

1 **Two new Malagasy species of genus *Piper* L. (Piperaceae): *Piper malgassicum* and**
2 ***Piper tsarasotrae* and their phylogenetic position**

3

4 Enrico PALCHETTI¹, Stefano BIRICOLTI¹, Massimo GORI¹, Giulia ROTA
5 NODARI¹, Nicola GANDOLFI², Alessio PAPINI^{3,*}

6 ¹Department of Agrifood and Environmental Science (DISPAA), University of
7 Florence. Piazzale delle Cascine n. 18 – 50144 - Florence, Italy

8 ²Association Tsyriparma: Volunteer association for protection of Malagasy forests.
9 Ambositra, Madagascar

10 ³Dept. of Biology, Univ. of Florence. Via Micheli n. 3, Firenze, Italy

11 *Correspondence: alpapini@unifi.it

12

13 **Abstract:** We describe here two new species of genus *Piper* L. from Madagascar: *Piper*
14 *malgassicum* and *Piper tsarasotrae*, the species names referring to the currently known
15 distribution areas. These two species contribute, at least in part, to the production the
16 local voatsiperifery pepper, probably mixed together with *P. borbonense* and are,
17 therefore, economically relevant as spices. We used a selected set of characters (those
18 more easily observable on herbarium samples) for a Principal Component Analysis, to
19 assess the relative distance between both species, including in the analysis the
20 autochthonous species of *Piper* known from Africa and Madagascar. In order to check
21 the identity and to assess the phylogenetic position of the two species, we also
22 sequenced the chloroplast gene *ndhF* and the *trnL* intron and the nuclear gene G3PDH.
23 On the basis of these results, we show here the relationships between these two new
24 *Piper* taxa and the most closely related species within the genus (excluding *P. heimii*
25 and *P. pachyphyllum*, for which only morphological characters were available).

26

27 **Keywords:** DNA sequencing, Madagascar, Principal Component Analysis, *Piper*, *Piper*
28 *malgassicum*, *Piper tsarasotrae*, Piperaceae,

29

30 **1. Introduction**

31

32 Most recently, treatments of the pantropical genus *Piper* L. (Piperaceae) included more
33 than 2000 species (Quijano-Abril et al., 2006). The phylogenetic position of *Piper* L.
34 and of family Piperaceae, was inserted within the complex basal group of dicots termed
35 “paleoherbs” (Loconte and Stevenson, 1991). More recently, APG IV (2016) inserted
36 Piperaceae in order Piperales, nested within Magnoliids.

37 The distribution of *Piper* is pantropical and the genus develops highly variable growth
38 forms and biomechanical organization (Isnard et al., 2012). The highest number of
39 species can be found in America, where earlier 500 species were listed (Burger, 1972;
40 Tebbs, 1993), then increased to at least 1100 (Jaramillo et al., 2008) and most recently
41 up to 1804 (Ulloa Ulloa et al., 2017).

42 The exact number of *Piper* species and their exact distribution is not easy to ascertain,
43 particularly due to the high number of taxa, some of which are difficult to distinguish
44 one from the other, resulting in many synonyms (Suwanphakdee et al., 2016).
45 Furthermore, some species are widespread, such as *P. umbellatum*, while others,
46 actively cultivated, escaped by accident and may have been naturalized, such as *P.*
47 *auritum*, *P. nigrum* or *P. methysticum* (Smith et al., 2008). Most species show a
48 restricted distribution area (Marquis, 2004; Quijano-Abril et al., 2006). As a matter of
49 fact, new species were recently described also from old herbarium collections (Görts-
50 Van Rijn and Callejas Posada, 2005).

51 Only two endemic species are currently known for the African continent, *P. guineense*
52 and *P. capense*. *Piper guineense* is a dioecious vine, relatively similar to the majority of
53 southwestern Asian species, whereas *P. capense* is a shrub with bisexual flowers, hence
54 resembling many species of the American continent (Smith et al., 2008). The
55 knowledge of the genus in Madagascar is far from being complete. Currently *P. heimii*
56 C. DC and *P. pachyphyllum* Baker are indicated for the island, while *P. borbonense*
57 (Miq.) C. DC. was described for the island called at that time Île Bourbon, currently La
58 Reunion (Weil et al., 2017), belonging to Mascarenes Islands, 600 km east of
59 Madagascar. Its presence in Madagascar is a matter of debate, even if De Candolle (De
60 Candolle, 1923; 1869) already assigned some samples from Madagascar and Mauritius

61 to this species (see appendix 1 about herbarium samples by the site
62 <http://www.caryologia.unifi.it/tjb/Appendix1.pdf>). However, this species is cultivated,
63 which makes it more difficult to assess its natural distribution.

64 Here, we describe two new species of *Piper* L. from Madagascar on the base of their
65 morphology and supported by molecular data. Both species are mixed with *P.*
66 *borbonense*, in the so-called high-quality spice voatsiperifery pepper.

67

68 **2. Materials and methods**

69 **2.1. Morphological characters analysis and PCA**

70 Herbarium samples were prepared, among which the type specimens were chosen (see
71 appendix 1 with herbarium samples saved at the permanent link
72 <http://www.caryologia.unifi.it/tjb/Appendix1.pdf>). A number of characters were
73 observed and measured (where necessary) with a stereomicroscope. The herbarium
74 samples were stored by the Tropical Herbarium of Florence (FT, Centro Studi Erbario
75 Tropicale, Università degli Studi di Firenze).

76 21 characters (those that showed variation) were coded in a matrix (Table 1) used as
77 input for the Principal Component Analysis with the software PAST 3.16 (Hammer et
78 al., 2001).

79 **2.1. Anatomical characters**

80 Inflorescence stems were cut with a blade, stained with 1% phloroglucinol (w/v) in 12%
81 HCl for 5 min and observed with a bright field light microscope to stain lignin (as in
82 Mosti et al. 2012).

83

84 **2.3. DNA extraction**

85 For the DNA extraction, leaf samples were collected from the tropical forest of Vohiday
86 (samples PNsv1-10, table 2) and from the Tsarasotra area (samples PNst1-10, table 2).
87 Plant tissue samples were conserved and transported inside 20 mL plastic tubes filled
88 with ethanol 96% (after Murray and Pitas, 1996; Bressan et al., 2014).

89 For DNA extraction, 40 mg of dry leaf sample were placed into a 2mL tube, together
90 with tungsten carbide beads, frozen in liquid nitrogen and finely ground in a tissue
91 homogenizer (Tissue Lyser ®, Qiagen). DNA was extracted using Invisorb Spin Plant
92 Mini kit (Stratec molecular®). Amplification of the *trnL* intron and the low copy
93 nuclear gene *G3pdh* followed respectively the protocols by Taberlet et al. (1991) and
94 Strand et al. (1997). A set of four primer pairs were designed using the chloroplast
95 genome sequence of *Piper kadsura* (GenBank: KT223569.1) to cover the entire *ndhF*
96 gene.

97 The InsTAclone PCR Cloning Kit was used to clone *G3pdh* (Thermo Scientific®). Ten
98 samples for each provenience were amplified using the universal primers GPD9R2 and
99 GPD9R4 (Olsen and Schaal, 1999). Up to 15 colonies for single cloned sample were
100 amplified using M13 primers. PCR products were purified using the QIAquick PCR
101 Purification Kit (Quiagen) and sent to the University of Florence internal sequencing
102 service CIBIACI. Manual correction and assembly of the sequences was performed
103 using the software programmes Multaline (Corpet, 1988) and MEGA7 (Kumar et al.,
104 2016).

105 The new DNA sequences produced during our investigation were deposited in Genbank
106 (Genbank accession numbers are indicated in Table 2).

107

108 **2.3. Phylogenetic analysis**

109 Together with the here produced new sequences, other sequences used are available in
110 Genbank, more specifically those of genus *Piper* used by Smith et al., (2008). We used
111 one species of *Peperomia* (*Peperomia pellucida*) and *Houttuynia cordata* as outgroups
112 on the basis of the phylogenetic analysis on Piperaceae by Jaramillo and Manos (2001)
113 and Wanke et al. (2007) showing that *Peperomia* is sister group to *Piper s.l.*, while
114 *Houttuynia* is more distantly related to both of these genera (see, for instance, Figure 5
115 in Wanke et al., 2007). Optimal multiple alignment was obtained with CLUSTALW
116 1.81 (Thompson et al., 1994). The matrices for each of the three gene sequences
117 employed were combined with the Python (Python version 2.6.4; Biopython 1.57)
118 program combinex2_0.py, written by A. Papini (Lewke Bandara et al., 2013; Simeone
119 et al., 2016), released under GPL licence and available at
120 www.unifi.it/caryologia/PapiniPrograms.html.

121 A maximum likelihood (Felsenstein, 1981) search was done by preliminarily using
122 MrMODELTEST 2.0 (Nylander, 2004) to evaluate the best likelihood model on the
123 basis of the Akaike information criterion (Akaike, 1974). The model was used as
124 settings for Bayesian Inference with the program MrBayes 3.4b4 (Huelsenbeck and
125 Ronquist, 2001; Ronquist et al., 2012). A maximum likelihood (ML) phylogenetic
126 analysis was done with RaxML (Stamatakis et al., 2012) and the resulting trees were
127 edited with Figtree (Rambaut, 2009). We mapped the support on the tree branches with
128 the results of the Bayesian phylogenetic analysis as follows: after the "burn-in" trees
129 were removed from the data set as in Papini et al. (2007; 2011). The remaining trees
130 were used to produce a 50% majority-rule consensus tree (with PAUP) in which the
131 percentage support was considered equivalent to Bayesian posterior probabilities.

132

133 **3. Results**

134 **3.1 Morphological characters analysis and PCA**

135 The characters used for the species description and for comparison were observed and
136 measured with a stereomicroscope on the herbarium samples of the two new species and
137 of the most closely related species of *Piper* (images of the samples and original
138 protologues in appendix 1: herbarium samples). In appendix 1
139 (<http://www.caryologia.unifi.it/tjb/Appendix1.pdf>) also a list with the investigated
140 samples (scanned samples, in the majority of cases) of other species is reported. The
141 characters were coded as numeric states (table 1) and analysed with PAST. The
142 Principal Component Analysis (PCA) was based on a set of characters, those most
143 variable and easily observable on herbarium samples. The results of the PCA analysis
144 are shown in Figure 1. The samples from Tsarasotra (from now on *P. tsarasotrae*) were
145 quite isolated, even if quite close to *P. guineense* and to the samples from Vohiday
146 (from now on *P. malgassicum*) and *P. heimii* (Figure 1). Figure 1 shows also that *P.*
147 *pachyphyllum* and *P. borbonense* are relatively close.

148 **3.2 Phylogenetic analysis**

149 The phylogenetic analysis (Figure 2) showed that *P. malgassicum* and *P. tsarasotrae*
150 are strictly related to each other and to *P. borbonense*, this last clustering together with
151 *P. malgassicum* with 100% Bayesian Support (BS). These two species formed a
152 monophyletic group with *P. tsarasotrae* with 70% BS. The sister group of this cluster
153 was the group of 5 sequences of the African species *P. guineense* (85% BS), while the
154 Asian species *P. caninum* formed the sister group to the former species, but with BS
155 less than 50% (Figure 2).

156 **3.3 Microscopy observations**

157 The observation with a stereomicroscope was useful for the observation of
158 micromorphological character of the flowers, necessary for the following description. In
159 *P. tsarasotrae* male flowers, usually three (very rarely four) stamens are present (Figure
160 3A) with two anthers with lateral apertures (Figure 3A1). In the female flowers, the
161 number of stigmas may vary from 3 to 4. In Figure 3B a case with three lobed stigmas
162 is shown. In *P. malgassicum* male flowers, stamens are sometimes solitary (Figure 3C)
163 and show two anthers with lateral apertures (Figure 3C1). In female flowers, stigmas are
164 most frequently three, sometimes four, still visible on the enlarged fertilized ovary
165 (Figure 3D). The stigmas are sessile (Figure 3D1).

166 The observation with the light microscope of cross sections of the stem showed that in
167 *P. tsarasotrae* two circles of bundles are present (Figure 4A): a group of larger more
168 internal bundles and an external group of smaller bundles (Figure 4B).

169 Also in *P. malgassicum* the inflorescence stem in cross section showed two circles of
170 bundles (Figure 4C): a group of larger more internal bundles and an external group of
171 smaller bundles (Figure 4D). In this species a continuous layer of sclerenchyma
172 enclosed the smaller bundles. (Figure 4C).

173

174 **3.4 Description of the two new species**

175 The morphological nomenclature here employed followed Simpson (2010).

176 *Piper tsarasotrae* Papini, Palchetti, Gori, Rota Nodari spec. nov.

177

178 Typus: Collectors Enrico Palchetti and Nicola Gandolfi for samples 1.1 A (female
179 samples, holotype) and 1.1.D (male sample, paratype, as defined in 9.6, ex. 5 of the
180 International Code of Nomenclature for algae, fungi, and plants: McNeill, 2012);

181 locality Tsarasotra, Ambositra region (Madagascar); localization: S20° 27' E47° 10'.

182 Deposited by Centro Studi Erbario Tropicale, Università degli Studi di Firenze (FT).

183

184 Similar to *Piper guineense* Schumach. & Thonn., but differing since the foliar basis is

185 uneven and acuminate instead of cordate. Dioecious. Shrub, sometimes epilithic and

186 sometimes creeping on the ground, swollen stem nodes. Leaves alternate. Shape oblong

187 ovate, 4.5-8 cm long and only 0.5-2 cm wide. Leaf apex acuminate, while the leaf base

188 is uneven and acuminate. Inflorescence leaf opposite, cylindrical and erect. Female

189 spikes 4-6 cm long, with a peduncle 1-2 cm long, with small sessile spirally arranged

190 flowers. Single ovary, 4 (rarely 3) lobed white stigma, covered by short appendages.

191 Male spikes 3-5 cm long, with a peduncle 1-2 cm long and stamens organized in groups

192 of three. Ripe fruit reddish and rounded, 0.4-0.7 mm long, fruit pedicel 0.8-1.2 cm.

193 Each fruit gives off a single rounded-shaped seed. Inflorescence stem in cross section

194 with two circles of bundles: a group of larger more internal bundles and an external

195 group of smaller bundles.

196 Living in arid forest. The environment of the species is shown in Figures 5A and 5B,

197 while the female cones are shown in Figures 5C and 5D. Fruits in Figure 5E. In Figure

198 5F, both the inflorescence and the fruits can be observed on the same individual.

199 Usually three (very rarely four) stamens present with two anthers with lateral apertures.

200 Number of stigmas from 3 to 4.

201

202 *Piper malgassicum* Papini, Palchetti, Gori, Rota Nodari spec. nov.

203

204 Typus: Collectors Enrico Palchetti and Nicola Gandolfi: PS9a (female sample,
205 holotype) sample and PS8 sample (male sample, designed as paratype, as defined in art.
206 9.6, ex. 5 of the International Code of Nomenclature for algae, fungi, and plants:
207 McNeill, 2012); locality Vohiday forest, Ambositra region (Madagascar); localization
208 S20° 32' E47° 35'. Deposited by Centro Studi Erbario Tropicale, Università degli Studi
209 di Firenze (FT).

210 Similar to *Piper borbonense* (Miq.) C. DC. but differing since its foliar basis is uneven
211 and roundish instead of cordate. Dioecious. Liana climbing up to 10-15 meters. Leaves
212 alternate. Shape ovate-elliptic, 6.5-8 cm long and 3-5 cm wide. Presence of adventitious
213 roots for climbing at the nodes. Heterophylly: the lower part of the stem showing
214 cordate leaves. Leaf apex acuminate, while the leaf base is uneven and rounded.
215 Inflorescence leaf opposite, cylindrical and erect. Female spikes 3-8 cm long, with a
216 peduncle 1-2 cm long, with small sessile spirally arranged flowers. Single ovary, 3-4
217 lobed white stigma. Male spikes 6-10 cm long, with a peduncle 2-3 cm long and
218 stamens organized mainly in groups of two. Ripe fruit reddish and oval, 0.4 cm long,
219 fruit pedicel 0.8-1.2 cm. Each fruit gives-off a single rounded-shaped seed.

220 In Figure 6A the cordate leaves of the lower part of the stem are shown. Figure 6B
221 shows the collection of the plant climbing trees up to 10-12 meters. The fruits are
222 shown in Figures 6C and 6D. The male inflorescence is shown in Figure 6E and the
223 adventitious roots are visible in Figure 6F. The female inflorescence is shown in Figure
224 7A, the fruits in Figure 7B and the number of stigmas in Figure 7C. Stamens
225 sometimes solitary with two anthers with lateral apertures . Stigmas most frequently
226 three, sometimes four, still visible on the enlarged fertilized ovary. Stigmas are sessile.
227 Inflorescence stem in cross section with two circles of bundles: a group of larger more

228 internal bundles and an external group of smaller bundles. Continuous layer of
229 sclerenchyma enclosing the smaller bundles. Living in humid forest.

230

231 **4. Discussion**

232 The morphological results show that the description of *P. tsarasotrae* does not overlap
233 with the description of the other species known to be indigenous of Madagascar such as
234 *P. heimii* (quite close to *P. malgassicum*) and *P. pachyphyllum*. *Piper heimii* appears to
235 be very close to *P. malgassicum*, but the first has lanceolate-ovate leaves 12.5 cm long
236 according to the protologue, whereas the second has ovate leaves, 6.5-8 cm long.
237 However, *P. pachyphyllum* and *P. heimii* have not been recently sampled and should be
238 further investigated.

239 The phylogenetic analysis of the two new species in the context of a subset of the
240 matrix used by Smith et al. (2008) showed that *P. malgassicum* and *P. tsarasotrae*
241 belong to a clade comprising *P. borbonense*, *P. guineense* and *P. caninum*. The same
242 clade was also identified by Smith et al. (2008) with higher Bayesian support with
243 respect to our phylogenetic analysis. Possibly the larger sampling in this group due to
244 the insertion of *P. malgassicum* and *P. tsarasotrae* decreased the robustness (however
245 considerably high, that is 85%). The placement in a monophyletic group formed by *P.*
246 *malgassicum*, *P. tsarasotrae* and *P. borbonense* (relatively close to *P. pachyphyllum* in
247 Figure 1) in the phylogenetic analysis is also corroborated by the biogeographical
248 position of these entities, since the first two species are endemic of Madagascar, while
249 *P. borbonense* originates from La Reunion Island and Mauritius (but also present in
250 Madagascar according to De Candolle (1923). The possible presence of *P. borbonense*

251 in Madagascar, not only in cultivated form but also as spontaneous species, also as
252 possible further component of the voatsiperifery pepper, should be ascertained.

253 The two new entities appear to belong to *Piper* s. s. in the sense of Jaramillo et al.
254 (2008).

255 *Piper malgassicum* is probably more closely related to *P. borbonense* and to *P. heimii*
256 than *P. tsarasotrae*, even if *P. malgassicum* appears to be wild in Madagascar, while *P.*
257 *borbonense* may have been introduced in this Island for spice production. *Piper*
258 *tsarasotrae* has a completely different ecological niche (creeping on the soil and on the
259 rocks, sometimes lianous, but on low plants) with respect to *P. malgassicum*, which is a
260 more typical forest lianous species of *Piper*. These three species appear to be
261 phylogenetically related to *P. guineense*, endemic of the African mainland. This genetic
262 affinity was already indicated by Jaramillo et al. (2008)• and appears to be more strict
263 between *P. tsarasotrae* and *P. guineense*.

264 The presence of two circles of vascular bundles in the stem of many *Piper* species was
265 defined as polystelic organization by Isnard et al. (2012) and was considered by these
266 authors as a synapomorphy of the family Piperaceae with the exception of genus
267 *Verhuellia*. This character was observed in detail, for instance, in some American
268 species such as *P. amalago* L. (Dos Santos et al., 2015), in which also a continuous
269 layer of sclerenchyma was described as in *P. malgassicum*. This scheme is typical of the
270 investigated species of *Piper* (Dos Santos et al., 2015), but *P. tsarasotrae* shows some
271 difference, since the sclerenchymatic layer is not continuous.

272 In conclusion, the two new species of *Piper* here described concur to the production of
273 some of the locally produced voatsiperifery pepper, probably together with *P.*
274 *borbonense*, and are hence of economical importance as spices. The association of

275 species morphological identification with DNA sequences could be useful as a bar
276 coding method for identification of the components of spices and drugs in traditional
277 mixtures (Chaveerach et al., 2006).

278

279 **References**

280

281 Akaike H (1974). A New Look at the Statistical Model Identification. IEEE T Automat
282 Contr 19: 716–723.

283 APG (Angiosperm Phylogeny Group), Chase MW, Christenhusz MJM, Fay MF, Byng
284 JW, Judd WS, Soltis DE, Mabberley DJ, Sennikov AN, Soltis PS et al. (2016).
285 An update of the Angiosperm Phylogeny Group classification for the orders and
286 families of flowering plants: APG IV. Bot J Linn Soc 181: 1–20.

287 Bressan E A, Rossi ML, Lee TSG, Figueira A (2014). Extraction of high-quality DNA
288 from ethanol-preserved tropical plant tissues. BMC Research Notes 7: 268.

289 Burger W (1972). Evolutionary trends in the Central American Species of *Piper*
290 (Piperaceae). Brittonia 24: 356–362.

291 Chaveerach A, Piya M, Runglawan S, Tawatchai T (2006). Ethnobotany of the genus
292 *Piper* (Piperaceae) in Thailand. Ethnobotany Research & Applications 4: 223–231.

293 Corpet F (1988). Multiple sequence alignment with hierarchical clustering. Nucleic
294 Acids Res 16: 10881–10890.

295 De Candolle C (1869). Piperaceae. In: De Candolle C, editor. Prodrromus Systematis
296 Naturalis Regni Vegetabilis Vol. 16 Part 1. Paris, France: Masson, pp. 235–471.

297 De Candolle C (1923). Piperacearum clavis analytica. Candollea 1: 65–415.

298 Dos Santos VLP, Franco CRC, Amano E, Messias-Reasond IJ, Budele JM (2015).
299 Anatomical investigations of *Piper amalago* (jaborandi-manso) for the quality
300 control. Rev Bras Farmacogn 25: 85-91.

301 Felsenstein J (1981). Evolutionary trees from DNA sequences: a Maximum Likelihood
302 approach. J Mol Evol 17: 368–376.

303 Görts-Van Rijn ARA, Callejas Posada R (2005). Three new species of *Piper*
304 (Piperaceae) from the Guianas. Blumea 50: 367–373.

305 Hammer O, Harper D, Ryan PD (2001). PAST: Paleontological Statistics Software
306 Package for Education and Data Analysis. Palaeontologia Electronica 4: 1–9.

- 307 Huelsenbeck JP, Ronquist F (2001). MRBAYES: Bayesian inference of phylogenetic
308 trees. *Bioinformatics* 17: 754–755.
- 309 Isnard S, Prosperi J, Wanke S, Wagner ST, Samain M-S, Trueba S, Frenzke L, Neinhuis
310 C, Rowe NP (2012). Growth form evolution in Piperales and its relevance for
311 understanding Angiosperm diversification: an integrative approach combining
312 plant architecture, anatomy, and biomechanics. *Int J Plant Sci* 173: 610-639.
- 313 Jaramillo MA, Callejas R, Davidson C, Smith JF, Stevens AC, Tepe EJ (2008). A
314 phylogeny of the tropical genus *Piper* using ITS and the chloroplast intron psbJ-
315 petA. *Syst Bot* 33: 647–660.
- 316 Jaramillo MA, Manos PS (2001). Phylogeny and patterns of floral diversity in the
317 Genus *Piper* (Piperaceae). *Am J Bot* 88: 706–716.
- 318 Kumar S, Stecher G, Tamura K (2016). MEGA7: Molecular Evolutionary Genetics
319 Analysis version 7.0 for bigger datasets. *Mol Biol Evol* 33: 1870–1874.
- 320 Lewke Bandara N, Papini A, Mosti S, Brown T, Smith LMJ (2013). A phylogenetic
321 analysis of genus *Onobrychis* and its relationships within the tribe Hedysareae
322 (Fabaceae). *Turk J Bot* 37: 981-992.
- 323 Loconte H, Stevenson DW (1991). Cladistics of the Magnoliidae. *Cladistics* 7: 267–
324 296.
- 325 Marquis RJ (2004). Biogeography of Neotropical *Piper*. In: Palmer ADN, Dyer LA,
326 editors. A model genus for studies of phytochemistry, ecology and evolution. New
327 York, USA: Kluwer, pp. 78–96.
- 328 McNeill J, Barrie FR, Buck WR, Demoulin V, Greuter W, Hawksworth DL, Herendeen
329 PS, Knapp S, Marhold K, Prado J et al. (2012) International Code of Nomenclature
330 for algae, fungi, and plants (Melbourne Code). *Regnum Veg* 154. A.R.G. Gantner
331 Verlag KG, Ruggell, Liechtenstein. ISBN 978-3-87429-425-6. Available at
332 <http://www.iapt-taxon.org/nomen/main.php>
- 333 Mosti S, Ross Friedman C, Piccolin F, Di Falco P, Papini A (2012). The unusual
334 tegumental tissues of the *Lunaria annua* (Brassicaceae) seed: a developmental
335 study using light and electron microscopy. *Flora* 207: 828-837.
- 336 Murray MG, Pitas JW (1996). Plant DNA from alcohol-preserved samples. *Plant Mol*
337 *Biol Rep* 14: 261–265.
- 338 Nylander JAA (2004). MrModeltest v2. Program distributed by the author. Uppsala,
339 Sweden: Evolutionary Biology Centre, Uppsala University.
- 340 Olsen KM, Schaal BA (1999). Evidence on the origin of Cassava: phylogeography of
341 *Manihot Esculenta*. *P Natl Acad Sci USA* 96: 5586–5591.

- 342 Papini A, Banci F, Nardi E (2007). Molecular evidence of polyphyletism in the plant
343 genus *Carum* L. (Apiaceae). *Genet Mol Biol* 30: 475-482.
- 344 Papini A, Simeone MC, Bellarosa R, Spada F, Schirone B (2011). *Quercus*
345 *Macranthera* Fisch. & Mey. Ex Hohen. and *Quercus Iberica* M. Bieb.:
346 taxonomic definition and systematic relationships with European oaks inferred
347 from nuclear Internal Transcribed Spacer (ITS) data. *Plant Biosyst* 145: 37-49.
- 348 Quijano-Abril MA, Callejas-Posada R, Miranda-Esquivel DR (2006). Areas of
349 endemism and distribution patterns for Neotropical *Piper* species (Piperaceae). *J*
350 *Biogeogr* 33: 1266-1278.
- 351 Rambaut A (2009). FigTree, a graphical viewer of phylogenetic trees. Edinburgh, UK:
352 Institute of Evolutionary Biology University of Edinburgh. Website
353 <http://tree.bio.ed.ac.uk/software/figtree/> [accessed 10thOctober 2017].
- 354 Ronquist F, Teslenko M, Van Der Mark P, Ayaccesseres DL, Darling A, Höhna S,
355 Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012). Mrbayes 3.2: efficient
356 bayesian phylogenetic inference and model choice across a large model space. *Syst*
357 *Biol* 61: 539-42.
- 358 Simeone MC, Grimm GW, Papini A, Vessella F, Cardoni S, Tordoni E, Piredda R,
359 Franc A, Denk T (2016). Plastome data reveal multiple geographic origins of
360 *Quercus* Group Ilex. *PeerJ* 2016: 4. doi:10.7717/peerj.1897.
- 361 Simpson MJ (2010). 9 Plant morphology. In: Simpson MJ, editor. *Plant Systematics*
362 (Second Edition). San Diego, USA: Academic Press, pp. 451-513.
- 363 Smith JF, Stevens AC, Tepe EJ, Davidson C (2008). Placing the origin of two species-
364 rich genera in the late Cretaceous with later species divergence in the Tertiary: a
365 phylogenetic, biogeographic and molecular dating analysis of *Piper* and
366 *Peperomia* (Piperaceae). *Plant Syst Evol* 275: 9-30.
- 367 Stamatakis A, Aberer AJ, Goll C, Smith SA, Berger SA, Izquierdo-Carrasco F (2012).
368 RAxML-Light: a tool for computing terabyte phylogenies. *Bioinformatics* 28:
369 2064-2066.
- 370 Strand AE, Leebens-Mack J, Milligan BG (1997). Nuclear DNA-based markers for
371 plant evolutionary biology. *Mol Ecol* 6: 113-118.
- 372 Suwanphakdee C, Simpson DA, Hodkinson TR, Chantaranonthai P (2016). Taxonomic
373 notes on the genus *Piper* (Piperaceae). *Nord J Bot* 34: 605-618.
- 374 Taberlet P, Gielly L, Pautou G, Bouvet J (1991). Universal primers for amplification of
375 three non-coding regions of chloroplast DNA. *Plant Mol Biol* 17: 1105-1109.

- 376 Tebbs MC (1993). Piperaceae. In: Kubitzki K, Rohwer JG, Bittrich V, editors.
377 Flowering Plants Dicotyledons. The Families and Genera of Vascular Plants, vol 2.
378 Berlin, Heidelberg, Germany: Springer, pp. 516–520.
- 379 Thompson JD, Higgins DG, Gibson TJ (1994). CLUSTAL W: improving the sensitivity
380 of progressive multiple sequence alignment through sequence weighting, position-
381 specific gap penalties and weight matrix choice. Nucl Acids Res 22: 4673–4680.
- 382 Ulloa UC, Acevedo-Rodriguez P, Beck S, Belgrano MJ, Bernal R, Berry PE, Brako L,
383 Celis M, Davidse G, Forzza RC et al. (2017). An integrated assessment of the
384 vascular plant species of the Americas. Science 358: 1614-1617.
- 385 Wanke S, Jaramillo MA, Borsch T, Samain MS, Quandt D, Neinhuis C (2007).
386 Evolution of Piperales—matK gene and trnK intron sequence data reveal lineage
387 specific resolution contrast. Mol Phylogenet Evol 42: 477-97.
- 388 Weil M, Shum Cheong Sing A, Méot JM, Boulanger R, Bohuon P. (2017). Impact of
389 blanching, sweating and drying operations on pungency, aroma and color of *Piper*
390 *borbonense*. Food Chem 219: 274-281.

391

392

393

394

395

396

397

398

399

400

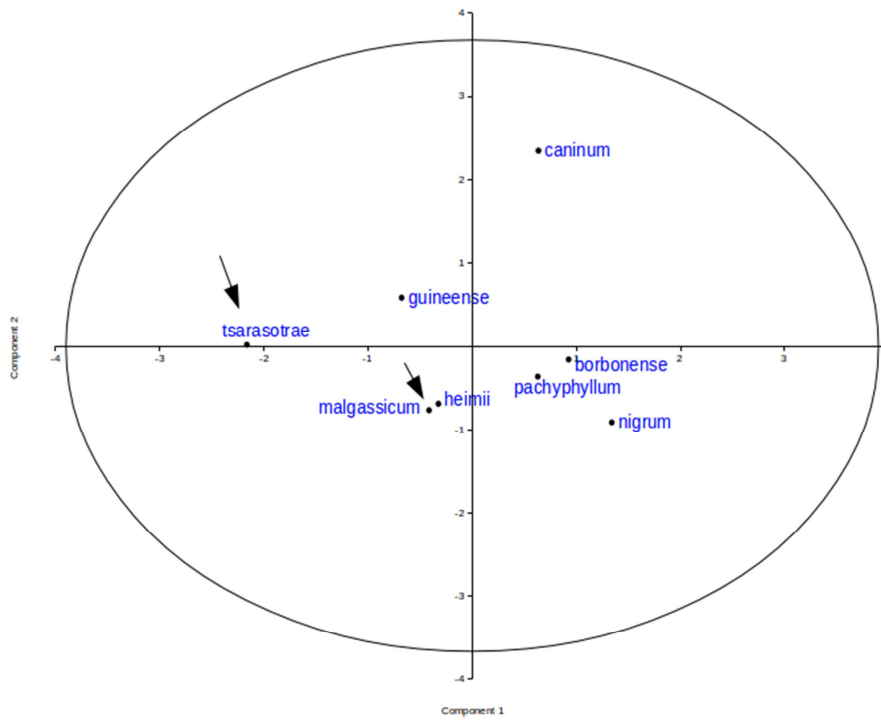
401

402

403

404

405 **Figure legends**



406

407 Figure 1. PCA analysis of the two new species of *Piper* together with the more strictly
408 related species. The position of *Piper tsarasotrae* (tsarasotrae in the figure) and *Piper*
409 *malgassicum* (malgassicum in the figure) are indicated by arrows.

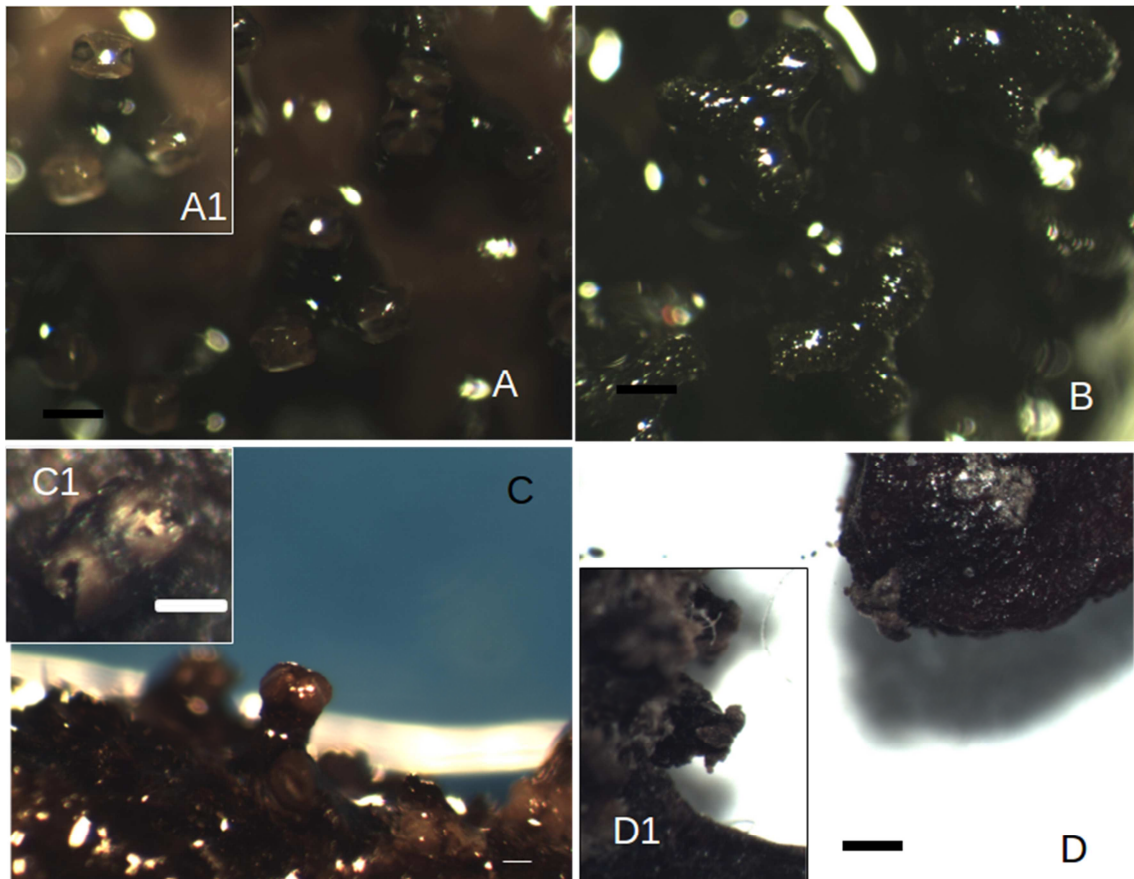


410

411 Figure 2. Phylogenetic analysis with maximum likelihood based on trnL intron, ndhF
 412 and G3pdH genes. Bayesian support reported on branches. The position of *Piper*
 413 *tsarasotrae* (SPN Tsarasotra in the figure) and *Piper malgassicum* (SPN Vohiday in the
 414 figure) are evidentiated in green, together with the genetically strictly associated *P.*
 415 *borbonense*. The phylogenetically close *P. guineense* accessions are evidentiated in red.
 416 All the names refer to species of genus *Piper* L. with the exception of *Peperomia*
 417 *pellucida* and *Hottuyinia cordata*, whose names are reported entirely (together with the
 418 genus name) with the provenence on the right.

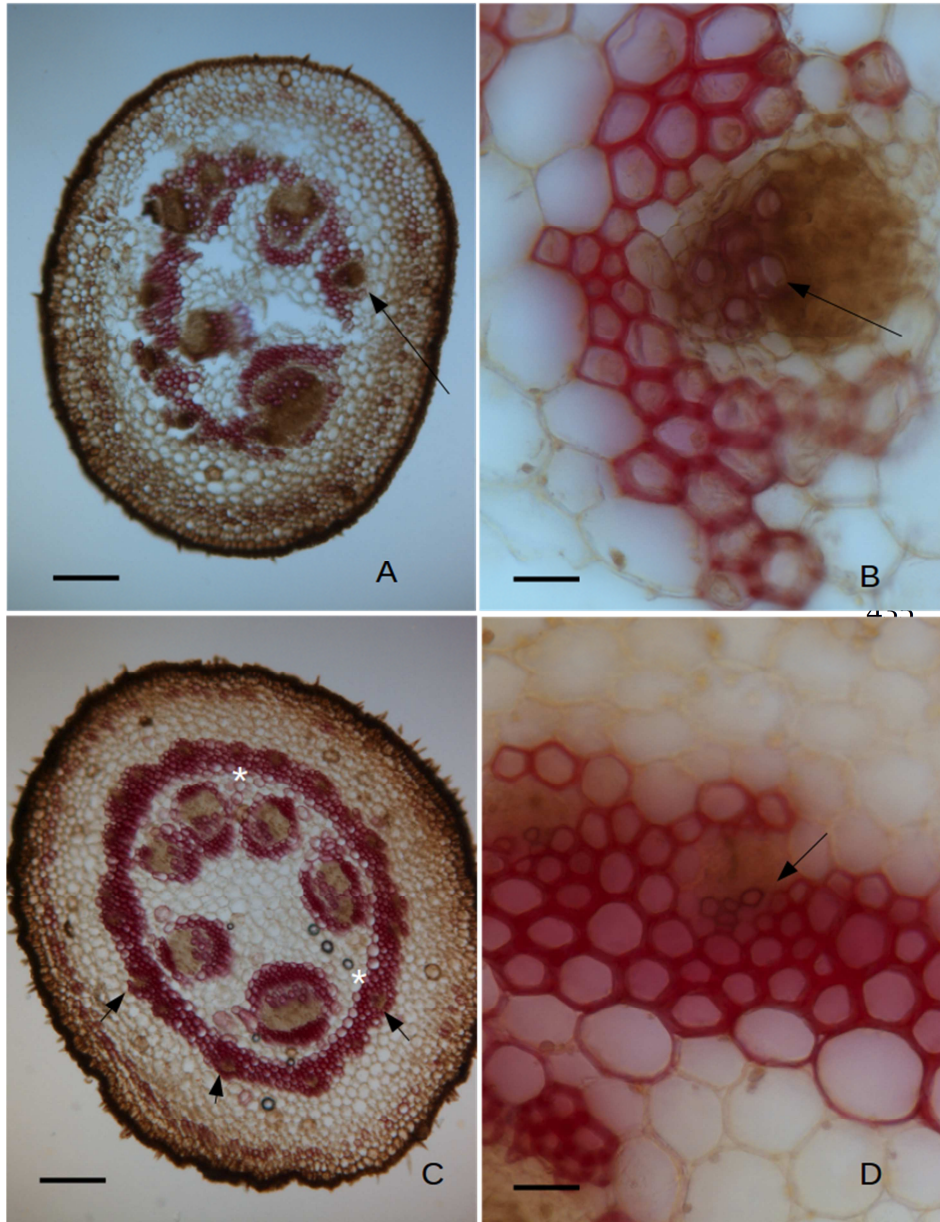
419

420



421

422 Figure 3 – Observation with stereomicroscope. *Piper tsarasotrae*. A: A group of 3
 423 stamens is visible. In A1 a detail of the anthers. B: Shape of the stigmas. The surface
 424 appears to be covered by appendages. *Piper malgassicum*. C: Stamen in lateral view.
 425 C1: detail of the anthers. D: Stigmas on an already grown ovary. D1: lateral view of an
 426 ovary with three stigmas.



444

445

446 Figure 4 - *Piper tsarasotrae*. Cross section of the inflorescence stem. A: general aspect
 447 of the eustele with a group of larger more internal bundles and an external group of
 448 smaller bundles. The arrow indicates the smaller bundle of Figure 7B. Bar = 400µm. B:
 449 Detail of Figure 7A. One of the smaller bundles. The arrow indicates one of the tracheal
 450 elements. Bar = 50µm. *Piper malgassicum*. Cross section of the inflorescence stem. C:

451 general aspect of the eustele with a group of larger more internal bundles and an
452 external group of smaller bundles. The arrows indicate the smaller bundles. The white
453 asterisks indicate a continuous layer of sclerenchymatic cells. 7D. Bar = 400µm. D:
454 Detail of Figure 7C. One of the smaller bundles. The arrow indicates one of the tracheal
455 elements. Bar = 50µm.

456



472

473

474

475 Figure 5 - *Piper tsarasotrae*. A: General habitus of the species. B: typical environment
476 of the species.
477 FC: female cones, with details of the stigmas. D: Position of the female cones on the
478 female plant. E: Ripe fruits. F: both the female inflorescence and the fruits at different
479 level of ripeness can be observed on the same individual.



480

481 Figure 6 - *Piper malgassicum*. A: the cordate leaves of the lower part of the stem are

482 shown. B: Method of collection of the fruits from plants climbing trees (up to 10-12

483 meters). C: Ripe fruits in the context of the plant. D: Infructescence with fruits at
484 various stages of ripeness. E: Detail of the male inflorescence. F: Adventitious roots
485 growing from nodes.



486

487 Figure 7 - *Piper malgassicum*. A: Female inflorescence on the plant. B: fruits at various
488 stages of ripeness. C: Female inflorescences at various stages of maturation with a detail
489 of the stigmas.

490

491 Table 1 – Morphological characters obtained from herbarium samples coded for the
 492 Principal Component Analysis (PCA). Characters used in the table and codification of
 493 character states: 1) stem nodes: swollen = 1; not swollen = 0; 2) habitat: harid forest =
 494 1; humid forest = 1; shady forest = 0; 3) leaf shape: lance-ovate = 1; ovate = 0; cordate
 495 = 2; 4) leaves of the low part of the stem: presence of cordate leaves = 0; never presence
 496 of cordate leaves = 1; 5) leaf length in cm: mimum 6 cm = 1; minimum 6,5 = 0; 6)
 497 maximum leaf length: less than 10 = 0; more than 10 = 1; 7) Minimal leaf width in cm:
 498 less than 3 = 0; more = 1; 8) Maximal leaf width in cm: less than 6.5 = 0; more = 1; 9)
 499 leaf apex: not acuminate = 0; acuminate = 1; 10) leaf base: Iniquil narrowly cuneate =
 500 0; iniquil cuneate = 1; cordate = 2; 11) leaves: alternate = 0; non alternate = 1; 12) leaf
 501 petiole: max length <= 2,5 = 0; more = 1; 13) petiole minimal length in cm: < 1 cm = 0;
 502 more or equal than 1 = 1; 14) leaf nerves: palmate = 0; pinnate = 1; 15) minimum
 503 number of stigmas: 2 = 2; 3 = 3; 4 = 4; 16) flower color: red = 1; not red = 0; 17) flower
 504 spike dimension: max length less than 5 = 0; more = 1; 18) fruit spike dimension in cm:
 505 maximum less than 4 cm = 0; more than 4 = 1; 19) flower spikes opposite to leaves: yes
 506 = 0; no = 1; 20) dioicy: dioecious = 0; not dioecious = 1; 21) growth form: liane = 0;
 507 shrub = 1; 22) vegetative dimension m: more than 6 = 0; up to 6 = 1. The interrogative
 508 mark “?” means either that the character is variable in the species or that the character
 509 state is not known. Characters obtained through direct measurements for the first two
 510 species; from Verdcourt (1996) for species 3 and 4; De Candolle (1869 C. DC.
 511 Prodrum Systematis Naturalis Regni Vegetabilis 16(1): 339. 1869) for species 5; De
 512 Candolle (1911 C. DC. 1911. Notul. Syst. (Paris) 2: 51) for species 6; Baker (1885
 513 Baker J. G. (1885) Further controbutions to the Flora of Central Madagascar. - Second
 514 and final part. Journal of the Linnean Society, Botany 21: 436. 1885.) for species 7 and
 515 Blume (1826 C. L. Blume (1826 Monographie der Oost-indische Pepersorten/diir.
 516 Verh. Batav. Genootsch. Kunst. 11: 214, f. 26) for species 8. Botanical nomenclature
 517 after Simpson (2010).

Ref.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
tsarasotrae	0	2	1	1	1	0	0	0	1	0	0	0	0	0	3	?	0	?	?	0	0	0
malgassicum	0	1	0	0	0	0	0	0	1	1	0	0	0	0	3	?	1	?	?	0	1	1
nigrum	0	1	0	?	0	1	1	1	1	1	0	0	1	1	4	?	1	?	?	1	1	1
guineense	?	0	0	?	1	1	0	1	1	?	0	1	0	0	3	?	0	1	yes	0	0	0
borbonense	?	0	0	?	0	1	1	0	1	1	0	0	1	1	3	?	?	?	?	1	0	0
heimii	?	?	1	?	0	1	1	0	?	0	0	0	0	1	4	?	0	?	?	0	?	?
pachyphyllum	?	?	0	?	0	1	1	0	1	1	0	0	0	1	3	1	?	?	?	yes	?	1
caninum	?	1	2	?	0	1	1	1	1	2	0	1	1	?	2		0	?	?	?	?	?

518

519

520

521

522 Table 2 – Geographical coordinates of the samples collected for DNA extraction.
 523 Genbank accession numbers of the corresponding g3pdH, trnL and ndhF are reported on
 524 the right side of each accession. All the samples of *P. tsarasotrae* come from the
 525 locality Tsarasotra, Ambositra region (Madagascar) while all the samples of *P.*
 526 *malgassicum* come from the Vohiday forest, Ambositra region (Madagascar). Latitude
 527 and longitude of collection places are indicated underneath the species name.

528

	<i>P. tsarasotrae</i> (<i>Tsarasotra</i>)	Genbank g3pdh	Genbank trnL	Genbank ndhf	<i>P. malgassicum</i> (<i>Vohiday</i>)	Genbank g3pdh	Genbank trnL	Genbank ndhf
	S20° 26.716'				S20° 31.899' E47°			
PNS1	E47° 11,157'	MH234634	MH234638	MH234636	PNSv1	27.492'	MH234633	MH234637
	S20° 27.146'					S20° 32.278' E47°		
PNS2	E47° 10.948'	not variable	not variable	not variable	PNSv2	35.298'	not variable	not variable
	S20° 27.150'					S20° 32.310' E47°		
PNS3	E47° 10,961'	not variable	not variable	not variable	PNSv3	35.281'	not variable	not variable
	S20° 27.165'					S20° 32.367' E47°		
PNS4	E47° 10,999'	not variable	not variable	not variable	PNSv4	29.198'	not variable	not variable
	S20° 27.165'					S20° 32.615' E47°		
PNS5	E47° 10,999'	not variable	not variable	not variable	PNSv5	35.498'	not variable	not variable
	S20° 27.169'					S20° 32.661' E47°		
PNS6	E47° 10,993'	not variable	not variable	not variable	PNSv6	35.301'	not variable	not variable
	S20° 27.941'					S20° 32.704' E47°		
PNS7	E47° 11,401'	not variable	not variable	not variable	PNSv7	35.146'	not variable	not variable
	S20° 27.941'					S20° 32.896' E47°		
PNS8	E47° 11,401'	not variable	not variable	not variable	PNSv8	27.492'	not variable	not variable
	S20° 95.647'					S20° 32.963' E47°		
PNS9	E47° 11,456'	not variable	not variable	not variable	PNSv9	35.403'	not variable	not variable
	S20° 98.747'					S20° 45.224' E47°		
PNS10	E47° 11,392'	not variable	not variable	not variable	PNSv10	28.428'	not variable	not variable

529