



Cedarville University
DigitalCommons@Cedarville

Pharmacy and Nursing Student Research and Evidence-Based Medicine Poster Session

Fall 12-6-2013

Natural Products as Therapeutic Agents in Cancer Treatment

Eric Huseman

Cedarville University, ehuseman@cedarville.edu

Lauren P. Williams

lpwilliams@cedarville.edu

Brittany Santee

Cedarville University, brittanysantee@cedarville.edu

Trevor Stump

Cedarville University, tstump@cedarville.edu

Chelsae Ward

Cedarville University, chelsaeeward@cedarville.edu

See next page for additional authors

Follow this and additional works at: [http://digitalcommons.cedarville.edu/
pharmacy_nursing_poster_session](http://digitalcommons.cedarville.edu/pharmacy_nursing_poster_session)

 Part of the [Nursing Commons](#), and the [Pharmacy and Pharmaceutical Sciences Commons](#)

Recommended Citation

Huseman, Eric; Williams, Lauren P.; Santee, Brittany; Stump, Trevor; Ward, Chelsae; Kunze, Rachel; Simpson, Denise S.; and Amos, Samson, "Natural Products as Therapeutic Agents in Cancer Treatment" (2013). *Pharmacy and Nursing Student Research and Evidence-Based Medicine Poster Session*. 55.

http://digitalcommons.cedarville.edu/pharmacy_nursing_poster_session/55

This Poster Session is brought to you for free and open access by DigitalCommons@Cedarville, a service of the Centennial Library. It has been accepted for inclusion in Pharmacy and Nursing Student Research and Evidence-Based Medicine Poster Session by an authorized administrator of DigitalCommons@Cedarville. For more information, please contact digitalcommons@cedarville.edu.



Authors

Eric Huseman, Lauren P. Williams, Brittany Santee, Trevor Stump, Chelsae Ward, Rachel Kunze, Denise S. Simpson, and Samson Amos

Natural products as therapeutic agents in cancer treatment

Eric Huseman, Lauren Williams, Brittany Santee, Trevor Stump, Chelsae Ward,
Rachel Kunze, Denise Simpson, Samson Amos

School of Pharmacy, Cedarville University, Cedarville, OH 45314



STATEMENT OF THE PROBLEM

Background

Cancer accounts for 25% of deaths in the United States.¹ Brain tumors play a large role in this percentage. The brain tumor incidence rate was 6.5 per hundred thousand between the years 2006 and 2010 in the United States.² Statistics indicate that there is an age adjusted mortality rate of 4.3 per hundred thousand persons across all age groups diagnosed with brain and other nervous system invasive cancers.²

Flavonoids and curcuminoids have shown potential for treating brain tumors in past studies. Genistein, a flavonoid, has shown promise for treating brain tumors since it inhibits the growth of glioblastoma and medulloblastoma cells by stopping the cell cycle at the G2/M checkpoint.³ Another study found that the curcuminoid curcumin binds to the cell surface membrane and later goes into the cytoplasm to induce cell death.⁴ A separate study found that curcumin downregulates various IGF (insulin-like growth factor) ligands to inhibit glioblastoma and medulloblastoma cells.⁵ Despite these promising results, there are some major gaps in the research. Specifically, there has not been a significant amount of research done on the effectiveness of flavonoids in the treatment of brain tumors.

Significance of the Problem

Relative to other types of cancers, brain tumors are difficult to treat. They prove resistant to chemotherapy and radiotherapy and show an increased tendency for infiltration. In response, scientists have conducted research to reveal the nature of brain tumor cells and the relationship they have with their *in vivo* environment. In one study, researchers found that brain tumor stem cells live in a unique environment. They depend on perivascular cells within the brain which provide an ideal growth environment for these stem cells.⁶ These researchers also discovered that brain tumor cells depend on microRNAs for growth, proliferation, and overall survival.⁶ Some avenues of current brain tumor research attempt to target these microRNAs in hopes of discovering a more effective clinical treatment.⁶

Past research has studied the effects of certain plant compounds such as curcuminoids and cyanidins on various types of cancer cells. These compounds were found to inhibit metastasis in breast and lung cancer cells.^{7,8} Our work will attempt to build and expand upon prior research examining flavonoids' effect on brain tumors in an attempt to find an effective treatment.

OBJECTIVES

Objective 1: To determine the effectiveness of flavonoids as therapeutic agents in brain tumor treatment.

Objective 2: To identify the signaling mechanism by which flavonoids mediate their therapeutic effects on brain tumor cell lines.

HYPOTHESES

Alternative Hypothesis 1 for Objective 1: There is a difference in cell death between cells treated with flavonoids and control cells not given any treatment.

Alternative Hypothesis 2 for Objective 1: There is a difference in cell proliferation between cells treated with flavonoids and control cells not given any treatment.

Alternative Hypothesis 3 for Objective 1: There is a difference in brain tumor invasion between cells treated with flavonoids and control cells not given any treatment.

PROPOSED METHODS

Study Design

Controlled experimental study

Sample

Human brain tumor cell lines: U-1242, U-251, and U-87

Treatment

Flavonoids at 10, 20, 40, and 80 μ M

Data Collection

Experiments will be performed in triplicate unless further trials are needed to resolve discrepancies found in initial three experiments. Data from flow analyses and Western Blot analyses will be saved as JPEG files and printed. Data from MTT assay and cell count will be recorded in laboratory notebooks. Notebooks will be checked and witnessed by fellow research participants. Data will be inputted into IBM SPSS 21[®] as appropriate.

Measurements

Cell count

Cells will be stained with trypan blue and counted using the grid count method and a hemocytometer.⁹

Cell proliferation

MTT Assays will be utilized to determine how flavonoids influence the proliferation of brain tumor cells.¹⁰ Western blot analyses will be conducted to identify proliferation markers such as MAPK, Akt, EGFR, and PKCs.¹¹

Cell death

Samples will be prepared and analyzed using flow cytometry to measure cell growth.¹⁰ Western blot analyses will be conducted to identify apoptotic markers such as cleaved Poly ADP ribose polymerase (PARP) and cleaved caspases.¹¹

PROPOSED ANALYSES

IBM SPSS 21[®] will be used to conduct all statistical analyses. Unpaired *t*-tests will be run to compare results from treated cells to control cells at 95% confidence. If necessary, one-way ANOVA with multiple comparisons will be used to compare multiple treatment groups and a control at 95% confidence. The Tukey test will be used to run post-hoc analyses if needed.

PROJECT TIMELINE

January 2014-December 2015: Data collection and analysis

LIMITATIONS

- While the study examines multiple brain tumor cell lines, the generalizability of the study is limited as there are many more cell lines found in brain tumors.
- Results obtained in the *in vitro* environment may not translate to the *in vivo* environment.

FUTURE DIRECTIONS

The goal of this study is to provide the basis for future *in vivo* translational studies in animal models.

ACKNOWLEDGEMENTS

We would like to thank Dr. Aleda Chen (Cedarville University) for her intellectual contributions to this project.

REFERENCES

1. Division of Cancer Prevention and Control, National Center for Chronic Disease Prevention and Health Promotion. Cancer statistics by cancer type. Center for Disease Control and Prevention Web site. Available at <http://www.cdc.gov/cancer/dccp/data/types.htm>. Updated December 19, 2012. Accessed September 22, 2013.
2. National Cancer Institute. Table 3.7: Cancer of the Brain and Other Nervous System (Invasive). U.S. National Institutes of Health. Available at: <http://seer.cancer.gov/csr/>. Accessed September 22, 2013.
3. Khaw A, Yong J, Kalthur G, Hande M. Genistein induces growth arrest and suppresses telomerase activity in brain tumor cells. *Genes Chromosomes And Cancer* [serial online]. October 1, 2012;51(10):961-974. Available from: Scopus[®], Ipswich, MA. Accessed September 22, 2013.
4. Hande M, Khaw A, Hande M, Kalthur G. Curcumin inhibits telomerase and induces telomere shortening and apoptosis in brain tumour cells. *Journal Of Cellular Biochemistry* [serial online]. June 1, 2013;114(6):1257-1270. Available from: Scopus[®], Ipswich, MA. Accessed September 22, 2013.
5. Lim K, Bisht S, Bar E, Maitra A, Eberhart C. A polymeric nanoparticle formulation of curcumin inhibits growth, clonogenicity and stem-like fraction in malignant brain tumors. *Cancer Biology And Therapy* [serial online]. March 1, 2011;11(5):464-473. Available from: Scopus[®], Ipswich, MA. Accessed September 22, 2013.
6. Koshkin A, Chistiakov D, Chekhonin VP. Role of microRNAs in Mechanisms of Glioblastoma Resistance to Radio and Chemotherapy. *BIOCHEMISTRY (Moscow)* [0006-2979]. 2012; 78(4). Available at Russian National Research Medical University. Accessed September 18, 2013. 1975_2010/browse_csr.php?section=3&page=sect_03_table.07.html. Accessed September 22, 2013.
7. Chendril D, Ranga R, Meigooni D, Sathishkumar S, Ahmed M. Curcumin confers radiosensitizing effect in prostate cancer cell line PC-3. *Oncogene* [serial online]. February 26, 2004;23(8):1599-1607. Available from: Academic Search Complete, Ipswich, MA. Accessed September 7, 2013.
8. Chen P, Chu S, Chiou H, Kuo W, Chiang C, Hsieh Y. Mulberry anthocyanins, cyanidin 3-rutinoside and cyanidin 3-glucoside, exhibited an inhibitory effect on the migration and invasion of a human lung cancer cell line. *Cancer Letters*. 2005;10.1016. Available from: Science Direct, The Netherlands. Accessed September 12, 2013.
9. Amos S et al. Farnesylthiosalicylic acid induces capsase activation and apoptosis in glioblastoma cells. *Cell Death and Differentiation*. 2006;13:642-651.
10. Cagle P et al. Effect of rosehip (*Rosa canina*) extract on human brain tumor cell proliferation and apoptosis. *Journal of Cancer Therapy*. 2012;3:534-545.
11. Amos S, Martin P, Polar G, Parsons S, Hussaini I. Phorbol 12-myristate 13-acetate induces epidermal growth factor receptor transactivation via protein kinase Cdelta/c-Src pathways in glioblastoma cells. *The Journal Of Biological Chemistry* [serial online]. March 4, 2005;280(9):7729-7738. Available from: MEDLINE with Full Text, Ipswich, MA. Accessed November 3, 2013.