- 1 This Accepted Author Manuscript (AAM) is copyrighted and published by Elsevier. It is posted here
- 2 by agreement between Elsevier and the University of Turin. Changes resulting from the publishing
- 3 process such as editing, corrections, structural formatting, and other quality control mechanisms -
- may not be reflected in this version of the text. The definitive version of the text was subsequently
 published in
- 6 Use of reagent test strips for diagnosis of endometritis in dairy cows
- 7 THERIOGENOLOGY 77(5) 858-864
- 8 http://dx.doi.org/10.1016/j.theriogenology.2011.09.009
- 9 You may download, copy and otherwise use the AAM for non-commercial purposes provided that
- 10 your license is limited by the following restrictions:
- (1) You may use this AAM for non-commercial purposes only under the terms of the CC-BY-NC-NDlicense.
- (2) The integrity of the work and identification of the author, copyright owner, and publisher must bepreserved in any copy.
- 15 (3) You must attribute this AAM in the following format: Creative Commons BY-NC-ND license
- 16 (http://creativecommons.org/licenses/by-nc-nd/4.0/deed.en),
- 17 http://dx.doi.org/10.1016/j.theriogenology.2011.09.009

18

19

20 Use of reagent test strips for diagnosis of endometritis in dairy cows

21 S.H. Cheong^a,*, D.V. Nydam^b, K.N. Galvão^a,1, B.M. Crosier^a, A. Ricci^c, L.S. Caixeta^a, R.B. Sper^a, M. Fraga^a, R.O.

- 22 Gilbert^a
- 23 a Department of Clinical Sciences, College of Veterinary Medicine, Cornell University, Ithaca, New York, USA
- 24 b Department of Population Medicine and Diagnostic Sciences, College of Veterinary Medicine, Cornell University, Ithaca, New
- 25 York, USA
- 26 c Faculta di Medicina Veterinaria, Universita di Torino, Torino, Italy
- 27 E-mail address: cs344@cornell.edu (S.H. Cheong) or rob.gilbert@cornell.edu (R.O. Gilbert).

28 Abstract

- The use of leukocyte esterase (LE), protein, and pH tests were evaluated on widely available 29 30 urinary test strips (Multistix 10 SG; Bayer Corporation, Elkart, IN, USA) on uterine lavage samples as a potential cow-side test for the diagnosis of cytologic endometritis. Uterine lavage samples of 31 32 563 lactating Holstein cows between 40 and 60 days postpartum from 28 herds were evaluated. Endometrial cytology was used as the reference for endometritis, with a cutoff point of $\geq 10\%$ 33 neutrophils. All three (LE, protein, and pH) were increased in cows with cytologic endometritis and 34 the associations were highly significant. Optimal cutoff points determined by receiver operating 35 characteristic analysis for LE, protein, and pH were $\geq ++, \geq 300 \text{ mg/dL}$, and ≥ 7.0 , respectively. 36 Combining the results for LE and pH improved the performance of the test strip, but this resulted in 37 a group of cows (20.6% of cows) which were approximately equally likely (46% with endometritis 38 and 54% without endometritis) to have cytologic endometritis or not, and therefore could not be 39 accurately classified. The direct relationship between reagent strip test and reproductive 40 performance was also evaluated. Reproductive impairment due to endometritis was restricted to 41 multiparous cows; significantly decreased reproductive performance was observed for multiparous 42 cows with lavage fluid LE \geq +++ (154 vs. 115 median days not-pregnant), as well as cows with pH 43 \geq 7.0 (150.5 vs. 111.5 median days not-pregnant), but not in cows with high protein, even at the 44 highest cutoff point. In conclusion, reagent strip test results were strongly associated with cytologic 45
- endometritis and reproductive impairment; however, in comparison with conventional cytology, the
 performance of reagent strip as an alternative test was relatively poor and may require further
 refinement.
- 49
- 50
- 51
- **-** -
- 52
- 53
- 54

55 1. Introduction

Endometritis is an inflammatory uterine disease that persists beyond normal uterine involution and 56 impairs reproductive performance [1], [2], [3] and [4]. Affected cows frequently have no external 57 symptoms [5] and [6]. Diagnostic methods, such as ultrasonographic evaluation of the reproductive 58 tract and uterine content are inferior to cytologic examination of uterine content [1] and [3], which 59 lead to the proposed disease definition based on cytology as the presence of >18% neutrophils in 60 uterine samples collected between 21 and 33 days postpartum, or >10% neutrophils between 34 and 61 47 days postpartum, in the absence of purulent vaginal discharge [5]. Cytologic evaluation of 62 63 uterine samples is currently the best method to diagnose inflammatory disease of the uterus. In a farm setting, however, this method is inconvenient, as it involves collection of the sample, 64 preparation of the slides and staining, followed by microscopic examination and identification and 65 enumeration of cells. Uterine samples collected using the cytobrush method [1] and [7] allow easier 66 67 slide preparation compared with samples collected using low-volume uterine lavage, but still require the time-consuming cell evaluation step. The lack of a practical cow-side test is a major 68 69 reason endometritis is not monitored or managed in commercial herds. A candidate cow-side test for the diagnosis of endometritis from uterine lavage fluid is leukocyte 70 71 esterase (LE), for example on a reagent strip intended for urinalysis, such as Multistix 10 SG (Bayer 72 Corporation, Elkart, IN, USA). In a smaller study, Santos et al. [8] reported high sensitivity (83%) and specificity (94%) when using the LE strip to diagnose endometritis. Multistix 10 SG (Bayer 73 Corporation) is a reagent strip of 10 tests, namely: LE, nitrite, urobilinogen, protein, pH, blood, 74 specific gravity, ketone (acetic acid), bilirubin, and glucose. The LE compound is present in 75 neutrophils; therefore, a positive result of this test is the most direct indicator of inflammatory cells 76 77 in urine using reagent strips. In addition, protein and pH reagent tests may be useful in the diagnosis of endometritis, as well as providing insight into the pathogenesis of the condition. Fluid 78 accumulation in the uterine lumen is used as an indicator of inflammation [1] and [3] which, if 79 80 present, could elevate the protein content of the recovered fluid of low volume uterine lavage, making protein concentration a potential diagnostic test. Furthermore, inflammation of the udder or 81 82 vesicular glands elevates the pH of milk [9] and seminal fluid [10], respectively, but it is unknown if inflammation of the uterus is associated with an elevation of pH in uterine fluid. 83 The objectives of this study were to: (1) determine if LE, protein, pH, or a combination of reagent 84 strip tests were associated with cytologic endometritis; (2) identify cutoff points for associated 85 reagent tests based on cytology and reproductive outcome; and (3) identify other factors associated 86

87 with LE, protein, and pH in uterine lavage samples.

88

89 2. Materials and methods

90 **2.1. Sample collection**

Uterine lavage samples used in this experiment were part of a larger study [4]. The present study 91 was initiated after 10 herds had already been sampled. All samples collected from that point on 92 were included in the present study. Selection of herds for the study was from a convenience sample 93 of herds that were willing to participate in the study. The inclusion criteria for herds sampled were: 94 located in New York State, large herd size (minimum of 400 milking cows), and used DairyComp 95 305 (Valley Ag Software, Tulare, CA, USA) for maintaining herd records. The inclusion criteria for 96 97 cows sampled were: between 40 and 60 days postpartum, apparently healthy (by cursory visual examination), no external vaginal discharge observed by visual examination, not inseminated, and 98 at least 2 days before the end of the voluntary waiting period for that specific farm (average 59 99 days; range 50 to 70). Herd records were obtained at the time of sampling and reproductive 100 101 outcomes were obtained by follow-up herd records collected 4 and 6 mo after sampling. Animal procedures were approved by the Cornell University Institutional Animal Care and Use 102 103 Committee. Uterine lavage samples were obtained as previously described [2]. Briefly, paper towels were used to cleanse the perineum of the cow, then a 63.5 cm sterile flex tip infusion pipette 104 105 (Exodus Breeders Corporation, York, PA, USA) was introduced into the uterus through the cervix, and 20 mL sterile saline solution (0.9% Sodium Chloride Injection USP; Baxter Healthcare Corp., 106 Deerfield, IL, USA) was infused into the uterus. Approximately 5 to 8 mL of fluid was recovered 107 by aspiration. The samples were put on ice and transported to the laboratory for analysis. One drop 108 of uterine lavage sample was added to each test on the Multistix 10 SG (Bayer Corporation) reagent 109 strip. Protein and pH results were evaluated after 1 min and the LE result evaluated after 2 min, as 110 per manufacturer instructions. Protein results were recorded in six categories which were: negative, 111 trace, + (30 mg/dL), ++ (100 mg/dL), +++ (300 mg/dL), and ++++ (>2000 mg/dL); pH results were 112 recorded in seven categories: 5.0, 6.0, 6.5, 7.0, 7.5, 8.0, and 8.5; and LE results were recorded in 113 five categories: negative, trace, + (small), ++ (moderate), and +++ (large). Cytologic evaluation of 114 the uterine lavage samples was performed after cytocentrifugation ($105 \times g$ for 3 min) and staining 115 116 using Camco stain Pak stain (Cambridge Diagnostic Products, Inc., Fort Lauderdale, FL, USA) by counting 200 cells (neutrophils, lymphocytes, macrophages, and uterine epithelial cells, excluding 117 erythrocytes) and results were expressed as the percentage of total cells. Cows were considered 118 positive for endometritis if neutrophils were >10% of total cells [5]. 119

120 **2.2. Data management and statistical analysis**

121 2.2.1. Association of reagent strip results with cytologic endometritis

All three end points (LE, protein, and pH) were recorded as ordered categories. The categories of 122 LE 'negative' and protein 'negative' had less than five observations that were positive for 123 endometritis and these categories were combined with 'trace' for all analyses. To test the hypothesis 124 that the reagent strip results were associated with endometritis, a multivariable logistic regression 125 model was produced using PROC GLIMMIX of SAS, Version 9.2 (SAS Institute, Cary, NC, USA) 126 with cytologic endometritis as the dependent variable. Herd was included as a random effect. The 127 128 association between LE, protein, and pH were tested individually with cytologic endometritis and a final model was built for each reagent strip test. In addition to reagent strip test results, fixed effects 129 130 considered were: parity (primiparous or multiparous), body condition score (\geq 3.5 or <3.5), ketosis, metritis, retained placenta (disease or no disease), Log (first test-day somatic cell count), days 131 132 postpartum at sampling, and two-way interactions. Disease occurrence data for ketosis, metritis, and

retained placenta were according to herd records with the diagnosis made by the herdsman. The

final model was built using manual backwards stepwise variable selection and variables were 125 matrixed if $\mathbf{P} < 0.05$

135 retained if P < 0.05.

136 To evaluate reagent strip as a diagnostic test for endometritis, the reagent strip test results were

dichotomized (positive test or negative test) at all possible cutoff levels. A series of 2×2 tables was

138 created for reagent strip test against cytologic evaluation. True positive, true negative, false positive

and false negative results were recorded, and sensitivity (Se), specificity (Sp), positive predictive

value (PPV), and negative predictive value (NPV) were calculated using those values at the

apparent prevalence in these herds. The cutoff point with the highest sum of sensitivity and

specificity was selected as optimal. Receiver operating characteristic (ROC) analysis was performed

using MedCalc version 11.5 (MedCalc Software, Mariakerke, Belgium) and the area under thecurve and P values are reported.

145 2.2.2. Association of reagent strip results with reproduction

Reproductive outcome was examined by Cox proportion hazards model using days from calving to subsequent pregnancy as the event of interest. Incomplete observations were right censored when the cows were culled, designated "Do-Not-Breed" by the herdsman, or at 210 days-in-milk. Reagent strip results were dichotomized at all levels and were tested individually and sequentially from the lowest threshold to the highest for effect on calving-to-conception interval using Stata version 10 (Stata Corp.) including herd as a random (shared frailty) effect. The model controlled for the effects of parity (primiparous or multiparous), body condition score (\geq 3.5 or <3.5), ketosis, metritis,

retained placenta, displaced abomasum (disease or no disease), Log (first test day somatic cell

- 154 count), days postpartum at first-insemination, and two-way interactions. Models were built by
- 155 manual backward stepwise exclusion of variables. If the interaction with parity was P < 0.05, the
- analysis was stratified by parity. Variables were retained in the final model if P < 0.05.

157 2.2.3. Factors associated with reagent strip tests

The purpose of this analysis was to determine if there were factors that affected LE, protein, and pH results, other than cytologic endometritis. A multivariable logistic regression model was produced with PROC GLIMMIX of SAS, Version 9.2 (SAS Institute), with cumulative logit function and reagent strip results as the dependent variable. The variables tested methods for model building were the same as described in section 2.2.1, except endometritis was tested as a fixed effect, in addition to the described list of variables.

164 2.2.4. Determination of combination cutoff points

165 Reagent strip possible results were dichotomized to all possible combinations and evaluated for Se,

- Sp, PPV and NPV to detect cytologic endometritis. The optimal combination was evaluated for
- 167 first-service conception rate and calving-to-conception interval, using the PROC GLIMMIX of
- SAS, Version 9.2 (SAS Institute) and Stata Version 10 (Stata Corp.), respectively, as described for
- 169 individual reagent strip test.
- 170 **3. Results**

171 **3.1. Descriptive statistics**

172 In total, 563 cows from 28 herds were included in the study. The median herd size was 895 (range,

- 173 540 to 3000) milking cows and the average projected 305 days mature-equivalent milk production
- was 12 562 (SD = 620) kg of milk. Overall prevalence of cytologic endometritis was 27.7%
- 175 (156/563), whereas the average within-herd prevalence was 27.8% (range, 5.3% to 52.6%).

176 **3.2. Reagent strip compared with cytology**

- 177 All three, LE, protein, and pH were strongly associated with cytologic endometritis and retained in
- the respective final models. Ketosis was retained in all three final models, and metritis was retained
- in the final model for protein reagent test association with cytologic endometritis. The proportions
- 180 of cows with endometritis increased as LE, protein, and pH values increased. Herd was not
- significant (P > 0.15) as a random variable for any of the reagent strip test models.
- 182 Dichotomized reagent strip results categorized by endometritis disease status determined using
- 183 lavage are summarized (Table 1). Based on receiver operator characteristics analysis, the optimal
- 184 cutoff points were \geq ++ for LE (area under the receiver operating characteristic curve [AUC] =
- 185 0.69; P < 0.0001), $\ge +++$ for protein (AUC = 0.60; P < 0.001), and ≥ 7.0 for pH (AUC = 0.64; P < 0.001)
- 0.001) to be used for the diagnosis of endometritis. At the optimal cutoff point, the LE test had Se =
- 187 76.9%, Sp = 51.8%, PPV = 38.0%, and NPV = 85.4%. At the optimal cutoff point, the protein test

had Se = 58.3%, Sp = 55.8%, PPV = 33.5%, and NPV = 77.7%. At the optimal cutoff point, the pH

```
test had Se = 44.9\%, Sp = 78.4\%, PPV = 44.3, and NPV = 78.8.
```

Performance of Multistix 10 SG (Bayer Corporation) LE, protein, and pH reagent strip for diagnosis of endometritis from uterine lavage samples collected from postpartum dairy cattle. TP (N) TN (N) FN (N) PPV (%) NPV (%) Test Cutoff FP (N) Se (%) Sp (%) LE 152 28 97.4 370 4 6.9 28.6 87.5 ≥ Trace 11 92.9 18.2 ≥+ 145 74 333 30.3 87.1 211 196 76.9 51.8 38.0 85.4 ≥++ 120 36 ≥+++ 53 367 40 103 34.0 90.2 57.0 78.1 147 94.2 11.5 29.0 83.9 Protein 47 360 9 $\geq +$ ≥++ 133 98 309 23 85.3 24.130.1 81.0 91 227 180 65 58.3 55.8 33.6 77.7 $\geq +++$ 125 19.9 31 367 40 90.2 43.7 74.6 $\geq ++++$ 55 pН ≥6.5 101 219 188 64.7 53.8 34.9 79.9 86 44.9 319 88 78.4 44.3 78.8 ≥7.0 70 ≥7.5 49 349 58 107 31.4 85.7 45.8 76.5 30 373 34 19.2 46.9 74.7 ≥8.0 126 91.6 ≥8.5 18 392 15 138 11.5 96.3 54.5 74.0

Protein results were recorded in six categories which were: negative, trace, + (30 mg/dL), ++ (100 mg/dL), ++ (300 mg/dL), and +++ (>2000 mg/dL); and LE results were recorded in five categories: negative, trace, + (small), ++ (moderate), and +++ (large). FN, false negative; FP, false positive; LE, leukocyte esterase; NPV, negative predictive value; PPV, positive predictive value; Se, sensitivity; Sp, specificity; TN, true negative; TP, true positive.

191 **3.2. Reagent strip and reproduction**

Table 1

190

862

Elevated LE was significantly associated with decreased hazard of pregnancy (increased calving-to-192 conception interval) only at the highest cutoff point of $\geq +++$ (144 median days not-pregnant for LE 193 \geq +++ and 117 median days not-pregnant for LE < +++ cows; hazard ratio [HR] = 0.76; 95% 194 confidence interval [CI] 0.57 to 1.00; P = 0.05). The interaction of parity and LE was significant (P 195 = 0.04), therefore, further analyses were stratified by parity. Stratification by parity showed the 196 effects to be limited to multiparous cows where LE > +++ (154 median days not-pregnant) had 39 197 longer median days not-pregnant compared with LE < +++ (HR = 0.66; 95% CI, 0.45 to 0.97; P = 198 0.03). Body condition score (HR = 1.52; 95% CI, 1.16 to 1.20; P = 0.002) and days postpartum at 199 200 first-insemination (HR = 0.98; 95% CI, 0.97 to 0.99; P = 0.003) were retained in the final model. In primiparous cows, the HR for LE was not significant (P > 0.20) at the highest cutoff with any 201

202 combination of covariates. The pregnancy survival curves for $LE \ge +++$ effect stratified by parity



Fig. 1. Kaplan-Meier survival curves for leukocyte esterase (LE) $\geq +++$ (thick line) effects on calving-to-conception interval. (A) Primiparous cows with LE $\geq +++$ did not show impaired reproductive performance compared with cows with LE < +++ (median days not-pregnant 111 and 117 days; P = 0.57). (B) Multiparous cows with LE $\geq +++$ had longer median days not-pregnant (154 days) compared with cows with LE < +++ (116 days; P = 0.02). LE results were recorded in five categories: negative, trace, + (small), ++ (moderate), and +++ (large).

203 are shown (Fig. 1).

- Protein concentration was not significant at any cutoff point, combined or stratified by parity (P > 0.20) on calving to conception interval.
- A significant effect on hazard of pregnancy was found for $pH \ge 7.0$ and the interaction between pH
- \geq 7.0 and parity was significant (P = 0.04). Stratification by parity revealed the effects to be limited
- 208 once again to multiparous cows where the median calving-to-conception interval increased from
- 209 111.5 days in pH < 7 to 150.5 days in pH \ge 7.0 (HR = 0.68; 95% CI, 0.50 to 0.92; P = 0.01) with
- body condition score (HR = 1.50; 95% CI, 1.14 to 1.96; P = 0.003) and days postpartum at first-
- insemination (HR = 0.98; 95% CI, 0.97 to 0.99; P = 0.001) also retained in the final model. In
- primiparous cows, pH was not significant (P > 0.20) at any cutoff and covariate combination.

3.3. Factors associated with reagent strip tests

Cytologically diagnosed endometritis was significantly associated with the reagent strip tests and

- was retained in the final models of LE (odds ratio [OR] = 4.49; 95% CI, 3.07 to 6.56; P < 0.0001),
- 216 protein (OR = 1.84; 95% CI, 1.26 to 2.67; P = 0.0015) and pH (OR = 2.67; 95% CI, 1.88 to 3.79; P
- < 0.0001). Herd was significant in all the final models. The final model for LE did not retain any
- other fixed variables, whereas the final model for pH retained parity (primiparous OR = 1.79; 95%)
- CI, 1.29 to 2.49; P = 0.001) and in the final model for protein, retained placenta was retained (OR =
- 220 1.94; 95% CI, 1.02 to 3.69; P = 0.045).

221 **3.4. Reagent tests used in combination**

- Although protein was associated with cytologic endometritis, the use of this test in combination did not improve the overall performance of the other reagent strip tests. The cutoff point of \geq +++ for LE in combination with pH \geq 7.0 had the best performance. Using this combination of cutoff points, NPV was 75.6% (394/521), whereas the PPV was 69.0% (29/42) with 18.6% Se and 96.8% Sp. A closer examination of the combined performance of LE and pH at each category to diagnose cytologic endometritis found the test to be much more accurate at the high and low end, whereas
- 228 cows that had LE = ++ but $pH \ge 7.0$ and cows that had LE = +++ but pH < 7.0 could not reliably be
- assigned an endometritis classification. Therefore, the results were stratified into the positive group
- 230 (LE \geq ++ and pH \geq 7.0), negative group (LE < ++ and pH < 7.0), and the undetermined group (LE
- 231 = ++ but pH \ge 7.0; or LE = +++ but pH < 7.0). The PPV for positive group was 69.0% and the NPV
- for the negative group was 82.5%; however, there was a large group of undetermined cows (20.6%
- 233 of cows).
- 234 Kaplan-Meier survival curves for calving-to-conception interval stratified by parity using combined
- LE and pH are shown (Fig. 2). Once again, reproductive impairment was restricted to multiparous
- cows where combination positive cows had 184 median days not-pregnant, whereas undetermined

cows had 129 median days not-pregnant and combination negative cows had 113 median days not-

238 pregnant (P < 0.001).



Fig. 2. Kaplan-Meier survival curves for combining leukocyte esterase (LE) and pH results on calving-to-conception interval. (A) Primiparous cows in the positive group (LE \geq +++ and pH \geq 7.0; thick solid line) were not different from the undetermined group (LE = ++ and pH \geq 7.0 or LE \geq +++ and pH < 7.0; interrupted thick line) and negative group (LE < ++ and pH < 7.0; thin line). (B) Multiparous cows in the positive group (184 median days not-pregnant) were different from the undetermined group (129 median days not-pregnant) and negative group (113 median days not-pregnant; P < 0.001). LE results were recorded in five categories: negative, trace, + (small), ++ (moderate), and +++ (large).

240 4. Discussion

239

Reagent strip results of LE, protein, and pH reagent strip were highly associated with cytologic 241 endometritis. However, the performance of LE in this study was not as accurate as described by 242 Santos et al. [8]. There are three important differences between Santos et al. and the present study. 243 First, the Santos study set the LE cutoff point at + and endometritis cutoff point at 5.5% neutrophils. 244 The analysis was repeated using data from this study at these cutoff points and found the LE test to 245 have a Se of 48.8% and Sp of 73.3%, which was still lower than the 83% Se and 94% Sp reported 246 by Santos et al. Secondly, cows were sampled between 1 and 7 wk postpartum by Santos et al., 247 whereas cows were sampled later postpartum between 40 and 60 days in the present study. Cows in 248 249 the early postpartum period tend to have very high proportions of neutrophils in endometrial cytology [2], which may have improved the performance of the reagent strip test. Finally, the 250 uterine lavage samples for the current study were only tested at the laboratory and the time interval 251 between sampling and testing was large in some cases, as the farms sampled were an average 3 h 252 drive from the laboratory. The reactivity of LE in human urine with reagent strip is decreased in 253 254 approximately 25% of samples after 24 h of refrigeration [11] increasing false negative results. In the present study, the false negative results remained low for LE, despite the long transport time. 255 Inflammatory conditions in cattle have been reported to cause an increase in the pH of fluid 256 excretions [9] and [10]. In the present study, pH of uterine fluid was increased in cows with 257 endometritis relative to normal cows and the correlation with endometritis was even higher than the 258 259 LE test. The optimal cutoff point of $pH \ge 7.0$ was similar to the recommended cutoff point for semen from bulls with seminal vesiculitis [10]. To our knowledge, this is the first report of 260 261 increased pH in uterine fluid of cows with endometritis. The Se and Sp of pH to diagnose

endometritis at the optimal cutoff point ≥7.0 was still relatively poor, at only 44.9% and 78.4%,
respectively.

The protein reagent strip test had the weakest association with cytologic endometritis and was not predictive of future reproductive performance. Storage of refrigerated human urine samples for 24 h increased the false positive results in reagent strips [11]. The observed PPV for the protein test was low, even at the highest cutoff point (43.7%), which may be attributed to the long transport time between sample collection and testing.

269 Multistix 10 SG (Bayer Corporation) is a reagent strip designed as a rapid test for human urinalysis.

270 The LE reagent strip is a highly sensitive test for human pyuria and is an excellent screening test

[12]. Conversely, the LE reagent strip test is not very sensitive and is not recommended as a

screening test for pyuria in small animal veterinary medicine [13] and [14]. The different source of

samples tested could affect the performance of the test. There are no published studies evaluating

the repeatability of LE, protein, or pH on the Multistix 10 SG (Bayer Corporation) reagent strip.

275 The LE and protein reagent strips were not designed for uterine lavage and the available increments

of reference results at the critical concentrations for endometritis are too large. The overall

prevalence of endometritis by cytology was 27.7%, but the prevalence at the optimal cutoff points

for LE and protein were 56.1% and 48.1%, respectively, which is a gross overestimation. The next

category for LE and protein grossly underestimated the prevalence at 16.5% and 12.6%,

respectively. At the optimal cutoff point for pH, the overall prevalence was 28.1%, which was close

to the cytologically derived prevalence in this study. The categorical increment for pH around the

critical point for cytologic endometritis was 0.5, which appeared to be narrow enough, whereas the available categories for LE and protein did not appear to have sufficient resolution for optimal

performance. Combining LE and pH results improved the PPV to 69.0% and the NPV to 82.5%

however; there was a group of undetermined cows (20.6%).

In summary, reagent strip results were significantly associated with cytologic endometritis and
predicted poor reproductive performance. However, the Se and Sp of reagent strip tests were
relatively poor. Modification of the test strips to optimize diagnostic categories for bovine

endometritis seems to offer the potential for an accurate and convenient cow-side diagnostic tool.

290 Acknowledgments

291 The authors thank Drs. Michael Capel, Mark Thomas, John Rath, Thomas Gill, and Robert

292 Ceglowski for their contributions in recruiting and maintaining contacts with the study herds,

293 Thomas Linden for technical help in sampling and sample processing, and the herdsmen and farm

294 owners who generously allowed access to their animals and records for this study. This project was

funded, in part, by the Cornell University Agricultural Experiment Station federal formula funds

- 296 (Animal Health) Project Number NYC-480861 received from the Cooperative State Research,
- 297 Education and Extension Service, US Department of Agriculture. Any opinions, findings,
- 298 conclusions, or recommendations expressed in the publication are those of the authors and do not
- 299 necessarily reflect the view of the USDA.

300 **References**

- 301 1] Barlund CS, Carruthers TD, Waldner CL, Palmer CW. A com- parison of diagnostic techniques
- for postpartum endometritis in dairy cattle. Theriogenology 2008;69:714 –23.
- 303 [2] Gilbert RO, Shin ST, Guard CL, Erb HN, Frajblat M. Preva- lence of endometritis and its effects
 304 on reproductive perfor- mance of dairy cows. Theriogenology 2005;64:1879 88.
- 305 [3] Kasimanickam R, Duffield TF, Foster RA, Gartley CJ, Leslie KE, Walton JS, et al. Endometrial
- 306 cytology and ultrasonography for the detection of subclinical endometritis in postpartum dairy
- 307 cows. Theriogenology 2004;62:9 –23.
- 308 [4] Cheong SH, Nydam DV, Galvão KN, Crosier BM, Gilbert RO. Cow-level and herd-level risk
- factors for subclinical endome- tritis in lactating Holstein cows. J Dairy Sci 2011;94:762–70.
- [5] Sheldon IM, Lewis GS, LeBlanc S, Gilbert RO. Defining postpartum uterine disease in cattle.
- 311 Theriogenology 2006; 65:1516 30.
- [6] Dubuc J, Duffield TF, Leslie KE, Walton JS, LeBlanc SJ. Definitions and diagnosis of
- postpartum endometritis in dairy cows. J Dairy Sci 2010;93:5225–33.
- 314 [7] Kasimanickam R, Duffield TF, Foster RA, Gartley CJ, Leslie KE, Walton JS, et al. A
- comparison of the cytobrush and uterine lavage techniques to evaluate endometrial cytology in
- clinically normal postpartum dairy cows. Can Vet J 2005;46: 255–9.
- [8] Santos NR, Roman HB, Gilbert RO. The use of leukocytei esterase reagent strips for diagnosis
- of subclinical endometritis in dairy cows. Theriogenology 2006;66:666 –7.
- [9] Marschke RJ, Kitchen BJ. Detection of bovine mastitis by bromothymol blue pH indicator test.
- 320 J Dairy Sci 1985;68: 1263–9.
- 321 [10] Juneja NL, Faulkner LC, Hopwood ML. Biochemical aspects of semen in bovine seminal
- vesiculitis. Fertil Steril 1965;16: 361–9.
- 323 [11] Froom P, Bieganiec B, Ehrenrich Z, Barak M. Stability of common analytes in urine
- refrigerated for 24 h before auto- mated analysis by test strips. Clin Chem 2000;46:1384 6.
- 325 [12] St John A, Boyd JC, Lowes AJ, Price CP. The use of urinary dipstick tests to exclude urinary
- tract infection: a systematic review of the literature. Am J Clin Pathol 2006;126:428 –36.
- 327 [13] Vail DM, Allen TA, Weiser G. Applicability of leukocyte esterase test strip in detection of
- canine pyuria. J Am Vet Med Assoc 1986;189:1451–3.

- 329 [14] Holan KM, Kruger JM, Gibbons SN, Swenson CL. Clinical evaluation of a leukocyte esterase
- test-strip for detection of feline pyuria. Vet Clin Pathol 1997;26:126 –31