

Faculty of Resource Science and Technology

GENETIC DIVERSITY OF ANTIBIOTIC RESISTANT ESCHERICHIA COLI FROM CULTURED CATFISH (CLARIAS BATRACHUS) AND THE ENVIRONMENT

Marian Madonna Mobilik (18950)

QR 82 E6 M687 2010

Bachelor of Science with Honours (Resource Biotechnology) 2010

UNIVERSITI MALAYSIA SARAWAK

		THESIS STA	ATUS ENDORSEMENT I	FORM
TITLE	ESCHERUCHIA LOI		TIBIOTIC SUSCEPTIBI ULTURED (ATPISH (
		ACADEM	IIC SESSION: 2007/20	0
L	MARIAN MADO		CAPITAL LETTERS)	
	nereby agree that this The Sarawak, subject to the fo			ormation Services, Universiti Malaysia
	1. The Thesis is solely	owned by Universiti	Malaysia Sarawak	
	2. The Centre for Acad	emic Information Ser	rvices is given full rights to pr	oduce copies for educational purposes only
	 The Centre for Acad database 	emic Information Ser	rvice is given full rights to do	digitization in order to develop local content
4				oduce copies of this Thesis as part of its e purpose of interlibrary loan between HLIJ
	5. ** Please tick (√)			
	CONFIDENTIAL			OFFICIAL SECRETS ACT 1972)
	RESTRICTED	(Contains restricted was conducted)	l information as dictated by th	e body or organization where the research
	UNRESTRICTED			
Al	eu (ma)			Validated by Sauce
	Ř'S ŠIGN ATŪRE)			(SUPERVISOR'S SIGNATURE)
LOT 8 TMN C 88300	HADDRONG II PHS LORONG II PRIENTAL KOTA KINABAW 2.06.2010			Date: 04-07-2010.
				Date.

Notes * Thesis refers to PhD, Master and Bachelor Degree
** For Confidential or Restricted materials, please attach relevant documents from relevant organizations/ authorities

UNIVERSITI MALAYSIA SARAWAK

P.KHIDMAT MAKLUMAT AKADEMIK

1000212240

Genetic Diversity of Antibiotic Resistant Escherichia coli from Cultured Catfish (Clarias batrachus) and the Environment

Marian Madonna Mobilik (18950)

A thesis submitted in partial fulfillment of the requirements for the degree of Bachelor of
Science with Honours
(Resource Biotechnology)

Supervisor: Dr. Samuel Lihan Co-Supervisor: Dr. Lesley Maurice Bilung

> Resource Biotechnology Program Department of Molecular Biology

Faculty of Resource Science and Technology Universiti Malaysia Sarawak 2010 I dedicate this work to my parents, Julius Mobilik and Mariana Norman and to my whole family who have given trust and have supported me in my education and to Eddy Jamilih for giving the strength and faith for me to believe in myself. I would not make it through if it was not because of friends who have dearly journeyed together.

Thank you from the bottom of my heart.

ACKNOWLEDGEMENT

Many thanks to all those, too numerous to mention individually here, who generously

contributed their knowledge and experience in completion of this study. I would like to

extend my appreciation and thanks to all the Resource Biotechnology Program lecturers

and course mates who provided guidance and support, especially to Prof Dr Edmund

Sim, Dr. Awang Ahmad Salihin, Assoc Prof Dr. Kasing Apun and Dr. Lesley Maurice

Bilung, Particular thanks to all Masters Students and lab assistants who gave advice and

relevant personal observation. Most of all, my heartfelt gratitude to Dr. Samuel Lihan for

his assistance, patience and advice in teaching me many aspects of this study which

includes the significance of microbiology in improving human health

Marian Madonna Mobilik

Kuching

5 May 2010

iii

TABLE OF CONTENTS

Content		Page
		30.00
Acknowled	gement	iii
List of App	endix	vi
List of Tabl	es	vii
List of Figu	res	viii
List of Abb	reviation	xi
Abstract		1
Abstrak		3
Chapter 1: Introduction		5
1.1	Objective	5
Chapter 1: Introduction 1.1 Objective Chapter 2: Literature Review 2.1 Aquaculture 2.2 E. coli		6
2.1	Aquaculture	6
2.2	E. coli	6
2.3	Antibiotic Resistant	7
2.4	PCR	9
2.5	RAPD-PCR	9
2.6	Catfish (Clarias batrachus)	11
2.7	Public Health	11
Chapter 3: Materials and Methods		12
3.1	Sample Collection	12
	3.1.1 Fish Sampling	12

	3.1.2	Water Sampling	12
	3.1.3	Sediment Sampling	12
3.2	Isolatio	n and Identification of Isolates	13
	3.2.1	Serial Dilution	13
	3.2.2	Plating of Sample	13
	3.2.3	Bacterial Identification	14
		3.2.3.1 IMViC test	14
		3.2.3.2 Gram Staining	15
3.3	Antibio	tic Resistance and Susceptibility	15
3.4	PCR		16
	3.4.1	DNA Preparation	16
	3.4.2	Detection of stx1 and VT2 gene	16
3.5	3.5 RAPD-PCR Analysis		17
3.6 RAPD-PCR Fragment Analysis		18	
Chapter 4: Result			19
4.1	Result		19
	4.1.1	IMViC test	21
	4.1.2	Gram Staining	22
	4.1.3	Antibiotic Susceptibility Test	23
	4.1.4	Detection of stx1 and VT2 gene	27
	4.1.5	RAPD-PCR Analysis	30
4.2	Discussi	on	39
Chapter 5: C	onclusion		45
Chapter 6: R	eferences		46

LIST OF APPENDIX

Appendix	Explanatory	Page	
1	Gram staining procedure	51	
2	Functions of Antibiotics	52	
3	Reagents for PCR and Primer for the PCR assay	53	
4	Oligonucleotide for RAPD-PCR assay and Reagents for	54	
	RAPD-PCR		
5	Oligonnucleotide sheet for PCR	55	
6	Oligonnucleotide sheet for RAPD-PCR	56	
7	100bp DNA ladder	57	
8	Go Taq ® Flexi DNA Polymerase	58	
9	dNTP Mix	59	

LIST OF TABLES

Table	Explanatory	Page
1	E. coli strains that were taken from several location and type	20
2	Resistance of E. coli strains towards types of antibiotics used in this study	23
3	Frequencies of resistant and susceptible of E. coli strains towards each	25
	antibiotic	
4	Genotypic diversity of <i>E.coli</i> amplified using RAPD-PCR profile obtained	33
	with primer GEN_1_50_09 and GEN_1_50_10.	
5	Genotypic diversity of E. coli using RAPD-PCR profile obtained with	36
	primer combinations of GEN_1_50_09 and GEN_1_50_10	

LIST OF FIGURES

Figure	Explanatory	Page
1	Positive and negative results of Simmon Citrate and Indole in IMViC	21
	test.	
2	Gram negative bacilli: one of the microscopic pictures of isolates that	22
	was taken after gram staining. The microscope magnification is	
	100x10 with the use of immersion oil.	
3	Percentage resistance and susceptibility according to type of antibiotic	26
4	The electrophoresis patterns of PCR detection of Escherichia coli	28
	isolates electrophoresed on 1.0 % agarose gel using stx1 primer. M,	
	Molecular weight sizes (base pairs, bp) are indicated by numbers on	
	the left; lane 1-11: EC1, EC2, EC3, EC4, EC5, EC6, EC7, EC8, EC9,	
	EC10, EC11. Lane M: 100bp DNA ladder marker, lane NC: Negative	
	control, lane PC: Positive control	
5	The electrophoresis patterns of PCR detection of Escherichia coli	28
	isolates electrophoresed on 1.0 % agarose gel using stx1 primer. M,	
	Molecular weight sizes (base pairs, bp) are indicated by numbers on	
	the left; lane 1-6: EC12, EC13, EC14, EC15, EC16 and EC17. Lane	
	M: 100bp DNA ladder marker, lane PC: Positive control	

The electrophoresis patterns of PCR detection of Escherichia coli isolates electrophoresed on 1.0 % agarose gel using VT2 primer. M, Molecular weight sizes (base pairs, bp) are indicated by numbers on the left; lane 1-11: EC1, EC2, EC3, EC4, EC5, EC6, EC7, EC8, EC9, EC10, EC11. Lane M: 100bp DNA ladder marker, lane NC: Negative control, lane PC: Positive control

29

31

- The electrophoresis patterns of PCR detection of Escherichia coli 29 isolates electrophoresed on 1.0 % agarose gel using VT2 primer. M, Molecular weight sizes (base pairs, bp) are indicated by numbers on the left; lane 1-6: EC12, EC13, EC14, EC15, EC16 and EC17. Lane M: 100bp DNA ladder marker, lane NC: Negative control, lane PC: Positive control
- 8 RAPD-PCR fingerprinting of *E. coli* isolates obtained with primer 30
 GEN_1_50_09 electrophoresed on 1.0% agarose gel. Lane M: 1kb
 DNA ladder (molecular weight in base pair, bp); lane 1-12: EC1, EC2,
 EC3, EC4, EC5, EC6, EC7, EC8, EC9, EC10, EC11 and EC12
- 9 RAPD-PCR fingerprinting of *E. coli* isolates obtained with primer

 GEN_1_50_09 electrophoresed on 1.0% agarose gel. Lane M: 1kb

 DNA ladder marker (Molecular weight in kilo base pair, bp); lane 1-5:

 EC13, EC14, EC15, EC16 and EC17. Lane NC: Negative Control.
- 10 RAPD-PCR fingerprinting of *E. coli* isolates obtained with primer

 GEN_1_50_10 electrophoresed on 1.0% agarose gel. Lane M: 1kb

 DNA ladder marker; lane 1-12: EC1, EC2, EC3, EC4, EC5, EC6,

 EC7, EC8, EC9, EC10, EC11 and EC12

11	RAPD-PCR fingerprinting of E. coli isolates obtained with	32
	combination of primer GEN_1_50_09 and GEN_1_50_10	
	electrophoresed on 1.0% agarose gel. Lane M: 1kb DNA ladder	
	marker (molecular weight in base pair, bp); lane 1-12: EC1, EC2,	
	EC3, EC4, EC5, EC6, EC7, EC8, EC9, EC10, and EC11.	
12	RAPD-PCR fingerprinting of E. coli isolates obtained with primer	32
	GEN_1_50_09 electrophoresed on 1.0% agarose gel. Lane M: 1kb	
	DNA ladder marker (molecular weight in base pair, bp); lane 1-6:	
	EC12, EC13, EC14, EC15, EC16 and EC17. Lane PC: Positive control	
13	Dendogram generated from the RAPD-PCR fingerprinting among 17	35
	isolates of the E. coli with primer GEN_1_50_09	
14	Dendogram generated from the RAPD-PCR fingerprinting among 17	38
	isolates of the E. coli with combination of primer GEN_1_50_09 and	
	GEN 1 50 10.	

LIST OF ABBREVIATIONS

CDC -Centre for Disease Control and Prevention

DNA -Deoxyribonucleic acid

E. coli -Escherichia coli

EC -Escherichia coli

EDTA - Ethylenediaminetetraacetic acid

EHEC -Enterohemorrhagic E. coli

EMBA - Eosine Methylene Blue Agar

EPEC -Enteropathogenic E. coli

ETEC -Enterotoxigenic E. coli

EtOH - Ethyl alcohol

IMViC test -Indole, Methyl red, Voges-Proskauer, Citrate test

MHA -Muellër Hinton Agar

NA - Nutrient Agar

NCCLS -National Committee for Clinical Laboratory Standards

OD -optical density

PCR -Polymerase Chain Reaction

RAPD-PCR -Random Amplified Polymorphic DNA - PCR

TBE - Tris/Borate/EDTA

TSA - Typticase Soy Agar

TSB - Tryptic Soy Broth

UV -Ultraviolet

FAO - Food and Agriculture Organization

UNO - United Nations Organization

Genetic Diversity of Antibiotic Resistant Escherichia coli from Cultured Catfish (Clarias batrachus) and the Environment

Marian Madonna Mobilik (18950)

Resource Biotechnology Programme Faculty of Science and Technology Universiti Malaysia Sarawak

ABSTRACT

Human gastrointestinal illness caused by *E. coli* has been known to occur due to the presence of shiga toxin and verotoxin gene carried by this bacteria. This food borne pathogen contaminates food through improper handling of food and also caused by poor practice of hygiene. However, the spread of this bacterium is reaching the point in which it inhabits in our food source, fish. Fish (catfish) grown in fish farms are treated with supplemented feeds to enhance its growth and to avoid the fishes from being infected by disease. However, the undigested feed pollutes the environment and caused the *E. coli* to acquire genetic alteration that may be harmful to human. Samples of fish (catfish), water and sediement were collected from fish farms around Kuching area and were tested through a series of biochemical test, antibiotic resistance test, and specific PCR and RAPD-PCR. There were 12 isolates resistant towards at least one type of antibiotic tested. The highest percentage of resistance is towards ampicilin (23.5%), while the lowest percentage of resistance is towards nitfurantoin (11.8%). No *E. coli* strains were resistant towards norfloxacin, sulphametoxazole / trimethroprim and chloramphenicol. The PCR analysis revealed that 4 and 6 *E. coli* strains possessed shiga toxin gene and verotoxin gene, respectively. The RAPD analysis showed that 9 RAPD fingerprinting profile were detected with the use of

primer GEN 1 50 09, one RAPD fingerprinting profile with the use of primer GEN 1 50 10

and 12 RAPD fingerprinting profiles with the use of combined primers (GEN 1 50 09 and

GEN 1 50 10). This proves that the use of combined primers have higher power of

discrimination. The results of this study suggested that the E. coli strains have multiple

resistances towards commonly used antibiotics and these strains are genetically diverse. The

occurrence of antibiotic resistance and shiga toxin and verotoxin possession among the E. coli

may pose potential hazard to human health.

Keywords: E. coli, antibiotic resistance, RAPD-PCR

Pengagihan Genetik terhadap Ketahanan Antibiotik Escherichia coli daripada Kultur Ikan Keli (Clarias batrachus) dan Persekitarannya

> Marian Madonna Mobilik (18950)

Program Bioteknologi Sumber Fakulti Sains dan Teknologi Sumber Universiti Malaysia Sarawak

ABSTRAK

Penyakit berkaitan perut dan usus manusia yang disebabkan oleh E. coli adalah kerana ia memiliki gen untuk penghasilan toksin shiga dan toksin vero. Patogen yang disebarkan melalui makanan ini berlaku dengan cara pengendalian makanan yang salah dan tidak mengamalkan kebersihan semasa menyediakan makanan. Penyebaran bakteria ini kini diketahui mendiami salah satu punca makanan manusia iaitu ikan. Ikan keli yang dipelihara dan dikomersilkan di dalam kolam telah diberi makanan tambahan untuk mempercepatkan tumbesaran ikan dan meningkatkan ketahanan ikan tersebut terhadap serangan penyakit. Sisa makanan yang tidak dicerna mencemarkan habitat ikan tersebut dan menyebabkan E. coli dalam habitat itu mengalami perubahan gen yang membahayakan kepada manusia. Sampel ikan keli, air dan mendapan telah diambil daripada kawasan penternakan ikan keli di sekitar Kuching dan diuji melalui beberapa peringkat ujian biokimia, ujian daya tahan antibiotic dan PCR. Selain itu, sebanyak 12 pencilan E. coli menunjukkan daya tahan terhadap sekurang-kurangnya satu antibiotik. Peratus ketahan yang tertinggi adalah terhadap ampicilin (23.5%), manakala peratus ketahan yang paling rendah adalah terhadap nitrofurantoin (11.8%). Semua pencilan tidak mempunyai ketahan terhadap norfloxacin, sulphametoxazole / trimethroprim and chloramphenicol Sebanyak 4 dan 6 pencilan E. coli masing-masing didapati memiliki gen toksin shiga dan gen toksin vero. Kaedah profil dan amplifikasi DNA polimorfik secara rawak (RAPD-

PCR) telah digunakan untuk menganalisis perbezaan genetik 17 pencilan E. coli yang

dipencilkan daripada kolam ternakan ikan keli. Dalam kalangan pencilan tersebut, terdapat 9

profil RAPD daripada pencetus GEN 1 50 09, 1 profil RAPD daripada pencetus

GEN 1 50 10 dan 12 profil RAPD daripada gabungan dua pencetus (GEN 1 50 09 dan

GEN 1 50 10). Ini menunjukkan bahawa penggunaan gabungan pencetus ternyata lebih

sensitif. Hasil kajian ini mencadangkan bahawa pencilan E. coli mempunyai ketahanan terhadap

beberapa antibiotik yang biasanya digunakan dalam bidang perubatan dan pencilan ini juga

mempunyai kepelbagaian genetik. Ketahanan terhadap antibiotik dan kehadiran gen toksin shiga

dan toksin vero di dalam pencilan E. coli menunjukkan bahawa ia mempunyai potensi unutk

membahayakan kesihatan manusia.

Kata kunci: E. coli, ketahanan antibiotik, RAPD-PCR

4

UNIVERSITI MALAYSIA SARAWAK

CHAPTER 1

1.0 INTRODUCTION

Escherichia coli (E. coli) is a gram-negative, non-spore-forming, rod-shaped bacterium. It is also a facultative anaerobe that can be found in the large intestines of most warm-blooded animals (Leclerc et. al., 2001). Although most strain are harmless, others can cause serious food poisoning, urinary tract infection, respiratory illness, and pneumonia (Weinstein et. al., 1988). Food borne pathogen increases the risk of human consuming harmful toxins. The resident strain that lives in the human intestine is not dangerous but perturbs if there is an enteric infection (Nataro and Kaper, 1998). What differentiates the resident strains from other of its serotype is the virulence characteristic. Study shows that strains that causes food poisoning and other illness produces toxin that has an irreversible cytopathic effect on cultured vero cells (Grant, 2004). Antibiotics are widely used in aquaculture for therapeutic purposes and as prophylactic agents. However, the use of antibiotics will cause environmental organism, endogenous bacterial population of farmed species and pathogens to have transfer of resistance which jeopardizes human and livestock (FAO, 2006). It was recognized that the transfer of resistant microorganism from animals to human is through food chain (European Food Safety Authority, 2008). Therefore, if microorganism that infects human is readily resistant to the present medication, new drug must be discovered to overcome it (Sorum and Sunde, 2001).

1.1 OBJECTIVES

The objective of this study is to isolate *E. coli* from the intestine of catfish (*Clarias batrachus*), the water and sediment of the production pond and to characterize the isolates by antimicrobial susceptibility testing, Shiga toxin detection, and genetic diversity analysis.

CHAPTER 2

2.0 LITERATURE REVIEW

2.1 Aquaculture

According to De Silva and Anderson (1995), the FAO of the UNO explains aquaculture as the farming of aquatic organism that involves certain processes, including the usage of growth hormone to increase production. The rate of production of freshwater fish increased since the introduction of aquaculture. This is supported by De Silva and Anderson in 1995 in their research that reported the quantity of freshwater fish produced from 1985 to 1989 have increased by 15.8% per annum compared to only 5.9% in 1985.

2.2 E. coli

E. coli is a commensal organism that lives in mammals and it is usually isolated from the mammalian feces or large intestine (Russell et. al., 2000). At some degree, it produces vitamin K for the host that makes it a symbiont (Ingledew and Poole, 1984). Certain strains are harmless but other strain may cause fever in infants and food poisoning in adults (Koch, 1995). Hence, this made E. coli a pathogen but an opportunistic one (Koch, 1995). Other than that, E. coli is a suitable model organism because it can multiply rapidly and its multiplication can be controlled in culture plates (Kaper, 2005). Diarrhoeagenic E. coli pose a worldwide threat to human health. The detection of enterotoxigenic strains of this bacterium usually requires biochemical or immunological testing of isolated strains, which are obtained by cultivation from food or environmental samples. Since most of the genes encoding E, coli enterotoxins are located on plasmids and selective enrichment steps may lead to plasmid loss, such detection strategies can yield false negative results (Dreyfus et al. 1983; Candrian et al. 1991a). A protocol based on

PCR that allows the detection of enterotoxigenic *E. coli* with the genetic potential for production of heat-labile toxin type I (LTI) in minced meat has been described by Wernars *et al.* (1991a). However, this procedure requires several cultivation steps and may detect only the LT1 allele, which was originally found in an *E. coli* of porcine origin. In contrast, PCR test that specifically recognizes DNA sequences of the malB operon of *E. coli* (Candrian *et al.* 1991b) was used to develop a PCR-based analysis method without cultivation steps for surface water and soft cheese. The protocol was applied to a second PCR test specific for both known alleles of LTI enterotoxigenic *E. coli* (Furrer *et al.* 1990) to determine the presence of such strains in soft cheese samples containing high levels of *E. coli*,

2.3 Antibiotic Resistant

Resistance of bacteria towards antibiotics is caused by prolong use of the antibiotics for a certain period of time which causes the bacteria to acquire resistance through natural selection or by evolutionary stress when antibiotics are given to a particular population. Many bacteria transfer the resistance capability via horizontal gene transfer: conjugation, transformation and transduction. Previous study on antibiotic resistance shows that genes that carry gene for resistance are usually found in the plasmid (circular DNA molecule that is separated from genomic DNA). Some bacteria carries resistance towards several different antibiotics which are termed bacteria with multiple resistance. This is one of the basic factors that small organisms like bacteria and virus do to continue to exist and multiply. The rate of evolution in bacteria is very fast which is one of the reasons a person can be infected with flu several times a year; same virus but different strain. It is a global issue in which emergence of antibiotic resistance not only

contribute to public health risk and increase in treatment cost but worst, it results in morbidity and motility due to treatment failure.

There are three methods for antibiotic susceptibility test: diffusion, dilution as well as diffusion and dilution method. For the purpose of this study, Kirby-Bauer disk diffusion method was used in the antibiotic susceptibility test. In this method, small disk containing a known amount of antibiotic is placed on an agar plate. After incubation, zone of inhibition can be seen depending on the level of resistance towards the antibiotics. The zone of inhibition is a clear area around the disc which the bacteria are incapable to grow. Each agar plate contains a single type of bacteria and is tested using six different types of antibiotic discs. The type of antibiotic used in this study was selected based on the possibility of antibiotic used in fish farming.

E. coli is one of the bacterium that exists naturally in the large intestine of human and other animals which facilitates in suppressing the growth of other harmful bacteria and also aids in the synthesis of some vitamins. The presence of E. coli in food indicates that the food is contaminated with other harmful bacteria such as Salmonella and Hepititis A virus. Alteration of gene in E. coli causes these bacteria to acquire similar toxin produced by Shigella dysenteriea known as Verotoxin and Shiga toxin that causes damage in the lining of the intestine which results in diarrhea and urinary tract infection.

Feeds given to fishes may be technologically derived from chemical or other harmful substances that are used in order to have cheap feeding cost. The feeds that were used contained anti nutrients that could affect not only the health of fishes but also the health of consumers and the

environment (Francis et. al., 2001) by having the natural microflora in fish intestine to be resistant towards antibiotic that is being used in the feeds (Sorum and Sunde, 2001). When human consumes the fishes and experience diarrhea (due to the excessive amount of E. coli), the current antibiotic will not be able to kill the pathogenic E. coli because it is already resistant to it (Martinez, 2009). As a result, sick people cannot be cured by means of medication and new drugs or antibiotic will have to be generated.

2.5 PCR

PCR technology has been applied to detect a wide range of micro-organisms in a variety of situations and although it does not appear to be feasible to apply the technology directly with some types of sample, it does offer the possibility to reduce the time taken to obtain a result (Fricker and Fricker, 1994). There has been considerable interest in the application of PCR technology to the detection of bacteria (Bej et. al., 1991), protozoan cysts and viruses in water. This is particularly true for detection of pathogens in water where the procedures are often laborious, time-consuming and incentives. However, the water industry relies to a large extent on the use of indicator organism to monitor water quality and much of the microbiology effort required is concentrated in this area. In particular, E. coli and the coliform group are used to indicate faecal and a lack of system integrity.

2.6 RAPD-PCR

One way to determine the similarities or differences in strains that are being isolated in a study is to use RAPD-PCR. RAPD markers are a modification of PCR contrived in the late 1980's (Wiliams et al., 1990). The technique is one of the best available DNA-based tools for scoring

variations between cultivars within species (Lakshmikumaran and Bhatia, 1998). This is done by using arbitrary primers to detect regions in the genomic DNA that are complimentary to it.

Hence, the genetic diversity can be identified to in a certain population. Identifying the genetic diversity is an important key in analyzing specific bacteria because one type of bacteria may have a variety number of strain. These different strains, although the same type of bacteria, have different characteristics. One strain can be beneficial to other strains or may be more hazardous. The factor that differ strains of the same type of bacteria may be as simple as a sequence of gene that codes for protein or the genetic make up of the plasmid in which a strain carries. For example, different *E. coli* strain has different degree of pathogenicity to humans and other animals. Some of it may even be beneficial to human such as the *E. coli* strain that inhabits the large intestine of humans and some mammals but there is also strain that is hazardous to human health. Thus, RAPD-PCR is a good molecular typing for distinguishing genomic diversity and this has been frequently used in bacterial study to discriminate bacteria into genus, species and strain. Some arbitrary primers can distinguish different strains effectively; others may not even present any bands. Hence, optimization is required in order to get result that can be analyzed.

This can be done by using the randomly designed primer to detect Usage of certain primers will result in the formation of a variety of bands that can tell the similarities or differences in several bacterial strain. One probable disadvantage, however, is the degree of reproducibility of these markers which can sometimes be low (e.g., Muralidharan and Wakeland, 1993; Ellsworth et al., 1993; Skroch and Nienhuis, 1995). This is due to the RAPD banding patterns which is highly sensitive to reaction conditions. A slight change in PCR conditions will result in a different band result. Other than that, one may face difficulty in exactly replicating reaction conditions across laboratories, where different brands of thermocyclers may be used.

Despite of all of the disadvantages, RAPD-PCR result can be optimized by adjusting the PCR conditions. The process require much cheaper cost compared to other method, it requires minimal labour and does not involve hazardous chemicals.

2.7 Catfish (Clarias batrachus)

Clarias batrachus or also known as the walking catfish is widely distributed in Asia and it lives in muddy and marshy waters (Garg et. al., 1995). Some people regard this walking catfish as a delicacy but others regards this as invaders to aquaculture farms that feed on culture fish. This fish has been described as a benthic, nocturnal, tactile omnivore that consumes detritus and opportunistically forages on large aquatic insects, tadpoles, and fish (Smithsonian Marine Station, 2007). It is reared locally in Malaysia for people to cook as a local delicacy.

2.8 Public Health

The pathogenic strains of *E. coli* can cause distinct disease syndrome such as diarrheal diseases, wound infection, meningitis, septicemia, atherosclerosis, hemolytic uremic syndrome and immunological diseases such as reactive and rheumatoid arthritis (Olsvik *et al.*, 1991). The ETEC strains produce one or more of toxins from the heat-labile and the heat-stable enterotoxin families (Levine *et al.*, 1985). These strains possess specific adhesion fimbria for intestinal attachment and colonization (Olsvik *et al.*, 1991). Some EPEC strain produces one or more of the cytotoxins, but adheres also to intestinal cells interfering with the electrolyte transport system (Olsvik *et al.*, 1991). The EHEC strains have been identified and shown to produce one or more of the cytotoxins (vero-cytotoxins, shiga-like toxins) (Olsvik *et al.*, 1991).