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## Impaired surface $\alpha\beta\gamma$ GABA<sub>A</sub> receptor expression in familial epilepsy due to a *GABRG2* frameshift mutation

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### Abstract

The purpose of the study was to explore the pathogenic mechanisms underlying generalized epilepsy and febrile seizures plus (GEFS+) in a family with a novel  $\gamma 2$  subunit gene (*GABRG2*) frameshift mutation. Four affected and one unaffected individuals carried a c.1329delC *GABRG2* mutation resulting in a subunit [ $\gamma 2S(S443delC)$ ] with a modified and elongated carboxy-terminus that is different from that of the wildtype  $\gamma 2S$  subunit. We expressed the wildtype  $\gamma 2S$  subunit and the predicted mutant  $\gamma 2S(S443delC)$  subunit cDNAs in HEK293T cells and performed immunoblotting, flow cytometry and electrophysiology studies. The mutant subunit was translated as a stable protein that was larger than the wildtype  $\gamma 2S$  subunit and was retained in the ER and not expressed on the cell surface membrane, suggesting *GABRG2* haploinsufficiency. Peak GABA-evoked currents recorded from cells cotransfected with wildtype  $\alpha 1$  and  $\beta 2$  subunits and mutant  $\gamma 2S$  subunits were significantly decreased and were comparable to  $\alpha 1\beta 2$  receptor currents. S443delC is the first GABR epilepsy mutation predicted to abolish the natural stop codon and produce a stop codon in the 3' UTR that leads to translation of an extended peptide. The GEFS+ phenotype observed in this family is likely caused by  $\gamma 2S$  subunit loss-of-function and possibly to dominant-negative suppression of  $\alpha 1\beta 2\gamma 2$  receptors. Many *GABRG2* truncation mutations result in GEFS+, but the spectrum of phenotypic severity is wider, ranging from asymptomatic individuals to the Dravet syndrome. Mechanisms influencing the severity of the phenotype are therefore complex and difficult to correlate with its demonstrable functional effects.

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## Keywords

GABA<sub>A</sub> receptor; GABRG2; GABRG2(S443delC); GEFS+; Epilepsy

## Introduction

Mutations in GABA<sub>A</sub> receptor subunit genes have been associated with generalized epilepsy syndromes and with the genetic epilepsy with febrile seizures plus (GEFS+) spectrum, including Dravet syndrome, in rare families and in sporadic cases with *de novo* mutations (Macdonald et al., 2010). Epilepsy-associated mutations in *GABRG2* include three missense mutations in coding sequences (Audenaert et al., 2006; Baulac et al., 2001; Wallace et al., 2001), three nonsense mutations in coding sequences (Harkin et al., 2002; Hirose, 2006; Sun et al., 2008), and one mutation in an intron splice donor site (Kananura et al., 2002) that was shown to cause protein truncation (Tian and Macdonald, 2012). Missense mutations impaired GABA<sub>A</sub> receptor membrane trafficking (Eugene et al., 2007; Frugier et al., 2007; Kang and Macdonald, 2004; Sancar and Czajkowski, 2004b), decreased receptor currents (Bianchi et al., 2002; Eugene et al., 2007), or affected  $\gamma 2$  subunit biogenesis (Audenaert et al., 2006). Phenotypes associated with missense mutations are relatively mild and include familial childhood absence epilepsy and febrile seizures (Wallace et al., 2001), GEFS+ without Dravet syndrome (Baulac et al., 2001) and febrile seizures (Audenaert et al., 2006). Nonsense mutations in coding sequences result in a combination of degradation of unstable subunit mRNA and production of unstable truncated subunits that produce dominant-negative suppression of the biogenesis of wild type subunits (Kang et al., 2009a, 2009b) and cause considerable loss of inhibition, consistent with the more severe phenotypes (Kang et al., 2009a).

We studied a family with mild generalized epilepsy and febrile seizures in which affected individuals carried a novel frame shift mutation of *GABRG2*, resulting in a subunit predicted to lose the last 24 C-terminal amino acids (aas) and gain 50 aas different from those of the natural variant, with consequent lower hydrophobicity of the C-terminus. This is the first *GABR* epilepsy mutation predicted to abolish the natural stop codon and produce a stop codon in the 3' UTR, thus producing an extended subunit peptide. The subunit mRNA should be stable and should produce  $\gamma 2S$  subunits with a disrupted 4th transmembrane domain and an extended C-terminal tail. To explore the pathogenic mechanisms underlying this novel mutation, we coexpressed wildtype  $\gamma 2S$  or predicted mutant  $\gamma 2S(S443delC)$  subunit cDNA with wildtype  $\alpha 1$  and  $\beta 2$  subunit cDNAs in HEK293T cells.

## Materials and methods

### Patients

We studied a non-consanguineous Italian family comprised of 4 affected members and a healthy carrier (Fig. 1A). Clinical features observed in affected individuals are summarized in Table 1. The proband (III:4), a 5-year-old boy, was brought to medical attention at 9 months after a febrile seizure, lasting less than a minute. He experienced 7 subsequent seizures until age 3, always during fever. At 19 months, his neurological examination was normal, and the Griffiths developmental scale general quotient was 96. His EEG showed a normal background activity with rare bursts of generalized epileptiform abnormalities during sleep (Fig. 1B), and his brain MRI was normal. No treatment was assigned. The proband's sister (III:3) was a healthy 7-year-old girl. The proband's 35-year-old mother (II:3) had a single febrile and several nonfebrile generalized convulsive seizures starting at 6 months and recurring during infancy, especially in sleep. She was initially resistant to phenobarbital but responded to valproate and remained seizure free from age 5 to 8 while on

this drug. At age 8, therapy was discontinued. A single seizure occurred again at age 16. Her EEG, at age 20, showed generalized bursts of slow waves. She had normal cognitive abilities and her brain MRI was normal. The proband's 49-year-old uncle (II:2) had experienced a few febrile seizures in infancy. His 10-year-old son (III:1) had only had a nonfebrile generalized seizure while awake at age 9. The proband's 70-year-old grandmother (I:1) did not recall having ever been told that she experienced seizures. The overall family clustering of clinical features is consistent with generalized epilepsy with febrile seizures plus (GEFS+). After obtaining informed consent we extracted genomic DNA from peripheral blood of affected family members (II:2, II:3, III:1 and III:4) and their healthy relatives (I:1 and II:4). The study was approved by the Commission for Medical Ethics of the Meyer's University Hospital.

### GABRG2 mutation analysis

We initially analyzed the *SCN1A* gene in the proband (III:4) without identifying any mutation (identified SNPs are listed in Supplementary Table 1). We subsequently carried out mutation analysis of *GABRG2* in the proband (III:4) and extended the genetic study to available family members (I:1, II:2, II:3, II:4, III:1) (Fig. 1A). DNA was extracted from peripheral blood leukocytes using the QIASymphony automated DNA isolation robot (QIAGEN, Germany), according to the manufacturer's protocol. The 9 exons covering the coding regions of *GABRG2* (NCBI Reference Sequence: NM\_000816.3) and their respective intron-exon boundaries were amplified by PCR and cycle sequenced using the BigDye Terminator v.1.1 chemistry (LIFE Technologies, USA). The sequence reactions were analyzed on a 3130XL sequencer (LIFE Technologies, Carlsbad, CA, USA). Primer sequences and PCR/sequencing conditions are available on request. The identified *GABRG2* alteration was not found in a control population of 190 ethnically matched subjects and was described according to nomenclature using the cDNA sequence NM\_000816.3.

### Expression vectors with GABA<sub>A</sub> receptor subunits

The coding sequences of human  $\alpha 1$ ,  $\beta 2$  and  $\gamma 2S$  subunits from translation initiation to stop codon were cloned into pcDNA3.1 expression vectors (Invitrogen) as described (Gallagher et al., 2005). The HA peptide sequence, YPYDVPDYA, was introduced between 4th and 5th aas of the mature  $\gamma 2S$  subunit, a functionally silent position (Connolly et al., 1996), to create  $\gamma 2S^{HA}$  and  $\gamma 2S(S443delC)^{HA}$  subunits. The  $\gamma 2$  subunit 3' polyA site fragments were cloned from RP11-1035I20 (BACPAC Resources; <http://bacpac.chori.org>). PCR primer sequence is available upon request. We designated the positions of  $\gamma 2$  subunit mutations in the immature peptide.

### Cell culture and transfection

Human embryonic kidney cells (HEK 293T) (ATCC, CRL-11268) were incubated at 37 °C in humidified 5% CO<sub>2</sub>, 95% air and grown in Dulbecco's modified Eagle's medium (Invitrogen) supplemented with 10% fetal bovine serum, 100 IU/ml penicillin, and 100  $\mu$ g/ml streptomycin (Invitrogen). Cells were transfected using FuGENE 6 transfection reagent (Roche Applied Science) at a DNA:Transfection Reagent ratio of 1:3. Transfected cells were harvested after 36 h in culture.

### Flow cytometry

Flow cytometry was performed to evaluate total  $\gamma 2$  subunit levels as described previously (Lo et al., 2008). For each condition, 50,000 transfected cells were immunostained with Alexa-647 fluorophore conjugated anti-HA antibodies (clone 16B12, Covance). Data were acquired using FACSDiva 6.0 (BD Biosciences) and analyzed offline using FlowJo 7.5 (Treestar, Inc.). Mean fluorescence intensity of each sample was evaluated and normalized

to the control ( $\alpha 1\beta 2\gamma 2S^{HA}$ ). Normalized mean fluorescence intensity was represented as a percentage of control. Data were plotted as mean $\pm$ SEM. Pair-wise two-tailed Student's t-tests were used to compare conditions unless otherwise specified.

### Immunoblotting

HEK 293T cells were lysed in radioimmune precipitation assay (RIPA) buffers (Pierce) and a protease inhibitor cocktail (Sigma Aldrich). Proteins in total cell lysates were separated with the NuPage® SDS-PAGE system (Invitrogen), transferred to a Millipore Immobilon® FL PVDF Membrane (Millipore), and blotted for Odyssey infrared imaging system (Li-cor). Monoclonal anti-HA epitope tag antibodies (0.2  $\mu$ g/ml; clone 16B12, Covance) were used to detect HA epitope-tagged  $\gamma 2$  subunits. Anti-Na<sup>+</sup>/K<sup>+</sup>-ATPase antibodies (0.2  $\mu$ g/ml; clone ab7671, Abcam) were used to check loading variability, and IRDye® secondary antibodies were used at a 1:10,000 $\times$  dilution (Li-cor).

### Immunocytochemistry and confocal microscopy

Transfected HEK293T cells were fixed with 1% paraformaldehyde to stain surface proteins or permeabilized with CytoPerm (BD Biosciences) to stain total proteins. The fixed/permeabilized cells were stained with Alexa 488 conjugated mouse monoclonal HA antibodies (Covance) and Alexa 647 conjugated mouse monoclonal  $\alpha 1$  subunit antibodies (Millipore). Confocal experiments were performed in part using the VUMC Cell Imaging Shared Resource (supported by NIH grants CA68485, DK20593, DK58404, HD15052, DK59637 and EY08126). Images were obtained from 1  $\mu$ m optical sections from HEK293T cells using a Zeiss LSM 510 META inverted confocal microscope with 8-bit, 1024 $\times$ 1024 pixel resolution. An average of 4 scans was taken to decrease the background noise.

### Electrophysiology

Whole-cell voltage-clamp recordings from lifted HEK293T cells were performed at room temperature 24–72 h after subunit transfection as described previously (Hernandez et al., 2011). Cells were bathed in an external solution (in mM: NaCl 142; KCl 8; MgCl<sub>2</sub> 6; CaCl<sub>2</sub> 1; HEPES 10; glucose 10, pH 7.4). Recording electrodes were fire-polished to resistances of 1.0–1.5 M $\Omega$  and filled with an internal solution (in mM: KCl 153; MgCl<sub>2</sub> 1; MgATP 2; HEPES 10; EGTA 5, pH 7.3), resulting in an E<sub>Cl</sub> of  $\sim$ 0 mV. Cells were voltage clamped at  $-20$  mV. GABA (1 mM) was applied to cells for 4 s, and cells were then washed with external solution for 40 s. Zn<sup>2+</sup> (10  $\mu$ M) was then pre-applied for 8 s followed by co-application of GABA (1 mM) and Zn<sup>2+</sup> (10  $\mu$ M) for 4 s. Whole-cell currents were low-pass filtered at 2 kHz, digitized at 5–10 kHz, and analyzed using the pClamp9 software suite (Axon Instruments).

## Results

### The c.1329delC deletion in GABRG2 is predicted to cause an open-reading frame shift and generate a novel $\gamma 2$ subunit C-terminus

The proband (III:4) had a heterozygous c.1329delC deletion in the last exon of *GABRG2*, which was also present in individuals I:1, II:2, II:3 and III:1 (Fig. 1A). There are two *GABRG2* intron 8 alternative splice variants (Whiting et al., 1990). The  $\gamma 2S$  subunit is the default splice variant while expression of  $\gamma 2L$  subunits requires the neuron-specific RNA binding protein Nova (Dredge and Darnell, 2003). The mutation should affect both variants, which have similar physiological functions (Baer et al., 2000; Homanics et al., 1999; Quinlan et al., 2000). We introduced the c.1329delC mutation into the  $\gamma 2S$  variant and studied its function.

The c.1329delC mutation deleted a cytosine nucleotide in the Ser443 codon TCC, predicted to cause open-reading frame shift and result in loss of the natural stop codon and generation of a new stop codon in the 3' UTR (p.Tyr444MetfsX51). Specifically, the mutant  $\gamma$ 2S subunit, named  $\gamma$ 2S(S443delC), was predicted to lose the last 24 C-terminal aas and gain 50 aas that differed from those of the natural variant (Fig. 2). The TMPred-calculated hydrophobicity (Krogh et al., 2001) of the mutant subunit showed that its C-terminus hydrophobicity was lower compared to the same region of the wildtype subunit (Supplementary Fig. 2C). The TMHMM program (Sonnhammer et al., 1998) did not identify any transmembrane domain in the novel C-terminus (Supplementary Fig. 2D). The intrinsic disorder of  $\gamma$ 2S(S443delC) subunit C-terminus was significantly increased as well (Supplementary Fig. 2E) (Ishida and Kinoshita, 2007; Schlessinger and Rost, 2005). Thus, the S443delC mutation likely disrupted  $\gamma$ 2S subunit membrane topology.

The *GABRG2* genomic sequence carries two polyA sites, 800 bp and ~2.4 kbp downstream of the stop codon. We cloned genomic sequence containing either the proximal polyA site or both proximal and distal polyA sites to  $\gamma$ 2S subunit cDNA and introduced the S443delC mutation. It is unclear whether C-terminal extension of the subunit into the 3' UTR would alter polyA site usage. When wildtype or mutant  $\gamma$ 2S subunits were expressed in HEK293T cells, all four subunits had the same 3'-UTR, suggesting that the mutation did not interfere with polyA site recognition (Supplementary Fig. 1). Sequences of the 3'-UTR fragments showed that they all utilized the proximal polyA site. Sequence of the mutant  $\gamma$ 2S(S443delC) subunit showed that, as predicted, the mutation caused a frame shift in exon 9 and generated a novel 50 aas C-terminus. The mutant  $\gamma$ 2S(S443delC) subunit premature peptide was 493 aas while the wildtype  $\gamma$ 2S subunit premature peptide was 467 aas, a difference of 26 aas.

### **$\gamma$ 2S(S443delC) subunits were present intracellularly, but their total level was significantly lower than $\gamma$ 2S subunits**

The  $\gamma$ 2S(S443delC)<sup>HA</sup> subunit was translated to a stable protein in HEK293T cells when expressed alone, but its molecular size was larger than the wildtype  $\gamma$ 2S<sup>HA</sup> subunit (Fig. 3A, lanes 2 and 3). Coexpression with  $\alpha$ 1 and  $\beta$ 2 subunits altered the glycosylation pattern of wildtype  $\gamma$ 2S<sup>HA</sup> subunits (Fig. 3A, lanes 2 and 5, see double bands in lane 5 but not lane 2) but did not affect that of mutant  $\gamma$ 2S(S443delC)<sup>HA</sup> subunits (Fig. 3A, lanes 3 and 6). The epilepsy-associated mutation, Q390X, generated a  $\gamma$ 2S subunit that was truncated in the second intracellular loop (Harkin et al., 2002). The  $\gamma$ 2S(Q390X)<sup>HA</sup> subunit was an intracellular protein that only had endoplasmic reticulum (ER) core glycosylation (Kang et al., 2010). The  $\gamma$ 2S(Q390X)<sup>HA</sup> subunit protein bands were much smaller than wildtype  $\gamma$ 2S<sup>HA</sup> or mutant  $\gamma$ 2S(S443delC)<sup>HA</sup> subunit bands (Fig. 3A, lane 4). The glycosylation pattern of the  $\gamma$ 2S(Q390X)<sup>HA</sup> subunit was also unaffected by coexpression with  $\alpha$ 1 and  $\beta$ 2 subunits (Fig. 3A, lanes 4 and 7). These data suggested that both  $\gamma$ 2S(S443delC)<sup>HA</sup> and  $\gamma$ 2S(Q390X)<sup>HA</sup> subunits were post-translationally modified in the ER, but that neither subunit was trafficked to the Golgi apparatus.

The Western blot result also suggested that the  $\gamma$ 2S(S443delC)<sup>HA</sup> subunit had decreased protein levels with expression of either single subunits or with coexpression with  $\alpha$ 1 and  $\beta$ 2 subunits. We used flow cytometry to evaluate total levels of  $\gamma$ 2S(S443delC)<sup>HA</sup> subunits with both single subunit expression or coexpression with  $\alpha$ 1 and  $\beta$ 2 subunits (Fig. 3B). When expressed alone, the  $\gamma$ 2S<sup>HA</sup> subunit total level was  $86.6 \pm 1.6\%$ , and when coexpressed with  $\alpha$ 1 and  $\beta$ 2 subunits, the  $\gamma$ 2S<sup>HA</sup> subunit total level was increased slightly ( $p=0.01$ ). However, the total level of  $\gamma$ 2S(S443delC)<sup>HA</sup> subunits was only  $34.6 \pm 4.6\%$  when expressed alone as a single subunit or  $35.7 \pm 6.0\%$  when coexpressed with  $\alpha$ 1 and  $\beta$ 2 subunits. The total level of  $\gamma$ 2S(S443delC)<sup>HA</sup> subunits was not changed by coexpression with  $\alpha$ 1 and  $\beta$ 2



subunits ( $p=0.74$ ), but was significantly lower than the  $\gamma 2S$  subunit total level either with single subunit expression or  $\alpha 1\beta 2\gamma 2S$  subunit coexpression ( $p<0.01$  in both cases).

### **$\gamma 2S(S443delC)$ subunits were not expressed on the cell membrane**

GABA<sub>A</sub> receptor subunits are synthesized in the ER where they are assembled to heteropentameric receptors and then trafficked to the cell membrane (Connolly et al., 1996). Misfolded and unassembled subunits are retained in the ER and degraded by the ubiquitin–proteasome system (Gallagher et al., 2005; Jacob et al., 2008). Because  $\gamma 2S(S443delC)$  subunits might have four transmembrane domains similar to wildtype subunits, we asked if  $\gamma 2S(S443delC)$  subunits could assemble with  $\alpha 1$  and  $\beta 2$  subunits and traffick to cell membranes as functional receptors. The  $\alpha 1$  subunits can assemble with  $\beta 2$  subunits to form functional  $\alpha 1\beta 2$  receptors (Angelotti and Macdonald, 1993; Gunther et al., 1995). Confocal images from cells cotransfected with  $\alpha 1\beta 2\gamma 2S^{HA}$  subunits showed the HA tag signal in both permeabilized and unpermeabilized cells, suggesting that wildtype  $\gamma 2S^{HA}$  subunits were present intracellularly and on the cell surface (Fig. 4A). HA and  $\alpha 1$  subunit signals were colocalized in both total and surface conditions. With coexpression of  $\alpha 1\beta 2\gamma 2S(S443delC)^{HA}$  subunits, HA signal was only detected in permeabilized cells (Fig. 4B), suggesting that mutant subunits were retained in the ER.

### **GABA-evoked currents of $\alpha 1\beta 2\gamma 2S(S443delC)$ receptors were significantly decreased**

We characterized the effect of the S443delC mutation on GABA receptor function. Whole-cell currents were elicited from lifted HEK293T cells cotransfected with  $\alpha 1\beta 2$  subunits and wildtype  $\gamma 2S$  or mutant  $\gamma 2S(S443delC)$  subunits by applying a saturating GABA concentration (1 mM) for 4 s (Fig. 5). As expected, current densities with expression of  $\alpha 1\beta 2\gamma 2S(S443delC)$  subunits were similar ( $251\pm 32$  pA/pF,  $n=28$ ) to those with  $\alpha 1\beta 2$  subunits ( $193\pm 24$  pA/pF,  $n=16$ ,  $p>0.05$ ) (Fig. 5A, left traces) but were ~70% smaller than those with  $\alpha 1\beta 2\gamma 2S$  subunits ( $807\pm 26$  pA/pF,  $n=36$ ,  $p<0.001$ ) (Fig. 5B, left trace). Moreover, currents from cells cotransfected with  $\alpha 1\beta 2\gamma 2S(S443delC)$  subunits were significantly more sensitive to  $Zn^{2+}$  inhibition than those with  $\alpha 1\beta 2\gamma 2S$  subunits (Fig. 5C). Currents evoked by 1 mM GABA from cells cotransfected with  $\alpha 1\beta 2\gamma 2S(S443delC)$ ,  $\alpha 1\beta 2\gamma 2S$  and  $\alpha 1\beta 2$  subunits were differentially inhibited by co-application of 10  $\mu M$   $Zn^{2+}$  (Fig. 5A and B, right traces). The fractional  $Zn^{2+}$  inhibition of currents from cells with  $\alpha 1\beta 2\gamma 2S(S443delC)$  subunits was higher than those with  $\alpha 1\beta 2\gamma 2S$  subunits ( $90\pm 1\%$ ,  $n=28$ ;  $8\pm 2\%$ ,  $n=15$ , respectively,  $p<0.001$ ) and similar to currents obtained from cells with  $\alpha 1\beta 2$  subunits ( $93\pm 1\%$ ,  $n=16$ ,  $p>0.05$ ). These findings suggested that currents recorded from cells coexpressing  $\alpha 1\beta 2\gamma 2S(S443delC)$  subunits were mainly from binary  $\alpha 1\beta 2$  receptors (Angelotti and Macdonald, 1993).

## **Discussion**

We identified a novel *GABRG2* frame shift mutation, S443delC, in an Italian family with GEFS+. The resultant DNA sequence suggested that the mutant allele should be translated to a protein with the last 24 aas of the wildtype  $\gamma 2$  subunit that contains the 4th transmembrane domain replaced by a novel 50 amino acid C-terminus with decreased hydrophobicity. The mutation shifted the stop codon into the 3' UTR, thus shortening it, but it did not interfere with polyA site recognition. In HEK293T cells the mutant  $\gamma 2S(S443delC)$  subunits were detected, but their total level was decreased. When coexpressed with  $\alpha 1\beta 2$  subunits,  $\gamma 2S(S443delC)$  subunits were retained inside cells, and GABA-evoked currents were similar to those from  $\alpha 1\beta 2$  receptors. Thus, the  $\gamma 2$  subunit mutation, S443delC, might generate a

## **$\gamma 2$ subunit null allele and be associated with epilepsy, at least in part, through haploinsufficiency**

There have been 16 epilepsy-associated *GABR* mutations, 7 of which were in *GABRG2*, suggesting its strong association with epilepsy (Macdonald et al., 2010). Although the  $\gamma 2$  subunit is not required for receptor assembly or surface trafficking (Connolly et al., 1996; Gunther et al., 1995),  $\gamma 2^{-/-}$  knockout mice died within 2 weeks of birth (Gunther et al., 1995). Mutations in  $\gamma 2$  subunits affected receptor expression, trafficking, and function (Macdonald et al., 2010). However, no studies of possible seizure phenotypes in  $\gamma 2^{-/-}$  or  $\gamma 2^{+/-}$  mice have been reported.

We demonstrated that the  $\gamma 2$ (S443delC) subunit was retained in the ER and not expressed on the cell surface, suggesting a mechanism of haploinsufficiency. It might also be associated with epilepsy by dominant-negative effects on wildtype subunits assembly and membrane trafficking. The epilepsy-associated  $\gamma 2$ (R82Q) and  $\gamma 2$ (Q390X) subunit mutations also were shown to generate proteins that were retained in the ER (Kang and Macdonald, 2004; Kang et al., 2009a). The mutant  $\gamma 2$ (R82Q) subunit has decreased oligomerization with partnering subunits and is expressed on the cell membrane at low levels (Bianchi et al., 2002; Hales et al., 2005). It decreased surface  $\alpha$  and  $\beta$  subunit levels (Eugene et al., 2007; Kang and Macdonald, 2004; Sancar and Czajkowski, 2004a), as well as GABA-evoked whole-cell current in cultured neurons (Eugene et al., 2007). The  $\gamma 2$ (Q390X) subunit was retained in the ER but not expressed on the cell membrane (Kang et al., 2009a). It bound to  $\alpha 1$  and wildtype  $\gamma 2$  subunits when coexpressed in HEK293T cells and decreased the membrane level of these wildtype subunits. However, both  $\gamma 2$ (R82Q) and  $\gamma 2$ (Q390X) subunits are stable proteins with similar total levels as the wildtype  $\gamma 2$  subunits (Kang and Macdonald, 2004; Kang et al., 2010). The S443delC mutation resulted instead in decreased total protein levels when expressed either as a single subunit or coexpressed with  $\alpha 1$  and  $\beta 2$  subunits.

The  $\alpha 1$ (A322D) mutation is associated with juvenile myoclonic epilepsy (Cossette et al., 2002). The mutation impaired membrane topology of  $\alpha 1$ (A322D) subunits so that it was misfolded, retained in the ER, and degraded by the proteasome (Gallagher et al., 2004, 2005, 2007), resulting in low total levels (Ding et al., 2010; Gallagher et al., 2004). The  $\gamma 2$ (S443delC) subunit has a novel C-terminus that is less hydrophobic than the wildtype C-terminus and was predicted not to fold correctly. The  $\gamma 2$ (S443delC) subunit might have decreased total level because of ER retention and increased proteasomal degradation, although that must be confirmed. However, the  $\alpha 1$ (A322D) subunit associated with wildtype subunits in the ER and reduced both wildtype  $\alpha 1\beta 2\gamma 2$  and  $\alpha 3\beta 2\gamma 2$  receptor surface expression (Ding et al., 2010). It is possible that although  $\gamma 2$ (S443delC) subunits have reduced total levels, they could oligomerize with  $\alpha$  and  $\beta$  subunits and decrease wildtype receptor surface expression. Thus, the S443delC mutation could have a combination of haploinsufficiency and dominant-negative effects. The  $\gamma 2$ (R82Q) mutation also decreased GABA<sub>A</sub> receptor surface expression. Mutant  $\gamma 2^{\text{R82Q/+}}$  knock in mice had the same type of seizures as humans bearing the mutation (Tan et al., 2007), and thus the  $\gamma 2$ (S443delC) subunit could also induce epilepsy by a similar mechanism.

This report confirms that most GABA<sub>A</sub> receptor truncation mutations result in a combination of generalized and febrile seizures, also recognized as GEFS+ spectrum (Macdonald et al., 2010). However, it is becoming increasingly obvious that the spectrum of phenotypic severity is inexplicably wide, ranging from asymptomatic individuals (see individual I:1 in this report and the Q40X mutation carrier (Hirose, 2006) to patients with the Dravet syndrome (Hirose, 2006; Macdonald et al., 2010)). Mechanisms influencing the severity of the phenotype associated with a given mutation are therefore complex and difficult to correlate with its demonstrable functional effects. Phenotypic severity is likely

modulated by the individual genetic background through different and possibly multiple mechanisms, including the response to ER stress (Hirose, 2006). A comparably wide spectrum of phenotypes within the febrile seizures–GEFS+–Dravet Syndrome spectrum is also observed in patients with *SCN1A* mutations. In spite of the very high number of reported *SCN1A* mutations, the mechanisms that lead the mutant protein to cause a given phenotype, rather than any other phenotype within the spectrum, remain largely unknown.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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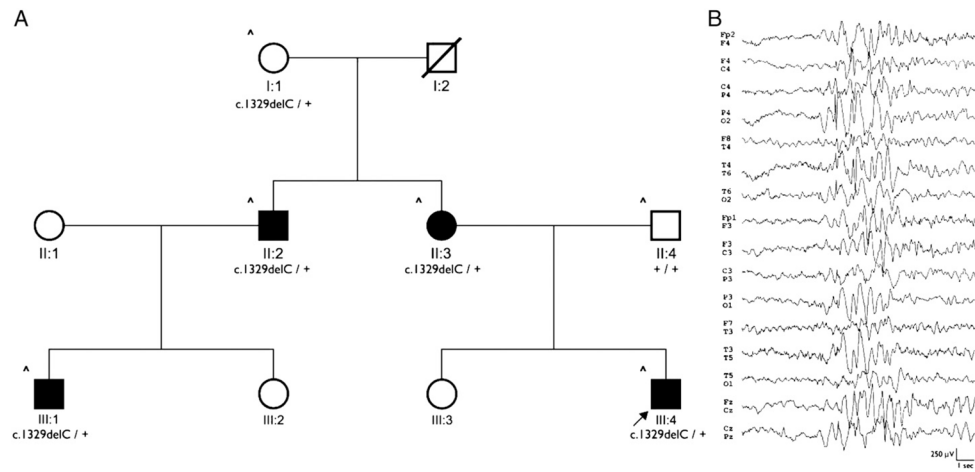
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**Fig. 1.** GABRG2(S443delC) mutation was identified in a GEFS+ family. (A) Pedigree of the family with GEFS+. The arrow points to the proband. (B) EEG from patient III:4 showing a burst of generalized epileptiform abnormalities.

A)  $\gamma 2S$  subunit: wildtype sequence

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1  MSSPNIWSTG SSVYSTPVFS QKMTVWILL LSLYPGFTSQ KSDDDYEDYA SNKTWVLT PK
61  VPEGDVTVIL NNLLEGYDNK LRPDIGVKPT LIHTDMYVNS IGPVNAINME YTIDIFFAQT
121 WYDRRLKFNS TIKVLRRLNSN MVGKIWIPDT FFRNSKKADA HWITTPNRML RIWNDGRVLY
181 TLRLTIDAEC QLQLHNFPM D EHSCPLEFSS YGYPREEIVY QWKRSSVEVG DTRSWRLYQF
241 SFVGLRNTTE VVKTTSGDYV VMSVYFDLSR RMGYFTIQTY IPCTLIVVLS WVSFWINKDA
301 VPARTSLGIT TVLTMTTLST IARKSLPKVS YVTAMDLFVS VCFIFVFSAL VEYGLHYFV
361 SNRKPSKDKD KKKKNPAPT I DIRPRSATI Q MNNATHLQER DEEYGYECLD GKDCASFFCC
421 FEDCRTGAWR HGRIHIRIAK MDSYARIFFP TAFCLFNLVY WVSYLVL*

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B)  $\gamma 2S(S443delC)$  subunit: predicted sequence

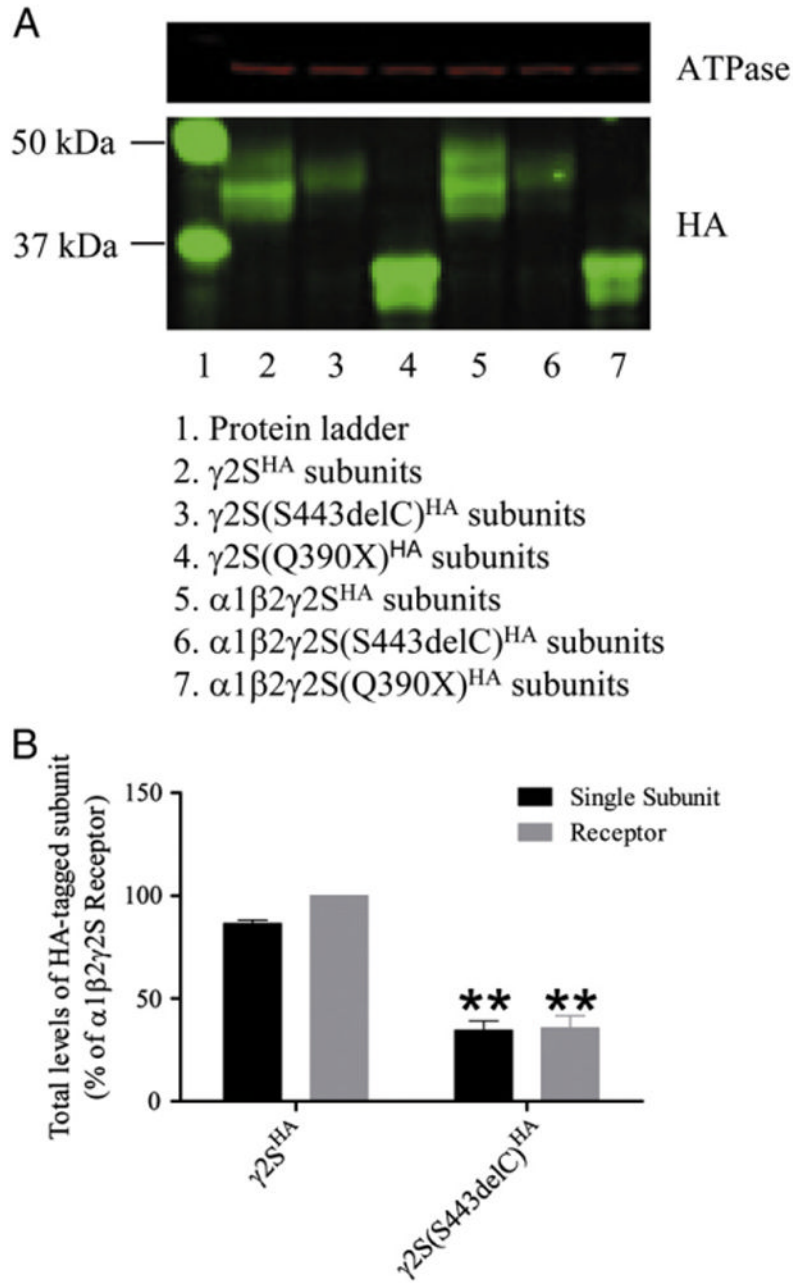
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1  MSSPNIWSTG SSVYSTPVFS QKMTVWILL LSLYPGFTSQ KSDDDYEDYA SNKTWVLT PK
61  VPEGDVTVIL NNLLEGYDNK LRPDIGVKPT LIHTDMYVNS IGPVNAINME YTIDIFFAQT
121 WYDRRLKFNS TIKVLRRLNSN MVGKIWIPDT FFRNSKKADA HWITTPNRML RIWNDGRVLY
181 TLRLTIDAEC QLQLHNFPM D EHSCPLEFSS YGYPREEIVY QWKRSSVEVG DTRSWRLYQF
241 SFVGLRNTTE VVKTTSGDYV VMSVYFDLSR RMGYFTIQTY IPCTLIVVLS WVSFWINKDA
301 VPARTSLGIT TVLTMTTLST IARKSLPKVS YVTAMDLFVS VCFIFVFSAL VEYGLHYFV
361 SNRKPSKDKD KKKKNPAPT I DIRPRSATI Q MNNATHLQER DEEYGYECLD GKDCASFFCC
421 FEDCRTGAWR HGRIHIRIAK MDSMLGSSSP LPSACLIWSI GSPTSTCEEV WVLLIWFLFT
481 ESHGEMSVLS PLK*

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**Fig. 2.**

The mutant  $\gamma 2S(S443delC)$  subunit sequence. The peptide sequence of mutant  $\gamma 2S(S443delC)$  subunit premature peptide is predicted to lose the last 24 C-terminal aas (underlined letters in A) and gain 50 aas different from those of the natural variant (bold letters in B).

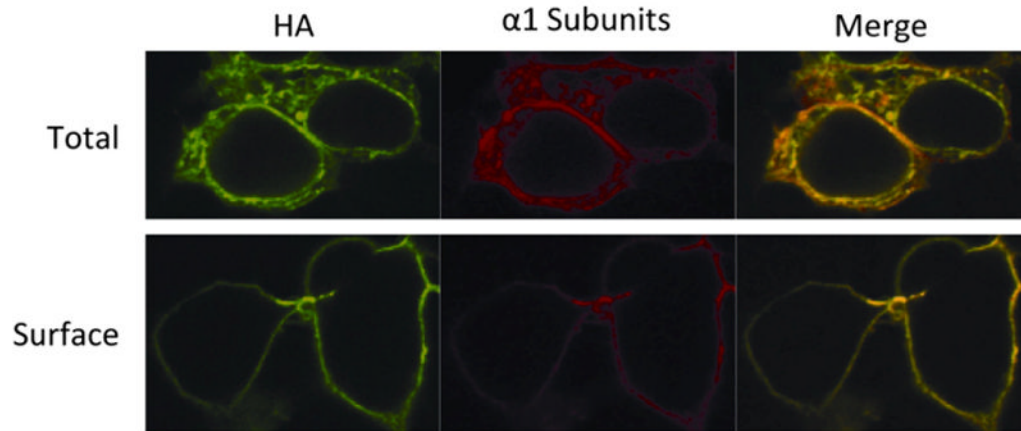
**Fig. 3.**

The  $\gamma 2(S443delC)$  subunits were stable, but their total level was decreased. (A) Western blot was performed on transfected HEK293T cells total cell lysates. We expressed the  $\gamma 2S^{HA}$  or  $\gamma 2S(S443delC)^{HA}$  subunit in HEK293T cells either alone or with  $\alpha 1$  and  $\beta 2$  subunits. The  $\gamma 2S(Q390X)^{HA}$  subunit was also expressed as truncated subunit control. The red channel shows the ATPase antibodies signal, and the green channel shows the HA-antibody signal. Lane 1, protein ladder; lane 2,  $\gamma 2S^{HA}$  subunits; lane 3,  $\gamma 2S(S443delC)^{HA}$  subunits; lane 4,  $\gamma 2S(Q390X)^{HA}$  subunits; lane 5,  $\alpha 1\beta 2\gamma 2S^{HA}$  subunits; lane 6,  $\alpha 1\beta 2\gamma 2S(S443delC)^{HA}$  subunits; lane 7,  $\alpha 1\beta 2\gamma 2S(Q390X)^{HA}$  subunits. (B) Total  $\gamma 2^{HA}$  levels quantified by flow cytometry. The transfected HEK293T cells were permeabilized and stained for HA-tagged  $\gamma 2S^{HA}$  or  $\gamma 2S(S443delC)^{HA}$  subunits using fluorescence conjugated

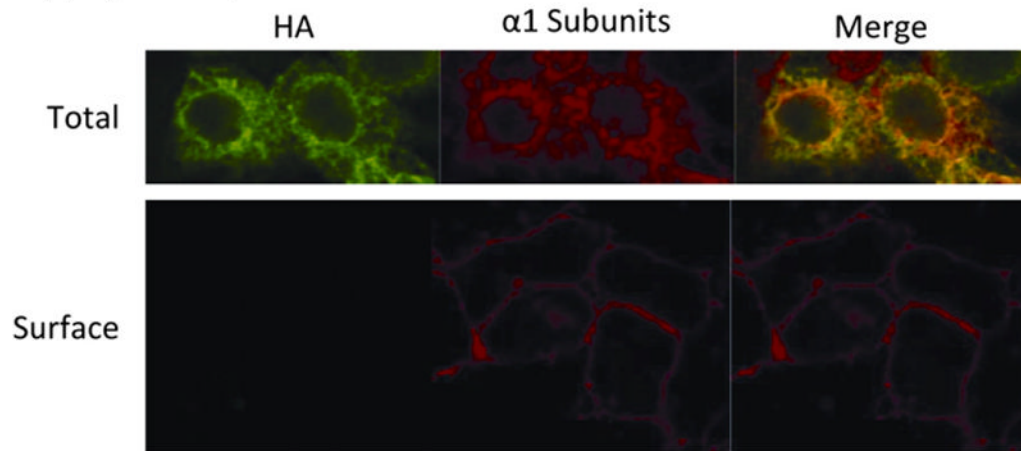


HA antibodies, and mean fluorescence intensities were evaluated by flow cytometry. The total level of HA tag in cells coexpressing  $\alpha 1$ ,  $\beta 2$  and  $\gamma 2S^{HA}$  subunits was set at 100%. The wildtype  $\gamma 2S^{HA}$  subunit and mutant  $\gamma 2S(S443delc)^{HA}$  subunit cDNAs were transfected either with pcDNA empty vector or with  $\alpha 1$  and  $\beta 2$  subunit cDNAs, and the results were expressed relative to the level obtained for the wild type  $\gamma 2S^{HA}$  subunit coexpressed with  $\alpha 1$  and  $\beta 2$   $\gamma 2S^{HA}$  subunits. The double stars correspond to  $p < 0.01$  compared to wildtype  $\alpha 1$   $\beta 2$  and  $\gamma 2S^{HA}$  subunit coexpression.

A) Wildtype  $\gamma 2^{HA}$  subunits were expressed on membrane.

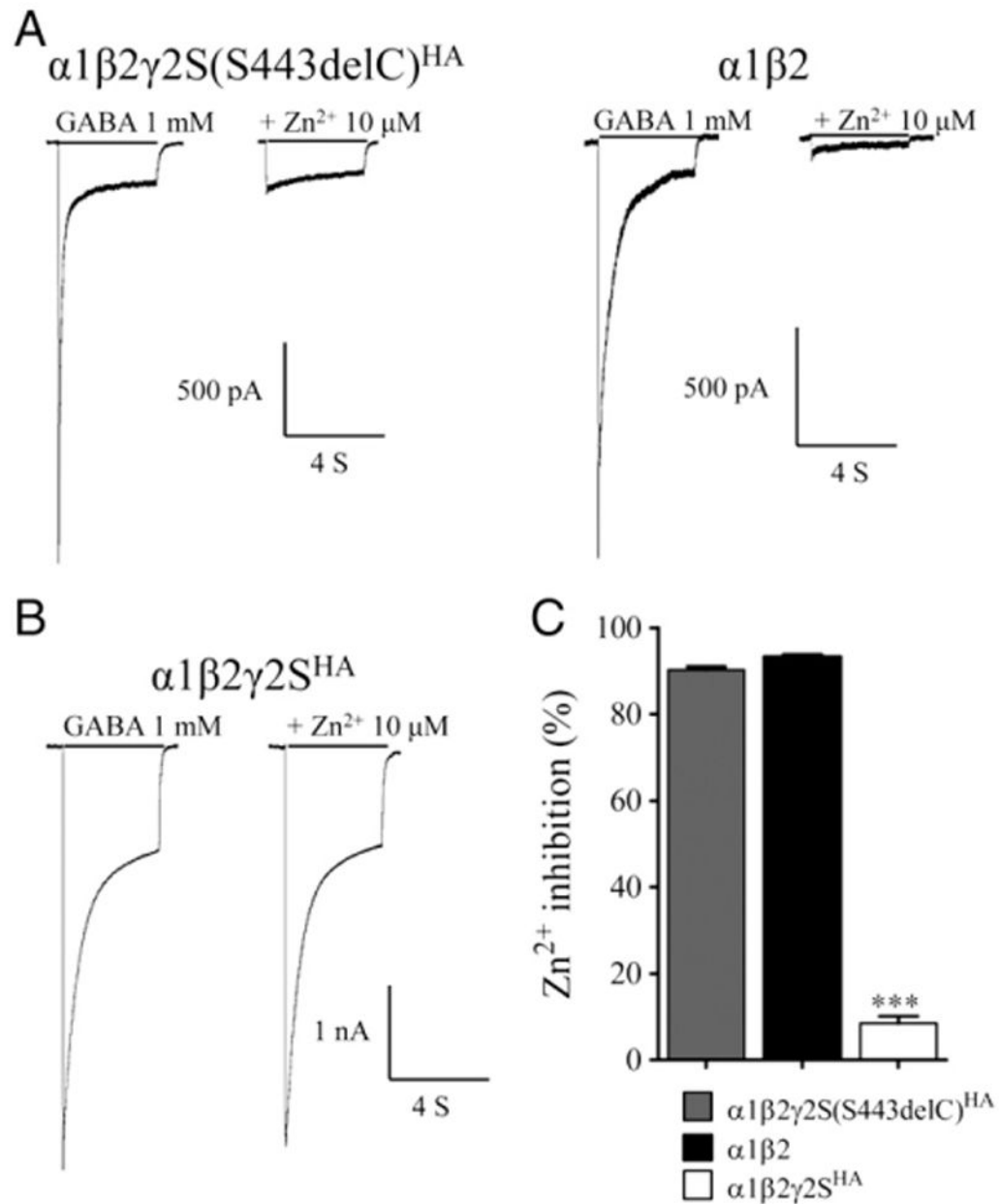


B)  $\gamma 2S(S443delC)^{HA}$  subunits were retained inside cells.



**Fig. 4.**

The  $\gamma 2(S443delC)$  subunits were retained in an intracellular compartment. Confocal images of wildtype  $\gamma 2^{HA}$  (A) and mutant  $\gamma 2S(S443delC)^{HA}$  (B) subunits coexpressed with  $\alpha 1$  and  $\beta 2$  subunits were obtained. We coexpressed  $\gamma 2^{HA}$  or  $\gamma 2S(S443delC)^{HA}$  subunits with  $\alpha 1$  and  $\beta 2$  subunits in HEK293T cells and stained both permeabilized and unpermeabilized cells with antibodies against the  $\alpha 1$  subunit and the HA tag. Total signals were evaluated by staining permeabilized cells, and surface signals were evaluated by staining paraformaldehyde fixed cells.



**Fig. 5.** The  $\gamma 2(S443delC)$  receptors decreased GABA-evoked currents. (A and B) Currents were recorded from lifted whole HEK293T cells coexpressing  $\alpha 1\beta 2\gamma 2S(S443delC)^{HA}$ ,  $\alpha 1\beta 2$  and  $\alpha 1\beta 2\gamma 2S^{HA}$  subunits. Cells were voltage clamped at  $-20$  mV and subjected to a 4 s pulse of either GABA alone (1 mM) (left traces), or co-applied with Zn<sup>2+</sup> (10  $\mu$ M) (right traces). Lengths of drug application (black line) are indicated above the current traces. (C) Summary graph displaying the Zn<sup>2+</sup> inhibition (%) from cells expressing  $\alpha 1$ ,  $\beta 2$ , and either  $\gamma 2S^{HA}$  or  $\gamma 2S(S443delC)^{HA}$  subunits was calculated. All data are presented as mean  $\pm$  S.E.M., and significance was determined using one-way ANOVA with Bonferroni's post-test. \*\*\* $p < 0.001$ , compared to wildtype.

Table 1

Clinical features in affected family members.

Patient ID/sex	Age	Age at seizure onset	Age at last seizure	Initial seizure(s)	Subsequent seizure(s)	Seizure frequency	Treatment	Other clinical features
II:2/M	49 y	1 y	3 y	FS	FS	3 episodes	No treatment	None
II:3/F	35 y	6 mo	16 y	FS	GTCS	6 episodes	PB, VPA; Drug free since age 8 y	None
III:1/M	10 y	9 y	9 y	GTCS	GTCS	1 episode	No treatment	None
III:4/M	5 y	9 mo	3 y	FS	FS	8 episodes	No treatment	None

M: male; F: female; y: years; mo: months; FS: febrile seizures; GTCS: Generalized Tonic-Clonic Seizures; PB: phenobarbital; VPA: valproate