J Cutan Pathol 2006: 33 (Suppl. 1): 6–9 Blackwell Munksgaard. Printed in Singapore Copyright © Blackwell Munksgaard 2006

Journal of Cutaneous Pathology

New aspects in the biology of cutaneous B-cell lymphomas

Immunologic and molecular genetic studies greatly contributed to a better understanding and interpretation of the distinct clinico-pathologic features of primary cutaneous B-cell lymphomas (CBCL), which are the basis for the consensus WHO-EORTC classification. There is increasingly accumulating evidence that these well defined clinicopathologic entities of CBCL have specific immunologic and molecular features, which further support their nosologic categorization as well as either interesting similarities with other extranodal B-cell lymphomas or definite peculiarities as compared to nodal B-cell lymphomas of similar histotype (specifically, follicle center lymphoma and diffuse large B-cell lymphoma).

Pimpinelli N. New aspects in the biology of cutaneous B-cell lymphomas. J Cutan Pathol 2006; 33 (Suppl. 1): 6–9. © Blackwell Munksgaard 2006.

Nicola Pimpinelli

Department of Dermatological Sciences, University of Florence Medical School, Florence, Italy

Nicola Pimpinelli, MD, Department of Dermatological Sciences, University of Florence Medical School, Via della Pergela, 58-64, 37, 50121 Florence, Italy Tel.: + 39 55 2344422 Fax: + 39 55 2758757 e-mail: pimpi@unifi.it

Accepted for publication November 3, 2005

The topic of primary cutaneous B-cell lymphomas (CBCL) in terms of diagnosis, classification, and historical perspective is extensively dealt with in another article of this JCP issue. As a general comment, the immunologic and molecular research in the field of CBCL considerably contributed—in addition to careful clinico-pathologic studies—to the categorization of these lymphomas first proposed in the EORTC classification¹ and recently recognized in the consensus WHO-EORTC classification.²

At present, there are many unanswered questions concerning the biology of CBCL and, more in general, of the homing of B cells into the skin and their predilection to stay localized and to proliferate in the skin in many instances. In fact, there still is no consistent information concerning the existence of a B-cell arm of the skin immune system. In the skin, there are no known equivalents to the Peyer's patches of the intestine or the Waldever's ring of the pharynx. In addition, different from T cells, B cells are not found in normal human skin. Currently, the only data concern the demonstration that sweat and sebaceous glands synthesize the secretory component of dimeric immunoglobulin (Ig)A in humans.³ Indeed, a possible speculation is that the skin and draining lymph nodes form an integrated system whereby a lymphoproliferative response to antigenic stimuli may be elicited in the skin.⁴ In addition, it has been hypothesized that the expression of skin-specific homing receptors by normal and/or neoplastic B lymphocytes in specific situations may explain the preferential circulation to and accumulation in the skin of lymphoid B cells. Another possibility is that a combination of etiological factors–infectious, immunologic, and genetic–may play a role in the development of lymphoproliferative B-cell disorders of the skin.

As a matter of fact, several antigenic stimuli are known to eventuate into a reactive B-cell response. These reactions, variously defined as cutaneous lymphoid hyperplasia, pseudo-B-cell lymphoma (P-CBCL), or lymphadenosis benigna cutis, may be caused by insect bites, in particular tick bites transmitting a Borrelia burgdorferi infection, acupuncture, antigen injections (drug eruptions, vaccinations, specific immune therapy), and tattoo pigments among others.⁵ Borrelia burgdorferi infection^{6–8} and tattoo pigments^{9,10} have also been suggested to be involved in the development of CBCL, in particular marginalzone B-cell lymphomas. Interestingly, B. burgdorferispecific DNA sequences have been demonstrated in a significant minority of CBCL from European areas with endemic Borrelia infections^{6,7} but could not be detected in CBCL from different areas in the United States¹¹ and Asia.¹² Different from *Borrelia* infection, no viral DNA sequences have been found in CBCL.¹³

Accumulating evidences suggest that these indolent CBCL (pcMZL, primary cutaneous marginal zone lymphoma; pcFCL, primary cutaneous follicle center lymphoma) and P-CBCL represent indeed a spectrum of cutaneous B-cell lymphoproliferative disorders with a step-wise progression from a reactive to a neoplastic state.^{10,14} This also explains why it can be so difficult to differentiate early stages of these indolent CBCL from P-CBCL. Apart from clinico-pathologic similarities and the above-mentioned shared association with Borrelia infection and tattoo pigments, clonal Ig gene rearrangements have been demonstrated not only in CBCL but also in a proportion of P-CBCL, as defined by immunohistochemical criteria.^{10,15} Interestingly, molecular analysis of CBCL recently documented intraclonal diversity with a high degree of somatic mutations, supporting the hypothesis of an antigen-driven process arising from transformed germinal-center B cells.¹⁶

Another intriguing approach to the biology of CBCL concerns the expression of chemokines (chemoattractanct cytokines), especially those regulating the localization of lymphoid cells during lymphopoiesis, antigen priming, and immune surveillance. referred to as "housekeeping" (rev. in 17). In fact, it has been demonstrated that both BCA-1 (also regarded as CXCL13, the most effective B-cell chemoattractant) and CXCR5 (a chemokine receptor with predominant expression on blood and tonsillar B cells, as well as on a subset of memory CD4⁺ Tcells) are highly expressed in Helicobacter pylori (HP)induced mucosa-associated lymphoid tissue (MALT) lymphoma with a distinctive pattern of expression,¹⁸ thus suggesting a possible pathogenic role of BCA-1 in the natural history of MALT lymphoma. We have recently investigated the possible involvement of BCA-1 and CXCR5 in the pathogenesis of CBCL and P-CBCL.¹⁹ First of all, no BCA-1 or CXCR5 expression was found at the immunohistochemical and molecular level in either clinically healthy skin or CTCL, thus confirming the B-cell-restricted specificity of these chemokines. The significant differences found between P-CBCL and indolent CBCL, marginal-zone lymphoma (MZL) type (CD35⁺/BCA-1⁺ dendritic cells found in lymphoid follicles of P-CBCL only; in CBCL virtually no CD35⁺ dendritic cells coexpressing BCA-1, with intensity of CXCR5 expression on CD22⁺ B cells lower than that of BCA-1),¹⁹ are in keeping with those evidenced in HP-induced MALT (gastritis) and gastric lymphoma, MALT type.¹⁸ In fact, the distribution and intensity of BCA-1 and CXCR5 in (secondary) lymphoid follicles of P-CBCL and indolent CBCL, MZL type parallel those found in HP-induced MALT (gastritis) and in gastric lymphoma, respectively. These findings add a further element to the close similarities between CBCL, MZL type, and MALT lymphoma, with

Biology of cutaneous B-cell lymphomas

chronic antigen stimulation as a possible common pathogenic denominator.^{18,19}

There is no doubt that immunologic and molecular genetic studies greatly contributed to a better understanding and interpretation of the distinct clinico-patholgic features of CBCL, which are the basis for the EORTC¹ and the consensus WHO-EORTC classifications.² Primary cutaneous MZL (pcMZL) neoplastic cells characteristically express bcl-2 protein and are negative for bcl-6, which is a very useful additional criterion to differentiate pcMZL from primary cutaneous follicle-center lymphoma (pcFCL).^{20,21} Recent studies show the presence of the t(14;18)(q32;q21) involving the IGH gene on chromosome 14 and the MLT gene on chromosome 18 in a proportion of pcMZL.^{22,23} Other translocations observed in gastric MALT lymphomas as well as other extranodal MZL, such as t(11;18)(q21;q21) and t(1;14)(p22;q32), have not been found in pcMZL.^{23,25} pcFCL consistently express bcl-6,^{20,21,26,27} whereas they-unlike nodal and secondary cutaneous follicular lymphomas-do not express bcl-2 protein or show faint bcl-2 staining in a minority of neoplastic B cells.^{27–30} Staining for MUM-1/IRF4 is negative.^{31,32} Somatic hypermutation of variable heavy and light chain genes has been demonstrated, which further supports the follicle-center-cell origin of these lymphomas.^{16,33} In most studies, pcFCL, including cases with a follicular growth pattern, do not show the t(14;18), which is characteristically found in systemic follicular lymphomas and a proportion of systemic diffuse large B-cell lymphomas.^{29,33,34} Inactivation of p15 and p16 tumor-suppressor genes by promotor hypermethylation has been reported in a relatively small proportion of pcFCL.³⁵ In a recent study using interphase fluorescence *in situ* hybridization, no evidence for translocations involving IgH, myc, or bcl-6 loci were found.²⁵ pcFCL have the gene expression profile of germinal center-like large B-cell lymphomas.^{31,32} In contrast to the group of pcFCL, primary cutaneous large B-cell lymphoma, leg type (pcLBCL-LT) show strong bcl-2 expression, also in the much less-frequent cases not located on the legs.^{21,29,36} Different from pcFCL, most pcLBCL-LT express MUM-1/IRF4 protein.31,32 The t(14;18) is not found in pcLBCL, although strong bcl-2 expression is common in this group.^{29,37} In some cases, bcl-2 overexpression may result from chromosomal amplification of the bel-2 gene.³⁷ Inactivation of p15 and p16 tumorsuppressor genes by promotor hypermethylation has been detected in a variable proportion of pcLBCL-LT.³⁵ Chromosomal imbalances have been identified in most pcLBCL-LT, with gains in 18q and 7p and loss of 6q as most common findings.^{37–39} Recent reports showed translocations

Pimpinelli

involving *myc*, *bcl-6*, and *IgH* genes in most pcLBCL-LT but not in patients with pcFCL with a diffuse infiltration of large centrocytes.²⁵ Finally, recent studies suggest that pcLBCL-LT have an activated B-cell gene expression profile, in parallel with MUM-1/IRF4 protein expression.^{31,32}

In conclusion, there is increasingly accumulating evidence that the well defined clinico-pathologic entities of CBCL have specific immunologic and molecular features, which further support their nosologic categorization as well as either interesting similarities with other extranodal B-cell lymphomas or definite peculiarities as compared to nodal B-cell lymphomas of similar histotype (specifically, follicle center lymphoma and diffuse large B-cell lymphoma).

References

- Willemze R, Kerl H, Sterry W, et al. EORTC classification for primary cutaneous lymphomas. Blood 1997; 90: 345.
- Willemze R, Jaffe E, Burg G, et al. WHO-EORTC classification for cutaneous lymphomas. Blood 2005; 105: 3768 (Epub 3 February 2005).
- Metze D, Jurecka W, Gebhart W, Schmidt J, Mainitz M, Niebauer G. Immunohistochemical demonstration of immunoglobulin A in human sebaceous and sweat glands. J Invest Dermatol 1988; 91: 13.
- Santucci M, Pimpinelli N, Arganini L. Primary cutaneous B-cell lymphoma. A unique type of low-grade lymphoma. Clinicopathologic and immunologic study of 83 cases. Cancer 1991; 67: 2311.
- van Vloten WA, Willemze R. The many faces of lymphocytoma cutis. J Eur Acad Dermatol Venereol 2003; 17: 3.
- Cerroni L, Zöchling N, Pütz B, Kerl H. Infection by Borrelia burgdorferi and cutaneous B-cell lymphoma. J Cutan Pathol 1997; 24: 457.
- Goodlad JR, Davidson MM, Hollowood K, et al. Primary cutaneous B-cell lymphoma and Borrelia burgdorferi infection in patients from the Highlands of Scotland. Am J Surg Pathol 2000; 24: 1279.
- Roggero E, Zucca E, Mainetti C, et al. Eradication of Borrelia burgdorferi infection in primary marginal zone B-cell lymphoma of the skin. Hum Pathol 2000; 31: 263.
- Sangueza OP, Yadav S, White CR, et al. Evolution of B-cell lymphoma from pseudolymphoma. Am J Dermatopathol 1992; 14: 408.
- Gilliam AC, Wood GS. Cutaneous lymphoid hyperplasias. Semin Cutan Med Surg 2000; 19: 133.
- Wood GS, Kamath NV, Guitart J, et al. Absence of Borrelia burgdorferi DNA in cutaneous B-cell lymphomas from the United States. J Cutan Pathol 2001; 28: 502.
- Li C, Inagaki H, Kuo T, et al. Primary cutaneous marginal zone B-cell lymphoma. A molecular and clinicopathologic study of 24 Asian cases. Am J Surg Pathol 2003; 27: 1061.
- Gellrich S, Schewe C, Sterry W, Lukowsky A. Absence of SV40 and other polyomavirus (JCV, BKV) DNA in primary cutaneous B Cell lymphomas. J Invest Dermatol 2005; 124: 278.

- 14. Goodlad JR, Hollowood K. Primary cutaneous B-cell lymphoma. Curr Diagn Pathol 2001; 7: 33.
- Nihal M, Mikkola D, Horvath N, et al. Cutaneous lymphoid hyperplasia. A lymphoproliferative continuum with lymphomatous potential. Hum Pathol 2003; 34: 617.
- 16. Gellrich S, Rutz S, Golembowski S, et al. Primary cutaneous follicle center cell lymphomas and large B cell lymphoma of the leg descend from germinal center B cells: a single cell polymerase chain reaction analysis. J Invest Dermatol 2001; 117: 1512.
- Loetscher P, Moser B, Baggiolini M. Chemokines and their receptors in lymphocyte traffic and HIV infection. Adv Immunol 2000; 74: 127.
- Mazzucchelli L, Blaser A, Kappler A, et al. BCA-1 is highly express in Helicobacter pylori-induced mucosa-associated lymphoid tissue and gastric lymphoma. J Clin Invest 1999; 104: R49.
- Mori M, Manuelli C, Pimpinelli N et al. BCA-1, a B-cell chemoattractant signal, is costantly expressed in cutaneous B-cell lymphoproliferative disorders. Eur J Cancer 2003; 39: 1625.
- 20. de Leval L, Harris NL, Longtine J, Ferry JA, Duncan LM. Cutaneous B-cell lymphomas of follicular and marginal zone types. Use of Bcl-6, CD10, Bcl-2, and CD21 in differential diagnosis and classification. Am J Surg Pathol 2001; 25: 732.
- Hoefnagel JJ, Vermeer MH, Janssen PM, Fleuren GJ, Meijer CJLM, Willemze R. Bcl-2, Bcl-6 and CD10 expression in cutaneous B-cell lymphoma: further support for a follicle centre cell origin and differential diagnostic significance. Br J Dermatol 2003; 149: 1183.
- Streubel B, Lamprecht A, Dierlamm J, et al. Ts(14;18) (q32;q21) involving IGH and MALT1 is a frequent chromosomal aberration in MALT lymphoma. Blood 2003; 101: 2335.
- Schreuder MI, Hoefnagel JJ, Jansen PM, van Keirken JH, Willemze R, Hebeda KM. FISH analysis of MALT lymphoma-specific translocations and aneuploidy in primary cutaneous marginal zone lymphoma. J Pathol 2005; 205: 302.
- 24. Gronbaek K, Ralfkiaer E, Kalla J, Skovgaard GL, Guldberg P. Infrequent somatic Fas mutations but no evidence of Bcl-10 mutations or t(11;18) in primary cutaneous MALT-type lymphoma. J Pathol 2003; 201: 134.
- Hallermann C, Kaune KM, Gesk S, et al. Molecular cytogenetic analysis of chromosomal breakpoints in the IGH, MYC, BCL6 and MALT1 gene loci in primary cutaneous B-cell lymphomas. J Invest Dermatol 2004; 123: 213.
- Goodlad JR, Krajewski AS, Batstone PJ et al. Primary cutaneous follicular lymphoma. A clinicopathologic and molecular study of 16 cases in support of a distinct entity. Am J Surg Pathol 2002; 26: 733.
- 27. Kodama K, Massone C, Chott A, Metze D, Kerl H, Cerroni L. Primary cutaneous large B-cell lymphomas: colinico-pathologic features, classification, and prognostic factors in a large series of patients. Blood 2005; 106: 2491.
- Cerroni L, Volkenandt M, Rieger E, Soyer HP, Kerl H. Bcl-2 protein expression and correlation with the interchromosomal (14;18) translocation in cutaneous lymphomas and pseudolymphomas. J Invest Dermatol 1994; 102: 231.
- Geelen FAMJ, Vermeer MH, Meijer CJLM, et al. Bcl-2 expression in primary cutaneous large B-cell lymphoma is site-related. J Clin Oncol 1998; 16: 2080.
- Cerroni L, Arzberger E, Pütz B, et al. Primary cutaneous follicular center cell lymphoma with follicular growth pattern. Blood 2000; 95: 3922.
- 31. Hoefnagel JJ, Dijkman R, Basso K, et al. Distinct types of primary cutaneous large B-cell lymphoma identified by gene

Biology of cutaneous B-cell lymphomas

expression profiling. Blood 2000; 105: 3671 (Epub 12 August 2004).

- 32. Sundram U, Kim Y, Mraz-Gernhard S, Hoppe R, Natkunam Y, Kohler S. Expression of the bcl-6 and MUM1-IRF4 proteins correlates with overall and diseasespecific survival in patients with primary cutaneous large Bcell lymphoma: a tissue microarray study. J Cutan Pathol 2005, 32: 227.
- Aarts WM, Willemze R, Bende RJ, Meijer CJLM, Pals ST, van Noessel CJ. VH gene analysis of primary cutaneous B-cell lymphomas: evidence for ongoing somatic hypermutation and isotype switching. Blood 1998; 92: 3857.
- Child FJ, Russell-Jones R, Woolford AJ, et al. Absence of the t(14,18) chromosomal translocation in primary cutaneous B-cell lymphoma. Br J Dermatol 2001; 144: 735.
- 35. Mao X, Lillington D, Child FJ, Russell-Jones R, Young B, Whittaker S. Comparative genomic hybridization analysis

of primary cutaneous B-cell lymphomas: identification of common genomic alterations in disease pathogenesis. Genes Chromosomes Cancer 2002; 35: 144.

- Goodlad JR, Krajewski AS, Batstone PJ, et al. Primary cutaneous diffuse large B-cell lymphoma. Prognostic significance and clinicopathologic subtypes. Am J Surg Pathol 2003; 27: 1538.
- Grange F, Petrella T, Beylot-Barry M, et al. Bcl-2 protein expression is the strongest independent prognostic factor of survival in primary cutaneous large B-cell lymphomas. Blood 2004; 103: 3662.
- Hallermann C, Kaune K, Siebert R, et al. Cytogenetic aberration patterns differ in subtypes of primary cutaneous B-cell lymphomas. J Invest Dermatol 2004; 122: 1495.
- 39. Wiesner T, Streubel B, Huber D, Kerl H, Chott A, Cerroni L. Genetic aberrations in primary cutaneous large B-cell lymphoma: a fluorescence in situ hybridization study of 25 cases. Am J Surg Pathol 2005; 29: 666.