

TECHNICAL ADVANCE

Characterization of the ethanol-inducible *alc* gene-expression system in *Arabidopsis thaliana*

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Summary

Controlled expression of transgenes in plants is key to the characterization of gene function and the regulated manipulation of growth and development. The *alc* gene-expression system, derived from the filamentous fungus *Aspergillus nidulans*, has previously been used successfully in both tobacco and potato, and has potential for use in agriculture. Its value to fundamental research is largely dependent on its utility in *Arabidopsis thaliana*. We have undertaken a detailed function analysis of the *alc* regulon in *A. thaliana*. By linking the *alcA* promoter to β -glucuronidase (GUS), luciferase (*LUC*) and green fluorescent protein (*GFP*) genes, we demonstrate that *alcR*-mediated expression occurs throughout the plant in a highly responsive manner. Induction occurs within one hour and is dose-dependent, with negligible activity in the absence of the exogenous inducer for soil-grown plants. Direct application of ethanol or exposure of whole plants to ethanol vapour are equally effective means of induction. Maximal expression using soil-grown plants occurred after 5 days of induction. In the majority of transgenics, expression is tightly regulated and reversible. We describe optimal strategies for utilizing the *alc* system in *A. thaliana*.

Keywords: *Arabidopsis thaliana*, ethanol, chemically inducible expression, plant-expression system, *Aspergillus nidulans*.

Introduction

A key tool in plant molecular biology is the development of effective gene-expression systems. The expression of a transgene can be achieved by using constitutive promoters such as the viral CaMV35S promoter (Odel *et al.*, 1985). However, a constitutive promoter is unsuitable when dealing with genes for which inappropriate expression is either highly deleterious or lethal. Additionally, such expression systems are unsuitable for studies where precise temporal regulation is required, for example, where expression of a gene is desired at a specific stage of plant development, or for analysis of mRNA decay properties. Biotechnological application of a regulated

expression system may also be important for crop plants; for instance, the conditional expression of pesticides or herbicide resistance; the induction of synchronous flowering of plants; and the production of a conditional male sterility system. In such cases an effective regulated gene-expression system is required.

The optimal system would employ an inexpensive, non-toxic inducer whose application can be fully controlled, and would lead to a reversibly dose-dependent expression with the potential to achieve high levels of gene expression but with negligible basal activity. To these ends, a number of regulated gene-expression systems have been