



Nanotechnologies for the restoration of alum-treated archaeological wood

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Abstract The project Saving Oseberg is funded by the Norwegian State with the aim to preserve the Viking Age wooden objects from the Oseberg burial mound. They were excavated in 1904 near Tønsberg, Norway, and many have been treated in the past with alum salts ($\text{KAl}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}$). Alum was widely used during the early 1900s as a treatment for archaeological wood to prevent shrinkage and impart strength. In the 1990s, conservators observed an alarming condition of the objects. Initial investigations showed that the alum treatment has initiated a slow but ongoing deterioration process, attacking the wood for over 100 years. Today, the artefacts are highly acidic and have significantly reduced mechanical strength. In the last decade, the use of non-aqueous alkaline nanoparticle dispersions has provided successful results for the protection of cellulose-based materials. Alum-treated archaeological wood samples from Oseberg, with a $\text{pH} \leq 2$, have been treated with alkaline nanoparticle dispersions, and the effects of the treatment have been evaluated by thermal analysis (TG-DTG), infrared spectroscopy (ATR-FTIR) and X-ray microtomography (micro-CT) analyses. In this contribution, the preliminary results will be presented.

1 Introduction

The Oseberg finds, a selection of which is on display at the Viking Ship Museum in Oslo, Norway, represent a unique collection of Viking Age objects. The objects—many of which are made of wood—were excavated in 1904 from a ship burial constructed for two important women in 834 AD [1]. Object types include those used in religious ceremonies as well as in everyday life, and many bear intricate carvings [2].

The objects were waterlogged and highly fragmented upon recovery. Some wood types were so well preserved—such as the oak from the Oseberg ship—that conservation treatment was not required before drying; other objects, however, many of which were made of diffuse porous hardwoods—such as the ceremonial sleds—were highly degraded and required stabilization before drying could take place. The method used to conserve these objects involved immersion in hot (ca 90 °C), concentrated solutions of alum salts ($\text{KAl}(\text{SO}_4)_2 \cdot 12\text{H}_2\text{O}$, aluminium potassium sulphate dodecahydrate). This treatment both strengthened and maintained the shape of the fragments: the salt replaced the water in the wood cells, and when removed from the hot bath the salt recrystallized immediately upon cooling. Before reconstruction into whole objects, fragments were impregnated with linseed oil and then pieced together using metal screws, pins, fills and modern wood. The reconstructed object was finally varnished. Fragments which were not reconstructed into objects are in the museum stores and may or may not contain linseed oil.

Although the alum method was mainly used in Scandinavia, it gained popularity in other parts of the world and was in use from the mid-nineteenth century to the 1960s, when it gradually was replaced by, for example, treatments

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using polyethylene glycol. After ca 1910, glycerol became a common addition to the alum treatment, which significantly increased the hygroscopicity of the treated wood relative to wood treated only with alum [3]. Thus, thousands of objects have been conserved with alum and alum/glycerol during the period the treatment was in active use.

Today, the majority of the alum-treated objects are in poor condition: the wood is mechanically weakened and highly acidic ($\text{pH} \leq 2$). The source of chemical deterioration is attributed to the alum treatment itself, which generates significant amounts of sulphuric acid when heated [4]. It is also hypothesized that the presence of reactive transition metal ions, such as iron, enhances deterioration by oxidation, likely involving Fenton reactions [5].

As alum-treated wood is a highly complex material and little understood, the research project *Saving Oseberg* was established in 2013 to further elucidate the deterioration mechanisms of these archaeological wooden artefacts and to propose a conservation treatment to strengthen and chemically stabilize the wood. One of the research topics involves measures to deacidify the wood.

1.1 Neutralization using nanoparticles

One method to neutralize acidified wood involves surface applications of poultices with sodium bicarbonate solution. This method was used on the seventeenth-century Swedish warship *Vasa*, which temporarily increased the surface pH to 9–10. However, the effect did not last as there was no alkaline reserve, such that pH gradually returned to acidic values over a short period of time [6]. Another treatment using ammonia vapours darkened the wood, and pH was difficult to control [7]. Strong alkali could be harmful to the residual cellulose in wood [8].

Another potential method for the deacidification of acidic wood is the application of non-aqueous dispersions of nanoparticles, mainly $\text{Ca}(\text{OH})_2$, as proposed by the Department of Chemistry & CSGI Consortium at the University of Florence [6]. The size of the nanoparticles used in this case ranges from 80 to 220 nm for $\text{Ca}(\text{OH})_2$. Alkaline earth metal hydroxide nanoparticles dispersions have demonstrated to be efficient for the preservation of cellulose-based artefacts, providing a stable neutral environment and, if present in excess, turning into mild alkaline species [9]. New formulations tailored for specific conservation issues have been recently obtained via a solvothermal reaction, starting from bulk metal to short-chain alcohols. Using this synthetic procedure, stable and highly concentrated calcium hydroxide nanoparticles dispersions may be obtained.

This method, also used to neutralize acidic paper and canvas, was applied to the *Vasa* ship [6, 10]. Nanoparticle

penetration inside the wood bulk is effected through a non-aqueous solvent, such as 2-propanol, which has a lower surface tension and is less viscous than water. Furthermore, when the nanoparticles have penetrated into the fibres, hydroxyl groups are released. In this way, it is possible to neutralize the acidity of the treated wood, and the alkaline reserve is expected to grant long-term protection to wood. Experiments performed at CSGI showed that archaeological wood treated with nanoparticles exhibited pH values granting long-lasting protection [11–13]. Furthermore, differential thermogravimetric analysis (DTG) carried out on wood samples treated with nanoparticles from *Vasa* has shown that nanoparticles increased the resistance of wood towards thermal degradation. That is, results showed that the pyrolysis temperature of cellulose increased relative to that of the untreated degraded wood.

Given its potential, this method can be used to neutralize the acidity of the alum-treated *Oseberg* finds. The deacidification efficacy was assessed by pH, DTG analysis, infrared spectroscopy (ATR-FTIR) and X-ray microtomography (micro-CT) measurements.

2 Materials and methods

2.1 Chemicals

Absolute ethanol (99.9 % Fluka) and metallic granular calcium (99 % Sigma Aldrich) were used for nanoparticle synthesis. Sulphuric acid (96 % Carlo Erba) was used for the acidification of wood samples. Alum (potassium aluminium sulphate dodecahydrate, 98 % Sigma Aldrich) and glycerol (99.5 % Sigma Aldrich) were used to treat samples.

2.2 Sample description

The samples used in this study included recently excavated waterlogged archaeological wood (both softwood and hardwood), alum-treated archaeological wood from the *Oseberg* find and sound oak.

The hardwood samples were excavated in 2005 and stored in water. They were radiocarbon dated to 975–1150 AD (AaHW 0001, AaHW 0002) and were identified by light microscopy to be aspen (*Populus* spp.). The softwood archaeological samples (ApSW 0003, ApSW 0004, ApSW 0005) were retrieved in the 1990s and stored in water. They were identified by light microscopy to be pine (*Pinus* spp.). In 2013, these samples were treated in a solution of 2 parts alum to 1 part water (w/w) heated to 90 °C for 24 h. The archaeological samples were documented by X-radiography before and after the alum treatment (Fig. 1).

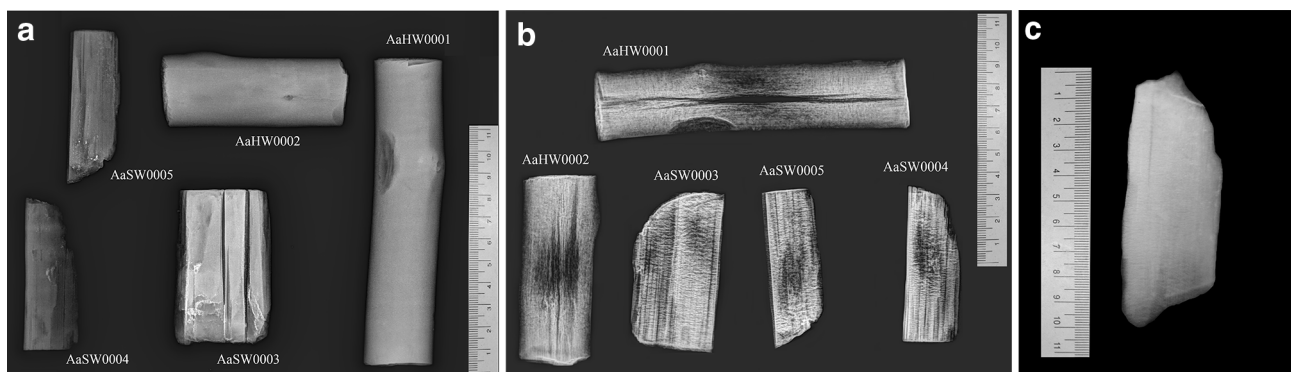


Fig. 1 X-rays of archaeological samples. Archaeological samples before treatment with alum (in the waterlogged state) (a) and after treatment with alum (b); alum-treated wood from Oseberg (c)

The sample from Oseberg was treated with alum without the addition of linseed oil (cat. no. C55000/229-232). It was classified as diffuse porous, since it could not be identified further due to its high degree of deterioration. An overview of samples is given in Table 1.

2.3 Nanoparticle preparation

Calcium hydroxide nanoparticles were synthesized, as reported in the literature [9] in an autoclave system working at high temperature and pressure, via a solvothermal process that can be described as a two-step reaction. In the first step of the synthesis, calcium metal is oxidized by short-chain alcohol, in this case ethanol, leading to the production of the corresponding alkoxide. The subsequent hydrolysis of this compound generates colloidal calcium hydroxide, which is dispersed in the solvent for the application.

2.4 Wood deacidification

Each sample was treated with nanoparticles dispersed in ethanol at a concentration of 5 g/L. The treatments were applied by directly dripping the solution onto the sample surface using a syringe, to wet them as homogeneously as possible, until complete saturation. The treated samples were then left to dry in a small chamber maintained at 50 % RH and 21 °C for 13 days.

Table 1 Sample overview

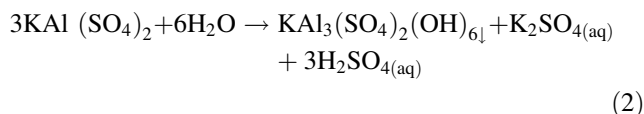
Sample name	Treatment
AaH0001, AaHW0002	Archaeological aspen, alum-treated 2013
ApSW0003, ApSW 0004, ApSW0005	Archaeological pine, alum-treated 2013
Oseberg (C55000/229-232)	Diffuse porous. Alum-treated early 1900s

A solution of glycerol and water was adjusted to a specific gravity (SG) of 1.204 to maintain the chamber equilibrium RH at 50 %, based on Eq. (1) [14].

$$SG = [-0.189 (RH) + 19.9]^{0.0806} \quad (1)$$

2.5 Measurement of wood pH

Alum solutions which have not been heated exhibit pH levels between 3.5 and 4 [4]. Solutions which have been heated to the temperature used in the alum treatment (90 °C) have a pH of about 2 after cooling [4]. Thus, one source of the high acidity measured in alum-treated wood ($pH \leq 2$) originated from the original thermal treatment (90 °C). This resulted in the formation of aluminium hydroxide compounds and sulphuric acid, shown by reaction (2) [4].

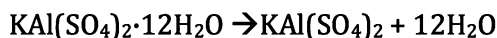


Potassium aluminium sulphate hydroxide (alunite) was identified in the precipitate. Considerable amounts are formed only after several hours of heating [4]. Moreover, according to Baty and Sinnott, aluminium ion Al^{3+} plays an important role as a catalyst in cellulose hydrolysis through coordination of aluminium by the glycosidic oxygen and C6' hydroxyl [15].

In order to verify the effects of deacidification treatments, it was necessary to measure the pH of wood samples before and after the treatment with nanoparticles. According to a standard TAPPI procedure (Technical Association of the Pulp and Paper Industries), 125 mg of wood powder was suspended in 9 mL of distilled water (cold extraction method). The extracting suspension was kept at room temperature. The pH was measured after 1 h

a

1. Between 50 and 350 °C (endothermic):
dehydration in several steps



2. Solid phasedecomposition



3. Between 750 and 950 °C:
decomposition of $\text{Al}_2(\text{SO}_4)_3$ (3 mol of SO_3)



4. At about 1100 °C (endothermic):
decomposition of K_2SO_4

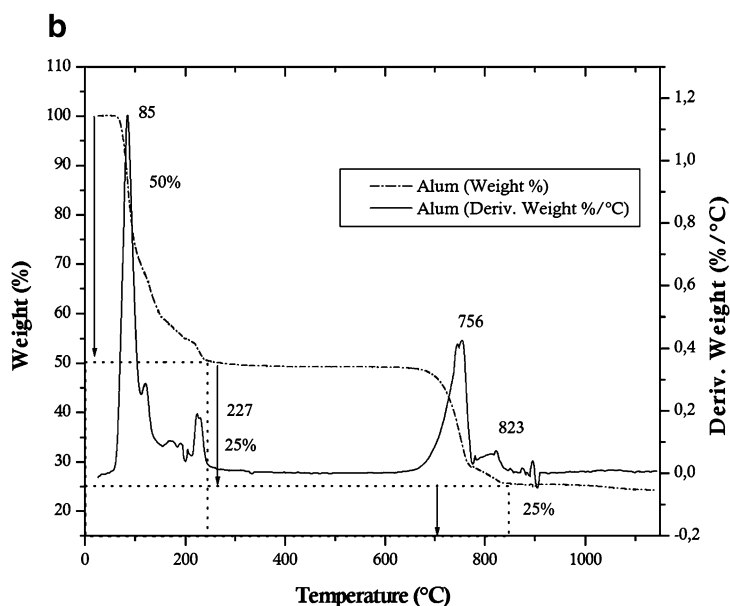


Fig. 2 Main thermal reactions of alum (*left*). TG and DTG curves of alum (*right*)

and 24 h using a digital pH meter Crison Basic 20 equipped with a glass electrode.

2.6 Thermal analysis

The differential thermogravimetric (DTG) curves of the samples were recorded in dynamic, non-isothermal conditions, using a TA SDT Q600 instrument. The experimental conditions were: temperature range: 30–1150 °C; sample weight: 6–8 mg; heating rate: 10 °C/min; N_2 flow: 100 ml/min. The error is ± 1.5 °C. Thermogravimetric analysis (TGA) was also carried out to investigate the percentages of mass loss as a function of the temperature.

2.7 Infrared spectroscopy

Samples were analysed by infrared spectroscopy, using Thermo Fisher FTIR spectrometer (Nicolet iS50) ATR-FTIR unit, containing a ZnSe/diamond hybrid crystal, at a resolution of 4 cm^{-1} , 32 scans per spectrum. Three spectra were taken from each sample before and after treatment with nanoparticles. The spectral range was $4000\text{--}400\text{ cm}^{-1}$.

2.8 Microtomography

Micro-CT is an experimental technique used to obtain three-dimensional microstructural information. In a typical experimental setup, the sample is placed on a rotating stage between an X-ray generator and a detector. For a specific exposure time, X-rays are attenuated

proportionally to the material's density and reach the detector to form a shadow projection containing information about the microstructure. The sample is then rotated a fractional amount, and a second shadow projection is obtained from a slightly offset perspective. A 180° rotation generates a sufficient set of such projections which are reconstructed into parallel microslices using a mathematical algorithm.

For the micro-CT imaging analysis, a SkyScan 1172 system was used at the CRIST center, Centro di Cristallografia Strutturale, of the University of Florence.

3 Results and discussion

Measurements of pH were performed on deacidified samples after 1 and 24 h from application; no significant changes were detected. The pH of untreated samples was found to be around 2.5–3, whereas after deacidification the pH of wood samples was raised to 5, quite close to the original pH of fresh hardwood. After 13 days at 50 % RH of storage, deacidified wooden samples still maintain the same level of acidity. All samples treated with nanoparticles showed an increase in pH value of 2–3 units. After 1 month of the treatment with nanoparticles, pH values were still stable.

The effects of deacidification treatment on degraded wood were also monitored by DTG. The relevant content of alum within wood porosity has required an accurate analysis of all the thermal events involving this material, in order to facilitate the interpretation of thermal analysis

Fig. 3 Comparison between DTG curves of Fresh oak and AaHW 0001 (alum-treated archaeological aspen). The DTG curve of fresh oak shows main thermal degradation peaks of wood in good condition (*left*). TG and DTG curves of sample AaHW 0001 (*right*)

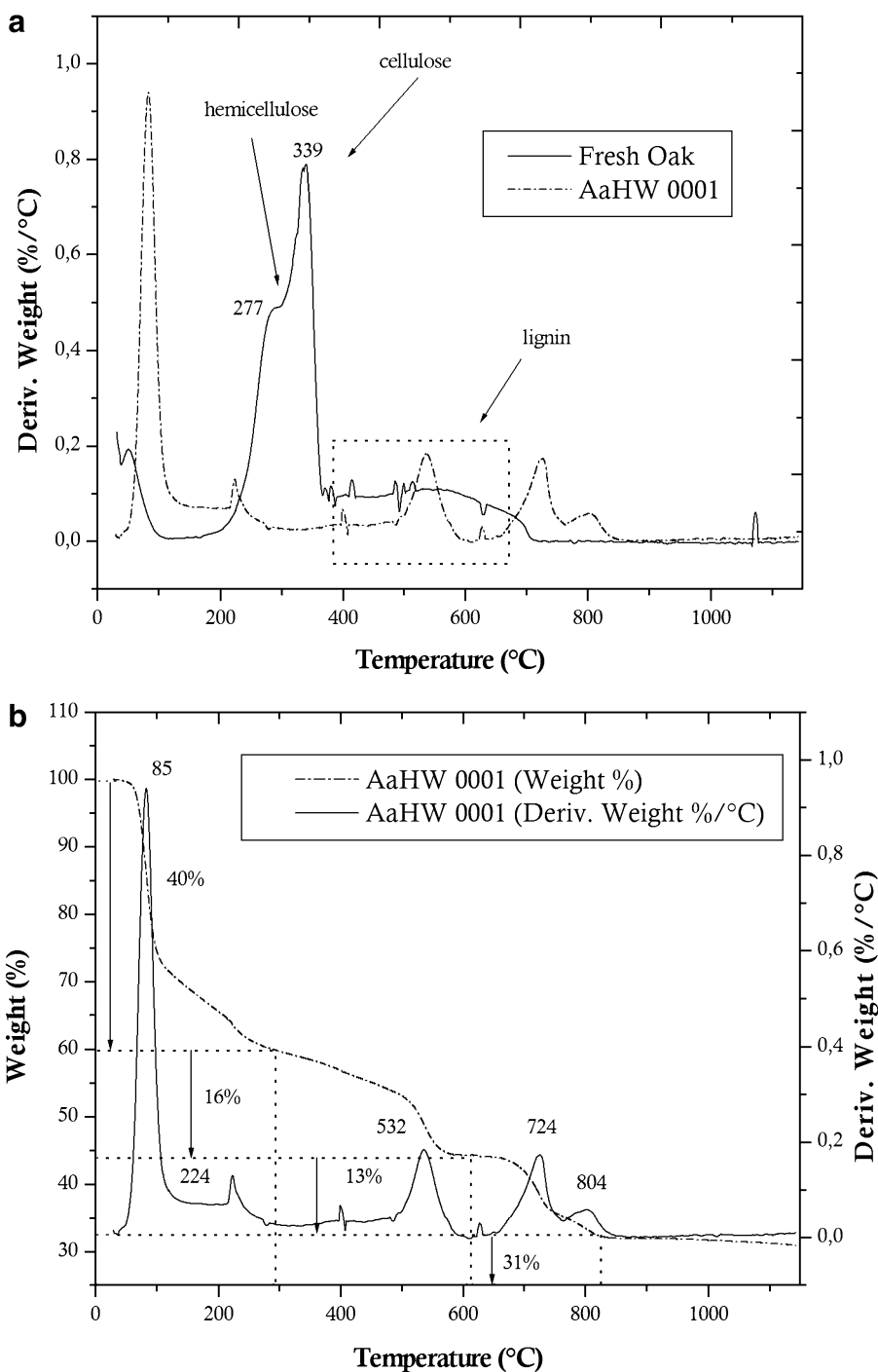


Table 2 Weight % of main products involved in the thermal degradation of AaHW0001

	Initial weight (mg)	Final weight (mg)	Water content	Lignin content	SO ₃ mol of Al ₂ (SO ₄) ₃	Al ₂ O ₃
AaHW 0001	8.962	2.769	40 %	16 %	13 %	31 %

data. The characteristic thermal curve for alum is shown in Fig. 2, and the main thermal reactions involved are highlighted [16]. As already reported in the literature [9], the

pyrolysis temperature of cellulose (T_p) can be used for monitoring the efficacy of deacidification treatment, where it was shown that higher T_p values correspond to samples

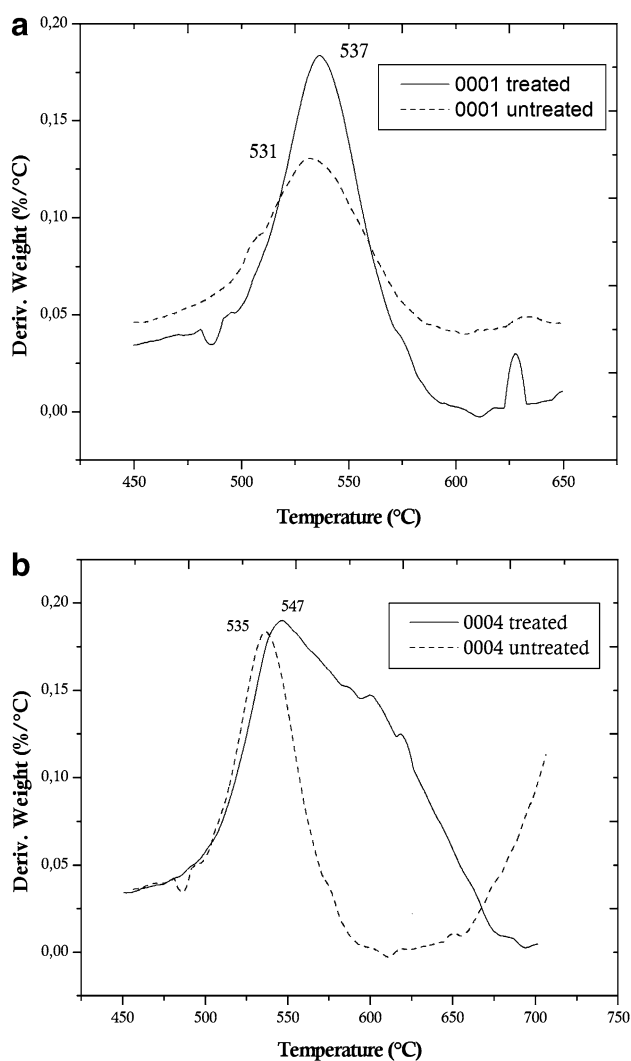


Fig. 4 Comparison between DTG curves of AaHW 0001 and AaHW 0001 samples before and after deacidification treatment (*left*). Comparison between DTG curves of ApSW 0004 and ApSW 0004 samples before and after deacidification treatment (*right*)

more resistant to thermal degradation in accordance with higher pH values [13]. Unfortunately, the wood samples in this study are highly degraded, and the signals associated with the pyrolysis of cellulose are negligible. The only thermal event that can be clearly seen in the derivative thermogravimetric curves is associated with the pyrolysis of lignin; the temperature corresponding to the maximum rate of lignin mass loss will be reported as T_1 . The release of CO is characteristic of the last stages of pyrolysis at higher temperatures at around 400 °C. The process of devolatilization of lignin is practically completed above 600 °C. In Fig. 3, the thermal profile for the fresh oak sample and sample AaHW 0001 is reported. Table 2 shows the % weight of the main pyrolysis products evolved from AaHW 0001.

The archaeological samples (AaHW 0001, AaHW 0002, ApSW 0003, ApSW 0004, ApSW 0005) were characterized by DTG after the application of nanoparticles. The deacidified samples showed a significant increase in T_1 , with differences ranging from 6 to 12 °C when compared to the untreated wooden samples (see Fig. 4). The observed increase in T_1 can partly be explained as an effect of the introduction of calcium hydroxide nanoparticles within wood porosity. In fact, it has been hypothesized that the bivalent positive calcium ions from the deacidification treatment could also contribute to increase the resistance of treated wood to thermal degradation [17] due to the chelating complex formed with hydroxyl and methoxy groups of lignin.

The distribution of alkaline nanoparticles within the wooden samples porosity was investigated with micro-CT-imaging. Two small splinters of wood (2 mm in size) were taken from AaHW 0001 and ApSW 0004 samples. Each splinter was mounted vertically on the SkyScan 1172 scanner sample chamber, for analysis with micro-CT imaging technique. Data were collected before and after the treatment with alkaline nanoparticles of $\text{Ca}(\text{OH})_2$ dispersed in ethanol (at concentration of 5 g/L). From cross sections and 3D reconstruction, the penetration of nanoparticles within the wood was evaluated. Figure 5a–d shows that alum (brighter areas in the pictures) is evenly distributed within wood, and apparently it fills almost all vessels. The darker areas represent the wood structure, mainly composed of lignin, as was demonstrated by DTG analyses. Denser white clusters, evidenced with boxes in the pictures, may be attributed, according to differences in attenuation, to alunite ($\text{KAl}_3(\text{SO}_4)_2(\text{OH})_6$), presumably formed, before the application, after the heating treatment of alum. Figure 5b, d shows images after treatment where brighter and less contrasted areas are clearly visible, indicating greater attenuation of the X-ray beam due to the presence of nanoparticles. In particular, it is quite evident that particles are mainly located at the surface. The pictures show a fair penetration with a large gradient from the surface to the inner part. This is an obvious consequence of the large amount of alum still filling the pores (white bright areas in the figures).

The analysis of FTIR spectra of alum-treated wood samples clearly shows that alum and wood presented overlapping bands in the region 1700–1500 and 1200–800 cm^{-1} . The high level of degradation of wood is further confirmed by the lack of the characteristic peaks from cellulose and hemicelluloses, in particular, the signals at 898 cm^{-1} (C1 group frequency in cellulose + hemicellulose; C–H deformation cellulose; glucose ring stretch, cellulose [18]), 1158 cm^{-1} (asymmetric C–O–C vibration carbohydrate [19]), 1371 cm^{-1} (symmetric C–H deformation carbohydrate + C–H bending

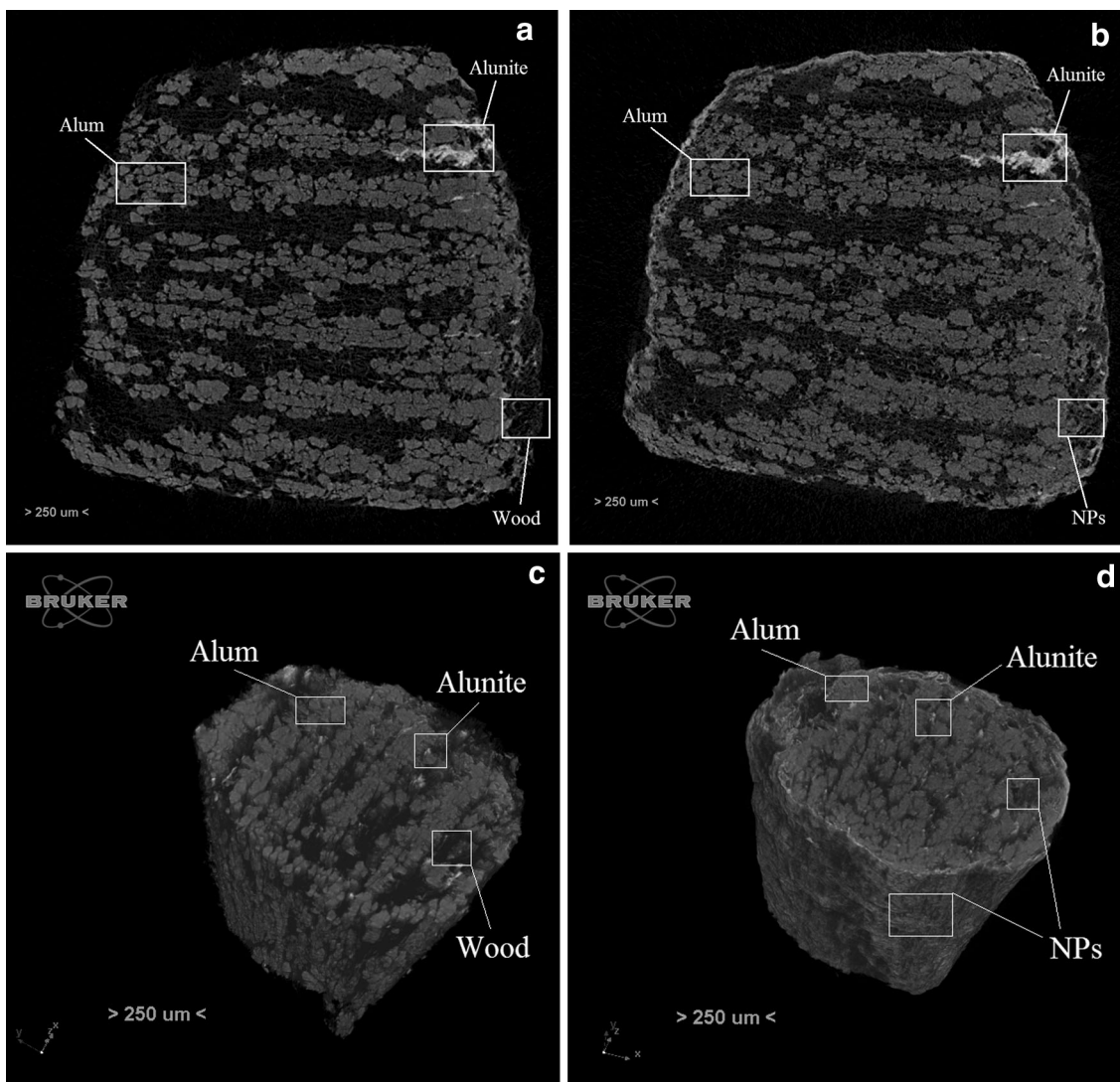


Fig. 5 Sample AaHW 0001 cross section before (a) and after deacidification treatment (b). 3D reconstruction of wooden splinter taken from AaHW 0001 sample before (c) and after deacidification treatment (d)

carbohydrate [20]) and 1738 cm^{-1} (COO stretch of ester from hemicellulose [20]). Despite the overlapping bands from alum and wood, aromatic skeletal vibration attributed to lignin at 1505 cm^{-1} [18] was observed. However, the presence of peak at 1713 cm^{-1} (assigned to carbonyl stretching [18]) suggests that the lignin has been subjected to oxidation.

Figures 6 and 7 show that alkaline particles were capable of neutralizing most of the acidity in the alum-treated Oseberg sample (Fragment 1). Spectra from the Oseberg sample treated with nanoparticles show the presence of a band at 659 cm^{-1} that can be attributed to antisymmetric bending vibration mode of SO_4^{2-} [21], which results from the neutralization reaction of sulphuric acid with alkaline

nanoparticles. The neutralization reaction is fast and complete, and this accounts for the lack of signals from calcium hydroxide at 3621 cm^{-1} .

Calcium hydroxide also reacts spontaneously with CO_2 forming CaCO_3 , as evidenced by FTIR analysis in Fig. 8, where the asymmetric stretching of CO_3^{2-} at 1409 cm^{-1} is shown for treated sample ApSW 0004. According to FTIR data, a complete carbonation, at room conditions, has occurred in 2 weeks.

In principle, the application of alkaline nanoparticles can be repeated with the purpose to further increase the pH of the Oseberg wooden samples. A slight excess of particles may provide the artefact with an alkaline buffer against recurring acidity [22].

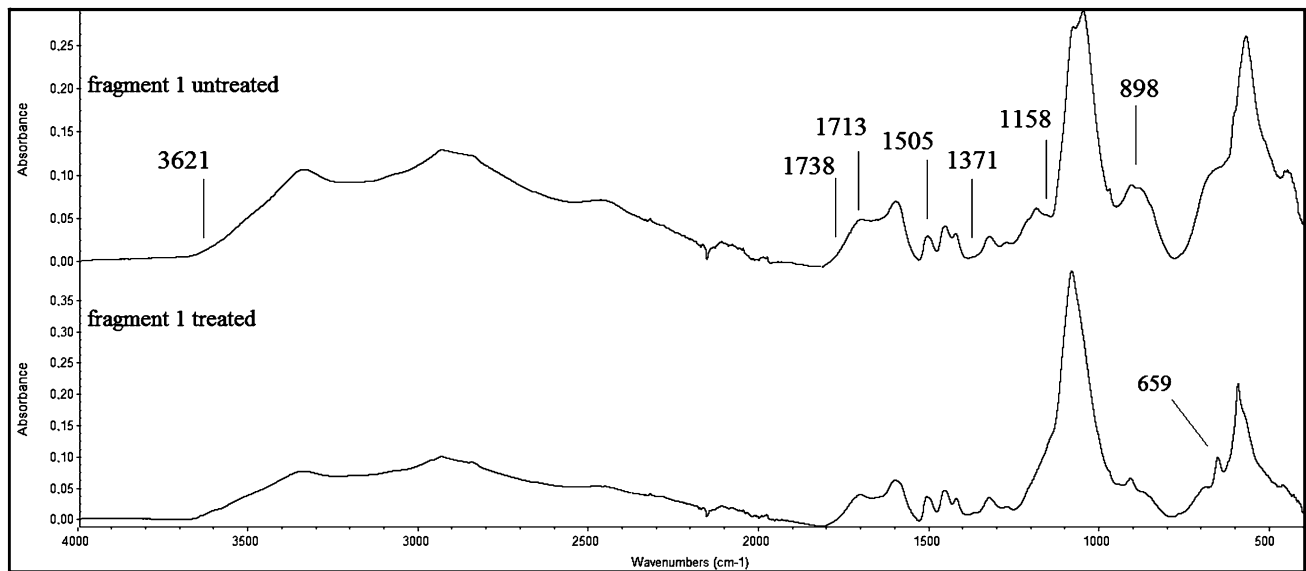


Fig. 6 Infrared spectra of the region 4000–400 cm^{-1} of the Oseberg sample (fragment 1) before and after deacidification treatment. Lacking of peaks attributed to carbohydrates modes explains the high grade of degradation of the sample

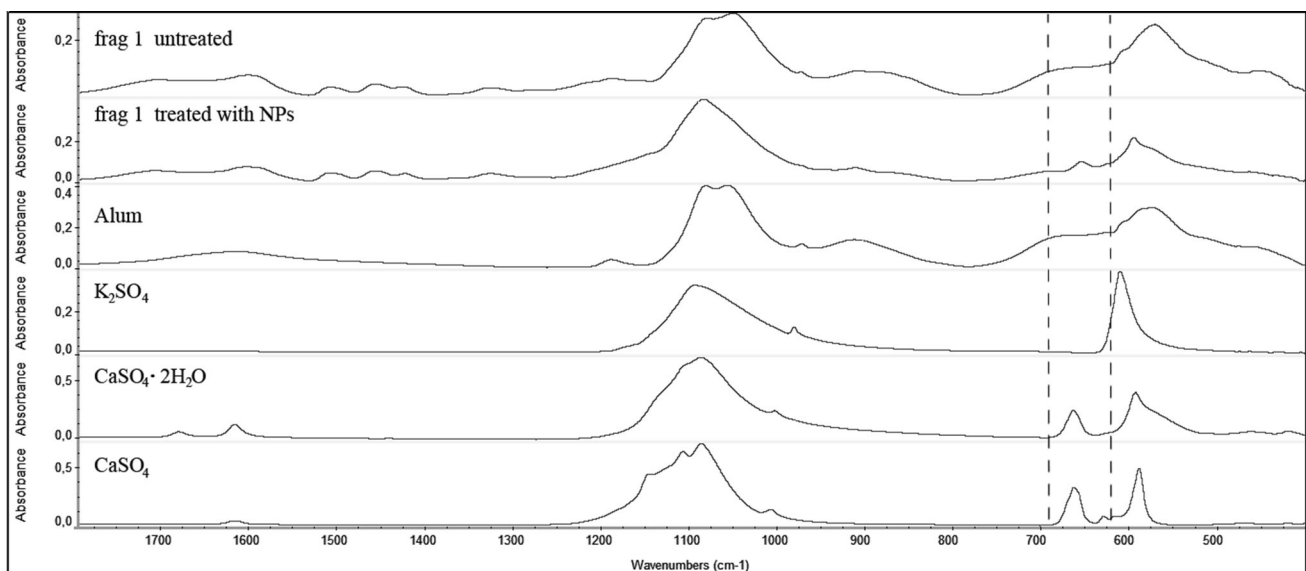


Fig. 7 Infrared spectra of the fingerprint region (1800–400 cm^{-1}) of Oseberg sample (fragment 1) before and after deacidification treatment, Alum, K_2SO_4 , $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$ and CaSO_4 . Treated fragment 1 shows the presence of calcium sulphate anhydrous (Anhydrite)

4 Conclusion

The application of alkaline nanoparticles dispersed in alcohol on some selected archaeological wooden finds has produced a pH increase (2–3 units) in line with the expectations. However, the investigations carried out on the archaeological samples have shown a dramatic scenario, where cellulose is almost lost and the wood samples are composed of lignin and alum only. The penetration of particles is partially inhibited by alum that is filling the

pores. However, final pH at 5–5.5 that is quite close to the original pH of hardwood may provide some benefit to the selected samples, as evidenced by thermal analysis. This study has provided important information about the composition of archaeological alum-treated wood and about the distribution of alum inside. At the moment, alum is playing a key role in the mechanical stabilization of wood; however, it is also the main source of acidity, and for that reason, it is mandatory to remove as much alum as possible, in order to allow a deeper and more homogeneous

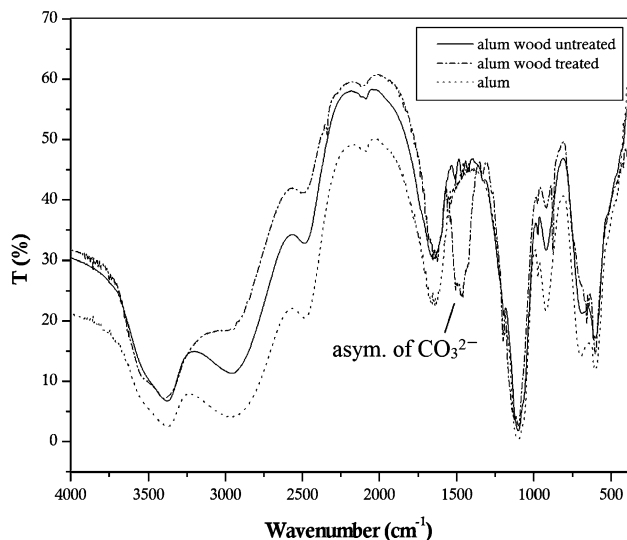


Fig. 8 Infrared spectra of the region 4000–400 cm^{-1} of alum and ApSW 0004 sample. Infrared spectra of wooden sample were acquired before and after treatment with slight excess of nanoparticles

penetration of alkaline nanoparticles and possibly to facilitate the injection of hybrid organic/inorganic materials, such as alkaline particles with functionalized cellulose, to produce a more compatible strengthening network within the wood porosity.

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References

1. N. Bonde, A.E. Christensen, *Antiquity* **67**, 575 (1993)
2. A.W. Brøgger, H. Shetelig, H. Falk, *Osebergfundet* (Distribuert ved Universitetets Oldsaksamling, Oslo, 1917)
3. B.B. Christensen, *Studies in Museum Technology I: The Conservation of Waterlogged Wood in the National Museum of Denmark* (The National Museum of Denmark, Copenhagen, 1970)
4. S. Braovac, H. Kutzke, *J. Cult. Herit.* **13**, 203 (2012)
5. G. Almkvist, I. Persson, *Holzforschung* **62**(6), 694 (2008)
6. R. Giorgi, D. Chelazzi, P. Baglioni, *Langmuir* **21**, 10743 (2005)
7. Y. Fors, M. Sandstrom, *Chem. Soc. Rev.* **35**, 399 (2006)
8. P. Calvini, V. Grosso, M. Hey, L. Rossi, L. Santucci, *Pap. Conserv.* **12**, 35 (1988)
9. G. Poggi, N. Toccafondi, L.N. Melita, J.C. Knowles, L. Bozec, R. Giorgi, P. Baglioni, *Appl. Phys. A* **114**, 685 (2014)
10. P. Baglioni, D. Chelazzi, *Nanoscience for the Conservation of Works of Art* (RSC Publishing, London, 2013)
11. P. Baglioni, R. Giorgi, *Soft Matter* **2**, 293 (2006)
12. P. Baglioni, D. Chelazzi, R. Giorgi, G. Poggi, in *Encyclopedia of Surface and Colloid Science*, 2nd edn., ed. by P. Somasundaran (Taylor & Francis, New York, 2012), pp. 1–16
13. D. Chelazzi, R. Giorgi, P. Baglioni, *Macromol. Symp.* **238**, 30 (2006)
14. C.F. Forney, D.G. Brandl, *HortTechnology* **2**, 52 (1992)
15. J.W. Baty, W. Minter, S. Yong Lee, *Book Pap. Group Annu.* **29**, 113 (2010)
16. M. Földvári, in *Handbook of Thermogravimetric System of Minerals and its Use in Geological Practice*, Occasional Papers of the Geological Institute of Hungary, vol 213 (Geological Institute of Hungary, Budapest, 2011), pp. 216–217
17. H.A. Shnawa, *Mater. Sci. Appl.* **2**, 692 (2011)
18. K.K. Pandey, *J. Appl. Polym. Sci.* **71**, 1969 (1999)
19. K.K. Pandey, A.J. Pitman, *Int. Biodeterior. Biodegradation* **52**, 151 (2003)
20. B. Mohebbi, *Int. Biodeterior. Biodegradation* **55**, 247 (2005)
21. Y. Liu, A. Wang, J.J. Freeman, in *Raman, MIR, and NIR Spectroscopic study of calcium sulfates: gypsum, bassanite, and anhydrite*, 40th Lunar and Planetary Science Conference (2009)
22. F.A. Andersen, L. Brečević, *Acta Chem. Scand.* **45**, 1018 (1991)