



FLORE

Repository istituzionale dell'Università degli Studi di Firenze

Synthesis of a constrained bicyclic Gly-Asn dipeptide isostere

Questa è la Versione finale referata (Post print/Accepted manuscript) della seguente pubblicazione:

Original Citation:

Synthesis of a constrained bicyclic Gly-Asn dipeptide isostere / Trabocchi, Andrea; Menchi, Gloria; Danieli, E.; Guarna, Antonio. - In: AMINO ACIDS. - ISSN 0939-4451. - STAMPA. - 35(2008), pp. 37-44. [10.1007/s00726-007-0636-7]

Availability:

This version is available at: 2158/253005 since: 2017-11-08T19:37:24Z

Publisher: Springer-Verlag Wien:Sachsenplatz 4-6, PO Box 89, A-1201 Vienna Austria:011 43 1 3302415, EMAIL:

Published version: DOI: 10.1007/s00726-007-0636-7

Terms of use:

Open Access

La pubblicazione è resa disponibile sotto le norme e i termini della licenza di deposito, secondo quanto stabilito dalla Policy per l'accesso aperto dell'Università degli Studi di Firenze (https://www.sba.unifi.it/upload/policy-oa-2016-1.pdf)

Publisher copyright claim:

(Article begins on next page)

Amino Acids

Synthesis of a bicyclic δ -amino acid as a constrained Gly-Asn dipeptide isostere

A. Trabocchi, G. Menchi, E. Danieli, and A. Guarna

Dipartimento di Chimica Organica "Ugo Schiff" and Laboratorio di Progettazione, Sintesi e Studio di Eterocicli Biologicamente Attivi (HeteroBioLab), Università degli Studi di Firenze, Firenze, Italy

Received September 18, 2007 Accepted November 6, 2007 Published online January 31, 2008; © Springer-Verlag 2008

Summary. δ -Amino acids are very attractive in drug discovery, especially in the peptidomimetic area, because of their capability to act as dipeptide isosteres and reverse turn mimetics. Herein we report the synthesis of a rigid δ -amino acid constrained by a 3-aza-6,8-dioxabicyclo[3.2.1]octanebased scaffold, which can be considered as a Gly-Asn dipeptide mimetic. Key steps are the condensation of glycidol and tartaric acid derivatives, and the intramolecular *trans*-acetalization of the oxidized adduct to give the bicyclic δ -amino acid. Starting from L-tartaric acid derivative, it was achieved the corresponding Gly-D-Asn isostere, whereas from the enantiomeric D-tartaric acid derivative the corresponding Gly-L-Asn isostere could be obtained, thus giving access to both enantiomeric dipeptide sequences.

Keywords: Amino acids – Peptides – Peptidomimetics – Bicyclic compounds – Chiral pool

Introduction

A field of interest in biomedical research is oriented towards the development of therapeutics based on peptides and proteins, by introducing both structural and functional specific modifications, and maintaining the features responsible for biological activity. In this context, the area of peptidomimetics provides new dimensions to the field of molecular diversity and drug discovery (Giannis and Kolter, 1993; Gante, 1994; Adessi and Soto, 2002). In particular, the development of new leads for drug discovery has taken advantage of the use of unnatural amino acids in different fields of medicinal chemistry, for the development of peptidomimetics possessing high molecular diversity and the capability to generate high-ordered structures (Cole, 1994; Juaristi and Lopez-Ruiz, 1999; Trabocchi et al., 2005; Trabocchi et al., 2007). The possibility of having a large collection of δ -amino acids, both linear and cyclic, is of special interest in the generation of different molecular entities able to interact with biological targets. The most interesting feature displayed by δ -amino acids is their structural analogy with dipeptides, and many δ -amino acids as peptidomimetics have been constructed to mimic dipeptide sequences with added structural constraints (Trabocchi et al., 2005). Relevant examples of δ-amino acids include linear (Wipf et al., 1998; Gardner et al., 1999; Shankaramma et al., 1999; Casimir et al., 2000), cyclic (Poitout et al., 1995; Graf von Roedern et al., 1996; Suhara et al., 1996; Wattersom et al., 2003; Durrat et al., 2004), bicyclic (Hanessian et al., 1997; Van Well et al., 2003), and spiro (Genin and Johnson, 1992; Khalil et al., 1999; Alonso et al., 2001) compounds, and different templates have been applied for the generation of such amino acids. Moreover, since the β -turn is a common structural feature of proteins associated with the dipeptide unit, much research about δ -amino acids has been concentrated on the creation of reverse turn mimetics, where the central amide bond is replaced by a rigid moiety (Kim and Germanas, 1997a, b; Belvisi et al., 1999; Estiarte et al., 2000; Trabocchi et al., 2002). Also, the folding properties of δ -peptides have been investigated (Szabo et al., 1998), as they proved to generate stable secondary structures. Among this class, sugar amino acid-based scaffolds gathered interest in both oligomer synthesis (Smith et al., 1999; Locardi et al., 2001) and peptidomimetics, due to the attractive feature of presenting many functional groups anchored in a rigid scaffold, and in well-defined positions.

Our last achievements in the generation of constrained bicyclic amino acids using derivatives from the chiral

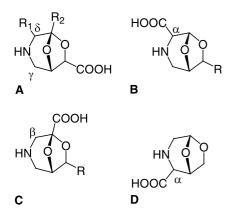


Fig. 1. 6,8-Dioxa-3-azabicyclo[3.2.1]octane-based amino acids: $\tilde{\gamma}/\delta$ (A), α (B and D), β (C)

pool were based on a general synthetic approach consisting in a coupling reaction between sugar and amino carbonyl derivatives, followed by intramolecular cyclization to give 6,8-dioxa-3-azabicyclo[3.2.1]octane-based scaffolds. This method allowed us to obtain a new class of bicyclic α - (Trabocchi et al., 2003a; Lalli et al., 2006), β -(Danieli et al., 2005), and γ/δ -amino acids (Guarna et al., 1999, 2002; Cini et al., 2002; Trabocchi et al., 2003b) by simply changing the starting material and tuning the reaction conditions according to the reactivity of the intermediate compounds (Fig. 1).

Recently, we reported the synthesis of bicyclic β -amino acids from glycidol and sugar derivatives (Fig. 1C) (Danieli et al., 2005). Starting from these molecules, by simply changing the sugar moiety with a tartaric acid derivative in the synthetic scheme, we designed a new type of δ -amino acid structure having the amino group at position 5 and the carboxyl function at position 7 (Fig. 2).

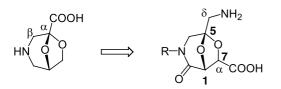


Fig. 2. Conceptual approach to a new δ -amino acid using the reported synthetic strategy for parent β -amino acids

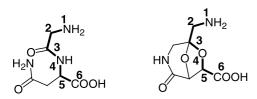


Fig. 3. Atom-by-atom correlation of the bicyclic δ -amino acid (*right*) with the Gly-Asn dipeptide sequence (*left*)

This new bicyclic amino acid shows a straightforward atom-by-atom correlation with respect to a general dipeptide sequence Gly-Asn, as shown in Fig. 3.

In particular, the carbonyl group at position 3 of the dipeptide Gly-Asn is mimicked by the acetal moiety in the bicyclic molecule, thus acting as a transition-state analogue, which is fundamental for peptidomimeticbased enzyme inhibitors (Giannis and Kolter, 1993; Gante, 1994). The replacement of the amide nitrogen atom at position 4 of the dipeptide by an oxygen atom, allows the maintenance of an heteroatom at such position. Moreover, the stereochemistry of Asn can be mimicked by proper choice of the tartaric acid stereoisomer, as the use of L-tartaric derivative 3 (see Fig. 4) forces the carbon atom at position 5 to have the same stereochemistry as for D-Asn, whereas the use of enantiomeric Dtartaric derivative provides the stereochemistry required for the L-Asn isostere.

Results and discussion

The synthesis of the new bicyclic δ -amino acid derivatives **1–3** consisted in amide bond formation between the tartaric acid derivative **4** with the amino alcohols **5** or **6** resulting from regioselective epoxide ring opening mediated by LiNTf₂, as reported (Cossy et al., 2002; Danieli et al., 2005). Subsequent alcohol oxidation and acid-catalyzed cyclization gave the corresponding bicyclic scaffold. The desired δ -amino ester **1**, deriving from amino alcohol **5**, was achieved after manipulation of the benzyloxy group of the parent bicyclic scaffold to give the amine, whereas more conveniently, δ -amino esters **2** and **3** where obtained directly with the Cbz-protected amino groups after acidic cyclization (see Fig. 4 for the retrosynthetic analysis).

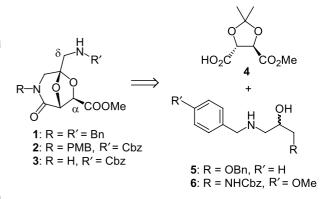


Fig. 4. Retrosynthetic strategy to δ -amino esters 1–3

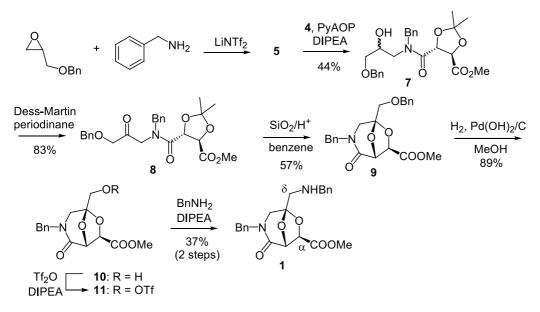




Table 1. COOH activators employed in the coupling reaction of acid 4 with amino alcohol ${\bf 5}$

Entry	Reagent system	Time (h)	Temperature (K)	Yield (%)
1	EDC, cat. DMAP	20	25	22
2	EDC-Cl, DIPEA	18	25	3
3	EDC-Cl, HOBt, NMM	24	25	18
4	1. DCC; 2. DIPEA	3 + 24	25	18
5	PyBrOP, DIPEA	16	25	26
6	PyAOP, DIPEA	12	25	44
7	HATU, TEA, DMF	16	25	15

Coupling reaction between amino alcohol **5** and tartaric acid derivative **4** to give **7** proved to be troublesome in terms of yield (Fig. 5).

Initially, EDC was added to acid **4** and the reaction was left in DCM for 20 h at room temperature and in the presence of catalytic DMAP, giving the desired amide in only 22% yield (Table 1, entry 1). Activation of COOH of **4** with EDC-Cl did not yield any improvement, nor the preactivation with DCC followed by addition of the amine and DIPEA as base (Table 1, entries 1–4). Use of other activating agents for the carboxylic function was thus investigated in order to obtain the adduct **7** in good yield. PyBrOP reagent, which is reported as a good coupling reagent for amide bond formation with relatively unreactive secondary amines Frérot et al., 1991), failed to give superior yields compared to carbodiimides (Table 1, entry 5). Finally, the use of PyAOP allowed to obtain compound **7** in higher yield (Table 1, entry 6), probably due to the effect of the nitrogen atom in the six-membered ring of the 1-hydroxy-7-aza-benzotriazolyl system in coordinating the amino group of **7** by virtue of hydrogen-bonding (Carpino, 1993). Interestingly, reaction conducted with the parent activator HATU and using DMF as solvent resulted in lower yield (Table 1, entry 7). Although the ester by-product was obtained in comparable quantity, the two products were separated easily by chromatography.

Subsequent secondary alcohol oxidation of the diastereomeric mixture of 7 using Dess-Martin periodinane gave the corresponding ketone 8 in 83% yield, and as a single stereoisomer, owing to the loss of the stereocenter at the carbon atom bearing the hydroxyl function. Acidcatalyzed cyclization was carried out in refluxing toluene, giving 9 in 40% yield. Better results were obtained when benzene was used as the solvent, in order to lower both the reaction temperature and the product degradation, giving 9 in 57% yield. Deprotection of the hydroxyl function of 9 was achieved by hydrogenolysis over $Pd(OH)_2/C$ in methanol, giving pure 10 after overnight stirring. Having in hands the bicyclic scaffold 10, the hydroxylic function at position 5 was converted to the corresponding triflate 11, and successively allowed to react with benzylamine, giving the final δ -amino acid derivative **1**, protected as Nbenzylamino ester, in 37% yield over two steps.

Following a modification of the synthetic scheme, we next devised a more convenient route to access the title Gly-Asn dipeptide isostere by introducing the amino group at the beginning of the synthesis rather than by hydroxylic group manipulation. Thus, oxiranyl-methana-

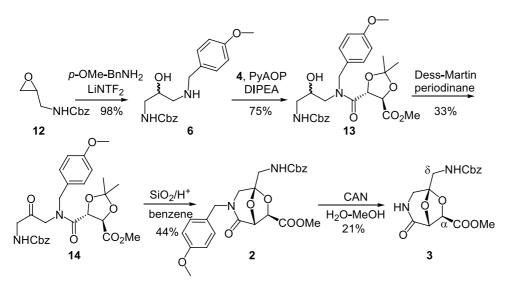


Fig. 6.

mine was taken into account as building block instead of O-benzyl-glycidol. Specifically, Fmoc-protected epoxide derivative was initially considered so as to achieve the corresponding Fmoc-derived dipeptide isostere for solidphase chemistry. According to reported procedures, allylamine was Fmoc-protected and successively treated with mCPBA in water – acetone to give the corresponding epoxide (Bischofberger et al., 1988; Volkmer-Engert et al., 1997). Unfortunately, when the resulting oxiranyl derivative was treated with a stoichiometric amount of p-methoxy-benzylamine in the presence of 0.5 equivalents of LiNTf₂, according to literature, substantial Fmoc-deprotection was observed, thus preventing the application of this protecting group in the new synthetic route. Thus, Cbz protecting group was selected, and after synthesizing the corresponding epoxide 12 in 72%, according to the above described method (Bischofberger et al., 1988; Volkmer-Engert et al., 1997), the amino alcohol 6 resulting from LiNTf₂-mediated epoxide ring-opening with *p*-methoxy-benzylamine was cleanly obtained in 98% overall yield (Fig. 6).

Successively, the construction of the bicyclic scaffold was performed in the usual way, by reacting the amino alcohol **6** with the protected monoester tartaric acid derivative **4** in the presence of PyAOP as carboxylic acid activator. Subsequent alcohol oxidation of **13** and acid-catalyzed cyclization of the resulting ketone **14** allowed to achieve the title bicyclic scaffold **2** in 11% overall yield. Finally, in order to achieve the bicyclic dipeptide isostere as Gly-D-Asn mimetic, the elimination of the *p*-methoxybenzyl derivative was required with the aim of hav-

ing the correct Asn side-chain mimetic, as shown in Fig. 3. Thus, the lactam moiety was treated with cerium ammonium nitrate (CAN), giving the molecule 3 with the deprotected amide function in 21% yield.

The same protocol was applied for the synthesis of the enantiomeric δ -amino acid as Gly-L-Asn isostere coming from the coupling between **6** and methyl di-*O*-isopropyl-idene-D-tartaric acid derivative *S*,*S*-**4**, giving the fully protected *S*,*S*-**2** in 25% overall yield. Particularly, the coupling and oxidation steps were performed in higher yields with respect to the other stereoisomer, indicating a more favourable stereochemical effect for the L-isostere. Also, coupling with PyAOP in both cases produced the title adduct in higher yields compared to **7**, corresponding to the other synthetic strategy.

Conclusion

In conclusion, the facile synthesis of a bicyclic δ -amino acid was achieved by coupling glycidol and tartaric acid derivatives, followed by alcohol oxidation and intramolecular acid *trans*-acetalization. Further derivatization of the hydroxymethyl function to give the aminomethyl group allowed to obtain the δ -amino acid structure. More interestingly, the application of oxiranyl-methanamine instead of glycidol allowed to simplify the overall synthesis, thus achieving the title Gly-Asn dipeptide isostere in few steps and avoiding the tedious manipulation of the glycidolderived hydroxylic function to achieve the corresponding amino group. This constrained Gly-Asn isostere may find application in the generation of peptidomimetics having this particular dipeptidic sequence, expecially where the conformational restriction is desired, and also, as δ -amino acids are often involved in β -turns, the application as a turn inducer is under investigation.

Experimental

Melting points are uncorrected. Chromatographic separations were performed on silica gel using flash-column techniques; R_f values refer to TLC carried out on 25-mm silica gel 60 F_{254} plates with the same eluant as indicated for column chromatography. ¹H and ¹³C NMR spectra of all compounds were recorded at 200 and 50.33 MHz, respectively, using CDCl₃ solutions. EI mass spectra were carried out at 70 eV ionizing voltage. Acid silica gel (H₂SO₄–SiO₂) was prepared as reported (Guidi et al., 1997).

(4R,5R)-5-[Benzyl-(3-benzyloxy-2(R/S)-hydroxy-propyl)-carbamoyl]-2,2-dimethyl-[1,3]dioxolane-4-carboxylic acid methyl ester (7)

Acid 4 (750 mg, 3.68 mmol) and PyAOP (1.19 g, 3.68 mmol, 1 eq) were dissolved in anhydrous CH2Cl2 (20 ml), and the resulting mixture was added to a solution of amine 5 (1.00 g, 3.68 mmol, 1 eq) in anhydrous CH₂Cl₂ (10 ml). After adding DIPEA (1.89 ml, 11.0 mmol, 3 eq), the mixture was left overnight stirring at room temperature, then it was successively washed with a 5% citric acid solution, a saturated NaHCO3 solution, and dried over Na₂SO₄. Purification by flash chromatography (EtOAc: petroleum ether = 1:2) yielded a yellow oil corresponding to 7 as a diastereometic mixture, each as a mixture of rotamers (740 mg, 1.62 mmol, 44%). ¹H-NMR (CDCl₃, 200 MHz) $\delta = 7.36-7.17$ (m, 10 H), 5.38–5.36 (m, 0.34 H), 5.33–5.31 (m, 0.64 H), 5.20 (d, J = 6.0 Hz, 0.17 H), 5.02 (d, J = 6.0 Hz, 0.17 H), 4.98–4.95 (m, 1.85 H), [4.77 (d, J = 16.0 Hz) and 4.73 (d, J = 16.1 Hz), 1 H], 4.51 (m, 2 H), 3.78, 3.77, and 3.75 (s, 3 H), 3.68– 3.56 (m, 1 H), 3.48–3.27 (m, 4 H), 1.46 and 1.45 (s, 6 H) ppm. ¹³C-NMR (CDCl₃, 50 MHz) Major diastereomer $\delta = 171.0$ (s), 170.5 (s), 137.9 (s), 136.1 (s), 128.8 (d, 2 C), 128.3 (d, 2 C), 127.7 (d, 4 C), 126.8 (d, 2 C), 113.1 (s), 76.2 (d), 73.5 (d), 72.0, 70.0, 69.0, 52.2, 50.5, 49.3, 26.4 (q, 2 C) ppm. MS m/z (%) 457 (M⁺, 2.1), 336 (16), 270 (22), 91 (100). Anal. Calcd for C25H31NO7: C, 65.63; H, 6.83; N, 3.06. Found: C, 65.71; H, 6.89; N, 2.89.

(4R,5R)-5-[Benzyl-(3-benzyloxy-2-oxo-propyl)-carbamoyl]-2,2dimethyl-[1,3]dioxolane-4-carboxylic acid methyl ester (8)

Compound 7 (116 mg, 0.253 mmol) was dissolved in CH₂Cl₂ (2.5 ml), then Dess-Martin periodinane (129 mg, 0.304 mmol) was added and the mixture was stirred at room temperature for 1 h. Successively, cold diethyl ether (3 ml) and a saturated NaHCO3 solution containing 25% Na2S2O3 (3 ml) were added, and the mixture was vigorously stirred for 10 min. Then it was diluted with additional diethyl ether, and the organic phase was separated, washed with a saturated NaHCO3 solution, brine and dried over anhydrous Na₂SO₄. Solvent evaporation afforded a crude product, that was purified by column chromatography (EtOAc:petroleum ether = 1:1) to give 8 (96 mg, 0.20 mmol, 83%) as a pale yellow oil. ¹H-NMR $(CDCl_3, 200 \text{ MHz})$ 2:3 mixture of rotamers A and B $\delta = 7.37 - 7.26 \text{ (m, 8 H)},$ 7.23–7.19 (m, 2 H), 5.38 (d, J = 5.1 Hz, 1 H, A), 5.34 (d, J = 5.5 Hz, 1 H, B), 5.03 (d, J = 5.5 Hz, 1 H, B), 4.87 (d, J = 15.0 Hz, 1 H, A), 4.78 (s, 2 H, A), 4.77 (s, 1 H, A), 4.53 (s, 2 H, B), 4.53–4.48 (d, J = 19.0 Hz, 1 H, B), 4.24–4.15 (m, 4 H, A + B), 4.07 (s, 2 H, B), 4.02 (s, 2 H, A), 3.80 (s, 3 H, A), 3.79 (s, 3 H, B), 1.48 (s, 3 H), 1.47 (s, 3 H), 1.41 (s, 3 H), 1.34 (s, 3 H) ppm. ¹³C-NMR (CDCl₃, 50 MHz) $\delta = 203.3$ (s, A), 202.5 (s, B), 170.6 (s), 168.6 (s), 136.7 (s, A), 136.5 (s, B), 136.0 (s, A), 135.6 (s, B), 128.8 (d, 2 C, A), 128.7 (d, 2 C, B), 128.5 (d, 2 C, A), 128.4 (d, 2 C, B), 128.2 (d, 2 C,

A), 128.0 (d, 2 C, B), 127.9 (d, 2 C, A), 127.9 (d, 2 C, B), 127.7 (d, 2 C, A), 127.3 (d, 2 C, B), 113.3 (s, A), 113.1 (s, B), 77.2 (d, A), 76.3 (d), 76.1 (d, B), 74.3 (t), 74.0 (t, A), 73.7 (t, B), 53.0 (t, A), 52.7 (q), 52.5 (t, B), 52.0 (t, A), 50.7 (t, B), 26.5 (q, 2 C) ppm. MS m/z (%) 455 (M⁺, 1.0), 334 (69), 306 (22), 91 (100). Anal. Calcd for C₂₅H₂₉NO₇: C, 65.92; H, 6.42; N, 3.08. Found: C, 66.01; H, 6.49; N, 3.00.

(1R,5R,7R)-3-Benzyl-5-benzyloxymethyl-2-oxo-6,8-dioxa-3-azabicyclo[3.2.1]octane-7-exo-carboxylic acid methyl ester (9)

Compound 8 (400 mg, 0.878 mmol) was dissolved in benzene (4 ml) to give a solution that was quickly added to a refluxing suspension of H₂SO₄/SiO₂ (141 mg) in benzene (7 ml). After 30 min, one-third of the solvent was distilled-off, the warm mixture was filtered through celite and the solvent was evaporated to give a crude oil that was purified by flash column chromatography (EtOAc:petroleum ether = 1:3), affording 9(200 mg, 0.50 mmol, 57%) as a viscous oil. $[\alpha]_D^{20} = -19.6 (c \ 1.0, \text{CHCl}_3).$ ¹H-NMR (CDCl₃, 400 MHz) $\delta = 7.36-7.26$ (m, 8 H), 7.22–7.20 (m, 2 H), 4.98 (s, 1 H), 4.78 (s, 1 H), 4.64 (d, J = 14.8 Hz, 1 H), 4.60 (d, J = 4.5 Hz, 2 H), 4.47 (d, J = 14.8 Hz, 1 H), 3.80 (d, J = 7.0 Hz, 2 H), 3.77 (s, 3 H), 3.57 (d, J = 12.3 Hz, 1 H), 3.19 (d, J = 12.3 Hz, 1 H) ppm. ¹³C-NMR (CDCl₃, 100 MHz) $\delta = 169.2$ (s), 165.4 (s), 137.7 (s), 135.4 (s), 128.8 (d, 2 C), 128.4 (d, 2 C), 127.9 (d. 3 C), 127.8 (d, 3 C), 107.4 (s), 78.8 (d), 78.0 (d), 73.8 (t), 70.8 (t), 52.7 (q), 51.7 (t), 48.7 (t) ppm. MS m/z (%) 397 (M⁺, 1.0), 306 (2), 291 (10), 91 (100). Anal. Calcd for C₂₂H₂₃NO₆: C, 66.49; H, 5.83; N, 3.52. Found: C, 66.55; H, 5.89; N, 3.49.

(1R,5R,7R)-3-Benzyl-5-hydroxymethyl-2-oxo-6,8-dioxa-3-azabicyclo[3.2.1]octane-7-carboxylic acid methyl ester (10)

To a solution of **9** (150 mg, 0.378 mmol) in MeOH (4 ml) was added 20% Pd(OH)₂/C (57 mg). The suspension was stirred under a hydrogen atmosphere at room temperature for 12 h, the it was filtered over celite and the solvent was evaporated to afford **10** (103 mg, 0.33 mmol, 89%) as a colourless oil and enough pure for the next step. ¹H-NMR (CDCl₃, 200 MHz) δ = 7.32–7.27 (m, 3 H), 7.21–7.17 (m, 2 H), 4.95 (s, 1 H), 4.78 (s, 1 H), 4.54 (s, 2 H), 3.81 (s, 2 H), 3.80 (s, 3 H), 3.31 (d, *J* = 11.7 Hz, 1 H), 3.10 (d, *J* = 11.7, 1 H) ppm. ¹³C-NMR (CDCl₃, 50 MHz) δ = 170.2 (s), 165.1 (s), 135.1 (s), 128.8 (d, 2 C), 127.9 (d, 3 C), 108.0 (s), 79.0 (d), 77.6 (d), 62.0 (t), 53.2 (q), 51.6 (t), 48.7 (t) ppm. MS *m*/*z* (%) 307 (M⁺, 8), 248 (2), 218 (3), 91 (100).

(1R,5R,7R)-3-Benzyl-5-benzylamino-methyl-2-oxo-6,8-dioxa-3-azabicyclo[3.2.1]octane-7-exo-carboxylic acid methyl ester (1)

Triflic anhydride (80 µl, 0.488 mmol) was dropwise added to a stirring solution of 10 (100 mg, 0.325 mmol) and triethylamine (99 µl, 0.715 mmol) in anhydrous CH₂Cl₂ (1.7 ml) at 0°C. The mixture was stirred at 0°C for 40 min, then it was washed with a saturated NaHCO₃ solution and brine. The organic phase was then dried over Na₂SO₄, and the solvent was evaporated to give 11 as a crude red oil that was used in the next step, without further purification. ¹H-NMR (CDCl₃, 200 MHz) $\delta =$ 7.35-7.31 (m, 3 H), 7.25-7.17 (m, 2 H), 5.04 (s, 1 H), 4.83 (s, 1 H), 4.67 (s, 2 H), 4.64 (d, J = 14.6 Hz, 1 H), 4.48 (d, J = 14.6 Hz, 1 H), 3.79 (s, 3 H), 3.46 (d, J = 11.7 Hz, 1 H), 3.20 (d, J = 11.7 Hz, 1 H) ppm. ¹³C-NMR $(CDCl_3, 100 \text{ MHz}) \delta = 168.2 \text{ (s)}, 164.4 \text{ (s)}, 150.1 \text{ (s)}, 134.8 \text{ (s)}, 128.9 \text{ (d, } 2 \text{ (s)})$ C), 128.1 (d), 127.9 (d, 2 C), 104.5 (s), 79.1 (d), 78.5 (d), 73.0 (t), 53.1 (d), 51.0 (t), 48.8 (t) ppm. To a solution of benzylamine (64 µl, 0.592 mmol) and diisopropylethylamine (51 μ l, 0.296 mmol) in anhydrous CH₂Cl₂ (0.5 ml), was dropwise added, at 0°C, a solution of 11 (130 mg, 0.296 mmol) in anhydrous CH₂Cl₂ (1 ml). The mixture was stirred under a nitrogen atmosphere, at room temperature for 12 h, then it was poured into a 5% aqueous solution of NaHCO3. The organic layer was separated, then it was washed with brine, dried over Na2SO4, and evaporated under vacuum to give a crude oil that was purified by flash column chromatography (EtOAc:petroleum ether = 2:1) to give **1** (40 mg, 0.10 mmol, 37% over two steps) as a pale yellow oil. $[\alpha]_D^{20} = -15.2$ (*c* 0.5, CHCl₃). ¹H-NMR (CDCl₃, 400 MHz) $\delta = 7.35-7.26$ (m, 8 H), 7.21–7.19 (m, 2 H), 4.97 (s, 1 H), 4.75 (s, 1 H), 4.59 (d, *J* = 14.6 Hz, 1 H), 4.49 (d, *J* = 14.6 Hz, 1 H), 3.91 (d, *J* = 13.5 Hz, 1 H), 3.83 (d, *J* = 13.5 Hz, 1 H), 3.77 (s, 3 H), 3.43 (d, *J* = 12.2 Hz, 1 H), 3.16 (d, *J* = 12.2 Hz, 1 H) 3.04 (s, 2 H), 2.17 (br, 1 H) ppm. ¹³C-NMR (CDCl₃, 100 MHz) $\delta = 169.5$ (s), 165.3 (s), 138.9 (s), 135.3 (s), 128.8 (d), 128.4 (d, 2 C), 128.1 (d, 2 C), 127.9 (d, 2 C), 127.8 (d), 127.2 (d), 108. 3 (s), 78.9 (d), 77.9 (d), 53.8 (t), 52.9 (q), 52.8 (t), 51.0 (t), 48.7 (t) ppm. MS m/z (%) 396 (M⁺, 1.0), 366 (1), 336 (2), 306 (1), 91 (100). Anal. Calcd for C₂₂H₂₄N₂O₅: C, 66.65; H, 6.10; N, 7.07. Found: C, 66.71; H, 5.99; N, 7.09.

[2-Hydroxy-3-(4-methoxy-benzylamino)-propyl]-carbamic acid benzyl ester (6)

p-Methoxy-benzylamine (1.01 ml, 7.71 mmol, 1.1 eq) and LiNTF₂ (1.07 g, 3.51 mmol, 0.5 eq) were added to a solution of epoxide 12 (1.452 g, 7.01 mmol) in anhydrous CH₂Cl₂ (3.5 ml) under a nitrogen atmosphere. The mixture was left stirring for 22 h at room temperature, then, a saturated NaHCO3 solution was added and the organic phase was separated and dried over Na₂SO₄, giving 3.30 g of crude product, which was further purified by flash chromatography, (EtOAc + 0.5% Et₃N, then MeOH), thus yielding pure 6 (2.37 g, 6.89 mmol, 98%) as a pale yellow oil. ¹H-NMR (CDCl₃, 200 MHz) $\delta = 7.30-7.20$ (m, 5 H), 7.17-7.13 (d, J = 8.5 Hz, 2 H), 6.81–6.77 (d, J = 8.5 Hz, 2 H), 5.92 (m, 1 H), 5.00 (s, 2 H), 3.77 (s, 2 H), 3.74 (m, 1 H), 3.71 (s, 3 H), 3.35-3.05 (m, 2 H), 2.70-2.45 (m, 2 H) ppm. ¹³C-NMR (CDCl₃, 50 MHz) $\delta = 179.1$ (s), 158.7 (s), 157.1 (s), 136.0 (s), 129.5 (d, 2 C), 129.3 (d, 2 C), 128.2 (d, 2 C), 127.8 (d), 127.6 (d, 2 C), 68.3 (d), 66.7 (t), 55.1 (q), 52.3 (t), 51.3 (t), 46.5 (t) ppm. Anal. Calcd for C₁₉H₂₄N₂O₄: C, 66.26; H, 7.02; N, 8.13. Found: C, 66.32; H, 7.11; N, 7.99.

(4R,5R)-5-[(3-Benzyloxycarbonylamino-2(R/S)-hydroxy-propyl)-(4methoxy-benzyl)-carbamoyl]-2,2-dimethyl-[1,3]dioxolane-4-carboxylic acid methyl ester (13)

Compound **13** was prepared as reported for **7**, starting from acid **4** (392 mg, 1.92 mmol) and amine **6** (661 mg, 1.92 mmol, 1 eq). Purification by flash chromatography (EtOAc, R_f 0.80) yielded a yellow oil corresponding to **13** as a diastereometic mixture, each as a mixture of rotamers (763 mg, 1.44 mmol, 75%). ¹H-NMR (CDCl₃, 200 MHz) δ = 7.26 (m, 5 H), 7.06 (d, J = 8.8 Hz, 2 H), 6.81 (d, J = 8.8 Hz, 2 H), 5.85 and 5.80 (br, 1 H), 5.23–5.05 (m, 1 H), 5.00 (s, 2 H), 4.84 (d, J = 5.9 Hz, 1 H), 4.69 (m, 2 H), 3.95 (m, 1 H), 3.75 and 3.72 (s, 3 H), 3.72 and 3.70 (s, 3 H), 3.68–2.95 (m, 4 H), 1.22–1.14 (s, 6 H) ppm. ¹³C-NMR (CDCl₃, 50 MHz) δ = 171.5, 169.0, 159.0, 143.4, 138.7, 136.6, 129.1, 128.3, 127.9, 127.7, 76.7, 76.0, 75.7, 69.1, 68.8, 67.8, 66.7, 55.1, 52.5, 51.4, 49.1, 44.2, 26.1, 25.9 ppm. MS m/z (%) 530 (M⁺, 0.2), 343 (22), 121 (100), 91 (41.7). Anal. Calcd for C₂₇H₃₄N₂O₉: C, 61.12; H, 6.46; N, 5.28. Found: C, 61.17; H, 6.51; N, 5.19.

(4R,5R)-5-[(3-Benzyloxycarbonylamino-2-oxo-propyl)-(4-methoxybenzyl)-carbamoyl]-2,2-dimethyl-[1,3]dioxolane-4-carboxylic acid methyl ester (14)

Compound 14 was prepared as reported for 8, starting from a solution of 13 (763 mg, 1.44 mmol) in CH₂Cl₂ (14.5 ml). Flash chromatography purification (EtOAc – petroleum ether 1:1, R_f 0.28) afforded pure 14 (254 mg, 0.48 mmol, 33%) as a pale yellow oil. [α]_D²⁴ = -5.2 (*c* 0.9, CDCl₃) ¹H-NMR (CDCl₃, 200 MHz) δ = 7.26–7.25 (m, 5 H), 7.13–7.06 (m, 2 H), 6.84–6.75 (m, 2 H), 5.56 (m, 1 H), 5.32 (m, 1 H), 5.29 (d, *J* = 12.8 Hz, 1 H), 5.05 (s, 2 H), 5.04 (d, *J* = 12.8 Hz, 1 H), 4.77–4.64 (m, 2 H), 4.39–4.17 (m, 1 H), 3.99–3.90 (m, 2 H), 3.72 (s, 3 H), 3.70 (s, 3 H), 1.42 (s, 6 H) ppm. ¹³C-NMR (CDCl₃, 50 MHz) δ = 200.0 (s), 170.2 (s), 168.0 (s),

158.9 (s), 155.8 (s), 135.8 (s), 129.5 (d, 2 C), 128.5 (d, 2 C), 127.7 (d, 2 C), 127.6 (d), 126.8 (d, 2 C), 76.0 (d), 75.7 (d), 66.6 (t), 60.3 (s), 55.0 (q), 52.5 (q), 52.3 (t), 51.2 (t), 50.0 (t), 26.3 (q), 26.1 (q) ppm. MS m/z (%) 528 (M⁺, 0.2), 364 (3.7), 322 (11.7), 136 (3.9). Anal. Calcd for C₂₇H₃₂N₂O₉: C, 61.35; H, 6.10; N, 5.30. Found: C, 61.42; H, 6.13; N, 5.09.

(1R,5R,7R)-5-(Benzyloxycarbonylamino-methyl)-3-(4-methoxy-benzyl)-2-oxo-6,8-dioxa-3-aza-bicyclo[3.2.1]octane-7carboxylic acid methyl ester (2)

Compound **2** was prepared as reported for **9**, starting from a solution of **14** (209 mg, 0.40 mmol) in benzene (3 ml). Flash chromatography purification (EtOAc – petroleum ether 2:1, R_f 0.53) yielded pure **2** (80 mg, 0.18 mmol, 44%) as a pale yellow oil. $[\alpha]_D^{27} = -17.4$ (*c* 4.0, CDCl₃). ¹H-NMR (CDCl₃, 200 MHz) $\delta = 7.33-7.31$ (m, 5 H), 7.14–7.10 (m, 2 H), 6.90–6.83 (m, 2 H), 6.05 (m, 1 H), 5.11 (s, 2 H), 4.94 (s, 1 H), 4.74 (s, 1 H), 4.48–4.44 (m, 2 H), 4.04 (m, 1 H), 3.78 (s, 6 H), 3.73 (s, 2 H), 3.47 (dd, J = 14.3, 3.3 Hz, 1 H), 3.30 (d, J = 11.8 Hz, 1 H), 3.10 (d, J = 12.0 Hz, 1 H) ppm. ¹³C-NMR (CDCl₃, 50 MHz) $\delta = 169.4$ (s), 168.3 (s), 164.6 (s), 159.0 (s), 156.6 (s), 145.8 (s), 129.8 (d, 2 C), 129.2 (d, 2 C), 128.3 (d, 2 C), 127.8 (d), 126.9 (d, 2 C), 106.8 (s), 78.8 (d), 77.7 (d), 76.0 (t), 66.9 (t), 66.6 (t), 55.2 (q), 52.8 (q), 47.9 (t) ppm. MS m/z (%) 456 (M⁺, 0.2), 335 (3.7), 322 (4.1), 136 (2.3). Anal. Calcd for C₂₄H₂₆N2O₈: C, 61.27; H, 5.57; N, 5.95. Found: C, 61.32; H, 5.61; N, 5.89.

(1R,5R,7R)-5-(Benzyloxycarbonylamino-methyl)- 2-oxo-6,8-dioxa-3aza-bicyclo[3.2.1]octane-7-carboxylic acid methyl ester (3)

Compound 2 (107 mg, 0.23 mmol) was dissolved in acetonitrile (1.2 ml), followed by addition of a solution of cerium ammonium nitrate (CAN) (252 mg, 0.46 mmol, 2 eq) in water (1.2 ml). After stirring at room temperature for 3 h, the mixture was diluted with EtOAc and the organic phase was separated. The aqueous phase was successively treated with EtOAc, and the organic extracts were combined and dried over Na2SO4, giving 126 mg of crude 3 after solvent evaporation. Flash chromatography purification (EtOAc:petroleum ether = 4:1, then CH₂Cl₂ - MeOH 10:1) afforded pure **3** (17 mg, 0.048 mmol, 21%) as a yellow oil. $[\alpha]_D^{22} = -16.3$ (c 3.4, CDCl₃). ¹H-NMR (CDCl₃, 200 MHz) $\delta = 7.31$ (m, 5 H), 6.11, 6.00 (br, 1 H), 5.10 (s, 2 H), 4.78 (s, 1 H), 4.73 (s, 1 H), 3.76 (m, 1 H), 3.72 (s, 3 H), 3.53 (dd, J = 14.4, 3.6 Hz, 1 H), 3.46 (d, J = 12.9 Hz, 1 H), 3.29 (d, J = 12.8 Hz, 1 H) ppm. ¹³C-NMR (CDCl₃, 50 MHz) $\delta = 169.4$ (s), 166.5 (s), 156.6 (s), 136.2 (s), 128.1 (d, 2 C), 127.7 (d), 127.6 (d, 2 C), 106.2 (s), 78.5 (d), 77.6 (d), 66.6 (t), 52.7 (q), 48.1 (t), 43.0 (t) ppm. MS m/z(%) 350 (M⁺, 2.7), 243 (17), 201 (44), 183 (27), 91 (100). Anal. Calcd for C₁₆H₁₈N₂O₇: C, 54.86; H, 5.18; N, 8.00. Found: C, 54.95; H, 5.23; N, 7.92.

(4S,5S)-5-[(3-Benzyloxycarbonylamino-2(R/S)-hydroxy-propyl)-(4methoxy-benzyl)-carbamoyl]-2,2-dimethyl-[1,3]dioxolane-4carboxylic acid methyl ester (**S,S-13**)

Prepared from *S*,*S*-4 (392 mg, 1.92 mmol) and **6** (661 mg, 1.92 mmol) as reported for **7**, giving 838 mg (1.58 mmol) of a yellow oil corresponding to (*S*,*S*-13) as a diastereomeric mixture in 82% yield, with identical NMR data as for 13. Anal. Calcd for $C_{27}H_{34}N_2O_9$: C, 61.12; H, 6.46; N, 5.28. Found: C, 61.16; H, 6.53; N, 5.23.

(4S,5S)-5-[(3-Benzyloxycarbonylamino-2-oxo-propyl)-(4-methoxybenzyl)-carbamoyl]-2,2-dimethyl-[1,3]dioxolane-4-carboxylic acid methyl ester (**S,S-14**)

Prepared from (*S*,*S*-13) (838 mg, 1.58 mmol) as reported for 8, giving 427 mg (0.81 mmol) of pure (*S*,*S*-14) as a pale yellow oil in 51% yield, with identical NMR data as for 14. $[\alpha]_D^{25} = +4.6$ (*c* 1.3, CDCl₃). Anal.

Calcd for $C_{27}H_{32}N_2O_9{:}$ C, 61.35; H, 6.10; N, 5.30. Found: C, 61.39; H, 6.15; N, 5.13.

(15,55,75)-5-(Benzyloxycarbonylamino-methyl)-3-(4-methoxybenzyl)-2-oxo-6,8-dioxa-3-aza-bicyclo[3.2.1]octane-7-carboxylic acid methyl ester (**S,S-2**)

Prepared from (*S*,*S*-14) (394 mg, 0.75 mmol) as reported for 9, giving 211 mg (0.46 mmol) pure (*S*,*S*-2) as a yellow oil in 62% yield, with identical NMR data as for 2. $[\alpha]_D^{25} = +19.1$ (*c* 1.1, CDCl₃). Anal. Calcd for C₂₄H₂₆N2O₈: C, 61.27; H, 5.57; N, 5.95. Found: C, 61.34; H, 5.63; N, 5.91.

Acknowledgments

The authors thank Università degli Studi di Firenze and MUR for financial support. Dr. Laura Bartali is gratefully acknowledged for carrying out some experiments. Brunella Innocenti is acknowledged for technical support.

References

- Adessi C, Soto C (2002) Converting a peptide into a drug: strategies to improve stability and bioavailability. Curr Med Chem 9: 963–978
- Alonso E, López-Ortiz F, del Pozo C, Peralta E, Macias A, Gonzaléz J (2001) Spiro β-lactams as β-turn mimetics. Design, synthesis, and NMR conformational analysis. J Org Chem 66: 6333–6338
- Belvisi L, Gennari C, Mielgo A, Potenza D, Scolastico C (1999) Conformational preferences of peptides containing reverse-turn mimetic bicyclic lactams: inverse γ-turns versus type-II' β-turns – insights into β-hairpin stability. Eur J Org Chem: 389–400
- Bischofberger N, Waldmann H, Saito T, Simon ES, Lees W, Bednarski MD, Whitesides GM (1988) Synthesis of analogs of 1,3-dihydroxyacetone phosphate and glyceraldehyde 3-phosphate for use in studies of fructose-1,6-diphosphate aldolase. J Org Chem 53: 3457–3465
- Carpino LA (1993) 1-Hydroxy-7-azabenzotriazole. An efficient peptide coupling additive. J Am Chem Soc 115: 4397–4398
- Casimir JR, Didierjean C, Aubry A, Rodriguez M, Briand J-P, Guichard G (2000) Stereoselective alkylation of *N*-Boc-protected-5-substituted δ -lactams: synthesis of α , δ -disubstituted δ -amino acids. Org Lett 2: 895–897
- Cini N, Machetti F, Menchi G, Occhiato EG, Guarna A (2002) Bicyclic compounds derived from tartaric acid and α-amino acids (BTAas): synthesis of new molecular scaffolds derived from the combination of (*R*,*R*)-tartaric acid and L-serine. Eur J Org Chem: 873–880
- Cole DC (1994) Recent stereoselective synthetic approaches to β -amino acids. Tetrahedron 50: 9517–9582
- Cossy J, Bellosta V, Hamoir C, Desmurs J-R (2002) Regioselective ring opening of epoxides by nucleophiles mediated by lithium bistrifluoromethanesulfonimide. Tetrahedron Lett 43: 7083–7086
- Danieli E, Trabocchi A, Menchi G, Guarna A (2005) Synthesis of glycidol- and sugar-derived bicyclic β and γ/δ -amino acids for peptidomimetic design. Eur J Org Chem: 4372–4381
- Durrat F, Xie J, Valéry J-M (2004) Synthesis of pyranoid δ -sugar amino acids and their oligomers from per-benzylated β -C-vinyl glucoside. Tetrahedron Lett 45: 1477–1479
- Estiarte MA, Rubiralta M, Diez A (2000) Oxazolopiperidin-2-ones as type II' β -turn mimetics: synthesis and conformational analysis. J Org Chem 65: 6992–6999
- Frérot E, Coste J, Pantaloni A, Dufour M-N, Jouin P (1991) PyBOP[®] and PyBroP: two reagents for the difficult coupling of the α , α -dialkyl amino acid, Aib. Tetrahedron 47: 259–270

- Gante J (1994) Peptidomimetics tailored enzyme inhibitors. Angew Chem Int Ed Engl 33: 1699–1720
- Gardner RR, Liang G-B, Gellman SH (1999) β -turn and β -hairpin mimicry with tetrasubstituted alkenes. J Am Chem Soc 121: 1806–1816
- Genin MJ, Johnson RL (1992) Design, synthesis, and conformational analysis of a novel spiro-bicyclic system as a type II β -turn peptidomimetic. J Am Chem Soc 114: 8778–8783
- Giannis A, Kolter T (1993) Peptidomimetics for receptor ligands discovery, development, and medical perspectives. Angew Chem Int Ed Engl 32: 1244–1267
- Graf von Roedern E, Lohof E, Hessler G, Hoffmann M, Kessler H (1996) Synthesis and conformational analysis of linear and cyclic peptides containing sugar amino acids. J Am Chem Soc 108: 10156–10167
- Guarna A, Guidi A, Machetti F, Menchi G, Occhiato EG, Scarpi D, Sisi S, Trabocchi A (1999) Synthesis and reactivity of bicycles derived from tartaric acid and α -amino acids: a novel class of conformationally constrained dipeptide isosteres based upon enantiopure 3-aza-6,8-dioxabicyclo[3.2.1]octane-7-carboxylic acid. J Org Chem 64: 7347–7364
- Guarna A, Bucelli I, Machetti F, Menchi G, Occhiato EG, Scarpi D, Trabocchi A (2002) Synthesis of a new enantiopure bicyclic γ/δ -amino acid (BTKa) derived from tartaric acid and α -amino acetophenone. Tetrahedron 58: 9865–9870
- Guidi A, Guarna A, Giolitti A, Macherelli M, Menchi G (1997) Condensation product between (R,R)-tartaric acid and a L-phenylalanine derivative as a new molecular scaffold. Arch Pharm Med Chem 330: 201–202
- Hanessian S, McNaughton-Smith G, Lombart H-G, Lubell WD (1997) Design and synthesis of conformationally constrained amino acids as versatile scaffolds and peptide mimetics. Tetrahedron 53: 12789–12854
- Juaristi E, Lopez-Ruiz H (1999) Recent advances in the enantioselective synthesis of β-amino acids. Curr Med Chem 6: 983–1004
- Khalil EM, Ojala WH, Pradhan A, Nair VD, Gleason WD, Mishra RK, Johnson RL (1999) Design, synthesis, and dopamine receptor modulating activity of spiro bicyclic peptidomimetics of L-prolyl-L-leucylglycinamide. J Med Chem 42: 628–637
- Kim K, Germanas JP (1997a) Peptides constrained to type VI β-turns. 1. Evidence for an exceptionally stable intramolecular hydrogen bond. J Org Chem 62: 2847–2852
- Kim K, Germanas JP (1997b) Peptides constrained to type VI β-turns. 2. Antiparallel β-ladder formation. J Org Chem 62: 2853–2860
- Lalli C, Trabocchi A, Guarna F, Mannino C, Guarna A (2006) Synthesis of a bicyclic proline analogue from L-ascorbic acid. Synthesis: 3122–3126
- Locardi E, Stöckle M, Gruner S, Kessler H (2001) Cyclic homooligomers from sugar amino acids: synthesis, conformational analysis, and significance. J Am Chem Soc 123: 8189–8196
- Poitout L, Le Merrer Y, Depezay J-C (1995) Synthesis of sugar-like amino-carboxylic acids from D-mannitol. Tetrahedron Lett 36: 6887–6890
- Shankaramma SC, Kumar Singh S, Sathyamurthy A, Balaram P (1999) Insertion of methylene units into the turn segment of designed β -hairpin peptides. J Am Chem Soc 121: 5360–5363
- Smith MD, Long DD, Martin A, Marquess DG, Claridge TDW, Fleet GWJ (1999) Absence of secondary structure in a carbopeptoid tetramer of a *trans*-5-aminomethyl-tetrahydrofuran-2-carboxylate. Tetrahedron Lett 40: 2191–2194
- Suhara Y, Ichikawa M, Hildreth JEK, Ichikawa Y (1996) Synthesis of sulfated β -1,6-linked oligosaccharide mimetics: a novel potent inhibitor of HIV replication. Tetrahedron Lett 37: 2549–2552
- Szabo L, Smith BL, McReynolds KD, Parrill AL, Morris ER, Gervay J (1998) Solid phase synthesis and secondary structural studies of $(1 \rightarrow 5)$ amide-linked sialooligomers. J Org Chem 63: 1074–1078

- Trabocchi A, Occhiato EG, Potenza D, Guarna A (2002) Synthesis and conformational analysis of small peptides containing 6-*Endo*-BT(t)L scaffolds as reverse turn mimetics. J Org Chem 67: 7483–7492
- Trabocchi A, Cini N, Menchi G, Guarna A (2003a) A new bicyclic proline-mimetic amino acid. Tetrahedron Lett 44: 3489–3492
- Trabocchi A, Menchi G, Rolla M, Machetti F, Bucelli I, Guarna A (2003b) Enantiospecific synthesis of 3-aza-6,8-dioxa-bicyclo[3.2.1]octane carboxylic acids from erythrose. Tetrahedron 59: 5251–5258
- Trabocchi A, Guarna F, Guarna A (2005) γ and δ -amino acids: synthetic strategies and relevant applications. Curr Org Chem 9: 1127–1153
- Trabocchi A, Menchi G, Guarna F, Machetti F, Scarpi D, Guarna A (2006) Design, synthesis, and applications of 3-aza-6,8-dioxa-bicyclo-[3.2.1]octane-based scaffolds for peptidomimetic chemistry. Synlett: 331–353
- Trabocchi A, Scarpi D, Guarna A (2007) Structural diversity of bicyclic amino acids. Amino Acids (in press)
- Van Well RM, Meijer MEA, Overkleeft HS, van Boom JH, van der Marel GA, Overhand M (2003) Synthesis of bridged sugar amino acids: a new entry into conformationally locked δ and ϵ -amino acids. Tetrahedron 59: 2423–2434

- Volkmer-Engert R, Hoffmann B, Schneider-Mergener J (1997) Stable attachment of the HMB-linker to continuous cellulose membranes for parallel solid phase spot synthesis. Tetrahedron Lett 38: 1029–1032
- Watterson MP, Edwards AA, Leach JA, Smith MD, Ichihara O, Fleet GWJ (2003) Synthesis of all diastereomeric methyl 2,5-anhydro-3-deoxyhexonates: precursors to C-2-deoxynucleosides and THF-templated γ and δ -amino acids. Tetrahedron Lett 44: 5853–5857
- Wipf P, Henninger TC, Geib SJ (1998) Methyl- and (trifluoromethyl)alkene peptide isosteres: synthesis and evaluation of their potential as β -turn promoters and peptide mimetics. J Org Chem 63: 6088–6089

Authors' address: Prof. Antonio Guarna, Dipartimento di Chimica Organica "Ugo Schiff" and Laboratorio di Progettazione, Sintesi e Studio di Eterocicli Biologicamente Attivi (HeteroBioLab), Università degli Studi di Firenze, Polo Scientifico e Tecnologico, Via della Lastruccia 13, I-50019 Sesto Fiorentino, Firenze, Italy, Fax: +39 055 4573569, E-mail: antonio.guarna@unif.it