolca, with
1019	description of a new genus and species, and a phylogeny of the family
1020	Brachionichthyidae (Teleostei: Lophiiformes). Zoological Journal of the Linnean
1021	Society, in press.
1022	Carnevale, G. & Pietsch, T. W. In Press. Batfishes from the Eocene of Monte Bolca.
1023	Geological Magazine.
1024	Carnevale, G., Pietsch, T. W., Takeuchi, G. T. & Huddleston, R. W. 2008. Fossil
1025	ceratioid anglerfishes (Teleostei: Lophiiformes) from the Miocene of the Los Angeles
1026	Basin, California. Journal of Paleontology, 82, 996-1008.
1027	Caruso, J. H. 1981. The systematics and distribution of the lophiid anglerfishes: I. A
1028	revision of the genus Lophiodes with the description of two new species. Copeia,
1029	1981 , 522-549.
1030	Caruso, J. H. 1983. The systematics and distribution of the lophiid anglerfishes: II.
1031	Revisions of the genera Lophiomus and Lophius. Copeia, 1983, 11-30.
1032	Caruso, J. H. 1985. The systematics and distribution of the lophiid anglerfishes: III.
1033	Intrageneric relationships. Copeia, 1985, 870-875.
1034	Caruso, J. H. & Bullis, H. R. Jr. 1976. A review of the lophiid anglerfish genus Sladenia
1035	with a description of a new species from the Caribbean Sea. Bulletin of Marine
1036	<i>Science</i> , 26 , 59-64.
1037	Cuvier, G. 1829. Le régne animal distribué d'après son organisation, pour servir de base à
1038	l'histoire naturelle des animaux et d'introduction à l'anatomie comparée. Nouvelle
1039	édition, Deterville, Paris, Poissons, 2, 122-406.
1040	D'Erasmo, G. 1922. Catalogo dei Pesci Fossili delle Tre Venezie. Memorie dell'Istituto
1041	Geologico della Reale Università di Padova, 6, 1-181.

1042	De Stefano, G. 1910. Osservazioni sulla ittiofauna pliocenica di Orciano e San Quirico in
1043	Toscana. Bollettino della Società Geologica Italiana, 28, 539-648.

- Eastman, C. R. 1905. Les types de poisons fossils de Monte Bolca au Muséum d'Histoire
 Naturelle de Paris. *Mémoires de la Sociétè géologique de France, Paléontologie*, 13,
 1-31.
- Eaton, T. H. Jr., Edwards, C. A., McIntosh, M. A. & Rowland, J. P. 1954. Structure and
 relationships of the anglerfish, *Lophius americanus*. *Journal of the Elisha Mitchell Scientific Society*, 70, 205-218.
- Edwards, J. L. 1989. Two perspectives on the evolution of the tetrapod limb. *American Zoologist*, 29, 235-254.
- Everly, A. W. 2002. Stages of development of the goosefish, *Lophius americanus*, and
 comments on the phylogenetic significance of the development of the luring apparatus
 in Lophiiformes. *Environmental Biology of Fishes*, 64, 393-417.

1055 Fariña, A. C., Azevedo, M., Landa, J., Duarte, R., Sampedro, P., Costas, G., Torres, M.

- A. & Cañas, L. 2008. *Lophius* in the world: a synthesis on the common features and
 life strategies. *ICES Journal of Marine Science*, 65, 1272-1280.
- Field, J. D. 1966. Contributions to the functional morphology of fishes. Part II. The feeding
 mechanism of the angler-fish, *Lophius piscatorius* Linnaeus. *Zoologica Africana*, 2,
 45-67.
- Freitas, R., Zhang, G. J. & Cohn, M. J. 2006. Evidence that mechanisms of fin
 development evolved in the midline of early vertebrates. *Nature*, 442, 1033-1037.
- 1063 Frickhinger, K. A. 1991. Fossilien atlas fische. Mergus, Melle, 1088 pp.
- Garman, S. 1899. Report on an exploration off the west coasts of Mexico, Central and South
 America, and off the Galapagos Islands, in charge of Alexander Agassiz, by the U.S.
- 1066 Fish Commission steamer 'Albatross', during 1891, Lieut. Commander Z. L. Tanner,

- 1067 U.S.N., commanding. XXVI. The fishes. *Memoirs of the Museum of Comparative*1068 *Zoology*, 24, 1-431.
- Geoffroy St. Hilaire, E. F. 1807. Sur le sac branchial de la baudroie et l'usage qu'elle en fait
 pour pêcher. *Annales du Muséum d'Histoire Naturelle de Paris*, 10, 480-481.
- Geoffroy St. Hilaire, E. F. 1824. Sur les parties de son organization que la baudroie emploie
 comme instruments de pêche. *Mémoires du Muséum d'Histoire Naturelle de Paris*,

1073 11, 117-131.

- 1074 Geoffroy St. Hilaire, E. F. 1827. Observations sur les usages acessoires des sacs branchiaux
 1075 chez la baudroie à l'occasione de la discussion rappelée dans le Bulletin, tome 9, n°
 1076 208. Bulletin des Sciences Naturelles (Férrussac), 10, 165-166.
- 1077 Gilbert, S. F. 2010. Developmental biology. Sinauer Associates, Inc., Sunderland, 711 pp.
- Gill, T. N. 1883. Supplementary note on the pediculati. *Proceedings of the United States National Museum*, 5:551–556.
- Gill, T. N. 1909. Angler fishes: their kinds and ways. *Annual Report of the Smithsonian Institution*, 1909, 565-615.
- **1082** Gregory, W. K. 1933. Fish skulls: a study of the evolution of natural mechanisms.

1083 *Transactions of the American Philosophical Society*, **23**, 74-481.

- Gregory, W. K. & Conrad, G. M. 1936. The evolution of the pediculate fishes. *The American Naturalist*, 70, 193-208.
- Gudger, E. W. 1945. The anglerfish, *Lophius piscatorius* et *americanus*, use the lure in
 fishing. *American Naturalist*, **79**, 542-548.
- **1088** Günther, A. C. L. G. 1861. Catalogue of the Acanthopterygian Fishes in the Collection of
- 1089 *the British Museum*. Trustees of the British Museum, London, Vol. 3, 586 pp.
- 1090 Hanow, M. C. 1768. Wahrscheinliche Erklärung eines soberbaren Fisches. (Lophius). Titius,
- 1091 *I. D. Gemeinnützliche Abhandlungen*, 1, 275-283.

1092 Hasegawa, Y., Tohida, Y., Kohno, N., Ono, K., Nokariya, H. & Uyeno, T. 1988.

- 1093 Quaternary vertebrates from Shiriya Area, Shimokita Peninsula, Northeastern Japan.
 1094 *Memoirs of the National Science Museum*, 21, 17-36. [in Japanese]
- Jordan, D. S. 1892. Relations of temperature to vertebrae among fishes. *Proceedings of the United States National Museum*, 1891, 107-120.
- Landini, W. 1977. Revisione degli "Ittiodontoliti pliocenici" della collezione Lawley.
 Palaeontographia Italica, 70, 92-134.

1099 Landini, W. & Sorbini, L. 1996. Ecological and trophic relationships of Eocene Monte

- 1100 Bolca (Pesciara) fish fauna. Pp. 105-112 in A. Cherchi (ed) Autecology of selected
- 1101 *fossil organisms: Achievements and problems*. Bolletino della Società Paleontologica
- 1102 Italiana, Special Volume 3.
- **Lawley, R.** 1876. *Nuovi studi sopra ai pesci ed altri vertebrati fossili delle colline toscane.*

1104 Tipografia dell'Arte della Stampa, Firenze, 122 pp.

- 1105 Le Danois, Y. 1974. Étude osteo-myologique et révision systématique de la famille des
- 1106Lophiidae (Pédiculates, Haploptérygiens). Mémoires du Muséum National d'Histoire

1107 *Naturelle, Paris, Série A, Zoologie*, **91**, 1-127.

- LeDouarin, N. M., Dupin, E. & Ziller, C. 1994. Genetic and epigenetic control in neural
 crest development. *Current Opinions in Genetics and Development*, 4, 685-695.
- 1110 Leriche, M. 1906. Contribution à l'étude des poissons fossils du Nord de la France et des
 1111 regions voisines. *Mémoires de la Société Géologique du Nord*, 5, 1-430.
- 1112 Leriche, M. 1908. Note préliminaire due des poissons nouveaux de l'Oligocène Belge.
- 1113 Bulletin de la Sociétè Belge de Gèologie, Paléontologie, Hydrologie, **21**, 378-384.
- 1114 Leriche, M. 1910. Les poissons Oligocènes de la Belgique. Mémoires du Musée Royal
- 1115 *d'Histoire Naturelle de Belgique*, **5**, 233-363.

- 1116 Leriche, M. 1926. Les poissons Néogènes de la Belgique. *Mémoires du Musée Royal*
- 1117 *d'Histoire Naturelle de la Belgique*, **32**, 365-472.
- 1118 Long, J. A. 1996. *The rise of fishes*. The Johns Hopkins University Press, Baltimore, 224 pp.
- 1119 Mabee, P. M., Crotwell, P. L., Bird, N. C. & Burke, A. C. 2002. Evolution of median fin
- 1120 modules in the axial skeleton of fishes. *Journal of Experimental Zoology (Mol Dev*
- 1121 *Evol*), **294**, 77-90.
- Matsuura, Y. & Yoneda, N. T. 1986. Early development of the lophiid anglerfish, *Lophius* gastrophysus. Fishery Bulletin, 84, 429-436.
- **Matsuura, Y. & Yoneda, N. T.** 1987. Osteological development of the lophiid anglerfish,
- 1125 *Lophius gastrophysus. Japanese Journal of Ichthyology*, **33**, 360-367.
- McDowall, R. M. 2008. Jordan's and other ecogeographical rules, and the vertebral number
 in fishes. *Journal of Biogeograpy*, 35, 501-508.
- 1128 Miya, M., Pietsch, T. W., Orr, J. W., Arnold, R. J., Satoh, T. P., Shedlock, A. M., Ho, H.
- 1129 –C., Shimazaki, M., Yabe, M. & Nishida, M. 2010. Evolutionary history of
- anglerfishes (Teleostei: Lophiiformes): a mitogenomic perspective. *BMC*
- 1131 *Evolutionary Biology*, **10**, 58.
- 1132 Montcharmont, U. 1950. Il cranio e lo scheletro io-branchiale di *Lophius piscatorius* L.
- 1133 (Teleostei Pediculati). Annuario dell'Istituto e Museo di Zoologia della Università
 1134 di Napoli, 2, 1-74.
- Montin, L. 1779. Beskrifning pa en fisk, *Lophius barbatus*. *Handlingar Svenska Vetenskaps Akademiens*, 187, 165-173.
- 1137 Monod, T. 1960. A propos du pseudobrachium des *Antennarius* (Pisces, Lophiiformes).
- 1138 Bulletin de l'Institut Français de l'Afrique Noire, **22**, 620-698.

- Morrow, R. 1882. On the bones of "Lophius piscatorius", angler fish, devil fish, goose fish,
 etc. ect. Proceedings and Transactions of the Nova Scotian Institute of Science, 5,
 340-357.
- Munk, O. 1992. Accessory escal gland (AEG) in some deep-sea anglerfishes. *Acta Zoologica*, 73, 33-37.
- 1144 Nagareda, B. H. & Shenker, J. M. 2009. Evidence for chemical luring in the polka-dot
 1145 batfish *Ogcocephalus cubifrons* (Teleostei: Lophiiformes: Ogcocephalidae). *Florida*1146 *Scientist*, 72, 11-17.
- 1147 Nelson, J. S. 2006. *Fishes of the world*. John Wiley & Sons, Hoboken, 601 pp.
- 1148 Neuhauss, S. C. F., Solnica-Krezel, L., Schier, A. F., Zwartkruis, F., Stemple, D. L.,
- 1149 Malicki, J., Abdelilah, S., Stainier, D. Y. R. & Driever, W. 1996. Mutations
- affecting craniofacial development in zebrafish. *Development*, **123**, 357-367.
- 1151 Neumann, C. J., Grandel, H., Gaffield, W., Schulte-Merker, S. & Nüsslein-Volhard, C.
- 1152 1999. Transient establishment of anteroposterior polarity in the zebrafish pectoral fin
 1153 bud in the absence of *sonic hedgehog* activity. *Development*, **126**, 4817-4826.
- **Nolf, D.** 1985. Otolithi Piscium. *Handbook of Paleoichthyology*, **10**, 1-146.
- 1155 Oliva, O., Skořepa, V. & Stokłosowa, S. 1968. Myodome in teleosts *Clupea harengus*,
- 1156 *Osmerus eperlanus, Perca fluviatilis, Stizostedion lucioperca, Lophius piscatorius.*
- 1157 *Věstník Českoslovenké Společnosti Zoologické*, **33**, 377-389.
- Parsons, I. 1750. Some account of the Rana piscatrix (*Lophius*). *Philosophical Transactions*of the Ray Society, 46, 126-131.
- **Patterson, C.** 1993. An overview of the early fossil record of acanthomorphs. *Bulletin of*
- 1161 *Marine Science*, **52**, 29-59.

1162 **Patterson, C. & Rosen, D. E.** 1977. Review of the ichthyodectiforms and other Mesozoic

- teleost fishes and the theory and practice of classifying fossils. *Bulletin of the American Museum of Natural History*, **158**, 81-172.
- **Patterson, C. & Rosen, D. E.** 1989. The Paracanthopterygii revisited: order and disorder. Pp.
- 11665-36 in D. M. Cohen (ed) Papers on the Systematics of Gadiform Fishes. Natural
- 1167History Museum of the Los Angeles County, 32.
- 1168 Pietsch, T. W. 1974. Osteology and relationships of ceratioid anglerfishes of the family
- Oneirodidae, with a review of the genus *Oneirodes* Lütken. *Natural History Museum*of the Los Angeles County Science Bulletin, 18, 1-113.
- **Pietsch, T. W.** 1981. The osteology and relationships of the anglerfish genus *Tetrabrachium*,

1172 with comments on lophiiform classification. *Fishery Bulletin*, 7, 387-419.

- **1173 Pietsch, T. W.** 2009. *Oceanic anglerfishes: extraordinary diversity in the deep-sea.*
- 1174 University of California Press, Berkley, 577 pp.
- 1175 Pietsch, T. W. & Carnevale, G. In Press. A new genus and species of lophiid anglerfish

1176 (Teleostei: Lophiiformes) from the Eocene of Monte Bolca, Italy. *Copeia*.

1177 Pietsch, T. W. & Grobecker, D. B. 1987. Frogfishes of the world: systematics,

1178 *zoogeography, and behavioral ecology*. Stanford University Press, Stanford, 406 pp.

- Pietsch, T. W. & Orr, J. W. 2007. Phylogenetic relationships of deep-sea anglerfishes of the
 suborder Ceratioidei (Teleostei: Lophiiformes) based on morphology. *Copeia*, 2007,
 1-34.
- 1182 Piotrowski, T., Schilling, T. F., Brand, M., Jiang, Y. –J., Heisenberg, C. –P., Beuchle, D.,
- 1183 Grandel, H., van Eeden, F. J. M., Furutani-Seiki, M., Granato, M., Haffter, P.,
- 1184 Hammerschmidt, M., Kane, D. A., Kelsh, R. N., Mullins, M. C., Odenthal, J.,
- 1185 Warga, R. M. & Nüsslein-Volhard, C. 1996. Jaw and branchial mutants in zebrafish
- 1186 II: anterior arches and cartilage differentiation. *Development*, **123**, 345-356.

1187	Potthoff, T. 1984. Clearing and staining techniques. Pp. 35-37 in H. G. Moser, W. J.
1188	Richards, D. M. Cohen, M. P. Fahay, A. W. Kendall Jr., & S. L. Richardson (eds)
1189	Ontogeny and systematic of fishes. American Society of Ichthyologists and
1190	Herpetologists Special Publication 1.
1191	Purdy, R. W., Schneider, V. P., Applegate, S. P., McLellan, J. H., Meyer, R. L. &
1192	Slaughter, B. H. 2001. The Neogene shark, ray, and bony fishes from Lee Creek
1193	Mine, Aurora, North Carolina. Smithsonian Contributions to Paleobiology, 90, 71-
1194	202.
1195	Raff, R. A. 1996. The shape of life. Chicago University Press, Chicago, 520 pp.
1196	Rafinesque, C. S. 1810. Indice d'ittiologia siciliana; ossia, catalogo metodico dei nomi
1197	latini, italiani, e siciliani dei pesci, che si rinvengono in Sicilia disposti secondo un
1198	metodo naturale e seguito da un'appendice che contiene la descrizione di alcuni
1199	nuovi pesci siciliani. Messina.
1200	Rasquin, P. 1958. Ovarian morphology and early embryology of the pediculate fishes
1201	Antennarius and Histrio. Bulletin of the American Museum of Natural History, 114,
1202	327-371.
1203	Ray, C. E., Wetmore, A., Dunkle, D. & Drez, P. 1968. Fossil vertebrates from the marine
1204	Pleistocene of Southern Virginia. Smithsonian Miscellaneous Collection, 153, 1-25.
1205	Regan, C. T. 1903. A revision of the fishes of the family Lophiidae. Annals and Magazine of
1206	Natural History, 7, 277-285.
1207	Regan, C. T. 1912. The classification of the teleostean fishes of the order Pediculati. Annals
1208	and Magazine of Natural History, 8, 279-289.
1209	Regan, C. T. & Trewavas, E. 1932. Deep-sea anglerfishes (Ceratioidea). Dana Report, 2, 1-
1210	113.

1211 Rosen, D. E. & Patterson, C. 1969. The structure and relationships of the

- paracanthopterygian fishes. *Bulletin of the American Museum of Natural History*, 141,
 357-474.
- 1214 Sakamoto, K., Onimaru, K., Munakata, R., Suda, N., Tamura, M., Ochi, H. & Tanaka,
- M. 2009. Heterochronic shift in *Hox*-mediated activation of *Sonic hedgehog* leads to
 morphological changes during fin development. *PloS ONE*, 4, e5121.
- 1217 Santini, F., Harmon, L. J., Carnevale, G. & Alfaro, M. E. 2009. Did genome duplication
- drive the origin of teleosts? A comparative study of diversification in ray-finned
 fishes. *BMC Evolutionary Biology*, 9, 194.
- 1220 Schilling, N. M., Piotrowski, T., Grandel, H., Brand, H., Heisenberg, C. –P., Jiang, Y. –
- 1221 J., Beuchle, D., Hammerschmidt, M., Kane, D. A., Mullins, M. C., van Eeden, F.
- 1222 J. M., Kelsh, R. N., Furutani-Seiki, M., Granato, M., Haffter, P., Odenthal, J.,
- 1223 Warga, R. M., Trowe, T. & Nüsslein-Volhard, C. 1996. Jaw and branchial mutants
- in zebrafish I: branchial arches. *Development*, **123**, 329-344.
- 1225 Schultz, O. 2006. An anglerfish, *Lophius* (Osteichthyes, Euteleostei, Lophiidae), from the
- Leitha Limestone (Badenian, Middle Miocene) of the Vienna Basin, Austria (Central
 Paratethys). *Beitrage zür Pälaontologie*, **30**, 427-435.
- 1228 Seilacher, A., Reif, W. E. & Westphal, F. 1985. Sedimentological, ecological and
- temporal patterns of fossil Lagerstätten. *Philosophical Transactions of the Royal Society of London B*, **311**, 5-24.
- 1231 Sorbini, L. 1972. *I fossili di Bolca*. Edizioni Corev, Verona, 133 pp.
- Sorbini, L. 1983. *La collezione Baja di pesci e piante fossili di Bolca*. Stamperia Valdonega,
 Verona, 119 pp.

- Sorbini, L. 1988. Biogeography and climatology of Pliocene and Messinian fossil fish of 1234 Eastern-central Italy. Bollettino del Museo Civico di Storia Naturale di Verona, 14, 1-1235 85.
- 1236
- Sorenson, M. D. 1999. TREEROT v. 2. Boston University, Boston. 1237
- Supino, F. 1908. Morfologia del cranio e note sistematiche e biologiche sulle famiglie 1238
- Trachinidae e Pediculati. Atti della Società Italiana di Scienze Naturali, 47, 100-116. 1239
- Swofford, D. L. 2002. PAUP*: Phylogenetic analysis using parsimony and other methods. v. 1240

4.0b10. Sinauer Associates, Inc., Sunderland. 1241

- 1242 Sychevskaya, E. K. & Prokofiev, A. M. 2010. On the occurrence of anglerfishes
- (Lophiidae) in the Lower Miocene of Transcaucasia. Journal of Ichthyology, 50, 205-1243 210. 1244
- Takeuchi, J. K., Koshiba-Takeuchi, K., Suzuki, T., Kanimura, M., Ogura, K. & Ogura 1245
- **T.** 2003. *Tbx5* and *Tbx4* trigger limb initiation through activation of the *Wnt/Fgf* 1246 signaling cascade. Development, 130, 2729-2739. 1247
- Tamura, K., Yonei-Tamura, S. & Izspisua Belmonte, J. 1999. Differential expression of 1248
- *Tbx4* and *Tbx5* in zebrafish fin buds. *Mechanisms of Development*, **87**, 181-184. 1249
- Valenciennes, A. 1837. Des Chironectes (Chironectes, Cuv., Antennarius, Comm.). Pp. 389-1250
- 437 in G. Cuvier & A. Valenciennes, Histoire Naturelle des Poissons, vol. 12. 1251
- Levrault, Paris & Strasbourg. 1252
- 1253 Volta, G. S. 1796. Ittiolitologia Veronese del Museo Bozziano ora annesso a quello del
- Conte Giovambattista Gazola e di altri Gabinetti Fossili Veronesi. Stamperia 1254 1255 Giuliani, Verona, 323 pp.
- 1256 Waterman, T. H. 1948. Studies on deep-sea anglerfishes (Ceratioidea). III. The comparative anatomy of Gigantactis longicirra Waterman. Journal of Morphology, 82, 81-149. 1257

1258	Wilson, D. P. 1937. The habits of the anglerfish, <i>Lophius piscatorius</i> L., in the Plymouth
1259	Aquarium. Journal of the Marine Biological Association of the United Kingdom, 21,
1260	447-496.
1261	Woodward, A. S. 1901. Catalogue of the Fossil Fishes in the British Museum (Natural
1262	History). Volume IV. Trustees of the British Museum, London, 636 pp.
1263	Yonei-Tamura, S., Abe, G., Tanaka, Y., Anno, H., Noro, M., Ide, H., Aono, H.,
1264	Kuraishi, R., Osumi, N., Kuratani, S. & Tamura, K. 2008. Competent stripes for
1265	diverse positions of limb/fins in gnathostome embryos. Evolution & Development, 10,
1266	737-745.
1267	Zigno, A. de 1874. Catalogo ragionato dei pesci fossili del calcare eoceno di Monte Bolca.
1268	Atti del Reale Istituto Veneto di Scienze, Lettere ed Arti, 4, 1-215.
1269	
1270	
1271	
1272	
1273	
1274	
1275	
1276	
1277	
1278	
1279	
1280	
1281	
1282	

1283 Captions of figures and tables

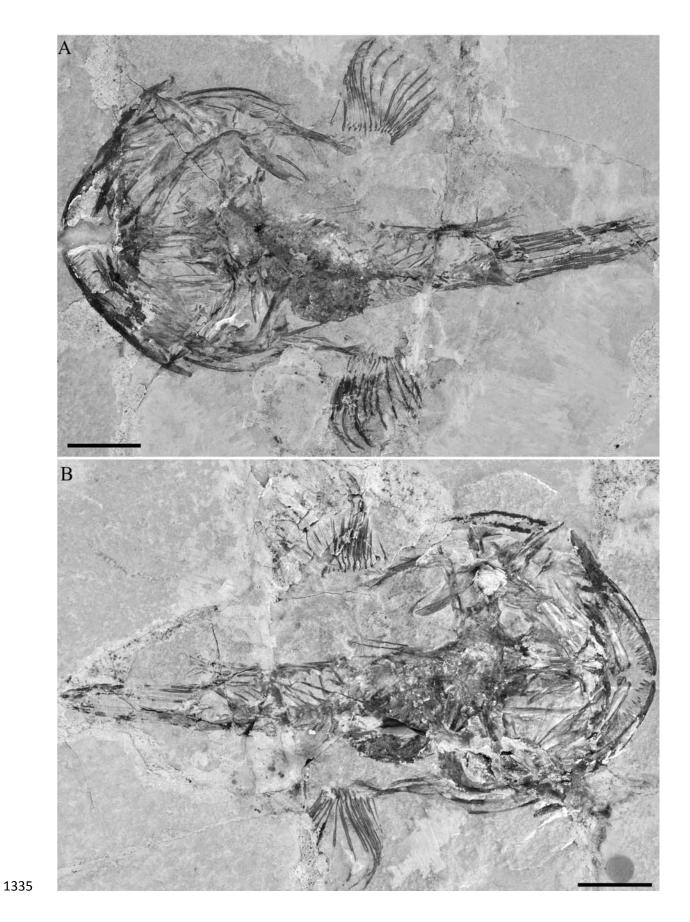
1284

Figure 1. †*Caruso brachysomus* (Agassiz, 1835). A, B, lectotype, MNHN Bol 42/43. Scale
bars = 20 mm. [planned for page width]

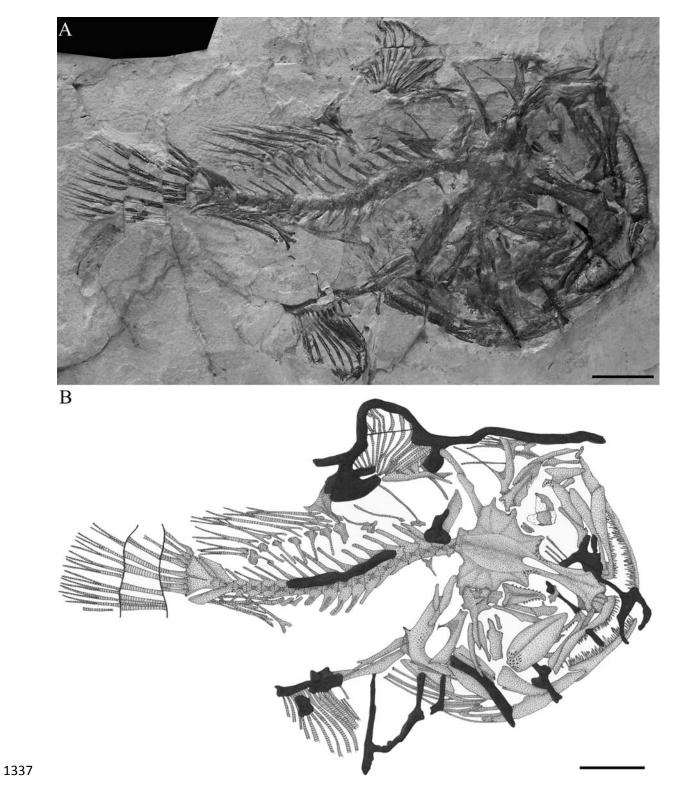
- **Figure 2.** *†Caruso brachysomus* (Agassiz, 1835). A, MCSNV T.978; B, complete skeleton
- drawn from specimen in A. Scale bars = 20 mm. [planned for page width]
- Figure 3. †*Caruso brachysomus* (Agassiz, 1835). MCSNV B.13. Scale bar = 20 mm.
 [planned for page width]
- Figure 4. †*Caruso brachysomus* (Agassiz, 1835). Reconstruction of the dorsal view of the
 neurocranium mainly based on MCSNV T.978 and MNHN Bol42/43. [planned for
 page width]
- 1293 page width]
- Figure 5. †*Caruso brachysomus* (Agassiz, 1835). Skull drawn from specimen MCSNV
 T.978. Scale bar = 20 mm. [planned for page width]
- Figure 6. *†Caruso brachysomus* (Agassiz, 1835). MNHN Bol 42, left lateral view of anterior
 portion of the premaxilla. Scale bar = 5 mm. [planned for column width]
- 1298 Figure 7. †*Caruso brachysomus* (Agassiz, 1835). MCSNV B. 13, right lateral view of
- 1299 opercle and subopercle. Scale bar = 5 mm. [planned for column width]
- Figure 8. †*Caruso brachysomus* (Agassiz, 1835). MCSNV B.13, left ceratobranchials and (?)
 second pharyngobranchial. Scale bar = 3 mm. [planned for column width]
- 1302 **Figure 9.** Dorsal view of neurocrania of (A) *Sladenia remiger*, CSIRO H.2559-02; *Lophiodes*
- 1303 monodi, MCZ 40928; (C) Lophiomus setigerus, TU 81104; (D) Lophius americanus,
- 1304 MCZ 51259. Scale bars = 5 mm. [planned for page width]
- 1305 Figure 10. Left lateral view of premaxillae and maxillae of (A) *Sladenia remiger*, CSIRO
- 1306 H.2559-02; Lophiodes monodi, MCZ 40928; (C) Lophiomus setigerus, TU 81104; (D)
- 1307 *Lophius americanus*, MCZ 51259. Scale bars = 5 mm. [planned for page width]

1308	Figure 11. Left lateral view of lower jaws, suspensoria and part of opercular apparatuses of
1309	(A) Sladenia remiger, CSIRO H.2559-02; Lophiodes monodi, MCZ 40928; (C)
1310	Lophiomus setigerus, TU 81104; (D) Lophius americanus, MCZ 51259. Bone in
1311	stipple, cartilage in black. Scale bars = 5 mm. [planned for page width]
1312	Figure 12. Left lateral view of opercles and subopercles of (A) Sladenia remiger, CSIRO
1313	H.2559-02; Lophiodes monodi, MCZ 40928; (C) Lophiomus setigerus, TU 81104; (D)
1314	<i>Lophius americanus</i> , MCZ 51259. Scale bars = 5 mm. [planned for page width]
1315	Figure 13. Branchial arches of (A) <i>Sladenia remiger</i> , CSIRO H.2559-02; <i>Lophiodes monodi</i> ,
1316	MCZ 40928; (C) Lophiomus setigerus, TU 81104; (D) Lophius americanus, MCZ
1317	51259. Bone in stipple, cartilage in black. Scale bars = 5 mm . [planned for page
1318	width]
1319	Figure 14. Left lateral view of vertebral columns, caudal skeletons, and median fins of (A)
1320	Sladenia remiger, CSIRO H.2559-02; Lophiodes monodi, MCZ 40928; (C)
1321	Lophiomus setigerus, TU 81104; (D) Lophius americanus, MCZ 51259. Bone in
1322	stipple, cartilage in black. Scale bars = 5 mm. [planned for page width]
1323	Figure 15. Left lateral view of pectoral girdles of (A) Sladenia remiger, CSIRO H.2559-02;
1324	Lophiodes monodi, MCZ 40928; (C) Lophiomus setigerus, TU 81104; (D) Lophius
1325	<i>americanus</i> , MCZ 51259. Bone in stipple, cartilage in black. Scale bars = 5 mm.
1326	[planned for page width]
1327	Figure 16. Cladogram of hypothesized relationships of the Lophiidae and two outgroups. The
1328	number above the base of a node is the Bremer value, and the number below the node
1329	indicates bootstrap support for the respective node. [planned for page width]
1330	Table 1. Measurements (in percent standard length) of <i>†Caruso brachysomus</i> (Agassiz,
1331	1835).

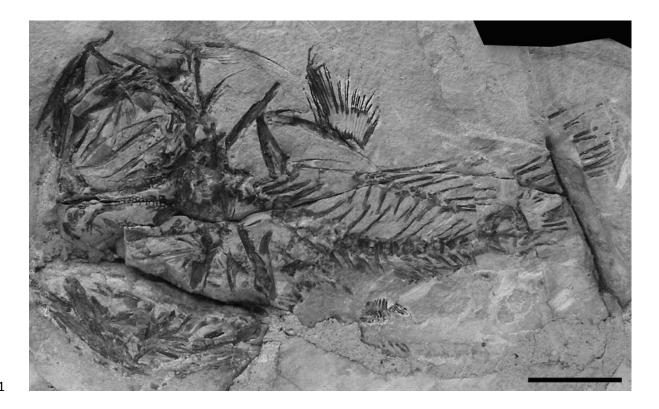
- 1332 Appendix. Character matrix of 38 morphological characters for genera of the Lophiidae and
- two outgroups.



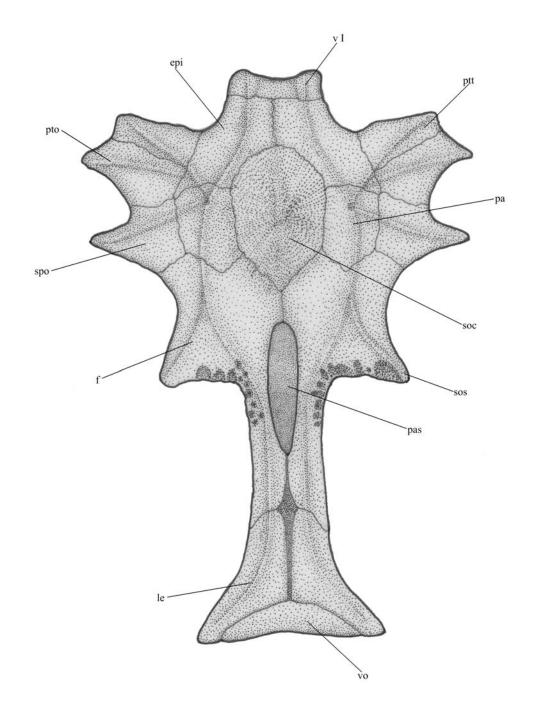
1336 Figure 1.



1338 Figure 2.

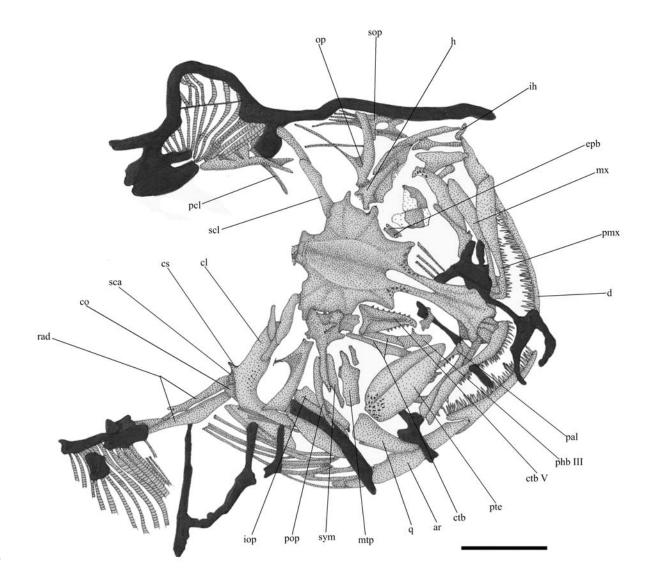


1342 Figure 3.



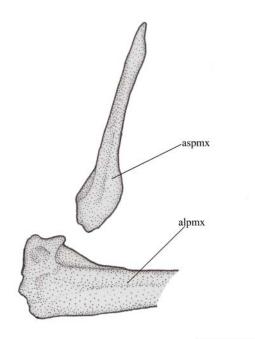


1344 Figure 4.





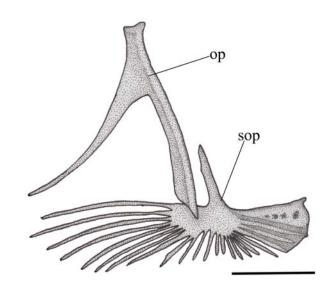
1346 Figure 5.





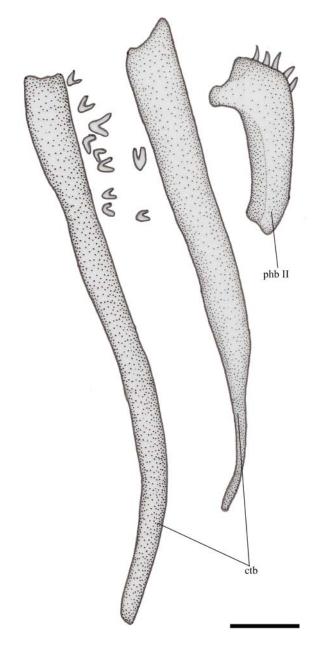
1348 Figure 6.

1349

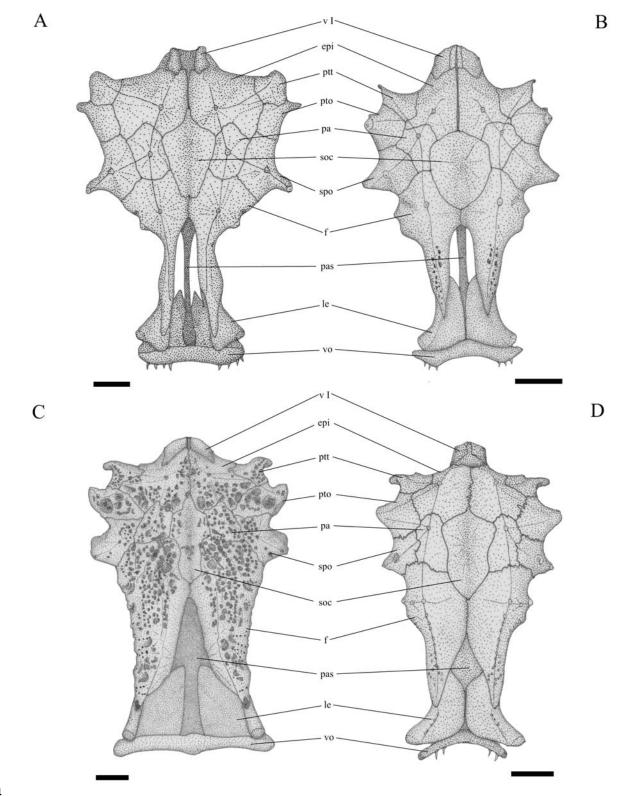


1350

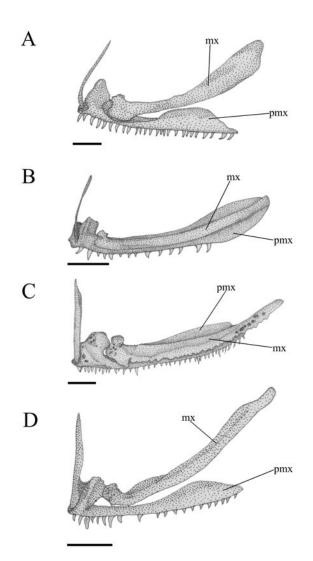
1351 Figure 7.



1353 Figure 8.

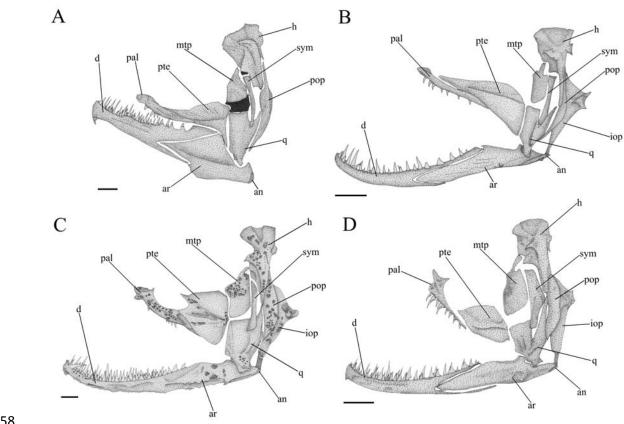


1355 Figure 9.

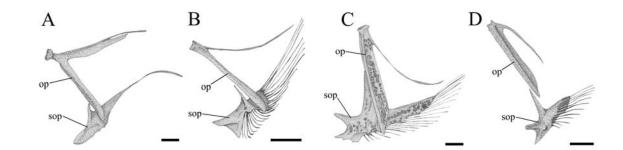




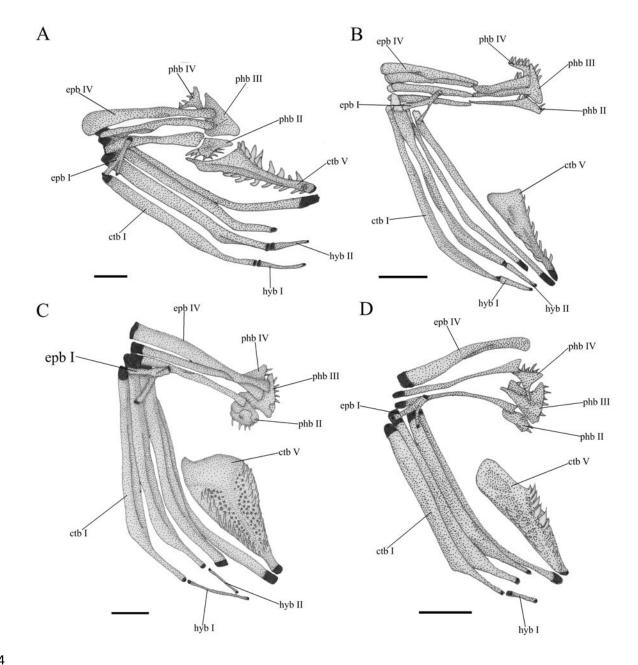
1357 Figure 10.



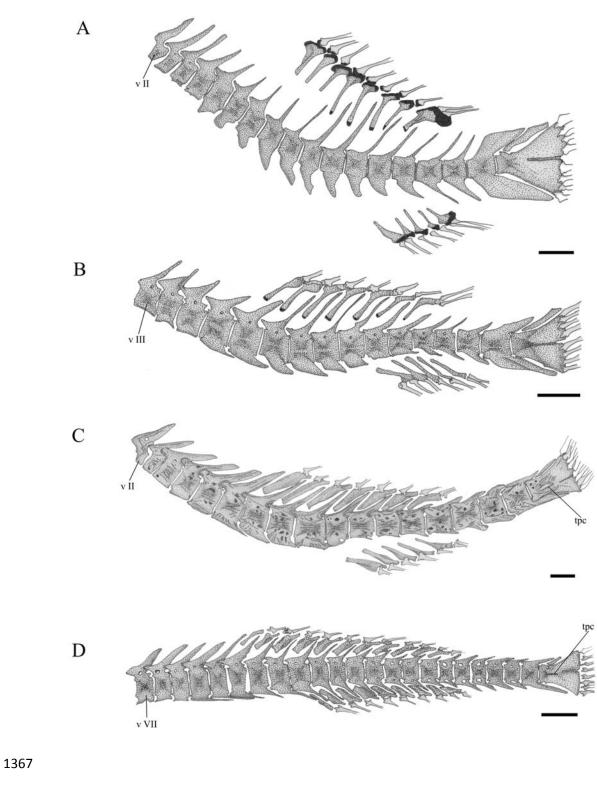
1359 Figure 11.



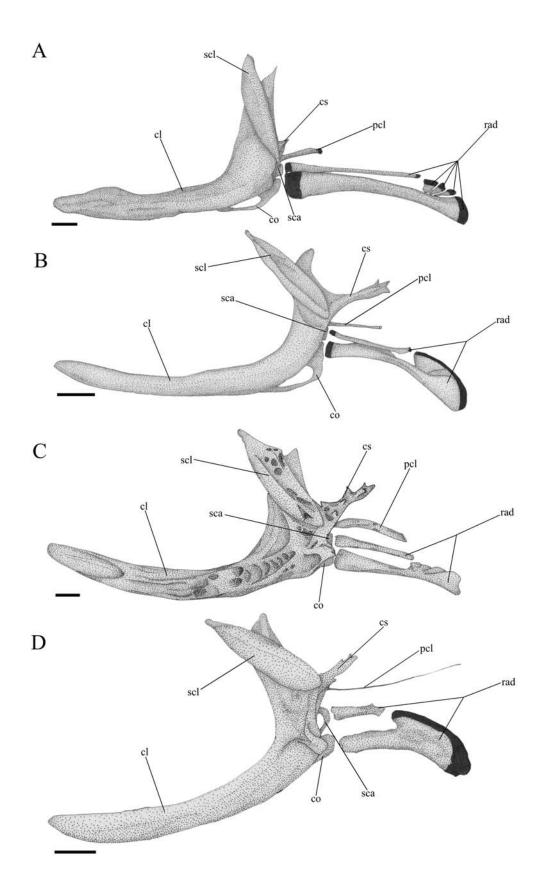
1362 Figure 12.



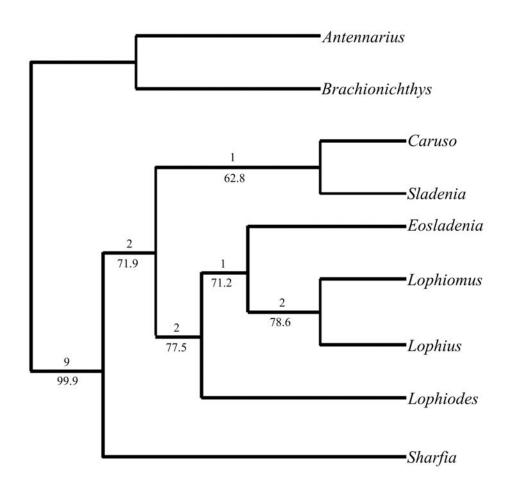
1365 Figure 13.



1368 Figure 14.



1371 Figure 15.



1374 Figure 16.

	MNHN Bol. 42/43	MCSNV T.978	MCSNV B.13
Total length	165.0 mm	164.1 mm	113.7 mm
Standard length	125.7 mm	136.5 mm	92.1 mm
Head length	43.3	40.2	39.0
Head width	24.9	23.0	21.1
Snout length	21.0	24.5	26.2
Snout width	5.7	5.6	6.8
Tail length	37.7	36.0	46.7

1383 Table 1.

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33
Antennarius	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Brachionichthys	0	0	0	0	0	0	0	0	0	0	0	0	0	?	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
†Caruso	1	0	1	0	1	0	1	0	1	1	1	1	1	?	?	?	1	0	0	0	0	0	0	1	0	0	0	1	0	1	?	1	1
†Eosladenia	0	0	1	0	1	0	1	0	1	?	1	1	1	?	?	?	1	0	1	1	0	1	1	1	?	?	?	?	?	?	?	2	0
Lophiodes	0	0	1	0	1	0	1	0	1	1	1	1	1	1	1	0	1	0	1	1	0	1	0	1	0	0	1	1	0	1	0	2	1
Lophiomus	0	1	1	1	0	1	1	1	1	1	1	1	1	1	1	0	1	0	1	1	0	1	1	1	1	1	0	1	0	1	0	2	0
Lophius	0	1	1	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	1	0	0	2	0
†Sharfia	0	0	1	0	1	0	1	0	1	1	0	0	1	?	?	?	1	0	0	0	1	0	0	1	0	0	0	1	?	?	?	1	0
Sladenia	1	0	1	0	0	0	0	0	1	1	1	0	1	1	1	0	1	0	0	0	0	0	0	1	0	0	0	1	0	1	1	1	1
100																																	

1385 Table 2.