

*Report***Prognostic significance of biologic markers in node-negative breast cancer patients: a prospective study**

Annalisa Volpi¹, Franca De Paola², Oriana Nanni², Anna Maria Granato², Paola Bajorko², Aldo Becciolini³, Emanuela Scarpi², Angela Riccobon¹, Manuela Balzi³, and Dino Amadori¹
¹Department of Medical Oncology, Pierantoni Hospital, Forlì; ²Istituto Oncologico Romagnolo, Forlì;
³Department of Radiobiology, University of Florence, Florence, Italy

Key words: biologic markers, node-negative breast cancer, prognosis

Summary

It is generally thought that future advances in the treatment and cure of breast cancer patients will be made possible through a deeper understanding of tumor biology and an improved capability to define the prognosis of each single patient. This will lead to the formulation of new, more selective, and patient-tailored therapies. It is therefore important, when studying potential prognostic factors, to follow methodologic requirements and guidelines which involve the carrying out of prospective studies as confirmatory steps. Repeatedly or recently investigated prognostic markers (tumor size, menopausal status, ER, PgR, ³H thymidine labeling index, c-erbB-2 and p27 expression) were evaluated on a series of 286 prospectively recruited node negative breast cancer patients who underwent loco-regional treatment alone and were closely followed. The individual and relative prognostic contribution of each variable with respect to other factors, as well as their ability to identify node negative patients at risk, were assessed by univariate and multivariate analysis. At a five-year follow-up, only tumor size ($p = 0.021$) and TLI ($p = 0.016$) individually proved to be significant prognostic indicators of relapse-free survival. Conversely, p27 expression was not related to RFS and c-erbB-2 expression appeared to have only a short-term effect on patient prognosis. TLI and tumor size, tested in multivariate analysis along with ER and menopausal status, maintained their independent prognostic relevance. The study, performed on a large series of node-negative patients given loco-regional treatment alone, for the first time prospectively recruited, showed the prognostic relevance of TLI and its independence from other clinico-pathologic and biologic factors over a five-year period.

Introduction

Public awareness and widespread information campaigns about cancer prevention programs and mammographic screenings have led, in the last decade, to an earlier detection of breast cancer. The proportion of women with newly-diagnosed small and node-negative disease is therefore increasing and represents a new reality to be approached.

Consolidated clinical experience has demonstrated that about a quarter of node-negative breast cancer patients are destined to relapse and die from the disease after loco-regional treatment alone. Recent studies have also shown that adjuvant therapy improves the clinical outcome of node-negative patients [1–4], but

the benefits and risks of therapy must be weighted in terms of both toxicity and cost. The clinical and biologic heterogeneity of breast cancer has led most clinicians to believe that the risk-to-benefit ratio does not favour treatment of all node-negative tumors. Consequently, an accurate prediction of the course of disease in individual patients is needed in order to identify those at risk who are candidates for adjuvant systemic treatment. It is also important for evaluating whether the patient could benefit from a particular type of therapy or should be spared the toxicity of ineffective treatment.

Over the past 20 years, new technologies have led to the proposal of an ever increasing number of potential prognostic factors for breast cancer patients.

As a result, clinicians are often faced with difficulties when trying to integrate the new and conventional prognostic factors in treatment decision-making. Furthermore, most studies have examined new biologic variables individually and almost always by retrospective analysis.

Current information in literature highlights the complexity of biologic processes underlying tumor transformation and progression such as gene alterations, cell differentiation and proliferation, apoptosis and invasiveness [5–18]. Moreover, much is now known of the multiplicity of cellular, biochemical and genetic markers representative of different biologic aspects. The large body of information derived from retrospective studies enabled us to select the markers with sufficient proof of prognostic power to be validated in a prospective study.

The aim of the present work was to assess the prognostic relevance of some biologic variables considered individually and their relative contribution with respect to pathologic factors in node negative breast cancer patients. Biologic variables repeatedly or recently investigated were determined in parallel on individual tumors in order to define their relative prognostic contribution and their ability to identify node-negative patients at risk.

Patients and methods

The clinic-biological study was conducted on a series of 286 women with node-negative breast cancer recruited by six clinical centers from 1989 to 1993. At least 10, and a median of 16, axillary lymph nodes were histologically examined. All patients had loco-regional treatment; more than 50% underwent quadrantectomy plus radiotherapy, and the remaining patients underwent mastectomy. Adjuvant systemic treatment was not given prior to relapse. The case series was consecutive on the basis of the availability of cell proliferation information. Patient distribution according to different age classes is reported in Table 1. Median age was 55 years and about 60% were postmenopausal patients, that is, more than 2 years had elapsed from spontaneous menopause or ovariectomy at the time of diagnosis.

More than two thirds of tumors were 2 cm or less in diameter and 18% were equal to or smaller than 1 cm. About 70% were ER-positive on the basis of the 10 fmol cut off, and about 55% were PgR-positive tumors on the basis of the 25 fmol cut off value.

Table 1. Clinico-pathologic and biologic characteristics of the case series

	Number of cases	Percentage
Age (years)		
≤45	74	25.9
46–50	44	15.4
51–60	85	29.7
>60	83	29.0
Menopausal status		
Premenopausal	113	39.5
Postmenopausal	173	60.5
Type of surgery		
Quadrantectomy + RT	161	56.9
Mastectomy	122	43.1
Missing	3	
Histotype		
Lobular	26	9.2
Ductal	225	80.1
Others	30	10.7
Missing	5	
Tumor size		
≤1.0 cm	49	18.2
1.1–2.0 cm	139	51.7
>2.0 cm	81	30.1
Missing	17	
Receptor status		
ER ≥ 10 fmol	198	72.0
<10 fmol	77	28.0
Missing	11	
PgR ≥ 25 fmol	132	54.5
<25 fmol	110	45.5
Missing	44	
TLI		
< 3.1%	143	50.0
≥ 3.1%	143	50.0
c-erbB-2		
Negative	131	57.2
Positive (any % of positive cells)	98	42.8
Missing	57	
p27		
Negative	71	28.0
Positive (any % of positive cells)	183	72.0
Missing	32	

Median TLI value was 3.1% which is in agreement with the value reported in larger series [19–23]. Only membrane staining was considered in scoring c-erbB-2 positive cells; cytoplasmic staining was ignored as aspecific immunoreaction. Clear p27 nuclear staining was defined as positive. Positivity (any percentage of positive cells) for c-erbB-2 and p27 was observed in about 40% and 70% of tumors, respectively. Estrogen v (ER) and progesterone (PgR) receptors were determined in the different centers, ³H-thymidine labeling index (TLI) in only two centers (Forlì and Florence), and c-erbB-2 and p27 expression was evaluated in the Forlì laboratory.

Postoperative follow-up was carried out in the outpatient clinic of all centers. All patients had a clinical and instrumental check-up at 3-month intervals for the first 2 years, every 6-months during the 3rd, 4th and 5th years, and subsequently once a year up to the 10th year. Median follow-up was 74 months (range 4–111 months). Seventeen patients were lost to follow-up (6%), of whom 11 after at least 40 months.

In vitro determinations

Immediately after surgery, part of the tumor material was incubated with ³H-thymidine and then processed for conventional histologic procedures for the determination of TLI, c-erbB-2 and p27 expression. The rest of the tumor material was frozen in liquid nitrogen and stored at –80°C for ER and PgR determination.

TLI

Fresh tumor samples were incubated in culture medium containing ³H-thymidine for 1 h at 37°C and fixed in formalin. The recent availability of a commercial kit (Euroframe, Asti, Italy) enabled all the participating centers to perform this first step of *in vitro* ³H-thymidine labeling in their own laboratory. Samples from all patients were then sent to the two referee centers (Forlì and Florence) for autoradiographic procedures and TLI determination. Histologic sections were dipped in a photographic emulsion (Ilford K5, Ilford Photographics, London, UK) and exposed in the dark for 3 days at 4°C. Autoradiograms were developed in Ilford Phenprint for 6 min at 19°C and fixed in Hypam compound for 10 min. Samples were stained with hematoxylin and eosin at 4°C. When the specimen was small enough to allow the radioactive precursor to penetrate completely, labeled cells were counted throughout the whole section; if not, counting was limited to the periphery of the section (up to

80 µm in depth). TLI, expressed as the ratio between thymidine-labeled cells and the total number of tumor cells, was determined independently by two observers, with 2,000–5,000 cells scored from different fragments of the same tumor. Quality control procedures were periodically repeated in the context of a National Quality Control Program promoted by the Italian Society of Basic and Applied Cell Kinetics (SICCAB) [24].

Steroid receptor content

ER and PgR were assayed by the dextran-coated charcoal method according to the European Organization for Research and Treatment on Cancer [25]. Quality control procedures for hormone receptor dosage were coordinated by the Italian ad hoc committee. Quantitative biochemical analysis was adopted to allow the use of different cut off values and to identify different steroid receptor content subgroups for future basic and clinical analyses.

p27 and c-erbB-2 expression

Tumor samples were fixed in 10% formalin. 3 µm sections from paraffin-embedded blocks were deparaffinized with xylene, rehydrated, and endogenous peroxidase activity blocked. p27 antigen retrieval was performed by means of microwaving at 750 W in 10 mM citrate buffer (pH 6.0) for 15 min followed by cooling at room temperature for 20 min. The sections were then treated for non-specific binding with 3% bovine serum albumin in PBS for 20 min. After this they were incubated for 1 h at room temperature with p27 monoclonal antibody clone 57 (Transduction Laboratories, Lexington, KY) diluted 1:300 in PBS, for 15 min in biotinylated anti-mouse secondary antibody, then rinsed and incubated with avidin–biotin conjugate (Dako, LSAB+kit). After washing in PBS, the peroxidase reaction was developed to a brown stain by 0.05% diaminobenzidine, which was enhanced with 0.07% imidazole and hydrogen peroxide. Cell nuclei were counterstained blue by Mayer's haemalum and the sections were mounted in Faramount (Dako).

For the determination of c-erbB-2 expression, the sections were incubated for 1 h at room temperature with the monoclonal antibody CB11, which recognizes the internal domain of the c-erbB-2 protein (HER-2/neu) (Biogenex, San Ramon, CA), diluted 1:50 in antibody diluent with Background Reducing Components (DAKO Corporation, Carpinteria, CA). At this concentration c-erbB-2 overexpressing tumor

Table 2. Relationship between biologic variables and clinico-pathologic factors

	TLI		c-erbB-2		p27	
	Median value % (range)	<i>p</i>	Median value of positive cells % (range)	<i>p</i>	Median value of positive cells % (range)	<i>p</i>
Age						
≤ 50 years	4.0 (0.01–16.8)	0.03	40 (5–90)	0.83	18.6 (1–93)	0.95
> 50 years	2.7 (0.01–17.0)		50 (5–100)		18.2 (1–100)	
Menopausal status						
Premenopausal	3.5 (0.01–15.0)	0.15	60 (5–90)	0.50	18.0 (1–93)	0.93
Postmenopausal	2.8 (0.01–17.0)		40 (5–100)		18.6 (1–100)	
Histotype						
Lobular	3.9 (0.2–15.0)	0.04	20 (5–50)	0.09	17.5 (1–97)	0.94
Ductal	2.8 (0.01–17.0)		50 (5–100)		18.6 (1–100)	
Others	4.4 (0.01–16.8)		62 (5–90)		12.6 (1–98)	
Tumor size						
≤ 1.0 cm	2.8 (0.2–11.0)	0.02	40 (5–90)	0.21	31.7 (1–100)	0.09
1.1–2.0 cm	3.2 (0.01–16.8)		35 (5–100)		13.9 (1–95)	
> 2.0 cm	4.2 (0.2–17.0)		70 (5–100)		19.4 (1–97)	
ER						
≥ 10 fmol	2.8 (0.01–15.0)	0.01	35 (5–100)	0.16	19.6 (1–100)	0.05
< 10 fmol	4.3 (0.1–17.0)		65 (5–100)		8.4 (1–94)	
PgR						
≥ 25 fmol	2.8 (0.01–15.0)	0.04	35 (5–90)	0.38	21.3 (1–98)	0.08
< 25 fmol	4.0 (0.01–17.0)		55 (5–100)		13.1 (1–100)	

cells showed a strong and focalized membrane staining.

At least 20 high-power fields were scored by two independent observers for c-erbB-2 (FDP and AMG) and p27 (FDP and PB). Immunoreactivity was expressed as the ratio between the percentage of stained cells and the total number of tumor cells, or the entire area of invasive neoplastic tissue for p27 and c-erbB-2, respectively.

Statistical methods

The relationship between TLI, c-erbB-2, p27 and clinico-pathologic or biologic factors was analysed using a non-parametric ranking statistic (Median test), and Spearman's correlation coefficient was used to investigate the relationship between the different biomarkers considered as continuous variables in individual tumors.

Relapse-free survival was calculated as the period from surgery until the date of the first documented

evidence of new disease manifestation in loco-regional or distant sites, or in the contralateral breast. Owing to the difficulty of distinguishing between a second breast carcinoma and contralateral recurrence, the latter lesion was considered as an event. In the case of a second primary cancer in a non-breast site, relapse-free follow-up data were censored at the time of the diagnosis of the second malignancy. All new disease manifestations were assessed by clinical, radiologic and, when feasible, histologic examination of the site of relapse. Univariate analysis was performed tracing Kaplan-Meier survival curves, and comparison of survival curves was based on the log-rank test [26].

The role of each of the putative prognostic variables (univariate analysis) and their joint effect (multivariate analysis) was evaluated using Cox proportional hazard models [27]. The analysis of the plot of $\ln(-\ln(S(t)))$, (where $S(t)$ is the Kaplan-Meier estimate of the relapse-free survival curves), against the logarithm of the time for each level of the factor studied, suggested that the assumption of proportional hazards was

Table 3. Correlation between biologic variables

	PgR		TLI		c-erbB-2		p27	
	r_s	p	r_s	p	r_s	p	r_s	p
ER	0.444	<0.001	-0.197	0.001	-0.180	0.008	0.128	0.046
PgR			-0.104	0.108	-0.269	<0.001	0.127	0.063
TLI					0.195	0.003	0.046	0.463
c-erbB-2							-0.042	0.535

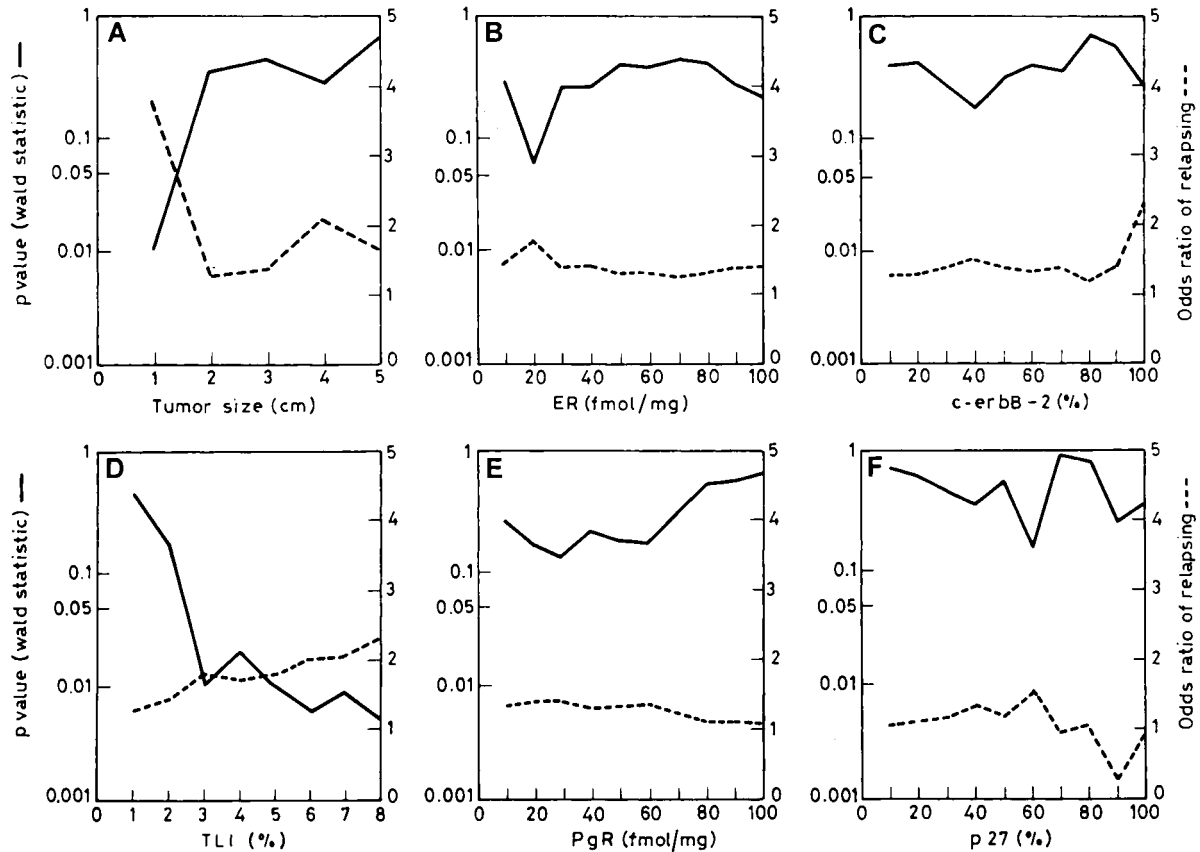


Figure 1. Cut off values for predicting 5-year relapse-free survival plotted against the relative p values (—) and Odds ratio of relapse (-----) (A: tumor size; B: ER; C: c-erbB-2; D: TLI; E: PgR; F: p27).

generally correct [28]. In this model, the exponential of each of the regression coefficients (β) is the odds ratio (OR), which is assumed to be constant in time. In univariate and multivariate analyses, the best putative prognosis categories were used as references. The null hypothesis $\beta_i = 0$ was tested by the Wald statistic. As the prognostic variables were categorised, one or more dummies were built for each of them.

The final model for multivariate analysis was obtained using a backward stepwise procedure. Variables

that did not contribute significantly to the multivariate Cox model ($p > 0.05$) were eliminated. A forward stepwise procedure was also performed, obtaining the same results as those of the backward procedure.

To evaluate the prognostic role of the different parameters analysed in the study we tested different cut off values ranging from the minimum to the maximum value observed in our case series. Each dichotomy was used as an independent variable in a Cox regression model to predict recurrence. Each model

Table 4. Five-year relapse-free survival as a function of clinico-pathologic-biologic variables

	Number of cases	RFS (%)	95% CI	Logrank	<i>p</i>
Menopausal status					
Premenopausal	113	76	68–84		
Postmenopausal	173	79	72–85	0.01	0.935
Tumor size					
≤ 1.0 cm	49	94	87–100		
1.1–2.0 cm	139	75	67–82		
> 2.0 cm	81	73	63–83	7.75	0.021
TLI					
<3.1%	143	83	76–89		
≥ 3.1%	143	72	65–80	5.83	0.016
ER					
≥ 10 fmol	198	76	70–82		
<10 fmol	77	79	70–88	1.21	0.271
PgR					
≥ 25 fmol	132	71	63–79		
<25 fmol	110	81	73–88	1.33	0.250
c-erbB-2					
<40%	175	80	74–86		
≥ 40%	54	70	57–82	1.73	0.188
p27					
≤ 60%	218	80	75–86		
> 60%	36	62	45–80	2.13	0.140

was evaluated with the Wald statistic and the one with the smallest *p* value was identified. All *p* values were based on two-sided testing and statistical analyses were carried out with SAS Statistical software [29].

Results

The analysis of the relationship between biologic and pathologic variables (Table 2) showed a significantly higher proliferative activity in younger patients and a suggestive, but not statistically significant, higher TLI in pre- rather than in postmenopausal women. Moreover, TLI median value increased as a function of tumor size and was significantly higher in ER or PgR negative tumors than in steroid receptor positive tumors.

c-erbB-2 immunoreactivity was observed in about 40% of tumors and the percentage of immunoreacting

cells ranged from 5% to 100%. Neither the percentage of c-erbB-2 positive tumors nor the percentage of positive cells were related to patient age or menopausal status, whereas a higher, albeit not significantly different, percentage of positive cells, was observed in larger or steroid receptor negative tumors than in smaller or steroid receptor positive tumors. Moreover, a lower percentage of positive cells was observed in lobular histotype than in ductal invasive histology. p27 positivity was observed in more than 70% of tumors and the percentage of positive cells ranged from 1% to 100%. A higher, but not significantly different (*p* = 0.09) frequency of immunoreacting cells was observed in tumors ≤ 1 cm. The percentage of positive cells was also higher in estrogen receptor (*p* = 0.05) or progesterone receptor positive (*p* = 0.08) tumors than in steroid receptor negative tumors.

The analysis of different markers analysed as continuous variables (Table 3) showed a significantly direct relation between c-erbB-2 and TLI, and an inverse

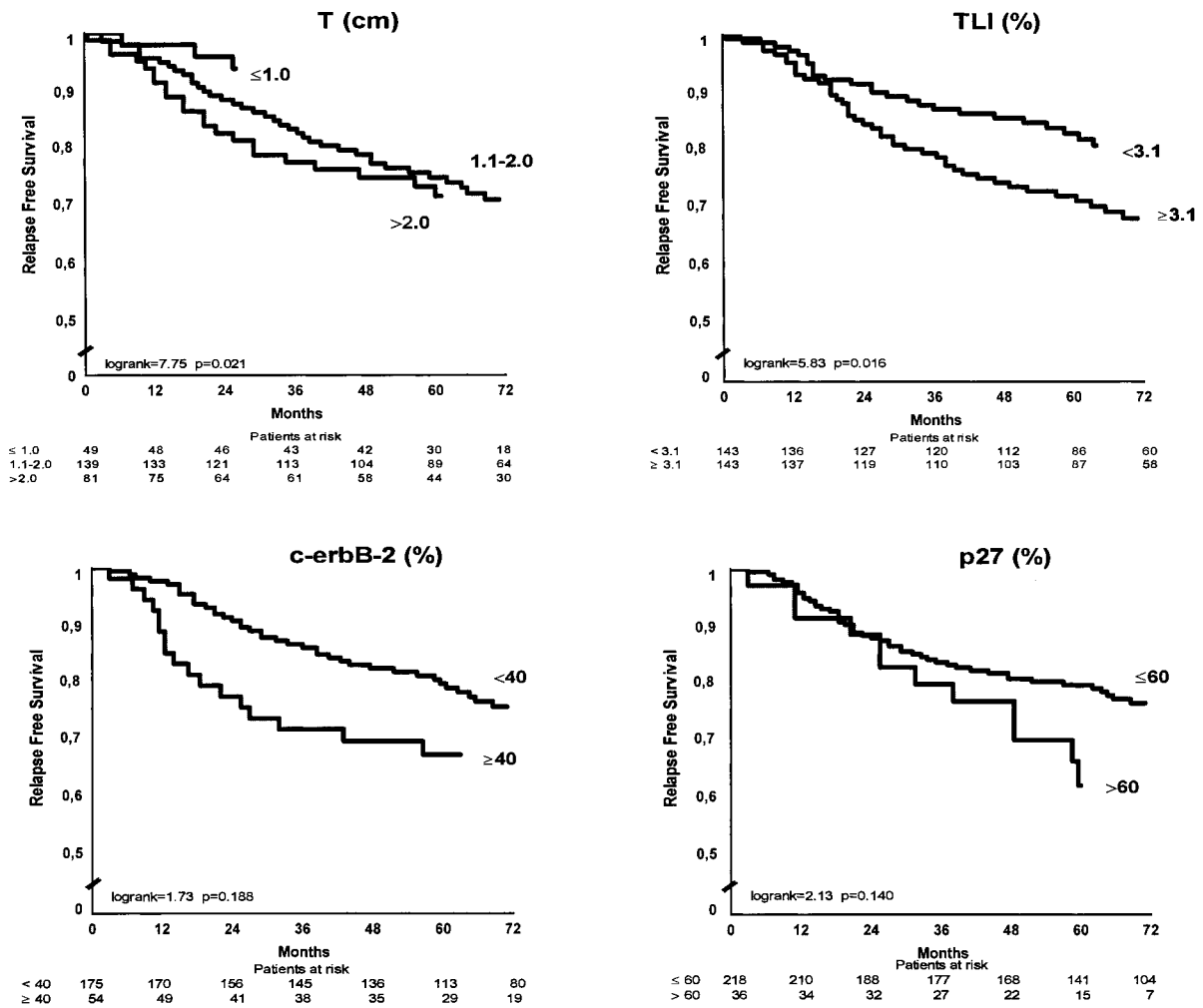


Figure 2. Relapse-free survival curves as a function of different prognostic variables.

relation between c-erbB-2 and ER and PgR. Moreover, p27 expression was significantly directly related to steroid receptor status but not to c-erbB-2 expression or proliferative activity. However, even when significant associations were observed, the correlation coefficients were very poor.

For the prognosis evaluation, tumor size, TLI, c-erbB-2 and p27 expression, and ER and PgR content were analysed as continuous or dichotomous variables. When considered as a continuous variable, pathologic tumor size showed the unique discriminant cut off value of 1 cm as predictor of risk of relapse at 5 years. TLI was able to identify subgroups of patients with a significantly different probability of relapse-free survival starting from the 3.0% of thymidine labeled cells. The odds ratio progressively increased as TLI value increased. Conversely, no value of

c-erbB-2 or p27 expression and ER or PgR content reached statistical significance as predictors of relapse (Figure 1).

In the present case series of node negative patients, 5-year relapse-free survival was 77% (95% CI 73–82%), in agreement with the results reported in most clinical studies. When biologic markers were analysed as dichotomous variables using conventional cut off values or the most discriminant values detected by the previous analyses on continuous variables, a significantly lower relapse-free survival at 5 years was observed for patients with tumors larger than 1 cm or rapidly proliferating tumors than in patients with smaller ($p = 0.021$) or slowly proliferating tumors ($p = 0.016$). Conversely, 5-year relapse-free survival was not related to c-erbB-2 or p27 expression when median values, other values or negativity versus

Table 5. Results from backward application of Cox model (260 patients, 68 events)

Step	Factor	Odds ratio	95% CI	p
<i>Relapse free survival</i>				
1	Tumor size (> 1 cm vs ≤ 1 cm)	3.34	1.21–9.18	0.019
2	TLI (≥ 3.1% vs <3.1%)	1.78	1.07–2.95	0.027
<i>Factors not entered</i>				
	ER status (≥ 10 fmol vs <10 fmol)	1.44	0.82–2.53	0.204
	Menopausal status (post vs pre)	1.05	0.64–1.71	0.847

positivity were considered as criteria. Similarly, the conventional ER and PgR cut off values, as well as any other value, did not identify subgroups of patients at different probabilities of 5-year relapse-free survival (Table 4).

The profile of relapse-free survival curves over time (Figure 2) showed significantly distinct curves for patients with small and large tumors. Similarly, significantly different relapse curves were observed for patients with slowly or rapidly proliferating tumors. In particular, diversification started from the first year and progressively increased up to the 5th year of follow-up.

Conversely, relapse-free survival analysed as a function of c-erbB-2 expression showed a higher risk in patients with tumors with a high rather than low c-erbB-2 expression. The difference increased starting from the 1st year up to the 3rd year and then tended to diminish progressively. The curves appeared to be getting closer together at the 5th year follow-up. Overall, the relapse-free survival risk was not significantly different for the two biologic subgroups. The relapse-free survival curves for patients with weakly or highly p27-expressing tumors were not significantly different and were superimposable, at least up to the 3rd year.

The variables which individually proved to be prognostic indicators, that is tumor size and TLI, in addition to ER status and menopausal status which have long been considered to be important indicators of the natural history of breast cancer, were tested in multivariate analysis. Only tumor size and, among the biologic variables investigated, cell proliferation, maintained their independent prognostic relevance on relapse-free survival (Table 5). Relapse-free survival curves for the four subgroups defined by the independ-

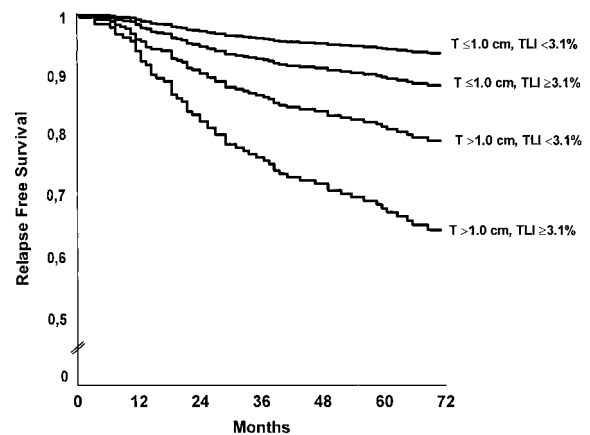


Figure 3. Relapse-free survival from the multivariate analysis.

ent prognostic variables identified by multivariate analysis are shown in Figure 3. For patients with tumors ≤ 1 cm, 5-year relapse-free survival was 94.5% (95% CI 89.1–100.0) when TLI was lower, and 89.9% (95% CI 80.7–100.0) when TLI was higher than ≥ 3.1%. In patients with tumors > 1 cm, 5-year relapse-free survival was 81.8% (95% CI 74.5–89.0) when TLI was low and 68.6% (95% CI 61.0–77.1) when TLI was high.

Discussion

Our study, performed on a large series of node-negative breast cancer patients given loco-regional treatment alone, showed, for the first time in a prospectively recruited series, the prognostic relevance of TLI and its independence from other clinicopathologic and biologic factors over a 5-year follow-

up period. These results confirm that tumor proliferative activity, evaluated as TLI, is a strong and reproducible indicator of tumor aggressiveness, as already proven by pilot and confirmatory retrospective studies on very large series of cases and with longer follow-up periods [30–41].

The reproducibility of TLI as an indicator of relapse-free survival and survival is favoured by the unequivocal autoradiographic image of labeled cells as compared to the modulation of immunohistochemical images, and guaranteed by an ongoing National Quality Control Program, which was activated in Italy almost 10 years ago [24]. Therefore, TLI represents an important variable for identifying patients with negative lymph-node tumors at risk who are thus candidates for adjuvant systemic therapy. Pathologic tumor size was also an independent prognostic indicator with a 94% disease-free probability at 5 years for patients with very small tumors. In contrast, in the present series of patients, hormonal receptor status did not affect the 5-year incidence of new disease manifestation. Moreover, the four subgroups defined according to TLI and pathologic tumor size, which were the only two independent prognostic variables identified by multivariate analysis, showed a significantly different risk of relapse. The risk was lowest for the subgroups of patients with the two favourable prognostic variables (about 5%) and more than six times higher for patients with the two unfavourable prognostic variables.

The fraction of c-erbB-2 expressing cells was not related to either patient age or menopausal status. In contrast, it was noticeably higher in ductal than in lobular histotype, in agreement with other results [42], and, as already reported, progressively increased as tumor size increased [43–46] and steroid receptor content decreased [42, 43, 47–51]. Moreover, we confirmed the relation between c-erbB-2 and cell proliferation. As far as we know, the only two papers which to date have investigated the relationship between c-erbB-2 expression and TLI in breast cancer found a significant association between high TLI and positive staining in both *in situ* [52] and invasive carcinoma [53].

The results on the prognostic value of c-erbB-2 are still somewhat contradictory, with the majority of studies reporting no prognostic relevance in node-negative breast cancers [42, 54–57]. The controversial results have been tentatively ascribed to the different follow-up periods considered in the studies [42]. The results from the present work appear to reinforce this

hypothesis, identifying, for the first time experimentally, c-erbB-2 expression as a short-term prognostic indicator. This finding further confirms the lack of long-term prognostic relevance of c-erbB-2, which, conversely, remains a potential predictor of response to hormonal therapy and to different antitumor drugs [58–63].

Similarly, the cell cycle inhibitor p27 was more frequently expressed in lobular than in ductal histotypes, but the difference did not reach statistical significance, probably due to unbalanced subgroups, and was inversely related to tumor size and directly related to steroid receptor content. In our case series of node-negative breast cancer patients treated with loco-regional therapy alone, p27 expression was not related to disease-free survival. This result cannot easily be compared with the findings obtained in other studies on large series of breast cancer patients. No information on the frequency of positive or negative p27 tumors is reported and the generally used semi-quantitative analysis is not a guarantee of equivalent evaluation criteria, even though similar cut off values of p27 positive cells were adopted in most of the studies for follow-up analysis. A prognostic relevance of p27 expression was observed in studies including both node-positive and node-negative patients considered as one group and treated with different types of chemotherapy or hormone therapy [64], or for whom treatment modalities are not specified [65–67]. The results on node negative patients are less consistent. In fact, a relation between the cell cycle inhibitor and clinical outcome was reported by Porter [67] and Wu [66] but not by Tan [64], who performed, as we did, a quantitative analysis of the percentage of p27 positive cells. However, it must also be pointed out that in all these studies, node-negative patients were treated with different types of systemic therapy, whereas our analysis was performed for the first time, as far as we know, on node-negative patients who underwent only loco-regional therapy. Our results are in agreement with those from a similar study recently published by Reed et al. [68].

In conclusion, TLI remains, in our experience, a variable of important clinical relevance by intrinsically containing both prognostic and predictive information on response to chemo- and hormone therapy [69–73]. Therefore, once node negative patients at risk have been identified on the basis of tumor size and cell proliferation, TLI, together with steroid receptors and c-erbB-2 expression, could open a new era of individually-tailored therapy for patients.

Acknowledgements

The co-authorship of all the following trial participants is acknowledged: P. Serra, A. Callea (Istituto Oncologico Romagnolo, Forlì), D. Casadei Giunchi, R. Maltoni (Department of Medical Oncology), A. Saragoni (Department of Pathology), A. Vio (Department of Surgery) – Morgagni Pierantoni Hospital, Forlì; A. Ravaioli, G. Drudi, L. Gianni (Department of Medical Oncology), R. Pozzuoli, P. Rinaldi, L. Bernardi (Department of Pathology) – Per gli Infermi Hospital, Rimini; F. Barbanti (Department of Surgery) – Santarcangelo Hospital, RN; A. Rossi, P. Turci (Medical Oncology Unit), B. Spada (Department of Surgery), V. Tison, F. Nuzzo, S. Naldi (Department of Histopathology) – Bufalini Hospital, Cesena, FO; L. Marri (Department of Histopathology), A. Gambi, L. Amaducci (Medical Oncology Unit) – Degli Infermi Hospital, Faenza, RA; G. Catalano, S. Luzi Fedeli (Department of Medical Oncology), F. Lungarotti (Department of Surgery), P. Mureto, A. Carnevali (Department of Histopathology) – S. Salvatore Hospital, Pesaro; A. Tienghi, M. Marangolo, B. Vertogen F. Zumaglini (Department of Medical Oncology), C. Morisi (Department of Pathology), G. Gaddoni (Department of Surgery) – S. Maria delle Croci Hospital, Ravenna; M. Indelli, A. Maestri (Department of Oncology) – S. Anna Hospital, Ferrara; P. Pacini, C. Fallai (Oncologic Day Hospital) – Careggi Hospital, Florence; V. Distante (Clinical Surgery I Institute), S. Bianchi (Histopathology Institute) – University of Florence, Florence.

The authors wish to thank Prof. Rosella Silvestrini for her invaluable scientific contribution and Ms. Grainne Tierney for editing the manuscript. This study was funded by the 'Consiglio Nazionale delle Ricerche' (CNR) and Istituto Oncologico Romagnolo.

References

1. Fisher B, Dignam J, Mamounas EP, Costantino JP, Wickerham DL, Redmond C, Wolmark N, Dimitrov NV, Bowman DM, Goss AG, Atkins JN, Abramson N, Sutherland CM, Aron BS, Margolese RG: Sequential methotrexate and fluorouracil for the treatment of node-negative breast cancer patients with estrogen receptor-negative tumors: eight-year results from National Surgical Adjuvant Breast and Bowel Project (NSABP) B-13 and first report of finding from NSABP B-19 comparing methotrexate and fluorouracil with conventional cyclophosphamide, methotrexate, and fluorouracil. *J Clin Oncol* 14: 1982–1992, 1996
2. Zambetti M, Valagussa P, Bonadonna G: Adjuvant cyclophosphamide, methotrexate and fluorouracil in node-negative and estrogen receptor-negative breast cancer. *Ann Oncol* 7: 481–485, 1996
3. The Ludwig Breast Cancer Study Group: Prolonged disease-free survival after one course of perioperative adjuvant chemotherapy for node-negative breast cancer. *N Engl J Med* 320: 491–496, 1989
4. Mansour EG, Gray R, Shatila AH, Osborne CK, Tormey DC, Gilchrist KW, Cooper MR, Falkson G: Efficacy of adjuvant chemotherapy in high-risk node-negative breast cancer. *N Engl J Med* 320: 485–490, 1989
5. Kerr JF, Wyllie AH, Currie AR: Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer* 26: 239–257, 1972
6. Fraser A, Evan G: A licence to kill. *Cell* 85: 781–784, 1996
7. Salvesen G, Dixit VM: Caspases: intracellular signaling by proteolysis. *Cell* 91: 443–446, 1997
8. Folkman J: What is the evidence that tumors are angiogenesis dependent? *J Nat Cancer Inst* 82(1): 4–6, 1990
9. Folkman J: Angiogenesis and breast cancer. *J Clin Oncol* 12(3): 441–443, 1994
10. Zhou P, Jiang W, Weghorst CM, Weinstein IB: Overexpression of cyclin D1 enhances gene amplification. *Cancer Res* 56(1): 36–39, 1996
11. King CR, Kraus MH, Aaronson SA: Amplification of a novel v-erbB related gene in a human mammary carcinoma. *Science* 229: 974–976, 1985
12. Akijama T, Sudo C, Ogawara H, Toyoshima K, Yamamoto T: The product of the human c-erbB-2 gene: a 185-kilodalton glycoprotein with tyrosine kinase activity. *Science* 232: 1644–1646, 1986
13. Kraus MH, Issing W, Miki T, Popescu NC, Aaronson SA: Isolation and characterization of ERBB3, a third member of the ERBB/epidermal growth factor receptor family: evidence for overexpression in a subset of human mammary tumors. *Proc Natl Acad Sci* 86: 9193–9197, 1989
14. Klÿn JGM, Berns PMJJ, Schmitz PIM, Foekeus JA: The clinical significance of epidermal growth factor receptor (EGF-R) in human breast cancer: a review on 5232 patients. *Endocr Rev* 13: 156–170, 1992
15. Gullick WJ, Srinivasan R: The type 1 growth factor receptor family: new ligands and receptors and their role in breast cancer. *Breast Cancer Res Treat* 52: 43–53, 1998
16. Stephens RW, Brÿnner N, Jänicke F, Schmitt M: The urokinase plasminogen activator system as a target for prognostic studies in breast cancer. *Breast Cancer Res Treat* 52: 99–111, 1998
17. Levine AJ, Momand J, Finlay CA: The p53 tumor suppressor gene (Review article). *Nature* 351: 453–456, 1991
18. Chang F, Syrjanen S, Syrjanen K: Implication of the p53 tumor suppressor gene in clinical oncology. *J Clin Oncol* 13: 1009–1022, 1995
19. Silvestrini R, Benini E, Daidone MG, Veneroni S, Boracchi P, Cappelletti V, Di Fronzo G, Veronesi U: p53 as an independent prognostic marker in node-negative breast cancers. *J Nat Cancer Inst* 85: 965–970, 1993
20. Silvestrini R, Daidone MG, Del Bino G, Mastore M, Di Fronzo G, Boracchi P: Prognostic significance of proliferative activity and ploidy in node-negative breast cancers. *Ann Oncol* 4: 213–219, 1993
21. Silvestrini R, Daidone MG, Di Fronzo G, Morabito A, Valagussa P, Bonadonna G: Prognostic implication of labeling index versus estrogen receptors and tumor size in node-negative breast cancer. *Breast Cancer Res Treat* 7: 161–169, 1986

22. Silvestrini R, Daidone MG, Mastore M, Di Fronzo G, Coradini D, Boracchi P, Squicciarini P, Salvadori B, Veronesi U: Cell kinetics as a predictive factor in node-positive breast cancer treated with adjuvant hormone therapy. *J Clin Oncol* 11: 1150–1155, 1993
23. Silvestrini R, Daidone MG, Valagussa P, Di Fronzo G, Mezzanotte G, Squicciarini P, Orefice S, Salvadori B: ³H-thymidine labeling index as a prognostic indicator in node-positive breast cancer. *J Clin Oncol* 8: 1321–1326, 1990
24. Silvestrini R, The SICCAB Group for quality control of cell kinetic determination: Feasibility and reproducibility of the ³H-TdR labeling index in breast cancer. *Cell Prolif* 21: 437–445, 1991
25. Piffanelli A, Pellizzola D, Giovannini G, Catozzi L, Faggioli L, Giganti M: Characterization of laboratory working standards for quality control of immunometric and radiometric estrogen receptor assays. *Clinical evaluation of breast cancer biopsies. Tumori* 75: 550–556, 1989
26. Kaplan EL, Meier P: Nonparametric estimation for incomplete observation. *J Am Stat Assoc* 53: 457–481, 1958
27. Cox DR: Regression models and life tables. *J Roy Stat Soc* 34: 187–220, 1972
28. Lawless JS: Statistical models and methods for life-time data. Wiley, New York, 1982
29. SAS Institute Inc: SAS/STAT User's Guide, version 6, 4th ed., vol. 1. Cary, NC: SAS Institute, 1989
30. Gentili C, Sanfilippo O, Silvestrini R: Cell proliferation in relation to clinical features and relapse in breast cancers. *Cancer* 48: 974–979, 1981
31. Silvestrini R, Daidone MG, Luisi A, Mastore M, Leutner, Salvadori B: Cell proliferation in 3800 node-negative breast cancers: consistency over time of biological and clinical information provided by ³H-thymidine labelling index. *Int J Cancer* 74: 122–127, 1997
32. Tubiana M, Pejovic MH, Chavandra G, Contesso G, Malaise EP: The long term prognostic significance of the thymidine labeling index in breast cancer. *Int J Cancer* 33: 441–445, 1984
33. Meyer JS, Friedman E, Mc Crote MM, Bauer WC: Prediction of early course of breast carcinoma by thymidine labeling. *Cancer* 51: 1879–1886, 1983
34. Hery M, Gioanni J, Lalanne CM, Namer M, Coudri A: The DNA labeling index: a prognostic factor in node-negative breast cancer. *Breast Cancer Res Treat* 9: 207–212, 1987
35. Meyer JS, Province M: Proliferative index of breast cancer by thymidine labeling index: prognostic power independent of stage, estrogen and progesteron receptors. *Breast Cancer Res Treat* 12: 191–199, 1988
36. Silvestrini R, Daidone MG, Valagussa P, Di Fronzo G, Mezzanotte G, Bonadonna G: Cell kinetics as a prognostic indicator in node-negative breast cancer. *Eur J Cancer Clin Oncol* 25: 1165–1171, 1989
37. Silvestrini R, Daidone MG, Di Fronzo G, Morabito A, Valagussa P, Bonadonna G: Prognostic implication of labeling index versus estrogen receptors and tumor size in node-negative breast cancer. *Breast Cancer Res Treat* 7: 161–169, 1986
38. Mangia A, Picciarelllo M, Tommasi S, Simone G, Barletta A, Longo S, De liso M, D'Errico D, Schittulli F, Catino AM, Paradiso A, De Lena M: Fattori prognostici nel carcinoma della mammella operabile N-: attività proliferativa e caratteristiche clinico-patologiche. *Folia Oncologica* 14: 247–253, 1991
39. Tubiana M, Pejovic MH, Kosciencly S, Chavandra N, Malaise E: Growth rate, kinetics of tumor cell proliferation and long-term outcome in human breast cancer. *Int J Cancer* 44: 17–22, 1987
40. Silvestrini R, Daidone MG, Gasparini G: Cell kinetics as a prognostic marker in node-negative breast cancer. *Cancer* 56: 1982–1987, 1985
41. Silvestrini R, Daidone MG, Luisi A, Boracchi P, Mezzetti M, Di Fronzo G, Andreola S, Veronesi U: Biologic and clinicopathologic factors as indicators of specific relapse types in node-negative breast cancer. *J Clin Oncol* 13: 697–704, 1995
42. Révillion F, Bonneterre J, Peyrat JP: ERBB2 oncogene in human breast cancer and its clinical significance. *Eur J Cancer* 34: 791–808, 1998
43. Kallionemi OP, Holli K, Visakorpi T, Koivula T, Helin HH, Isola JJ: Association of c-erbB-2 protein over-expression with high rate of cell proliferation, increased risk of visceral metastasis and poor-long term survival in breast cancer. *Int J Cancer* 49: 650–655, 1991
44. Wiltschke C, Kindas-Muegge I, Steininger A, Reiner A, Reiner G, Preis PN: Coexpression of HER-2/neu and p53 is associated with a shorter disease-free survival in node-positive breast cancer patients. *J Cancer Res Clin Oncol* 120: 737–742, 1994
45. Rilke F, Colnaghi MI, Cascinelli N, Andreola S, Baldini MT, Bufalino R, Della Porta G, Menard S, Pierotti MA, Testori A: Prognostic significance of HER-2/neu expression in breast cancer and its relationship to other prognostic factors. *Int J Cancer* 49: 44–49, 1991
46. Delarue JC, Terrier P, Terrier-Lacombe MJ, Mouriesse H, Gotteland M, May-Levin F: Combined overexpression of c-erbB-2 protein and epidermal growth factor receptor (EGR-R) could be predictive of early and long-term outcome in human breast cancer: a pilot study. *Bull Cancer* 81: 1067–1077, 1994
47. De Potter CR, Beghin C, Makar AP, Vandekerckhove D, Roels HJ: The neu-oncogene protein as a predictive factor for haematogenous metastases in breast cancer patients. *Int J Cancer* 45: 55–58, 1990
48. O'Reilly SM, Barnes DM, Camplejohn RS, Bartkova J, Gregory WM, Richards MA: The relationship between c-erbB-2 expression, S-phase fraction and prognosis in breast cancer. *Br J Cancer* 63: 444–446, 1991
49. Kommos F, Colley M, Hart CE, Franklin WA: *In situ* distribution of oncogene products and growth factor receptors in breast carcinoma: c-erbB-2 oncoprotein, EGFr, and PDGFr- β -subunit. *Mol Cell Probes* 4: 11–23, 1990
50. Lovekin C, Ellis IO, Locker A, Robertson JF, Bell J, Nicholson R, Gullick WJ, Elston CW, Blaney RW: c-erbB-2 oncoprotein expression in primary and advanced breast cancer. *Br J Cancer* 63: 439–443, 1991
51. Hartmann LC, Ingle JN, Wold LE, Farr GH Jr, Grill JP, Su JQ, Maihle NJ, Krook JE, Witzig TE, Roche PC: Prognostic value of c-erbB-2 overexpression in axillary lymph node positive breast cancer. Results from a randomized adjuvant treatment protocol. *Cancer* 74: 2956–2963, 1994
52. Barnes DM, Meyer JS, Gonzalez JG, Gullick WJ, Millis RR: Relationship between c-erbB-2 immunoreactivity and thymidine labeling index in breast carcinoma *in situ*. *Breast Cancer Res Treat* 18: 11–17, 1991
53. Contegiacomo A, Pizzi C, De Marchis L, Alimandi M, Delrio P, Di Palma E, Petrella G, Ottini L, French D, Frati L, Bianco AR, Mariani-Costantini R: High cell kinetics is associated with amplification of the int-2, bcl-1, myc and erbB-2 proto-oncogenes and loss of heterozygosity at the DF3 locus in primary breast cancers. *Int J Cancer* 61(1): 1–6, 1995

54. Slamon DJ, Godolphin W, Jones LA, Holt JA, Wong SG, Keith DE, Stuart SG, Udove J, Ullrich A, Press MF: Studies of the HER-2/neu proto-oncogene in human breast and ovarian cancer. *Science* 244: 707–712, 1989
55. Thor AD, Schwartz LH, Koerner FC, Edgerton SM, Skates SJ, Yin S, McKenzie SJ, Paniali DL, Marks PJ, Fingert HJ, et al.: Analysis of c-erbB-2 expression in breast carcinomas with clinical follow-up. *Cancer Res* 49: 7147–7152, 1989
56. Gusterson BA, Gelber RD, Goldhirsch A, Price KN, Säv-Söderborgh J, Anbazhagan R, Styles J, Rudenstam CM, Goulouh R, Reed R, Martínez-Tello F, Tiltman A, Torhorst J, Grigolato P, Bettelheim R, Neville AM, Bürki K, Castiglione M, Collins J, Lindtner J, Senn H-J for the International (Ludwig) Breast Cancer Study Group: Prognostic importance of c-erbB-2 expression in breast cancer. *J Clin Oncol* 10: 1049–1056, 1992
57. Gasparini G, Widner N, Bevilacqua P, Maluta S, Dalla Palma P, Caffo O, Barbareschi M, Boracchi P, Marubini E, Pozza F: Tumor microvessel density, p53 expression, tumor size, and peritumoral lymphatic vessel invasion are relevant prognostic markers in node-negative breast carcinoma. *J Clin Oncol* 12: 454–466, 1994
58. Muss HB, Thor AD, Berry DA, Kute T, Liu ET, Koerner F, Cirincione CT, Budman DR, Wood WC, Barcos M, Henderson IC: c-erbB-2 expression and response to adjuvant therapy in women with node-positive early breast cancer. *N Engl J Med* 330: 1260–1266, 1994
59. Allred DC, Clark GM, Tandon AK, Molina R, Tormey DC, Osborne CK, Gilchrist KW, Mansour EG, Abeloff M, Eudey L, Mc Guire WL: HER-2/neu in node-negative breast cancer: prognostic significance of overexpression influenced by the presence of *in situ* carcinoma. *J Clin Oncol* 10: 599–605, 1992
60. Arteaga CL, Winnier AR, Poirier MC, Lopez-Larraz DM, Shawver KL, Hurd SD, Stewart SJ: p185 c-erbB-2 signalling enhances cisplatin-induced cytotoxicity in human breast carcinoma cells: association between an oncogenic receptor tyrosine kinase and drug-induced DNA repair. *Cancer Res* 54: 3758–3765, 1994
61. Baselga J, Seidman AD, Rosen PP, Norton L: HER2 overexpression and paclitaxel sensitivity in breast cancer; therapeutic implications. *Oncology (Huntingt)* 11(Suppl 2): 43–48, 1997
62. Leitzel K, Teramoto Y, Konrad K, Chinchilli VM, Volas G, Grossberg H, Harvey H, Demers L, Lipton A: Elevated serum c-erbB-2 antigen levels and decreased response to hormone therapy of breast cancer. *J Clin Oncol* 13: 1129–1135, 1995
63. Stål O, Sullivan S, Wingren S, Skoog L, Rutqvist LE, Carstensen JM, Nordenskjöld B: c-erbB-2 expression and benefit from adjuvant chemotherapy and radiotherapy of breast cancer. *Eur J Cancer* 31A: 185–190, 1995
64. Tan P, Cady B, Wanner M, Worland P, Cukor B, Magi-Galluzzi C, Lavin P, Draetta G, Pagano M, Loda M: The cell cycle inhibitor p27 is an independent prognostic marker in small (T1a, b) invasive breast carcinomas. *Cancer Res* 57: 1259–1263, 1997
65. Catzavelos C, Bhattacharya N, Ung YC, Wilson JA, Roncari L, Sandhu C, Shaw P, Yeger H, Morava-Protzner I, Kapusta L, Franssen E, Pritchard KI, Slingerland JM: Decreased levels of the cell-cycle inhibitor p27^{Kip1} protein: prognostic implications in primary breast cancer. *Nat Med* 3: 227–230, 1997
66. Wu J, Shen Z-Z, Jiang M, Han Q-N, Fontana JA, Barsky SH, Shao ZM: Prognostic role of p27^{Kip1} and apoptosis in human breast cancer. *Br J Cancer* 79: 1572–1578, 1999
67. Porter PL, Malone KE, Heagerty PJ, Alexander GM, Gatti LA, Firpo EJ, Daling JR, Roberts JM: Expression of cell-cycle regulators p27^{Kip1} and cyclin E, alone and in combination, correlates with survival in young breast cancer patients. *Nat Med* 2: 222–225, 1997
68. Reed W, Florenes WA, Holm R, Hannisdal E, Nesland JN: Elevated levels of p27, p21 and cyclin D1 correlate with positive oestrogen and progesterone receptor status in node-negative breast carcinoma patients. *Virchows Arch* 435: 116–124, 1999
69. Sulkes A, Livingstone RB, Murphy WK: Tritiated thymidine labeling index and response in human breast cancer. *J Natl Cancer Inst* 62: 513–515, 1979
70. Daidone MG, Silvestrini R, Canova S, Valagussa P: Tumor cell kinetics and course of node-positive (N+) breast cancer (Abstract). *Proc Am Soc Clin Oncol* 8: 24, 1989
71. Zambetti M, Bonadonna G, Valagussa P, Daidone MG, Coradini D, Bignami P, Contesso G, Silvestrini R: Adjuvant CMF for node-negative and estrogen receptor-negative breast cancer. *NCI Monograph* 11: 77–83, 1992
72. Gardin G, Alama A, Rosso R, Campora E, Repetto L, Pronzato P, Merlini L, Naso C, Camoriano A, Meazza R: Relationship of variations in tumor cell kinetics induced by primary chemotherapy to tumor regression and prognosis in locally advanced breast cancer. *Breast Cancer Res Treat* 32: 311–318, 1994
73. Amadori D, Volpi A, Maltoni R, Nanni O, Amaducci L, Amadori A, Casadei Giunchi D, Vio A, Saragoni A, Silvestrini R: Cell proliferation as a predictor of response to chemotherapy in metastatic breast cancer: a prospective study. *Breast Cancer Res Treat* 43: 7–14, 1997

Address for offprints and correspondence: Prof. Dino Amadori, Department of Medical Oncology, Pierantoni Hospital, Via Forlanini 34, 47100 Forlì, Italy; *Tel.:* 01139 0543 731737; *Fax:* 01139 0543 731736; *E-mail:* divonco@ausl.fo.it; i.o.r@fo.nettuno.it