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# **PHENOLIC CONTENT, ANTIOXIDANT POTENTIAL AND ANTIMICROBIAL ACTIVITIES OF FRUIT AND VEGETABLE BY-PRODUCT EXTRACTS**

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## **ABSTRACT**

The use of fruit and vegetable by-products as natural food additives has recently been suggested, due to their richness in polyphenols. The aim of this research study was to determine polyphenolic content and the antioxidative and antimicrobial activities of thirteen fruit and vegetable by-product extracts obtained with three solvent mixtures. The Folin-Ciocalteu method was employed to calculate the total phenolic content (TPC) while antioxidant capacity (AC) was assessed with DPPH<sup>•</sup> and ABTS<sup>•+</sup>. The highest TPC and AC values were obtained for the acetic extracts. Pomegranate peels and hazelnut skins showed the highest values of TPC (212.3 and 166.3 mg GAE/g dw respectively) and AC (95.7 and 92.9 of inhibition percentage respectively for DPPH<sup>•</sup> assay). The antimicrobial activity against twelve foodborne pathogens and spoilage microorganisms was evaluated. Pomegranate and apple peels showed the highest inhibition of *Staphylococcus aureus* and *Pseudomonas fluorescens*. The results obtained demonstrated that by-products could be used as natural food additives with beneficial health properties.

**Keywords:** by-products, antioxidant capacity, antibacterial activity, DPPH, total phenols

**Running title :** Antioxidant activities of by-products

## **INTRODUCTION**

Polyphenols are a very important part of our everyday diet, since they are naturally present in fruit and vegetables. As free radical scavengers, they can potentially interact with biological systems and play a role in preventing human neurodegenerative diseases and cardiovascular disorders.<sup>[1,2]</sup> Besides having a strong antioxidant effect,<sup>[3,4]</sup> polyphenols often also exhibit antimicrobial activity.<sup>[5]</sup>

A large number of plants have been examined to define their polyphenolic content and profile.<sup>[6-14]</sup>

Recently, polyphenolic content was also examined in some plant by-products<sup>[15,16]</sup> which are available in large quantities and at low cost<sup>[17]</sup> but are currently used only as feedstuffs or fertilizers. Their use as food additives could help industries to solve the environmental problems related to the disposal of these materials,<sup>[18]</sup> and provide new sources of natural antioxidants.<sup>[19]</sup> Thus, the aim of this research study was to determine the polyphenolic content and related antioxidative and antimicrobial properties of extracts obtained from thirteen fruit and vegetable by-products produced in Italy (pomaces from pomegranate, apple, white grape and red grape; peels from pomegranate, apple, hazelnut, white potato and purple potato; seeds from dog rose and cornelian cherry; leaves from leek). An ultrasound-assisted liquid-solid extraction procedure with three different solvent mixtures (methanol/water/acetic acid, ethanol/water and acetone/water) was employed. The total phenolic content (TPC) was evaluated by the Folin-Ciocalteu method, while the antioxidant capacity (AC) was assessed by means of two *in vitro* assays, the DPPH radical scavenging assay (RSA) and the ABTS or TEAC (Trolox equivalent antioxidant capacity) assay. Antimicrobial activity was screened by the agar-well diffusion method, using twelve different foodborne pathogens and spoilage microorganisms.

## **MATERIALS AND METHODS**

### **Chemicals**

2,2-Diphenyl-1-picrylhydrazyl (DPPH), Trolox<sup>®</sup> (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid), potassium persulfate, (+)-catechin hydrate, gallic acid and HPLC grade methanol were purchased from Sigma-Aldrich (Milan, Italy). HPLC grade acetone and ethanol, 2,2'-azino-bis-(3-ethylbenzothiazolin-6-sulfonate) diammonium salt (ABTS), sodium carbonate and Folin-Ciocalteu reagent were purchased from Fluka (Milan, Italy). Acetone Brain Heart Infusion Broth and technical agar (Agar No. 3) were bought from Oxoid (Milan, Italy).

## Plant material collection and extraction

The thirteen by-products listed in Table 1 were purchased directly from producers in Piedmont (North-West Italy). For pomegranate, pomace was used, i.e. the solid remains left over after crushing arils for juice extraction. Pomace contains the pulp and seeds of the fruit. The peel was examined separately. For apple, both the pomace and the peel were examined. The “Grigia di Torriana” apple variety, a typical apple produced in Piedmont, was examined. This variety is generally used for juice and jam production, and is characterized by brown peel and high astringency. Potatoes are also typical of North Italy. The “Viola” (purple) potato is a typical cultivar, also known in France as “Violette noir” or “Truffle potato”. A sample of the “Nocciola Piemonte PGI” hazelnut kernels, namely “Tonda gentile Trilobata” *cultivar* was collected. Shortly prior to analysis, hazelnut skins were removed by roasting at 160°C for 20 min in a drying ventilated oven (Mazzali Moduvers, Monza, Italy). For dog rose, cornelian cherry and potato, the pulp was also examined.

Fresh samples were washed, frozen in liquid nitrogen and lyophilized (LIO-5P, Cinquepascal, Milan, Italy), while dried samples were simply ground by a high-speed mill (IKA A11 Basic, Germany). Each sample (1 g) was extracted with 50 ml of three solvent mixtures: methanol/water/acetic acid (90:9.5:0.5, v/v/v); ethanol/water (80:20, v/v) and acetone/water (70:30; v/v). Extraction was performed in darkness, by ultrasound bath (Branson<sup>®</sup> 220, Sigma-Aldrich, Milan, Italy) working at 48 kHz for 15 min at 20 °C. The extracts were centrifuged at 3500 rpm for 20 min. The supernatant was collected, stored at + 4°C and the sample was re-extracted twice using the same procedure. Finally, the extracts were combined, filtered through 0.45 µm filters (Sartorius Stedim Biotech, Florence, Italy) and used to evaluate the total phenolic content (TPC) and antioxidant capacity (AC). An aliquot of the extracts was concentrated to dryness by rotary evaporation at 35 °C under reduced pressure (Büchi Rotavapor<sup>®</sup> R-210, Flawil, Switzerland). The solid residue was

dissolved in distilled water and lyophilised. Powders thus obtained, (maintained in darkness and nitrogen atmosphere), were used in the antimicrobial activity evaluation.

### **Total phenols assay**

The total phenolic content (TPC) of the extracts was evaluated by the Folin-Ciocalteu colorimetric method.<sup>[20]</sup> Briefly, 500 µl of extract, or gallic acid standard solutions, and 2.5 ml of 1:10 diluted Folin-Ciocalteu phenol reagent were mixed in a 10 ml test tube. After exactly 3 min, 2 ml of 7.5 % (w/v) aqueous sodium carbonate were added, the mixture was mixed again and then left to stand at 45 °C in the dark for 15 min. The absorbance, against appropriate reagent blank, was read at 765 nm in disposable 1 cm path length polystyrene (PS) cuvettes (VWR International, Milan, Italy) with a UV-1700 Spectrophotometer (PharmaSpec, Shimadzu, Milan, Italy). Gallic acid standard solutions were prepared by dissolving gallic acid in water at concentrations ranging from 0 to 250 mg/L. The total phenolic content was expressed as mg of gallic acid equivalents (GAE) per g of dry extract (dw). All samples were analyzed in triplicate.

### **DPPH radical-scavenging assay (RSA)**

Free radical-scavenging ability of fruit, vegetable and by-product extracts was based on the reaction with the stable radical DPPH, in accordance with the procedure outlined by von Gadow et al.<sup>[21]</sup> In a 5 ml test tube, 75 µl aliquot of extract was added to 3 ml of DPPH<sup>•</sup> methanol solution ( $6.1 \times 10^{-5}$  M). The mixture was mixed and left to stand at room temperature in the dark for 60 min. The absorbance was read spectrophotometrically at 515 nm in disposable PS cuvettes (1 cm path length) against a control methanol solution of DPPH<sup>•</sup>. The inhibition percentage (IP) of DPPH<sup>•</sup> was calculated according to the following equation:

$$IP[\%] = (A_{\text{control}} - A_{\text{sample}}) / A_{\text{control}} \times 100$$

where  $A_{\text{sample}}$  and  $A_{\text{control}}$  are the absorbance values of the reaction mixture with and without samples, respectively. All samples were analyzed in triplicate.

### **TEAC assay**

The Trolox equivalent antioxidant capacity (TEAC) assay, which measures the reduction of the ABTS radical cation by antioxidants, was performed according to the modified method of Re et al.<sup>[22]</sup> The pre-formed radical monocation of 2,2-azinobis-3-ethylbenzothiazoline-6-sulfonic acid (ABTS<sup>•+</sup>) was generated by oxidation of ABTS aqueous solution (7 mM) with 2.45 mM potassium persulfate ( $K_2S_2O_8$ ), and allowing the mixture to stand in the dark at room temperature for 12-16 h before use. Just before analysis, the resulting blue-green ABTS<sup>•+</sup> stock solution was diluted with ethanol to an absorbance of 0.700 ( $\pm$  0.020) at 734 nm and equilibrated at 30 °C in the dark. A reagent blank reading was taken ( $A_{\text{blank}}$ ). In a 5 ml test tube, 30  $\mu$ l of extracts were added to 3 ml of diluted ABTS<sup>•+</sup> solution. The extinction at 734 nm (1 cm path length PS cuvettes, 30 °C) was measured exactly 6 min after the initial mixing. The ABTS<sup>•+</sup> scavenging effect (% Inhibition) was calculated as follow:

$$\% \text{ Inhibition} = (A_{\text{blank}} - A_{\text{sample}}) / A_{\text{blank}} \times 100$$

where  $A_{\text{blank}}$  and  $A_{\text{sample}}$  are the absorbances of ABTS<sup>•+</sup> working solution before and after the sample addition. A calibration curve was prepared with different concentrations of Trolox (0-350  $\mu$ M/l) and the antioxidant capacity was expressed as  $\mu$ M of Trolox equivalent (TE) per g of dry extract (dw). All samples were analyzed in triplicate.

### **Antibacterial assay**

An overnight culture of approximately  $10^8$  colony forming units (CFU)/ml was used for all the microorganisms. A range of microorganisms were used as indicators: *Listeria monocytogenes* NCTC 10527, *Staphylococcus aureus* ATCC12606, *Bacillus cereus* DSM 350, *Lactobacillus*

*sakei* DSMZ 6333, *Lactococcus lactis* DSM 4366, *Staphylococcus xylosum*, *Salmonella*, *Pseudomonas fluorescens*, *Escherichia coli* DH5 $\alpha$ , *Escherichia coli* ATCC 35150 (Shiga toxin-producing *Escherichia coli*), *Pseudomonas aeruginosa* and *Serratia marcescens*, from the collection of the Department of Exploitation and Protection of the Agricultural and Forestry Resources, University of Turin, Italy. Strains that did not originate from an international culture collection, were isolated from foodstuffs and their identification to the species level was performed by 16S rDNA sequencing. The agar-well diffusion method was used to determine antibacterial activity.<sup>[23]</sup> Sterile BHI (Brain Heart Infusion) agar was mixed with the indicator microorganism (final concentration 1% v/v) and poured into sterile standard Petri dishes (20 ml). After setting, medium cups of 6 mm diameter were prepared. For each test, 70  $\mu$ l of the different solutions with 10 and 20 mg of extract/ml concentration were added to the well.<sup>[5]</sup> Pure methanol was used as control. After incubation at 37 °C for 24 h, the resulting inhibition zone diameters were measured. All tests were performed in triplicate.

### **Statistical analysis**

Data, unless otherwise specified, were expressed as mean  $\pm$  standard deviation of triplicate experiments. Statistical analysis was performed with the SPSS software package (version 12.0 for Windows, SPSS Inc., Chicago, Illinois). The one-way analysis of variance (ANOVA) with Duncan's test was carried out to compare samples. The relationship amongst TPC and AC assays was described by the Pearson correlation coefficient  $r$ .

## **RESULTS AND DISCUSSION**

### **Total phenolic content**

The total phenolic content (TPC) of fruit and vegetable by-product extracts obtained using three



different solvent mixtures are reported in Table 2. Results showed that solvents had significantly different capacities in the extraction of polyphenols, and the most effective was the aqueous acetone solution. Taking into consideration only those extracts obtained with this solvent, the TPC ranged from 212.3 mg of GAE/g dw in pomegranate peel to 1.6 mg GAE/g dw in “Piatlina” potato pulp. The TPC of extracts could be subdivided into three groups, namely high ( $\geq 50$  mg GAE/g dw), medium ( $< 50 - \geq 20$  mg GAE/g dw) and low ( $< 20$  mg GAE/g dw). Pomegranate, hazelnut and apple skin, dog rose pulp, white marcs and cornelian cherry pulp all belong to the first group. Cornelian cherry seeds, red marcs and dog rose seeds belong to the second group, while potato peel and pulp, leek leaves, pomegranate and apple pomace pertain to the third.

The TPC of pomegranate peel aqueous acetone extracts (212.3 mg GAE/g dw) was nearly 16-fold higher than that of aril pomace extracts (13.2 mg GAE/g dw), according to data reported by Li et al.<sup>[24]</sup> where larger amounts of phenols were found in pomegranate peel with respect to arils (249.4 mg GAE/g and 24.4 mg GAE/g dry extract respectively). Nasr et al.<sup>[25]</sup> determined a similar TPC content (216.9 mg GAE/g dw) in pomegranate peel extract while, more recently, Vijaya Kumar Reddy et al.<sup>[13]</sup> reported a TPC of 2.2 mg GAE/g fresh weight for acidified aqueous methanol extract of pomegranate arils. Besides pomegranate peel, roasted hazelnut skin could also be considered a polyphenol-rich product, showing 166.3 mg GAE/g dw of TPC for aqueous acetone solvent. Contini et al.<sup>[26]</sup> reported a TPC of 466.8 mg GAE/g dw from skin waste of whole roasted hazelnut, but extraction figures differed with long maceration time of the defatted product.

The TPC of dog rose pulp aqueous acetone extracts was similar (85.5 mg GAE/g dw) to those reported by Wenzig et al. (82.2 mg GAE /g dw).<sup>[27]</sup> Our study also analysed the dog-rose seeds, a by-product obtained during jam production. Extracts of dog-rose seeds showed a low value of TPC (21.5 mg GAE/g dw). This value is lower than that of fruits but interesting when considering the use of seeds as a low-cost additive for functional foods. The TPC of cornelian cherry pulp fruits (50.1 mg GAE/g dw) was higher than that reported by Ju and Hsieh (20.9 – 33.4 mg GAE/g semi-dried fruits),<sup>[28]</sup> Marinova et al. (4.3 mg GAE/g fresh mass),<sup>[29]</sup> and Pantelidis et al. (15.9 mg GAE/g

dw)<sup>[30]</sup> but comparable with data published by Yilmaz et al. (26.6 – 74.8 mg GAE/g dw).<sup>[10]</sup> The TPC of cornelian cherry seeds (37.7 mg GAE/g dw) was higher than that of dog rose seeds and similar to that of fruits.

Apple fruits have been widely investigated as a good source of polyphenols. Suarez et al.<sup>[31]</sup> reported apple pomace to have a higher value for acetic extracts (6.5 g GAE/kg dw) than for methanolic extracts (3.6 g GAE/kg dw). Wolfe et al.<sup>[32]</sup> showed that extracts of apple peel exhibited significantly higher TPC than those of apple pulp. Similar results were obtained in this study with 61.3 mg GAE/g dw for apple peel and only 10.4 mg GAE/g dw for apple pulp. .

Among vegetables, the TPC of acetic extracts of potato peel and pulp from different Italian varieties (“Viola”, “Desirée” and “Piatlina”) showed very low values. The purple “Viola” variety, exhibited the highest TPC among potato samples (8.9 mg GAE/g dw). Al-Weshahy and Rao reported that the TPC of peel for six varieties ranged from 1.5 to 2.1 mg GAE/ g dw for black tuber and from 2.9 to 3.3 mg GAE/g dw for red tuber.<sup>[33]</sup> The differences in values of TPC were probably due to the colour and variety of potato tested<sup>[34]</sup> but also a result of the presence of anthocyanins in the skin of colored potato varieties, which was found to be 2.5- fold higher than in the tuber pulp.<sup>[35]</sup> The TPC of leek leaf aqueous acetone extract was 7.0 mg GAE/g dw. Few studies are available on polyphenolic content of leek and even fewer devoted to the edible part of this vegetable. Marinova et al.<sup>[29]</sup> and Dragović-Uzelac et al.<sup>[36]</sup> reported a TPC of 35.7 and 75.3 mg GAE/ 100 g fresh mass respectively but, in the absence of data about dry matter, these values are not comparable with those obtained in this study.

Data published by Turkmen et al.<sup>[37]</sup> was more comparable as they reported a TPC of 3.0 mg GAE/g dw for leek. As the edible part of leek is formed by modified leaves, it is possible that its polyphenolic composition is similar to that of non-modified leaves which are by-products. Regarding marc extracts, the TPC for marc obtained from white grapes was significantly higher (50.5 mg GAE/g dw) than that obtained from red grapes (24.1 mg GAE/g dw). These values were similar to those reported by Vatai et al. who registered a TPC for marc from red grapes between

17.3 and 20.2 mg GAE/g dw.<sup>[38]</sup> Differences in TPC between red and white marc are due to their different origins: white marc is produced during must production after crushing, while red marc is obtained during pressing after the alcoholic fermentation. Also, a high quantity of polyphenolic compounds of red grapes is dissolved in wine during wine-making.

### **Antioxidant capacity**

Several methods have been developed to assess the *in vitro* antioxidant capacity of plant extracts. Relationships between assays were regulated by the method applied but also by the structure of antioxidants analyzed. Therefore, the use of at least two different analytical approaches to test the antioxidant capacity of specific substrates is recommended.<sup>[39]</sup> Buenger et al.<sup>[40]</sup> reported that the DPPH<sup>•</sup> assay, followed by the ABTS<sup>•+</sup> assay, yield the best results (based on reproducibility and sensitivity). These tests, involving chromogen compounds of a radical nature, are also the most common antioxidant capacity assays, used for their ease, speed and sensitivity.<sup>[41]</sup> In this study the DPPH<sup>•</sup> and the ABTS<sup>•+</sup> assays were thus selected to evaluate the antioxidant potential of extracts obtained from fruit and vegetable by-products (Table 3).

All products showed a scavenging activity against DPPH radical but significant differences were highlighted among extraction solvents. Generally, the aqueous acetonic extracts showed the highest antioxidant capacities and IP values range between 95.73% in pomegranate peel and 4.01% in “Piatlina” potato peel (Table 3). According to Kaur and Kapoor these values could be subdivided into three antioxidant activity groups: high ( $\geq 50\%$ ), moderate (20-50%) and low ( $< 20\%$ ).<sup>[42]</sup> Among the examined extracts, eight belonged to the former and included pomegranate peel (95.62%), hazelnut skin (92.90%), cornelian cherry seeds (77.44%), dog rose pulp (74.37%), marcs from red grape (65.98%), apple peel (63.44%), marcs from white grape (58.34 %) and cornelian cherry pulp (54.44%). The group with moderate activity was represented by dog rose seeds (45.40%), pomegranate pomace (24.91%), and “Viola” potato peel (20.34%). Finally, the other extracts could be included in the low anti-oxidant activity ( $< 20\%$ ) group. The total antioxidant

capacity of peel was generally significantly higher than pulp and pomace ( $p < 0.05$ ).

The ABTS<sup>•+</sup> method was used to confirm the results from the DPPH<sup>•</sup> test since it is based on a similar antioxidant mechanism and the results are laid out in Table 3. The TEAC values ranged between 0.10 and 0.71  $\mu\text{M/g dw}$ , showing the same trend reported for IP. The highest TEAC values (0.70 – 0.71  $\mu\text{M TE/g dw}$ ) were detected in hazelnut skin, pomegranate peel, apple peel, cornelian cherry pulp and seeds, white and red grape marcs, and dog rose pulp extracts. The lowest TEAC values were observed for “Piatlina” and “Desirée” potato peel extracts. Therefore, the results for TEAC tests are well in line with those of the DPPH<sup>•</sup> assay.

For aqueous acetonc extracts, IP and TEAC values are directly correlated ( $r=0.93$ ;  $p<0.01$ ) according to their similar redox mechanism. These assays are also correlated with TPC ( $r=0.78$  and  $r=0.82$ ,  $p<0.01$  respectively). There is no unanimous opinion about the relationship between the content of phenols and their antioxidant activity. Some authors observed close or very close correlations,<sup>[43,44]</sup> but this hypothesis was frequently discussed and opposed. Adopting the Folin-Ciocalteu method, various phenolic compounds have different responses to this assay, proportionally due to the number and positioning of hydroxyl groups. Since these structural features of phenols are also responsible for antioxidant activity, measurements of phenols in natural products may be related to this potential. In addition, the Folin-Ciocalteu assay mechanism is an oxidation/reduction reaction and, as such, can be considered another antioxidant method.<sup>[45]</sup>

### **Antibacterial activity**

Employing the agar-well diffusion technique, the antibacterial activity of phenolic extracts against twelve food -related microorganisms was evaluated. The microorganisms selected belonged to pathogenic, spoilage or technologically important species, commonly found in foods. Only six products showed antimicrobial activity, and the highest values were highlighted for acetone and methanol extracts (Table 4). Higher antibacterial activity of acetone extracts was also reported by Negi and Jayaprakasha.<sup>[46]</sup> Two different extract concentrations were tested (10 and 20 mg of dry

extract/ml), but, as expected, higher activity was shown for the 20 mg/ml concentration. *Staphylococcus. marcescens*, *S. xylosus* and *Lb. sakei* were found to be the most resistant bacteria while *S. aureus* was the most sensitive. Pomegranate peel extracts were active against eleven bacterial species, and seed extracts against four. As reported by Negi and Jayaprakasha acetonetic and methanolic extracts from pomegranate peel showed antimicrobial activity against *B. cereus*, *S. aureus*, *E. coli* and *P. aeuroginosa*.<sup>[46]</sup> As a large quantity of tannins were identified in pomegranate extracts,<sup>[47]</sup> Cowan suggested that the antibacterial properties of these extracts could be related to tannins and their activity to inactivate microbial adhesions, enzymes, and cell envelope transport proteins, and to modify the morphology of microorganisms.<sup>[48]</sup>

Apple peel and pomace extracts were active against eight and five microorganisms respectively. Fattouch et al.<sup>[49]</sup> reported that acetonetic extracts from apple peel inhibited *S. aureus*, *B. cereus*, *P. aeuroginosa*, *E. coli*, and *Salmonella* spp. Peel extracts exhibited more antibacterial activity than pulp, according to their biochemical properties. Low activity was highlighted for cornelian cherry pulp extract, which showed activity for only four microorganisms, whereas dog rose pulp extract was active against an *E.coli* strain and *S. aureus*. Other extracts did not show any antimicrobial activity.

## CONCLUSION

The ultrasonic extraction method with aqueous acetone mixture was the most effective for polyphenols from fruit and vegetable by-products. These extracts showed the highest value of antioxidant capacity. High values of polyphenolic content and antioxidant capacity were identified in pomegranate peel, hazelnut skin, cornelian cherry seed, marc and apple peel extracts. High antioxidant capacities were also shown by some minor fruits such as dog rose and cornelian cherry pulp. For some of these products antimicrobial activity was also observed, in particular against *S. aureus*, an important foodborne pathogen, and *P. fluorescens*, a spoilage microorganism. Further

studies are needed to evaluate the possible use of these fruit and vegetable by-products as natural food additives to increase their safety and nutritional value.

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## REFERENCES

1. German, J.B.; Frankel, E.N.; Waterhouse, A.L.; Hansen, R.J.; Walmez, R.L. (1997). Wine phenolics and targets of chronic diseases. In *Wine: nutritional and therapeutic benefits*; Watkins, T.R., Eds.; American Chemical Society: Washington DC, **1997**; 196-214.
2. Poudel, P.R.; Tamura, H.; Kataoka, I.; Mochioka, R. Phenolic compounds and antioxidant activities of skin and seeds of five wild grapes and two hybrids native to Japan. *Journal of Food Composition and Analysis* **2008**, 21, 622-625.
3. Bors, W.; Werner, H.; Michel, C.; Saran, M. Flavonoids as antioxidants: determination of radical-scavenging efficiencies. *Methods in Enzymology* **1990**, 186, 343-355.
4. Hanasaki, Y.; Ogawa, S.; Fukui, S. The correlation between active oxygen scavenging and antioxidative effects of flavonoids. *Free Radical Biology and Medicine* **1994**, 16, 845-850.
5. Rauha, J.-P.; Remes, S.; Heinonen, M.; Hopia, A.; Kahkonen, M.; Kujala, T.; Pihlaja, K.; Vuorela, H.; Vuorela, P. Antimicrobial effects of Finnish plant extracts containing flavonoids and other phenolic compounds. *International Journal of Food Microbiology* **2000**, 56, 3-12.
6. Hvattum, E. Determination of phenolic compounds in rose hip (*Rosa canina*) using liquid chromatography coupled to electrospray ionisation tandem mass spectrometry and diode-array detection. *Rapid Communication in Mass Spectrometry* **2002**, 16, 655-662.

7. Imeh, U.; Khokhar, S. Distribution of conjugated and free phenols in fruits: antioxidant activity and cultivar variations. *Journal of Agricultural and Food Chemistry* **2002**, *50*, 6301-6306.
8. Seeram, N.P.; Schutzki, R.; Chandra, A.; Nair, M.G. Characterization, quantification, and bioactivities of anthocyanins in *Cornus* species. *Journal of Agricultural and Food Chemistry* **2002**, *50*, 2519–2523.
9. Demir, F.; Kalyoncu, I.H. Some nutritional, pomological and physical properties of cornelian cherry (*Cornus mas* L.). *Journal of Food Engineering* **2003**, *60*, 335-341.
10. Yilmaz, K.U.; Ercisli, S.; Zengin, Y.; Sengul, M.; Kafkas, E.Y. Preliminary characterisation of cornelian cherry (*Cornus mas* L.) genotypes for their physicochemical properties. *Food Chemistry* **2009**, *114*, 408-412.
11. Djordjevic, T.M.; Siler-Marinkovic, S.S.; Dimitrijevic-Brankovic, S.I. Antioxidant activity and total phenolic content in some cereals and legumes. *International Journal of Food Properties* **2011**, *14*, 175-184.
12. Pawlowska, A. M.; Camangi, F.; Braca A. Quali-quantitative analysis of flavonoids of *Cornus mas* L. (*Cornaceae*) fruits. *Food Chemistry* **2010**, *119*, 1257- 1261.
13. Vijaya Kumar Reddy, C.; Sreeramulu, D.; Raghunath, M. Antioxidant activity of fresh and dry fruits commonly consumed in India. *Food Research International* **2010**, *43*, 285-288.
14. Ghirardello, D.; Prosperini, S.; Zeppa, G.; Gerbi, V. Phenolic acid profile and antioxidant capacity of hazelnut (*Corylus avellana* L.) kernels in different solvent systems. *Journal of Food Nutrition Research* **2010**, *49*, 195-205.
15. Balasundram, N.; Sundram, K.; Samman, S. Phenolic compounds in plants and agri-industrial by-products: antioxidant activity, occurrence, and potential uses. *Food Chemistry* **2006**, *99*, 191–203.
16. Peschel, W.; Sánchez-Rabaneda, F.; Diekmann, W.; Plescher, A.; Gartzía, I.; Jiménez, D.; Lamuela-Raventós, R.; Buxaderas, S.; Codina, C. An industrial approach in the search of natural antioxidants from vegetable and fruit wastes. *Food Chemistry* **2006**, *97*, 137–150.

17. Schieber, A.; Stintzing, F.C.; Carle, R. By-products of plant food processing as a source of functional compounds — recent developments. *Trends in Food Science and Technology* **2001**, *12*, 401-413.
18. Virot, M.; Tomao, V.; Le Bourvellec, C.; Renard, C.M.C.G.; Chemat, F. Towards the industrial production of antioxidants from food processing by-products with ultrasound-assisted extraction. *Ultrasonics Sonochemistry* **2010**, *17*, 1066-1074.
19. Mour, A.; Cruz, J.M.; Franco, D.; Dominguez, J.M.; Sineiro, J.; Dominguez, H.; Núñez, M. J.; Parajó J.C. Natural antioxidants from residual sources. *Food Chemistry* **2001**, *72*, 145-171.
20. Singleton, V.L.; Rossi, J.A. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture* **1965**, *16*, 144–158.
21. von Gadow, A.; Joubert, E.; Hansamann, C.F. Comparison of antioxidant activity of aspalathin with that of other plant phenols of Rooibose tea (*Aspalathus linearis*),  $\alpha$ -tocopherol, BHT and BHA. *Journal of Agricultural and Food Chemistry* **1997**, *45*, 632-648.
22. Re, R.; Pellegrini, N.; Proteggente, A.; Pannala, A.; Yang, M.; Rice-Evans, C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Biology and Medicine* **1999**, *26*, 1231–1237.
23. Agnese, A.M.; Perez, C.; Cabrera, J.L. *Adesmia aegiceras*: antimicrobial activity and chemical study. *Phytomedicine* **2001**, *8*, 389–394.
24. Li, Y.; Guo, C.; Yang, J.; Wei, J.; Xu, J.; Cheng, S. Evaluation of antioxidant properties of pomegranate peel extract in comparison with pomegranate pulp extract. *Food Chemistry* **2006**, *96*, 254–260.
25. Nasr, C.B.; Ayed, N.; Metche, M. Quantitative determination of the polyphenolic content of pomegranate peel. *Zeitschrift für Lebensmittel-Untersuchung und -Forschung* **1996**, *203*, 374-378.



26. Contini, M.; Baccelloni, S.; Massantini, R.; Anelli, G. Extraction of natural antioxidants from hazelnut (*Corylus avellana* L.) shell and skin wastes by long maceration at room temperature. *Food Chemistry* **2008**, 110, 659-669.
27. Wenzig, E.M.; Widowitz, U.; Kunert, O.; Chrubasik, S.; Bucar, F.; Knaudera, E.; Bauer, R. Phytochemical composition and *in vitro* pharmacological activity of two rose hip (*Rosa canina* L.) preparations. *Phytomedicine* **2008**, 15, 826–835.
28. Ju, Z.Y.; Hsieh, Y.H.P. In *Nutraceuticals and functional foods: antioxidants and phytochemical analysis*, Proceedings of the IFT Annual Meeting, Las Vegas, NV, July 12–16, 33F-13, **2004**.
29. Marinova, D.; Ribarova, F.; Atanassova, M. Total phenolics and total flavonoids in Bulgarian fruits and vegetables. *Journal of University Chemical Technology and Metallurgy* **2005**, 40, 255–260.
30. Pantelidis, G.E.; Vasilakakis, M.; Manganaris, G.A.; Diamantidis, G. Antioxidant capacity, phenol, anthocyanin and ascorbic acid contents in raspberries, blackberries, red currants, gooseberries and Cornelian cherries. *Food Chemistry* **2007**, 102, 777–783
31. Suarez, B.; Alvarez, A.L.; Garcia, Y.D.; del Barrio, G.; Lobo, A.P.; Parra F. Phenolic profiles, antioxidant activity and *in vitro* antiviral properties of apple pomace. *Food Chemistry* **2010**, 120, 339-342.
32. Wolfe, K.; Wu, X.Z.; Liu, R.H. Antioxidant activity of apple peels. *Journal of Agricultural and Food Chemistry* **2003**, 51, 609-614
33. Al-Weshahy, A.; Rao, A.V. Isolation and characterization of functional components from peel samples of six potato varieties growing in Ontario. *Food Research International* **2009**, 42, 1062-1066.
34. Al-Saikhan, M.S.; Howard, L.R.; Miller, J.C. Antioxidant activity and total phenolics in different genotypes of potato (*Solanum tuberosum* L). *Journal of Food Science* **1995**, 60, 341-343.

35. Jansen, G.; Flamme, W. Coloured potatoes (*Solanum tuberosum* L.) – anthocyanin content and tuber quality. *Genetic Resources and Crop Evolution* **2006**, *53*, 1321-1331.
36. Dragović-Uzelac, V.; Bursać Kovačević, D.; Levaj, B.; Pedisić, S.; Mezak, M.; Tomljenović, A. Polyphenols and antioxidant capacity in fruits and vegetables common in the Croatian diet. *Agriculturae Conspectus Scientificus* **2005**, *74*, 175-179.
37. Turkmen, N.; Sari, F.; Velioglu, Y.S. The effect of cooking methods on total phenolics and antioxidant activity of selected green vegetables. *Food Chemistry* **2005**, *93*, 713-718.
38. Vatai, T.; Škerget, M.; Knez, Z. Extraction of phenolic compounds from elderberry and different grape marc varieties using organic solvents and/or supercritical carbon dioxide. *Journal of Food Engineering* **2009**, *90*, 246–254.
39. Schleiser, K.; Harwat, M.; Böhm, V.; Bitsh, R. Assessment of antioxidant activity by using different *in vitro* methods. *Free Radical Research* **2002**, *36*, 177-187.
40. Buenger, J.; Ackermann, H.; Jentsch, A.; Mehling, A.; Pfitzner, I.; Reiffen, K.A.; Schroeder, K.R.; Wollenweber, U. An interlaboratory comparison of methods used to assess antioxidant potentials. *International Journal of Cosmetic Science* **2006**, *28*, 135–146.
41. Arnao, M.B. Some methodological problems in the determination of antioxidant activity using chromogen radicals: a practical case. *Trends in Food Science and Technology* **2000**, *11*, 419-421.
42. Kaur, C.; Kapoor, H.C. Anti-oxidant activity and total phenolic content of some Asian vegetables. *International Journal of Food Science and Technology* **2002**, *37*, 153-161.
43. Kähkönen, M.P.; Hopia, A.I.; Vuorela, H.J.; Rauha, J.P.; Pihlaja, K.; Kujala, T.S.; Heinonen, M. Antioxidant activity of plant extracts containing phenolic compounds. *Journal of Agricultural and Food Chemistry* **1999**, *47*, 3954-3962.
44. Stratil, P.; Klejdus, B.; Kubáň, V. Determination of total content of phenolic compounds and their antioxidant activity in vegetables – Evaluation of spectrophotometric methods. *Journal of Agricultural and Food Chemistry* **2006**, *54*, 607-616.

45. Prior, R.L.; Wu, X.; Shaich, K. Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agricultural and Food Chemistry* **2005**, *53*, 4290-4302.
46. Negi, P.S.; Jayaprakasha, G.K. Antioxidant and antibacterial activities of *Punica granatum* extracts. *Journal of Food Science* **2003**, *68*, 1473–1477.
47. Gil, M.I.; Tomas-Barberan, F.A.; Hess-Pierse, B.; Holcroft, D.M.; Kader, A.A. Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing. *Journal of Agricultural and Food Chemistry* **2000**, *48*, 4581–4589.
48. Cowan, M.M. Plant products as antimicrobial agents. *Clinical Microbiology Reviews* **1999**, *12*, 564–582.
49. Fattouch, S.; Caboni, P.; Coroneo, V.; Tuberoso, C.; Angioni, A.; Dessi, S.; Marzouki, N.; Cabras, P. Antioxidant and antimicrobial activities of Tunisian pome fruit pulp and peel aqueous acetone extracts. *Journal of Agricultural and Food Chemistry* **2008**, *56*, 1084–1090.

## **Table captions**

**Table 1.** Fruit and vegetable by-products examined

**Table 2.** Total phenolic content (TPC) in the plant extracts obtained with the three solvents. Data are means  $\pm$  SD (n=3).

**Table 3.** Antioxidant capacity (AC) evaluated for extracts obtained with the three solvents. Data are means  $\pm$  SD (n=3).

**Table 4.** Antibacterial activity of extracts obtained with the three solvents and evaluated by the agar- well diffusion assay. The concentration was 20 mg of dry extract/ml.

**Table 1.** Fruit and vegetable by-products examined.

<b>Common name</b>	<b>Scientific name</b>	<b>Families</b>	<b>Variety</b>	<b>Examined products</b>
<i>Fruits</i>				
Pomegranate	<i>Punica granatum</i>	<i>Lythraceae</i>	Dente di cavallo	Pomace – Peel
Apple	<i>Malus domestica</i>	<i>Rosaceae</i>	Grigia di Torriana	Pomace – Peel
Dog Rose	<i>Rosa canina</i>	<i>Rosaceae</i>	--	Pulp – Seeds
Cornelian cherry	<i>Cornus mas</i>	<i>Cornaceae</i>	--	Pulp – Seed
Hazelnut	<i>Corylus avellana</i>	<i>Corylaceae</i>	Round the Kind Trilobata	Skin
White grapes	<i>Vitis Vinifera</i>	<i>Vitaceae</i>	Chardonnay	Marc
Red grapes	<i>Vitis Vinifera</i>	<i>Vitaceae</i>	Nebbiolo	Marc
<i>Vegetables</i>				
Potato	<i>Solanum tuberosum</i>	<i>Solanaceae</i>	Viola	Pulp - Peel
Potato	<i>Solanum tuberosum</i>	<i>Solanaceae</i>	Desirée	Pulp - Peel
Potato	<i>Solanum tuberosum</i>	<i>Solanaceae</i>	Piatlina	Pulp - Peel
Leek	<i>Allium porrum</i>	<i>Liliaceae</i>	Monstrueux di Carentan	Non edible leaves

**Table 2.** Total phenolic content (TPC) in the plant extracts obtained with the three solvents. Data are means  $\pm$  SD (n=3).

	TPC (mg GAE <sup>†</sup> /g dry weight)		
	Solvent A <sup>††</sup>	Solvent B <sup>††</sup>	Solvent C <sup>††</sup>
Hazelnut (skin)	124.6 $\pm$ 2.44 <sup>b,A</sup>	116.5 $\pm$ 5.25 <sup>e,A</sup>	166.3 $\pm$ 5.43 <sup>f,B</sup>
Pomegranate (peel)	197.1 $\pm$ 1.76 <sup>g,B</sup>	173.2 $\pm$ 3.54 <sup>f,A</sup>	212.3 $\pm$ 3.31 <sup>g,C</sup>
Apple (peel)	32.3 $\pm$ 1.14 <sup>e,B</sup>	20.1 $\pm$ 8.99 <sup>c,A</sup>	61.3 $\pm$ 4.65 <sup>e,C</sup>
“Viola” Potato (peel)	9.5 $\pm$ 0.29 <sup>bc,A</sup>	8.9 $\pm$ 1.40 <sup>ab,A</sup>	8.9 $\pm$ 0.08 <sup>a,A</sup>
“Desiree” Potato (peel)	5.9 $\pm$ 0.22 <sup>ab,A</sup>	7.4 $\pm$ 0.52 <sup>a,B</sup>	7.5 $\pm$ 0.08 <sup>a,B</sup>
“Piatlina” Potato (peel)	4.2 $\pm$ 0.09 <sup>a,A</sup>	4.2 $\pm$ 0.22 <sup>a,A</sup>	5.2 $\pm$ 0.07 <sup>a,B</sup>
Leek (leaves)	5.9 $\pm$ 0.08 <sup>ab,A</sup>	6.4 $\pm$ 0.50 <sup>a,AB</sup>	7.0 $\pm$ 0.31 <sup>a,B</sup>
Cornelian cherry (seed)	36.6 $\pm$ 5.51 <sup>f,A</sup>	33.3 $\pm$ 2.82 <sup>d,A</sup>	37.7 $\pm$ 3.32 <sup>c,A</sup>
Dog Rose (seeds)	21.2 $\pm$ 0.38 <sup>d,B</sup>	16.3 $\pm$ 0.31 <sup>bc,A</sup>	21.5 $\pm$ 0.33 <sup>b,B</sup>
White grape (marcs)	34.2 $\pm$ 1.25 <sup>ef,A</sup>	35.8 $\pm$ 9.76 <sup>d,A</sup>	50.5 $\pm$ 6.57 <sup>d,B</sup>
Red grape (marcs)	10.4 $\pm$ 1.68 <sup>c,B</sup>	6.6 $\pm$ 0.88 <sup>a,A</sup>	24.1 $\pm$ 0.75 <sup>b,C</sup>
Dog Rose (pulp)	19.9 $\pm$ 1.00 <sup>e,B</sup>	16.7 $\pm$ 0.64 <sup>d,A</sup>	85.5 $\pm$ 1.44 <sup>f,C</sup>
Cornelian cherry (pulp)	32.6 $\pm$ 1.32 <sup>d,B</sup>	26.8 $\pm$ 3.00 <sup>e,A</sup>	50.1 $\pm$ 2.77 <sup>e,C</sup>
Pomegranate (pomace)	9.9 $\pm$ 0.24 <sup>c,B</sup>	8.7 $\pm$ 0.45 <sup>c,A</sup>	13.2 $\pm$ 0.29 <sup>d,C</sup>
Apple (pomace)	8.9 $\pm$ 0.18 <sup>c,A</sup>	9.8 $\pm$ 1.55 <sup>c,A</sup>	10.4 $\pm$ 0.25 <sup>c,A</sup>
“Viola” Potato (pulp)	3.1 $\pm$ 0.20 <sup>b,B</sup>	2.4 $\pm$ 0.19 <sup>ab,A</sup>	2.6 $\pm$ 0.04 <sup>ab,A</sup>
“Desiree” Potato (pulp)	2.1 $\pm$ 0.42 <sup>b,A</sup>	4.2 $\pm$ 0.10 <sup>b,B</sup>	4.4 $\pm$ 1.04 <sup>b,B</sup>
“Piatlina” Potato (pulp)	0.5 $\pm$ 0.40 <sup>a,A</sup>	0.8 $\pm$ 0.43 <sup>a,A</sup>	1.6 $\pm$ 0.24 <sup>a,B</sup>

<sup>†</sup> Gallic acid equivalent.

<sup>††</sup> Solvent A = methanol (90%) in acidified water; solvent B = ethanol (80%) in water; solvent C = acetone (70%) in water.

<sup>a-h</sup> Values within column with the same letters are not significantly different at  $p \leq 0.05$ .

<sup>A-C</sup> Values within row with the same letters are not significantly different at  $p \leq 0.05$ .

**Table 3.** Antioxidant capacity (AC) evaluated for extracts obtained with the three solvents. Data are means  $\pm$  SD ( $n=3$ ).

	IP (%)			TEAC ( $\mu\text{M TE}^{\dagger}/\text{g dry weight}$ )		
	Solvent A <sup>††</sup>	Solvent B	Solvent C	Solvent A	Solvent B	Solvent C
Hazelnut (skin)	79.51 $\pm$ 5.18 <sup>e,A</sup>	86.58 $\pm$ 5.54 <sup>h,AB</sup>	92.90 $\pm$ 2.45 <sup>h,B</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>e,A</sup>
Pomegranate (peel)	95.73 $\pm$ 0.04 <sup>f,A</sup>	95.27 $\pm$ 0.57 <sup>i,A</sup>	95.62 $\pm$ 0.03 <sup>h,A</sup>	0.70 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>e,A</sup>
Apple (peel)	49.33 $\pm$ 2.19 <sup>d,B</sup>	20.89 $\pm$ 2.26 <sup>c,A</sup>	63.44 $\pm$ 2.58 <sup>ef,C</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>e,A</sup>
“Viola” potato (peel)	23.50 $\pm$ 0.60 <sup>b,B</sup>	19.26 $\pm$ 0.80 <sup>c,A</sup>	20.34 $\pm$ 0.02 <sup>c,A</sup>	0.26 $\pm$ 0.01 <sup>c,A</sup>	0.26 $\pm$ 0.01 <sup>c,A</sup>	0.30 $\pm$ 0.02 <sup>c,B</sup>
“Desirée” potato (peel)	10.15 $\pm$ 0.61 <sup>a,A</sup>	12.83 $\pm$ 0.88 <sup>b,A</sup>	14.87 $\pm$ 0.93 <sup>bc,A</sup>	0.15 $\pm$ 0.01 <sup>b,A</sup>	0.19 $\pm$ 0.01 <sup>b,B</sup>	0.22 $\pm$ 0.01 <sup>a,C</sup>
“Piatlina” potato (peel)	8.42 $\pm$ 0.43 <sup>a,A</sup>	7.76 $\pm$ 0.61 <sup>a,A</sup>	10.17 $\pm$ 1.21 <sup>ah,B</sup>	0.13 $\pm$ 0.01 <sup>a,A</sup>	0.13 $\pm$ 0.01 <sup>a,A</sup>	0.20 $\pm$ 0.01 <sup>a,B</sup>
Leek (leaves)	5.19 $\pm$ 0.87 <sup>a,A</sup>	5.79 $\pm$ 0.92 <sup>a,A</sup>	5.89 $\pm$ 0.22 <sup>a,A</sup>	0.13 $\pm$ 0.01 <sup>a,A</sup>	0.19 $\pm$ 0.01 <sup>b,B</sup>	0.25 $\pm$ 0.04 <sup>b,C</sup>
Cornelian cherry (seed)	81.95 $\pm$ 7.66 <sup>e,A</sup>	76.44 $\pm$ 2.48 <sup>g,A</sup>	77.44 $\pm$ 6.86 <sup>g,A</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>g,A</sup>	0.71 $\pm$ 0.01 <sup>e,A</sup>
Dog Rose (seeds)	36.16 $\pm$ 2.22 <sup>c,B</sup>	31.74 $\pm$ 2.09 <sup>d,A</sup>	45.40 $\pm$ 0.05 <sup>d,C</sup>	0.53 $\pm$ 0.01 <sup>d,B</sup>	0.44 $\pm$ 0.01 <sup>d,A</sup>	0.62 $\pm$ 0.01 <sup>d,C</sup>
White grape (marcs)	41.68 $\pm$ 5.25 <sup>c,A</sup>	40.91 $\pm$ 4.99 <sup>e,A</sup>	58.34 $\pm$ 6.18 <sup>e,B</sup>	0.67 $\pm$ 0.01 <sup>e,A</sup>	0.67 $\pm$ 0.01 <sup>e,A</sup>	0.70 $\pm$ 0.01 <sup>e,B</sup>
Red grape (marcs)	52.96 $\pm$ 9.96 <sup>d,A</sup>	62.84 $\pm$ 3.21 <sup>f,A</sup>	65.98 $\pm$ 4.03 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.71 $\pm$ 0.01 <sup>e,A</sup>
Dog Rose (pulp)	72.48 $\pm$ 1.11 <sup>d,A</sup>	73.79 $\pm$ 2.34 <sup>d,A</sup>	74.37 $\pm$ 2.57 <sup>d,A</sup>	0.51 $\pm$ 0.02 <sup>e,B</sup>	0.40 $\pm$ 0.02 <sup>c,A</sup>	0.71 $\pm$ 0.01 <sup>e,C</sup>
Cornelian cherry (pulp)	59.03 $\pm$ 10.22 <sup>c,B</sup>	41.98 $\pm$ 3.51 <sup>c,A</sup>	54.44 $\pm$ 4.88 <sup>c,A</sup>	0.71 $\pm$ 0.01 <sup>f,A</sup>	0.69 $\pm$ 0.01 <sup>d,A</sup>	0.70 $\pm$ 0.01 <sup>e,A</sup>
Pomegranate (pomace)	26.19 $\pm$ 1.23 <sup>b,B</sup>	16.71 $\pm$ 3.08 <sup>b,A</sup>	24.91 $\pm$ 3.50 <sup>b,B</sup>	0.37 $\pm$ 0.02 <sup>d,A</sup>	0.37 $\pm$ 0.07 <sup>c,A</sup>	0.46 $\pm$ 0.03 <sup>d,A</sup>
Apple (pomace)	7.67 $\pm$ 0.28 <sup>a,B</sup>	5.55 $\pm$ 0.66 <sup>a,A</sup>	8.73 $\pm$ 1.06 <sup>a,B</sup>	0.26 $\pm$ 0.03 <sup>c,A</sup>	0.28 $\pm$ 0.01 <sup>b,AB</sup>	0.32 $\pm$ 0.02 <sup>c,B</sup>
“Viola” potato (pulp)	8.48 $\pm$ 0.17 <sup>a,C</sup>	4.66 $\pm$ 0.24 <sup>a,A</sup>	6.93 $\pm$ 0.03 <sup>a,B</sup>	0.11 $\pm$ 0.01 <sup>b,A</sup>	0.14 $\pm$ 0.01 <sup>a,B</sup>	0.17 $\pm$ 0.01 <sup>b,C</sup>
“Desiree” potato (pulp)	4.88 $\pm$ 0.10 <sup>a,A</sup>	5.58 $\pm$ 1.29 <sup>a,A</sup>	5.81 $\pm$ 0.27 <sup>a,A</sup>	0.07 $\pm$ 0.01 <sup>a,A</sup>	0.11 $\pm$ 0.01 <sup>a,B</sup>	0.12 $\pm$ 0.01 <sup>a,B</sup>
“Piatlina” potato (pulp)	4.75 $\pm$ 0.08 <sup>a,A</sup>	4.01 $\pm$ 0.35 <sup>a,B</sup>	4.01 $\pm$ 0.13 <sup>a,A</sup>	0.08 $\pm$ 0.01 <sup>a,A</sup>	0.15 $\pm$ 0.07 <sup>a,A</sup>	0.10 $\pm$ 0.01 <sup>a,A</sup>

<sup>†</sup> Trolox Equivalent

<sup>††</sup> Solvent A = methanol (90%) in acidified water; solvent B = ethanol (80%) in water; solvent C = acetone (70%) in water.

<sup>a-h</sup> Values within column by the same letters are not significantly different at  $p \leq 0.05$ .

<sup>A-C</sup> Values within row by the same letters are not significantly different at  $p \leq 0.05$ .

**Table 4.** Antibacterial activity of extracts obtained with the three solvents and evaluated by the agar-well diffusion assay.

The concentration was 20 mg of dry extract/ml.

Solvent <sup>†</sup>	Pomegranate (peel)			Pomegranate (pomace)			Apple (peel)			Apple (pomace)			Cornelian cherry (pulp)			Dog rose (pulp)		
	A	B	C	A	B	C	A	B	C	A	B	C	A	B	C	A	B	C
<i>Salmonella</i> Enteritidis O:103	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+
<i>Pseudomonas fluorescens</i>	++	-	-	+	-	-	+	+	+	+	-	-	-	-	-	-	-	-
<i>Pseudomonas aeruginosa</i>	+	+	+	-	-	-	+	+	+	-	-	-	+	-	-	-	-	-
<i>Escherichia coli</i> ATCC 35150	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Escherichia coli</i> DH5 $\alpha$	+	-	+	+	-	-	+	-	+	+	-	-	-	-	-	+	+	-
<i>Serratia marcescens</i>	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Staphylococcus aureus</i> ATCC 10527	+	+	++	+	+	+	-	+	+	-	-	+	-	-	+	+	+	+
<i>Listeria monocytogenes</i> NCTC 10527	+	+	+	-	-	-	+	+	+	+	+	+	-	-	-	-	-	-
<i>Bacillus cereus</i> DSM 350	+	-	+	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-
<i>Lactococcus lactis</i> DSM 4366	+	-	+	+	-	+	+	+	+	+	-	+	+	+	+	+	+	+
<i>Lactobacillus sakei</i> DSMZ 6333	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Staphylococcus xylosus</i>	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

<sup>†</sup> Solvent A = methanol (90%) in acidified water; solvent B = ethanol (80%) in water; solvent C = acetone (70%) in water.

<sup>a</sup> -: inhibition zone  $\leq$ 1 mm, +: 1 mm < inhibition zone  $\leq$ 3mm, ++: inhibition zone >3 mm