



Technological University Dublin ARROW@TU Dublin

Articles

2019

# A review of the putative causal mechanisms associated with lower macular pigment in diabetes mellitus

Grainne Scanlon Technological University Dublin, grainne.scanlon@tudublin.ie

James Loughman Technological University Dublin, james.loughman@tudublin.ie

Donal Farrell Technological University Dublin

Daniel McCartney Technological University Dublin, Daniel.McCartney@tudublin.ie

Follow this and additional works at: https://arrow.tudublin.ie/otpomart

Part of the Optometry Commons

### **Recommended Citation**

Scanlon G, Loughman J, McCartney D. A review of the putative causal mechanisms associated with lower macular pigment in diabetes mellitus. Nutrition Research Reviews. 2019 Aug 14:1-18. doi: 10.1017/S095442241900012X

This Article is brought to you for free and open access by ARROW@TU Dublin. It has been accepted for inclusion in Articles by an authorized administrator of ARROW@TU Dublin. For more information, please contact yvonne.desmond@tudublin.ie, arrow.admin@tudublin.ie, brian.widdis@tudublin.ie.



This work is licensed under a Creative Commons Attribution-Noncommercial-Share Alike 3.0 License



# A review of the putative causal mechanisms associated with lower macular pigment in diabetes mellitus

Grainne Scanlon<sup>1\*</sup>, James Loughman<sup>1,2</sup>, Donal Farrell<sup>3</sup> and Daniel McCartney<sup>3</sup>

<sup>1</sup>Centre for Eye Research Ireland, School of Physics, Clinical & Optometric Sciences, Environmental Sustainability and Health Institute, Technological University Dublin, Dublin, Republic of Ireland <sup>2</sup>African Vision Research Institute, University of KwaZulu-Natal, Durban, South Africa <sup>3</sup>School of Biological and Health Sciences, Technological University Dublin, City Campus, Dublin, Republic of Ireland

#### Abstract

Macular pigment (MP) confers potent antioxidant and anti-inflammatory effects at the macula, and may therefore protect retinal tissue from the oxidative stress and inflammation associated with ocular disease and ageing. There is a body of evidence implicating oxidative damage and inflammation as underlying pathological processes in diabetic retinopathy. MP has therefore become a focus of research in diabetes, with recent evidence suggesting that individuals with diabetes, particularly type 2 diabetes, have lower MP relative to healthy controls. The present review explores the currently available evidence to illuminate the metabolic perturbations that may possibly be involved in MP's depletion. Metabolic co-morbidities commonly associated with type 2 diabetes, such as overweight/obesity, dyslipidaemia, hyperglycaemia and insulin resistance, may have related and independent relationships with MP. Increased adiposity and dyslipidaemia may adversely affect MP by compromising the availability, transport and assimilation of these dietary carotenoids in the retina. Furthermore, carotenoid intake may be compromised by the dietary deficiencies characteristic of type 2 diabetes, thereby further compromising redox homeostasis. Candidate causal mechanisms to explain the lower MP levels reported in diabetes include increased oxidative stress, inflammation, hyperglycaemia, insulin resistance, overweight/ obesity and dyslipidaemia; factors that may negatively affect redox status, and the availability, transport and stabilisation of carotenoids in the retina. Further study in diabeteic populations is warranted to fully elucidate these relationships.

Key words: HDL: Macular pigment: TAG: Oxidative stress: Inflammation

#### Introduction

The macula is an oval-shaped area at the centre of the retina which consists of a dense collection of light-sensitive cone cells responsible for central, high-resolution vision and colour perception<sup>(1)</sup>. At the centre of the macula lies the fovea, where macular pigment (MP) is located. MP is comprised of three carotenoids: lutein, zeaxanthin and the retinal metabolite of lutein, meso-zeaxanthin<sup>(2,3)</sup>. Lutein and zeaxanthin are not synthesised de novo in humans but are exclusively of dietary origin, typically derived from a diet rich in coloured fruit and vegetables<sup>(4)</sup>, while meso-zeaxanthin can be generated by conversion from retinal lutein<sup>(5)</sup> or may be obtained from dietary sources such as trout and salmon<sup>(6)</sup>. The highest concentration of MP is found in the receptor axon layer of the foveola, while in the parafovea, MP is located in the inner plexiform layers<sup>(7,8)</sup>, an area of the retina which is primed for the generation of reactive oxygen species (ROS) and consequently, oxidative damage<sup>(9)</sup>. These hydroxyl-carotenoids are selectively located in the macula to the exclusion of all other dietary carotenoids. Fig. 1 highlights the location of MP and the macula within the eye.

MP has a number of important functions in the eye. Evidence suggests that MP contributes to visual performance and/or experience, as it acts as an optical filter for blue light (peak absorption approximately 460 nm)<sup>(3,10)</sup>. The importance of MP's antioxidant and anti-inflammatory properties is supported by its capacity to protect against the cumulative damaging effects of oxidative stress<sup>(9)</sup>, and inflammation<sup>(11)</sup>, which affect the macula in conditions such as age-related macular degeneration (AMD)<sup>(12-15)</sup>. In light of this evidence, the relatively recent emphasis on MP as a possible ocular-protectant in diabetes<sup>(16)</sup>, a pathological condition similarly associated with inflammation, oxidative stress and progressive retinal damage, is logical and represents a natural extension of previous work in AMD<sup>(12-15)</sup>.

The underlying molecular mechanisms associated with the onset of diabetes and the potential protective effects of lutein and/or zeaxanthin against retinal oxidative damage, inflammation and visual function have been explored in diabetic murine

**Abbreviations:** AMD, age-related macular degeneration; CAT, catalase; CRP, C-reactive protein; DiVFuSS, Diabetes Visual Function Supplement Study; GPx, glutathione peroxidase; GSH, glutathione; MP, macular pigment; MPOD, macular pigment optical density; RCT, randomised controlled trial; RPE, retinal pigment epithelium; ROS, reactive oxygen species; SOD, superoxide oxidase; VEGF, vascular endothelial growth factor.

\* Corresponding author: Grainne Scanlon, fax +353 1 402 4915, email grainne.scanlon@dit.ie

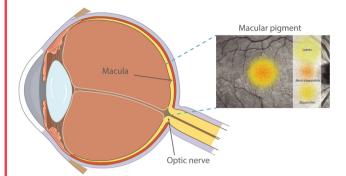


Fig. 1. Diagram highlighting the location of macular pigment and the macula within the eye. For a colour figure, see the online version of the paper.

models<sup>(17-23)</sup>. The onset of diabetes in alloxan and streptozotocin diabetic mice and rats was accompanied by an increase in markers of oxidative stress and inflammation including: malondialdehyde, 8-hydroxy-2'deoxyguanosine, NF-B and vascular endothelial growth factor (VEGF), with a concomitant decrease in local antioxidants including glutathione (GSH) and glutathione peroxidase (GPx)<sup>(17-19)</sup>. Importantly, treatment with antioxidants (lutein and/or zeaxanthin) has been shown to lower oxidative stress and inflammation and increase endogenous antioxidants; findings which in some<sup>(17,18)</sup> but not all<sup>(20)</sup> studies occurred independently of any effects on hyperglycaemia. The beneficial effects of these carotenoids on retinal function have also been observed in diabetic murine models, and these include the preservation of electro-retinogram b-wave amplitude and prevention of neurodegenerative effects on the inner retinal layers<sup>(17-19)</sup>. Findings from these studies provide important insights into the potential role that MP may have in protecting against diabetic retinal disease; however, the observed findings need to be interpreted with caution, as higher MP dosages have been used in animal studies compared with those used clinically in humans, and many of these studies have never conclusively demonstrated retinal uptake of the administered carotenoids beyond the retinal pigment epithelium (RPE) and choroid<sup>(24,25)</sup>. A summary of the experimental animal studies examining MP optical density (MPOD) in diabetes is outlined in Table 1.

A number of cross-sectional studies<sup>(16,26-29)</sup>, including findings from our own study group<sup>(28)</sup>, and randomised controlled trials (RCT)<sup>(30,31)</sup>, have explored the relationship between type 1 and type 2 diabetes and MPOD in humans. MPOD is generally found to be lower in diabetes<sup>(16,26,28)</sup>, with some studies suggesting that oxidative stress is implicated in its depletion<sup>(16,26)</sup>. Lower levels of MPOD have also been associated with raised glycated Hb (HbA1c) levels and the presence of retinopathy in patients with type 2 diabetes<sup>(16)</sup>. In another study, patients with diabetes (type 1 and 2) with grade 2 diabetic maculopathy had significantly lower MPOD compared with those with no maculopathy (P = 0.016)<sup>(26)</sup>.

Although oxidative stress is implicated as a causative factor, the exact reasons why MP is lower in diabetes, and type 2 diabetes in particular<sup>(16,28)</sup>, are not fully understood. One study found lower levels of MP in type 2 v. type 1 diabetes<sup>(28)</sup>, suggesting that other coincident metabolic and pathological abnormalities which are characteristic of type 2 diabetes (for example, overweight/obesity, dyslipidaemia) may explain the lower MPOD levels observed. Furthermore, lutein and zeaxanthin supplementation has been shown to exert positive and beneficial ocular effects in those with diabetic eye disease, including structural improvements in measures of macular oedema and functional improvements in visual acuity and other visual function measures<sup>(30,31)</sup>. Table 2 outlines a summary of the crosssectional studies and RCT examining the relationship between MPOD and diabetes (type 1 and 2).

Although the evidence exploring MP and diabetes is relatively sparse, the findings that have been reported are sufficient to suggest that MPOD may be adversely affected by the condition, and that there is a plausible rationale to explore the possible benefits of lutein and/or zeaxanthin supplementation for ocular health and visual function in diabetes, particularly type 2. The recent generation of such evidence has prompted this literature review, which is designed to explore and elucidate the metabolic perturbations and candidate causal mechanisms which may underpin the likely complex interdependency of MP and diabetic eye disease (references used in Tables 1 and 2<sup>(16–23,26–36)</sup>).

#### Literature search methods

For the purpose of the present review, partly because of the relative dearth of higher-level RCT evidence, we have appraised all forms of published research, even lower-level evidence sources, to ensure that the scientific and clinical implications of the available evidence might be synthesised into useful treatise, which accurately reflects what is known about this area, and what remains to be elucidated. The review was carried out in two stages. First, we identified all relevant published articles from human and animal studies which reported on the relationship between MP (lutein and/or zeaxanthin and/or meso-zeaxanthin) and diabetes (type 1 and type 2), up until the year 2019. The second part of the search involved identifying publications which investigated the relationship between the metabolic perturbations typically associated with diabetes, and type 2 diabetes in particular (for example, adiposity/dyslipidaemia) and MP. Preselected keywords including 'lutein', 'zeaxanthin', 'macular pigment', 'diabetes', 'diabetes AND MP', 'diabetes AND lutein/ zeaxanthin', 'BMI', 'body fat AND diabetes', 'adipose', 'high density lipoprotein AND diabetes', 'triglycerides AND diabetes', 'oxidative stress', 'inflammation', 'hypertension AND MP', 'insulin resistance' and 'hyperinsulinemia' were entered into academic databases and search engines including PubMed, Google Scholar, Mendeley, Scopus, Cochrane Library and the ISRCTN registry to define our search of the literature relating to MP and diabetes up until 2019. Although many supporting publications were retrieved, in total, only eight animal studies (Table 1) and eleven human studies (Table 2) were included to help clarify our current understanding of the links between diabetes and MP. A total of thirteen papers on adiposity and MP (Table 3) and seven papers on MP and dyslipidaemia (Table 4) were included to help analyse the relationship between MPOD and the metabolic correlates of type 2 diabetes (adiposity and dyslipidaemia). The overall findings suggest that MP is lower in diabetes, type 2 diabetes in particular,

Study	Design: animal model	Lutein and/or zeaxanthin	Outcome evidence and conclusion
Muriach <i>et al.</i> (2006) <sup>(18)</sup>	Albino mice (alloxan-induced	Lutein (70 % purity, 0.2 mg/kg body weight)	Decrease in oxidative stress (decrease MDA and NF-kB, increase GSH and GPx); FBG, h-wave restored
Kowluru <i>et al.</i> (2008) <sup>(19)</sup>	Rats (STZ-induced DM)	Zeaxanthin (0·02 or 0·1 % equivalent to 8·4 mg/kg). Placebo controlled	Decrease in work routing Decrease in oxidative stress (decrease 8-OHdG, nitrotyrosine, iNOS, VEGF and ICAM+1 levels)
Arnal <i>et al.</i> (2009) <sup>(23)</sup>	Rats (STZ-induced DM)	Lutein (in combination with DHA). Placebo controlled	Decrease in outstyle stress (MDA and nitrotyrosine, increase GSH and GPx). Restored ERG b-wave amplitude and latency time
Sasaki <i>et al.</i> (2010) <sup>(17)</sup>	Mice (STZ-induced DM)	Lutein (0.1 mg, w/w). Placebo controlled	Decrease in ROS and ERK activation and apoptosis. Protection of inner retina from visual impairment
Tang <i>et al.</i> (2011) <sup>(21)</sup>	Mice (db/db) spontaneous diabetes	<ol> <li>% (energy) Wolfberry (lutein and zeaxanthin). Placebo controlled</li> </ol>	Lower expression of endoplasmic reticulum stress biomarkers (BiP, PERK, ATF6, caspase-12) and restored AMPK. thioredoxin, Mn SOD, and FOXO3 $\alpha$ activities
Yu <i>et al.</i> (2013) <sup>(22)</sup>	Mice (db/db) spontaneous diabetes	Wolfberry diet (lutein and zeaxanthin). Placebo controlled	Activation of AMPK in mitochondria which caused retinoprotection in the retina of db/db mice
Kowluru <i>et al.</i> (2014) <sup>(20)</sup>	Rats (STZ-induced DM)	Multi-nutritional supplement (lutein 20 mg; zeaxanthin 40 mg/kg of powder diet, in addition to numerous antioxidants). Placebo controlled	Carotenoids ameliorated capillary cell apoptosis. Improved ERG, a- and b-wave amplitudes. Decreased oxidative damage and increased antioxidant activity. Inflammatory markers (decrease in VEGF. IL-18 and NF-kB)
Zhou <i>et al.</i> (2017) <sup>(32)</sup>	Rats (high-fat diet and low- dose STZ-induced DM)	Zeaxanthin (50 mg/kg; 200 mg dissolved in 10 ml maize oil). Placebo controlled	Supplementation with zeaxanthin reduced blood glucose, improved cognitive deficits, neural cell survival and increased p-AKT levels. Inhibited cleaved caspase-3 levels and NF-kB nuclear transcription in the hippocampus
MDA, malondialdehyde; GSH intercellular adhesion molec kinase; Mn SOD, Mn super	, glutathione; GPx, glutathione peroxida: cule; ROS, reactive oxygen species; ER roxide dismutase; FOXO3α forkhead O	DA, malondialdehyde; GSH, glurathione; GPx, glutathione peroxidase; ERG, electroretinogram; STZ- streptozotocin; 8-OHG, 8-hydroxy-2 <sup>-</sup> /d intercellular adhesion molecule; ROS, reactive oxygen species; ERK, extra-cellular receptor kinase; BIr, binding lg protein; PERK, protein k kinase; Mn SOD, Mn superoxide dismutase; FOXO3α forkhead O transcription factor 3 α; p-AKT; phosphorylated serine/threonine kinase.	MDA, malondialdehyde; GSH, glutathione; GPx, glutathione peroxidase; ERG, electroretinogram; STZ- streptozotocin; 8-OHdG, 8-hydroxy-2'deoxyguanosin; NOS, inducible NO synthase; VEGF, vascular endothelial growth factor; ICAM-1, intercellular adhesion molecule; ROS, reactive oxygen species; ERK, extra-cellular receptor kinase; BIP, binding Ig protein; PERK, protein kinase RNA-like ER kinase; ATF6, activating transcription factor 6; AMPK, AMP-activated protein kinase; Mn SOD, Mn superoxide dismutase; FOXO34 forkhead O transcription factor 3 4; p-AKT; phosphory/ated serine/threonine kinase.

Table 1. Summary of experimental animal studies examining the relationship between lutein and/or zeaxanthin and diabetes mellitus (DM)

and that supplementation with macular carotenoids and/or other antioxidants may confer protection against diabetic eve disease. To elucidate the causal mechanisms and metabolic perturbations which might possibly explain the lower MP levels observed in diabetes, we will first explore the condition diabetes mellitus itself. including its association with oxidative stress and inflammation; and subsequently present the evidence linking adiposity and dyslipidaemia with type 2 (or poorly controlled type 1) diabetes and MPOD.

#### **Diabetes mellitus**

Diabetes mellitus is a group of metabolic disorders caused by the complex interaction of genetics, environmental factors and lifestyle choices<sup>(37)</sup>. Diabetes is characterised by a deficiency of insulin and/or systemic insulin resistance. Over the past number of decades, the number of individuals with diabetes, particularly type 2 diabetes, has increased dramatically, making it a critical and universal public health challenge<sup>(38)</sup>. Type 1 diabetes usually develops in normal-weight children, teenagers and younger adults, and is an autoimmune condition involving the selective destruction of pancreatic  $\beta$ -cells, ultimately resulting in complete deficiency of insulin<sup>(39)</sup>. Conversely, type 2 diabetes is linked to a sedentary lifestyle and being overweight, and is characterised by systemic insulin resistance and subsequent pancreatic endocrine dysfunction, which results in attenuated insulin synthesis as well as inhibition of its cellular effects<sup>(37)</sup>. Whilst diabetes is characterised by having higher than normal blood glucose levels, the pathogenesis and development of type 1 and type 2 diabetes differ; therefore, the relationship between MPOD and the different forms of diabetes should not be generalised. At presentation, type 2 diabetes is most often accompanied by other comorbidities including overweight/obesity, insulin resistance, hypertension and dyslipidaemia, features which are less common in type 1 diabetes at diagnosis, but which may occur as complications of type 1 diabetes later in the course of the disease. Oxidative stress and inflammation are implicated in both type 1 and type 2 diabetes; however, research has shown that these metabolic disturbances are very pronounced in type 2 diabetes(40-44). The oxidative stress, inflammation, adiposity and dyslipidaemia which characterise diabetes may have independent relationships with MPOD, and therefore constitute plausible causal mechanisms in diabetic eye disease. These pathological mechanisms merit further exploration and will be discussed in more detail herein.

#### Oxidative stress and diabetes

Chronic hyperglycaemia induces oxidative stress in patients with diabetes<sup>(45)</sup>. Oxidative stress, defined as the excessive production of ROS, results in oxidative injury when the redox balance is upset; i.e. where the level of oxidative species exceeds the capacity of the antioxidant defence system to neutralise them<sup>(46,47)</sup>. ROS can include free radicals, which are partially reduced oxygen species containing one or more unpaired electrons (for example, superoxide anion or hydroxyl radical), and species with their full complement of electrons in an unstable or

Nutrition Research Reviews

4

Table 2. Summary of cro	Table 2. Summary of cross-sectional studies and randomised controlled trials (RCT) examining the relationship between macular pigment optical density (MPOD) and diabetes mellitus	ween macular pigment optical density (MPOD) and diabetes mellitus
Study	Design	Outcome, evidence and conclusion
Davies & Morland (2002) <sup>(26)</sup> Zagers <i>et al.</i> (2005) <sup>(27)</sup>	Cross-sectional study to assess MPOD in diabetes ( <i>n</i> 26) <i>v</i> . healthy, non-diabetic controls ( <i>n</i> 30) Cross-sectional study to assess MPOD in diabetes ( <i>n</i> 15) <i>v</i> . healthy non-diabetic subjacts ( <i>n</i> 14)	Those with maculopathy had lower levels of MP than those without maculopathy ( $P=0.016$ ) The density of macular pigment in diabetics was not different from that in controls ( $P=0.3$ )
Mares <i>et al.</i> (2006) <sup>(29)</sup> Lima <i>et al.</i> (2010) <sup>(16)</sup>	Cross-sectional study on subsample of women who participated in CAREDS ( <i>n</i> 1698). MPOD measured by HFP Cross-sectional study including: non-diabetic controls ( <i>n</i> 14), diabetics with ( <i>n</i> 12) and	Lower MOD was associated with diabetes. Relationship of dietary intake of lutein Lower MOD was associated between diabetic and non-diabetic women and zeaxanthin to MP differed between diabetic and non-diabetic women Type 2 diabetic patients, with or without retinopathy, had reduced MPOD v.
Scanlon <i>et al.</i> (2015) <sup>(28)</sup>	without retinopathy ( <i>n</i> 17) Cross-sectional study assessing MP in type 1 ( <i>n</i> 34) and 2 ( <i>n</i> 68) diabetics and	MPOD levels were lower in diabetics v. controls (P=0.04). MPOD was lower in type 2 v.
She <i>et al.</i> (2016) <sup>(33)</sup>	resulty controls (1 4-b) Cross-sectional study assessing MP in diabetic patients without retinopathy ( <i>n</i> 134), with early-stage non-proliferative retinopathy ( <i>n</i> 48) and non-diabetic controls ( <i>n</i> 210)	WPOT I (r=0.02) and type z v. controls (r=0.05) MPOD levels were not significantly different between the three groups (i.e. diabetics with or without retinopathy and controls; P=0.24). MPOD was positively associated with central forward threas (P=0.001)
Moschos <i>et al.</i> (2017) <sup>(34)</sup>	Retrospective study on type 2 diabetic patients ( <i>n</i> 60; 120 eyes) without DR. Patients received carotenoid supplements containing lutein (10 mg), zeaxanthin (2 mg) and <i>meso</i> -zeaxanthin (10 mg) once per d for 2 years	MPOD register to contract measured in this study. OCT showed an increase in the central foveal thickness ( $P$ <0.001) and mfERG revealed increased retinal response density within the central 13° surrounding the fovea (rings 1 to 3) ( $P$ <0.001) at 2 years after the onset of carninements.
Gonzalez-Herrero <i>et al.</i> (2018) <sup>(35)</sup>	Prospective controlled study assessing macular integrity and macular sensitivity by microperimetry in patients with non-proliferative DR. Patients were assigned (1:1) to the DHA + xanthophyll supplementation group ( $n$ 12; 24 eyes) or the control group ( $n$ 12: 24 eyes) or the control group ( $n$ 12: 24 eyes) or the control group	Significant improvements in macular markets are 90 d ( $P=0.030$ ) in the supplemented group. Total antioxidant capacity levels increased and plasma IL-6 levels decreased significantly in the DHA + xanthophyll-supplemented group but not in the control and more not more not in the control and the mace index of the black in the supplemented group but not in the control and plasma in
Cennamo <i>et al.</i> (2019) <sup>(36)</sup>	Prospective controlled study assessing MPOD and microvascular density on OCTA in a cohort of type 1 diabetes patients with retinopathy ( <i>n</i> 59, total of 82 eyes	MPOD was significantly lower in patients with retinopathy compared with controls $(P<0.001)$ . OCTA vessel density was significantly lower in patients than in controls $(P<0.001)$ .
Hu <i>et al.</i> (2011) <sup>(30)</sup>	emotion, control group (17 +0, total of oc eyes examined) RCT: controlled trial including non-diabetic retinopathy patients ( <i>n</i> 61) and non- diabetic controls ( <i>n</i> 60)	In controls (r < 0.001) Serum lutein/zeaxanthin concentrations in DR patients were lower than those in normal controls. Lutein/zeaxanthin intake can improve VA, CS and macular oedema in DR patients
Chous <i>et al.</i> (2015) <sup>(31)</sup> DiVFuSS	RCT: type 1 ( <i>n</i> 27) and type 2 ( <i>n</i> 40) diabetes with no DR or mild to moderate NPDR <i>v</i> . placebo. Intervention group received two daily doses of a xanthophyll, antioxidant and plant extract supplement	Significant improvement in MPOD ( $P=0.008$ to <0.0001); visual function on all measures ( $P=0.008$ to <0.0001); serum lipids ( $P$ values ranging from 0.01 to 0.0004), hs-CRP ( $P=0.01$ ) and DPN (Fisher's exact test; $P=0.0024$ ). These effects were observed without an effect on glycaemic control

Downloaded from https://www.cambridge.org/core. Dublin Institute of Technology, on 14 Aug 2019 at 08:15:21, subject to the Cambridge Core terms of use, available at https://www.cambridge.org/core/terms. https://doi.org/10.1017/S095442241900012X

MP, macular pigment; CAREDS, Carotenoids in Age-Related Eye Disease Study; HFP heterochromatic flicker photometry; HbA1c, glycated Hb; DR, diabetic retinopathy; OCT, optical coherence tomography; multifocal electroretinography; OCTA, optical coherence tomography; NA, visual acuity; CS, contrast sensitivity; DiVFuSS, Diabetes Visual Function Supplement Study; NPDR, non-proliferative diabetic retinopathy; hs-CRP, high-sensitivity C-reactive protein; DPN, diabetic peripheral neuropathy.

Nutrition Research Reviews

Table 3. Summary of studies linking macular pigment optical density (MPOD) and body fat

Table 3. Summary of studies	Table 3. Summary of studies linking macular pigment optical density (MPOD) and body fat		
Study	Design	Sample	Results
Johnson <i>et al.</i> (2000) <sup>(98)</sup>	Study 1: 15-week modified diet to increase lutein and zeaxanthin intake with serum, buccal mucosal cell and adipose samples taken at 0, 4, 8 and 15 weeks and 2 months post-intervention. Study 2: a cross-sectional analysis of relationships	Study 1: 7 subjects (4 women and 3 men); study 2: 21 subjects (13 women and 8 men)	<ol> <li>Only serum zeaxanthin increased from baseline and only at week 4. Adipose tissue lutein peaked at week 8.</li> <li>Significant negative correlations found between adipose tissue, lutein concentrations and MP for women. A positive effect was seen in men</li> </ol>
Broekmans <i>et al.</i> (2002) <sup>(124)</sup>	Cross-sectional design looking at serum and adipose tissue concentrations in relation to MP	376 Healthy adults. Serum ( <i>n</i> 376); adipose ( <i>n</i> 187)	Regression models showed a positive significant association between MP density and serum lutein, serum zeaxanthin, and adipose lutein concentrations in men after adjustment for age, but no relation in women. This effect remained in men after further correction
Hammond <i>et al.</i> (2002) <sup>(99)</sup>	Cross-sectional study investigating MPOD, BMI, body fat percentage, dietary intake and serum carotenoid concentrations	680 Healthy adults: body fat ( <i>n</i> 400); dietary ( <i>n</i> 280); serum ( <i>n</i> 280)	Inverse association between MPOD and BMI ( $P$ <0.0008) and body fat ( $P$ <0.01)
Nolan <i>et al.</i> (2004) <sup>(101)</sup>	Cross-sectional study investigating MPOD, body fat	100 Healthy adults: 45 males and 55 females	Significant inverse relationship between MPOD and body fat
Mares <i>et al.</i> (2006) <sup>(29)</sup>	processingly, detaily carboning many and servin reveal Cross-sectional study investigating MPOD, dietary intake of lutein and zeaxanthin and other predictors of MP levels	Substances Substample of women who participated in CAREDS. 1698 Women aced 53–86 vears	MPOD is directly related to dietary initiate and serum levels of lutein and zeaxanthin. Higher abdominal fat is related to lower MPOD
Moeller <i>et al.</i> (2009) <sup>(125)</sup>	Intervention in a subsample of the Women's Health Initiative to increase lutein and zeaxanthin through dietary modification	394 Healthy adults; intervention (n 1581); control/usual diet (n 236)	Lower MPOD was associated with above-median waist circumference. This explained 2.7 % of the variation
Kirby <i>et al.</i> (2011) <sup>(113)</sup>	Randomised controlled weight loss trial	104 Subjects with BMI >28 kg/m <sup>2</sup> ; Intervention group ( <i>n</i> 54); control group ( <i>n</i> 50)	Significant weight loss in the intervention group ( $P$ <0.0001) was related to serum increases in lutein
Gupta <i>et al.</i> (2012) <sup>(120)</sup>	Clinical study; MPOD was compared with various anthropometric measurements including waist:height ratio, waist circumference, waist:hip ratio and BMI	161 Healthy adults; 82 males and 79 females	Mean MPOD values did not differ significantly in various types of obesity when compared with normal subjects
Nolan <i>et al.</i> (2012) <sup>(115)</sup>	Prospective cohort study	Subset of TILDA participants with MPOD and BMI measures; 4281 adults aged over 50 vears	Obese individuals had significantly lower MPOD (mean 0.190) than either overweight individuals (mean 0.208, <i>P</i> <0.01) or those with normal weicht (mean 0.213; <i>P</i> <0.01)
Bovier <i>et al.</i> (2013) <sup>(116)</sup>	Cross-sectional study investigating MPOD, assessment of body composition and serum lutein/zeaxanthin status	100 Healthy adults; 39 males and 61 females	Significant inverse relationship between body fat percentage (total and recional) and MPOD ( $P<0.01$ )
Scanlon <i>et al.</i> (2015) <sup>(28)</sup>	Cross-sectional study to assess MPOD in type 1 and type 2 diabetics and BMI, dietary intake and serum carotenoid concentrations	102 Diabetics, of which type 1 ( <i>n</i> 34) and type 2 ( <i>n</i> 68), and 48 healthy controls	BMI was significantly higher and MPOD lower in type 2 v. type 1 diabetics and healthy controls. BMI was not directly correlated with MPOD
Khan <i>et al.</i> (2018) <sup>(126)</sup>	Cross-sectional study investigating MPOD, whole body adiposity (% fat) and intellectual ability	114 Adults with overweight and obesity (≥25 kg/m²)	MPOD was inversely related to % fat ( $P=0.04$ ) and positively associated with IQ ( $P=0.01$ ) and fluid intelligence ( $P=0.01$ )
Edwards <i>et al.</i> (2019) <sup>(127)</sup>	Cross-sectional study investigating MPOD, body fat % (DXA) and event-related brain potentials	101 Adults with overweight and obesity (≥25 kg/m²)	MPOD was not significantly associated with body fat % in this study ( $P$ >0.07). MPOD was associated with mean and peak amplitude (N2) ( $P$ <0.04) and latency (P3) ( $P$ <0.01) of neuroelectric indices

MP, macular pigment; CAREDS, Carotenoids in Age-Related Eye Disease Study; TILDA, The Irish Longitudinal Study of Ageing; IQ, intelligence quotient; DXA, dual-energy X-ray absorptiometry.

Mares at al (2006)(29) C	Design	Sample	Results
	Cross-sectional study on women. MPOD measured by HFP	Subsample of women who participated in the CAREDS ( <i>n</i> 1698)	Serum lutein and zeaxanthin correlated with serum cholesterol (r 0.17) but adjustment for this did not affect the direct association
Lima <i>et al.</i> (2010) <sup>(16)</sup> Ci	Cross-sectional study with three test groups: non-diabetic controls, diabetics without retinopathy and diabetics with	43 Type 2 diabetics with and without retinopathy matched to controls	between serum levels and MPOD No significant associations were observed with lipid levels and MP (P=0.08)
Loane <i>et al.</i> (2010) <sup>(139)</sup> Ci	Cross-sectional study on healthy men and women, without ocular disease, aged 20–70 years	302 Subjects: 181 with negative family history of AMD; 121 with positive family history of AMD	A significant and positive association between serum HDL and serum concentrations of lutein. A significant inverse association between serum TAG concentration and both serum lutein concentration and MDD is handow of AMD.
Renzi <i>et al.</i> (2012) <sup>(140)</sup> Oi	Cross-sectional study on young healthy subjects. MPOD measured by HFP	Relationships between serum lutein and zeaxanthin, MPOD and lipoprotein levels ( <i>n</i> 108)	HDL was significantly ( $P$ <0.05) related to MPOD ( $r$ 0.33), to serum lutein ( $r$ 0.36) and to serum zeaxanthin ( $r$ 0.26). MPOD was also significantly related to total cholesterol r( $r$ 0.19)
Olmedilla-Alonso <i>et al.</i> (2014) <sup>(138)</sup>	Cross-sectional study assessing serum concentrations and intake of lutein and zeaxanthin and MPOD using HFP	Healthy men and women aged 20–35 and 45–65 years ( <i>n</i> 108)	MP only correlated with serum lutein and zeaxanthin when expressed in terms of cholesterol + TAG ( $P$ =0.012) or LDL-C ( $P$ =0.017). In the older group relationships were also seen with LDL-C ( $P$ =0.002)
Nagai <i>et al.</i> (2015) <sup>(141)</sup> Gi	Cross-sectional study assessing serum concentrations of carotenoids and lipoproteins. MPOD was measured using HEP	55 Healthy volunteers (110 eyes) aged 23–53 years	MPOD was positively correlated with serum concentrations of lutein (P=0.00001), zeaxanthin (P=0.0005) and dietary lutein intake (P=0.002). Serum oxidised LDL was inversely correlated with MPOD (P=0.006) after adiustion for and sex and servin lutein
Chous <i>et al.</i> (2015) <sup>(31)</sup> Di	DiVFUSS RCT: type 1 and type 2 diabetes with no DR or mild to moderate NPDR v. placebo. Intervention group received two daily doses of a xanthophyll, antioxidant and plant extract supplement	Total group ( $n$ 67): type 1 ( $n$ 27); type 2 ( $n$ 40). Intervention DiVFuSS supplement ( $n$ 39): type 1 ( $n$ 16); type 2 ( $n$ 23); placebo ( $n$ 28): type 1 ( $n$ 11); type 2 ( $n$ 17)	Significant improvement in MPOD ( $= 0.003$ to <0.0001) and visual function on all measures ( $P = 0.008$ to <0.0001) and visual function on all measures ( $P = 0.008$ to <0.0001); mean LDL, HDL and TAG levels within the intervention group at 6 months were $-9$ , $+7$ , $-8.6$ % respectively v. $+1$ , $-3$ , $-2$ %, respectively in the placebo group

Downloaded from https://www.cambridge.org/core. Dublin Institute of Technology, on 14 Aug 2019 at 08:15:21, subject to the Cambridge Core terms of use, available at https://www.cambridge.org/core/terms. https://doi.org/10.1017/S095442241900012X

Nutrition Research Reviews

reactive state (for example, singlet oxygen or  $H_2O_2$ )<sup>(48)</sup>. The body's natural defence against oxidative damage is neutralisation by endogenous antioxidants<sup>(49)</sup>, which include enzymic antioxidants such as superoxide oxidase (SOD), catalase (CAT) and GPx, and non-enzymic antioxidants such as GSH. Our endogenous antioxidant defence system, however, is incomplete without exogenous antioxidant nutrients including vitamin C, vitamin E, carotenoids (β-carotene, lutein, zeaxanthin and meso-zeaxanthin) and polyphenols<sup>(50)</sup>. Exogenous antioxidants, such as those available in dietary fruit and vegetables, are therefore necessary to balance redox status. Through normal physiological processes, antioxidants inhibit or quench free radical reactions and can delay or inhibit cellular damage<sup>(46)</sup>. When these processes are overwhelmed, however, as in type 2 diabetes, ROS will readily react with lipids, proteins and nucleic acids, resulting in impaired cell function or cell death<sup>(51,52)</sup>.

The retina is already at an increased risk of oxidative stress and damage, given its high oxygen demand and consumption, its exposure to visible light irradiation and its high concentration of PUFA in the photoreceptor outer segments<sup>(53)</sup>. These risks increase when the generation of free radicals and ROS is stimulated by environmental factors<sup>(54,55)</sup>, short wavelength light exposure<sup>(56)</sup>, and other internal stresses including inflammation<sup>(57)</sup> and hyperglycaemia<sup>(52,58,59)</sup>. While PUFA in the photoreceptor outer segment are thought to protect against oxidative and inflammatory damage, they can potentially act as a substrate for the propagation of free radicals as they provide a readily available source of electrons<sup>(9,54)</sup>. With increasing age, systemic antioxidant levels decline and ROS levels increase in most tissues, and this is associated with a number of neurodegenerative diseases, including AMD and diabetes<sup>(9,26,60)</sup>.

Hyperglycaemia, oxidative stress and changes in redox homeostasis are fundamental events in the pathogenesis of diabetic retinopathy. Various mechanisms have been suggested to contribute to the formation of reactive oxygen-free radicals. Hyperglycaemia causes tissue damage through five major pathways, namely: increased flux of glucose and other sugars through the polyol pathway; increased intracellular formation of advanced glycation endproducts (AGE); increased expression of the receptor for AGE (RAGE) and its activating ligands; activation of protein kinase-C isoforms; and activity of the hexosamine pathway<sup>(61)</sup>. Over-activation of these pathways can lead to damage of cellular organelles and enzymes, increased lipid peroxidation and the development of insulin resistance<sup>(62,63)</sup>. Several lines of evidence indicate that all five mechanisms are activated by a single upstream event: mitochondrial overproduction of ROS<sup>(64)</sup>. Locally, the occurrence of these metabolic changes results in defective angiogenesis and the activation of a number of pro-inflammatory pathways which stimulate an influx of leucocytes which can alter vascular permeability in the retina $^{(21)}$ . The role of hyperglycaemia in diabetic microvascular complications in the eye has been established by large-scale prospective studies in type 2 diabetes (United Kingdom Prospective Diabetes Study)(65). Beyond elevations in blood glucose, however, ROS formation is also increased by elevations in NEFA (a common feature of type 2 diabetes), through their direct effect on mitochondria(66).

While oxidative stress is increased in diabetes, the activities of antioxidant defence enzymes responsible for scavenging free radicals and maintaining redox homeostasis (SOD, CAT, GPx) are concomitantly diminished in the retina of both type 1 and type 2 diabetes<sup>(47,49,67)</sup>. The activity of non-enzymic antioxidants is also depressed during hyperglycaemia-induced oxidative stress<sup>(67)</sup>. Redox balance is important, as its disruption can cause antioxidants to become pro-oxidative, potentially precipitating damage to the retina. It cannot be assumed, however, that a single reductive or antioxidant treatment would be sufficient to reverse the underlying pathophysiological mechanisms of diabetic retinopathy. Antioxidant protection in the retina is provided mainly from vitamins C and E, carotenoids, GPx, SOD and CAT enzymes, and GSH compound<sup>(68)</sup>, and many of these antioxidants are aided by micronutrients (for example, Cu, Zn, Se) which act as cofactors<sup>(69)</sup>. The carotenoids lutein and zeaxanthin are very effective quenchers of singlet molecular oxygen and lipid peroxy radicals; however, in the process, these carotenoids are themselves oxidised to their corresponding radical cations<sup>(70)</sup>. These cations must be reduced to regenerate the original carotenoid, thereby allowing their reuse as an antioxidant. Vitamin E ( $\alpha$ -tocopherol) reduces oxidised carotenoids, which leaves tocopherol oxidised<sup>(69)</sup>. However, oxidised tocopherol can be reduced and regenerated by vitamin C (ascorbic acid) and GSH, which are themselves further reduced by Cu and Zn<sup>(15,71)</sup>. Dietary lutein, zeaxanthin and vitamins E and C, as well as endogenous antioxidant enzymes, therefore act in concert with one another to lower the levels of ROS (via ROS detoxification) and to recycle oxidised lipid antioxidants (via re-reduction).

The presence of oxidation products of the macular carotenoids in human and primate retinal tissue indicates that lutein, zeaxanthin and meso-zeaxanthin play an important antioxidant role there<sup>(72)</sup>. Zeaxanthin appears to be a more potent antioxidant than lutein<sup>(73)</sup>, with meso-zeaxanthin even more efficacious, but only in conjunction with its binding protein<sup>(74)</sup>. Li *et al.*<sup>(75)</sup> demonstrated that a 1:1:1 ratio mixture of lutein, zeaxanthin and meso-zeaxanthin quenches more singlet oxygen than any of these carotenoids individually at the same total concentration<sup>(75,76)</sup>, suggesting that they are most effective when acting synergistically with one another. Diet is a major lifestyle factor which can greatly influence the incidence and progression of type 2 diabetes. Some dietary and lifestyle modifications associated with enhanced antioxidant supply could therefore be an effective prophylactic means to prevent or limit oxidative stress in diabetic eye disease. Type 2 diabetes is associated with a sedentary lifestyle and being overweight; therefore, patients with type 2 diabetes may not only be exposed to diets which are high in fat and low in antioxidants<sup>(77)</sup>, and to the increased oxidative stress imposed by their diabetes<sup>(16,26,28)</sup>, but may also be subject to greater chronic oxidative stress as a result of their obesity itself<sup>(78)</sup>. The co-occurrence of these phenomena consequently demands an even greater supply of exogenous antioxidants (vitamins C and E, carotenoids lutein, zeaxanthin and meso-zeaxanthin) to balance redox status and to combat this state of chronic oxidative stress to which the diabetic retina is exposed. While the control of blood glucose is critical in patients with diabetes, neuroprotective strategies and novel antioxidant treatments should also be

considered to target the specific aetiologies and underlying pathophysiological mechanisms of the condition, and to ultimately protect against the vascular pathologies of diabetes. It is logical that natural antioxidants (lutein, zeaxanthin and meso-zeaxanthin) may have beneficial implications for diabetic retinopathy in this context. However, larger trials involving intensive assessment of oxidative stress, antioxidant defence activities and their effects on one another in the diabetic state are necessary before empirical supplementation of these carotenoids can be recommended clinically.

#### Inflammation and diabetes

Diabetic retinopathy has traditionally been considered a disease of the retinal microvasculature, associated with oxidative stress induced by hyperglycaemia. More recently, however, inflammation has been cited as a deleterious factor in the development of diabetic complications<sup>(79)</sup>, and is now also implicated in the pathogenesis of many ocular diseases including AMD<sup>(80)</sup> and diabetic retinopathy<sup>(81)</sup>. Numerous interplays exist between inflammation and oxidative stress and vice versa<sup>(82)</sup>. Hyperglycaemia increases the levels of pro-inflammatory proteins<sup>(79)</sup>, and it is now believed that inflammatory processes underlie many of the functional changes in retinal vasculature observed histologically in early diabetic retinopathy, such as pericyte loss, microaneurysms, and occluded and degenerated capillaries(83).

While inflammation is normally a protective response to tissue stress, hyperglycaemia leads to a dysregulation of inflammation, which ultimately becomes chronic. Oxidative stress triggers inflammatory responses, and inflammation also enhances the production of ROS, which creates a self-perpetuating cascade towards oxidative tissue damage<sup>(84)</sup>. The increased expression of many inflammatory proteins in diabetic retinopathy is regulated through the activation of pro-inflammatory transcription factors including NF- $\kappa$ B<sup>(85)</sup>. Activation of NF- $\kappa$ B leads to the synthesis of pro-inflammatory molecules such as IL-1, IL-6, TNF- $\alpha$ , c-Jun N-terminal kinase-1 (JNK-1), Iĸ-B kinase-β (IKK-β) and growth factors such as VEGF<sup>(21,85)</sup>.

In the retina, the microvascular endothelium forms an effective barrier to control the movement of blood, fluid and proteins across the vessel wall. This barrier is formed by tight junctions, which consist of over forty different proteins and various inflammatory mediators including VEGF, TNF- $\alpha$ , protein kinase-C, IL-1 $\beta$  and IL-6<sup>(83,86)</sup>. Dysfunction of these proteins is highly correlated with the pathogenesis of blood retinal barrier breakdown and increased vascular permeability, which leads to diabetic macular oedema, a major cause of vision loss in diabetic retinopathy<sup>(87)</sup>. Inflammatory processes can induce oxidative stress, and similarly, oxidative stress can induce inflammation through the activation of multiple pathways. The coincident over-activation of inflammation and oxidative stress can therefore deplete cellular antioxidant capacity and reduce MP levels in the eye<sup>(9,88)</sup>.

The expression of various pro-inflammatory cytokines, such as TNF- $\alpha$ , IL-1 and IL-6, is also increased from visceral adipose tissue (more common in type 2 diabetes), and this has been linked to systemic inflammation and accompanying insulin resistance<sup>(89)</sup>. The inhibition of signalling downstream of the insulin receptor is a primary mechanism through which inflammatory signalling leads to insulin resistance<sup>(79)</sup>. Several cross-sectional studies have shown that insulin resistance and type 2 diabetes are associated with higher levels of C-reactive protein (CRP), IL-6 and TNF- $\alpha$ , all markers of subclinical inflammation<sup>(90-92)</sup>. Various longitudinal studies have also shown that elevated levels of CRP and IL-6 are linked to the later development of type 2 diabetes(93,94).

Inhibition of different inflammatory mediators, including NF-κB, IL-1β, VEGF and cyclo-oxygenase-2 (COX-2), has been shown to limit the degeneration of retinal capillaries characteristic of early-stage diabetic retinopathy<sup>(95)</sup>. There is evidence to suggest that lutein and zeaxanthin may prevent the development of diabetic retinopathy by suppressing ROS induced by inflammation<sup>(14)</sup>. Li et al.<sup>(11)</sup> investigated the anti-inflammatory effects of lutein on Muller cells in a murine model. Treatment with lutein led to lower production of the pro-inflammatory factors NF- $\kappa$ B, IL-1 $\beta$  and COX-2, suggesting an anti-inflammatory role for lutein in protection against retinal ischaemic/hypoxic injury<sup>(11)</sup>. In another study, the protective effects of lutein and zeaxanthin against photo-oxidative damage to RPE cells were investigated. Supplementation with carotenoids modulated the inflammatory responses in these cultured RPE cells in response to photo-oxidation<sup>(96)</sup>.

The effects of zeaxanthin on diabetes-induced retinal oxidative damage and inflammation have also been investigated in diabetic rats<sup>(19)</sup>. It was found that zeaxanthin significantly inhibits diabetes-induced retinal oxidative damage and elevations in VEGF and its adhesion molecule, abnormalities which are commonly associated with the pathogenesis of diabetic retinopathy<sup>(19)</sup>. Clinical findings have highlighted the inverse association between lutein and IL-6 in coronary artery disease patients<sup>(97)</sup>. A recent novel RCT, the Diabetes Visual Function Supplement Study (DiVFuSS)<sup>(31)</sup>, explored the effects of a nutritional supplement (which included lutein and zeaxanthin) on patients with type 1 and type 2 diabetes, and suggested that the preparation used mitigated the damaging effects of systemic inflammation on ocular function, and that these effects may have been mediated by enhancements in MPOD. Chronic low-grade inflammation and oxidative stress co-exist in diabetes, particularly type 2 diabetes but also poorly controlled type 1 diabetes. Therefore, findings from DiVFuSS, which achieved significant increases in MPOD (27 % mean increase in DiVFuSS group v. 2 % mean decrease in placebo group), as well as reductions in serum high-sensitivity CRP (60 % mean decrease in DiVFuSS group v. 11 % mean decrease in placebo group), may be due in part to the inclusion of compounds which, in combination, specifically target both inflammation and oxidative stress, and therefore, which block some of the oxidative and/or inflammatory pathways responsible for the progression of diabetic retinopathy<sup>(31)</sup>. The inclusion of many compounds in this test formula and the fact that both types of diabetic patients (type 1 and 2) were examined renders interpretation of the outcome more difficult; however, these findings are promising as they clearly show that MPOD can be augmented in patients with type 2 diabetes<sup>(31)</sup>.

While it is clear that oxidative stress and inflammation are implicated in the initiation and development of diabetic retinopathy, it remains unclear whether the pathological effects of this oxidative stress/inflammation are mediated through MPOD depletion in type 2 diabetes. The current body of evidence which directly links lower MP levels with other metabolic correlates of diabetes, namely adiposity and dyslipidaemia, will now be explored to elucidate the possible mechanisms by which these metabolic factors mediate their effects on MPOD. It is important to note that both increased adiposity and dyslipidaemia (primarily reduced HDL) may adversely affect MP by compromising the availability<sup>(98-101)</sup>, and transport<sup>(102,103)</sup>, of dietary carotenoids to the retina. It follows that a condition such as type 2 diabetes, which unfavourably affects these parameters, may have deleterious effects on MPOD, with clinically relevant outcome effects. An analysis of the literature is presented describing these putative mechanisms: (a) evidence in relation to MPOD and body fat/adiposity (n 13; Table 3); and (b) evidence concerning MPOD and dyslipidaemia (n 7; Table 4).

### Type 2 diabetes, macular pigment optical density and adiposity: the evidence

Diabetes mellitus is a chronic disorder which can alter carbohydrate, protein and fat metabolism. Weight gain, following excessive energy intake, results in the body becoming markedly resistant to the action of insulin. Despite initial impairment in insulin action, glucose tolerance can remain normal for some time because there is a compensatory increase in insulin secretion by pancreatic  $\beta$ -cells, which results in a well-compensated metabolic state, a condition known as hyperinsulinaemia. There is now mounting evidence to indicate that persistently elevated plasma insulin levels can contribute to the development of hypertension, plasma lipid abnormalities and atherosclerosis<sup>(104)</sup>. The escalating rates of insulin secretion as a result of advancing obesity, however, cannot be maintained and type 2 diabetes ensues.

Intra-abdominal fat appears to be an important determinant of insulin sensitivity compared with subcutaneous fat<sup>(105)</sup>. Visceral fat and subcutaneous fat differ in their phenotypic, physiological and functional aspects, as there are a greater number of macrophages, T lymphocytes and pro-inflammatory molecules in visceral v. subcutaneous fat in obese individuals. Abdominal adipose tissue produces a variety of adipocytokines including IL-1, IL-6, IL-8, TNF- $\alpha$ , leptin and resistin; therefore, the abdominal adipose mass acts as an important mediator of inflammation in diabetes<sup>(106)</sup>. Obesity also leads to an increased size of adipocytes. The death of adipocytes, which is rare in healthy individuals, is very common in obese individuals and has been linked to adipocyte hypoxia. The accumulation of macrophages in the adipose tissue of obese individuals not only eliminates the dead cells but also further increases their synthesis of inflammatory mediators IL-8, IL-6, IL-1 and TNF- $\alpha^{(106)}$ . The resultant inflammation becomes chronic; therefore, obesity is considered a low-grade inflammatory condition which in turn leads to an increase in oxidative stress. Serum 8-hydroxy guanosine (8-OhdG), a known sensitive marker of oxidative DNA damage and of total systemic oxidative stress *in vivo*, has been shown to be positively correlated with BMI in individuals with type 2 diabetes mellitus<sup>(107)</sup>. Over-expression of oxidative stress, and the concomitant reduction in antioxidant defence, damages cellular structures<sup>(108)</sup>, often resulting in further inflammatory responses. Antioxidant defences may be lower in overweight/type 2 diabetic patients, due to their lower intake of antioxidant-rich foods (for example, fruits and vegetables), their increased utilisation of these molecules (for example, increased inflammation/ROS in insulin-dependent tissue such as the retina) and their impaired generation of other supportive antioxidants<sup>(108)</sup>, factors which may collectively lead to MPOD depletion.

The diminished response to insulin in type 2 diabetes also leads to increased lipolysis in adipocytes. Enlarged adipocytes release NEFA and adipocytokines(67,104); therefore greater amounts of NEFA reach the liver via the portal vein, resulting in fatty liver infiltration. Mobilised NEFA are oxidised by vascular endothelium to generate ROS. Elevations of NEFA in serum also inhibit insulin suppression of hepatic glucose production, leading to an increase in glucose production by the liver<sup>(109)</sup>. This, in turn, results in both insulin resistance and inflammation in the major insulin target tissues (skeletal muscle, liver and endothelial cells). Abnormal levels of lipids, fatty acids and various adipocytokines from adipose tissue therefore initiate a vicious cycle of fat damage, inflammation, worsening insulin resistance, impaired  $\beta$ -cell insulin secretion and ultimately type 2 diabetes<sup>(104,110)</sup>. The chronic low-grade inflammation associated with obesity and type 2 diabetes leads to increased oxidative stress and further inflammation, putting a greater demand on antioxidant defences.

Apart from its pro-inflammatory and pro-oxidant effects, adipose tissue is also a major body store for carotenoids such as lutein and zeaxanthin. Higher levels of body fat have been shown to be related to lower levels of circulating carotenoids<sup>(111)</sup>, making these pigments less available to retinal tissue<sup>(112)</sup>. Furthermore, a randomised controlled weight loss trial found that a significant weight loss in the intervention group was related to increases in serum lutein<sup>(113)</sup>. Higher BMI<sup>(114)</sup> and higher body fat percentage<sup>(101)</sup> have both been linked with an increased risk of AMD. Studies have reported an inverse relationship between adiposity and BMI and MPOD<sup>(99,115)</sup>, with the observed relationships largely attributable to participants with a BMI > 29 kg/m<sup>2(99)</sup>. One study found that higher body fat percentage, even within relatively healthy limits, was associated with lower retinal tissue lutein and zeaxanthin status<sup>(116)</sup>, suggesting that adiposity may affect the nutrient status of the retina. Sex differences in MPOD have also been reported<sup>(98,101)</sup>, which may imply differences in metabolism of MP, or reflect differences in adipose tissue storage of these dietary carotenoids between men and women. Competitive absorption by a larger overall fat mass (percentage fat mass is generally higher in women)<sup>(117)</sup>, or specifically by greater abdominal fat mass (which is more common in men)<sup>(118)</sup>, may reduce lutein and zeaxanthin uptake by the retina. Concentrations of carotenoids differ according to body site, with levels demonstrably higher in abdominal fat<sup>(119)</sup>, which may explain the differences in MPOD currently observed between men and women<sup>(98,101)</sup>. Since adipose tissue may trap lutein and zeaxanthin<sup>(116)</sup>, individuals who are overweight or

obese (type 2 diabetes) may consequently have lower concentrations of the carotenoids lutein and zeaxanthin available at the macula.

Overall, the research appears to suggest that obesity has a negative impact on MPOD; however, there is also some evidence to the contrary. One study found that mean MPOD values did not differ significantly between subjects with various types of obesity and ideal-weight subjects<sup>(120)</sup>. Our previous study<sup>(28)</sup> is the only one to date which has examined the relationship between BMI and MPOD in a group with diabetes. This study found that MPOD was significantly lower and BMI was significantly higher in patients with type 2 diabetes, compared with type 1 and non-diabetic controls, highlighting the possibility that different relationships exist between MPOD status and type 1 and 2 diabetes. The higher BMI levels observed in type 2 diabetes are reflective of excess adiposity which acts as a store for lutein and zeaxanthin. However, as described previously, obesity is also independently associated with increased oxidative stress<sup>(108,121,122)</sup>, and with inflammation<sup>(123)</sup> which exacerbates oxidative stress, and may therefore have multiple depletive effects on MP. Unfortunately, the cross-sectional nature of this study<sup>(28)</sup> means that it cannot elucidate the extent or nature of any causal relationships between BMI and MPOD in these type 2 subjects.

The different presenting features of type 1 and 2 diabetes, and the interplay of their disease components with MPOD status (for example, adiposity), may mean that the mechanistic relationship between these conditions and MPOD status is not uniform across both. The fact that much of the research to date includes both diabetic groups may obfuscate any relationships present. Although the evidence to date is somewhat conflicting, there appears to be a theoretical basis for a relationship between the characteristic body fatness of type 2 diabetes and lower levels of MPOD. A summary of the studies investigating the association between MPOD and adiposity are outlined in Table 3<sup>(28,29,98,99,101,113,115,116,120,124–127)</sup>.

#### Type 2 diabetes, macular pigment optical density and dyslipidaemia: the evidence

Insulin resistance and obesity, metabolic abnormalities commonly associated with type 2 diabetes, may themselves lead to disturbances in the production and clearance of plasma lipoproteins<sup>(128)</sup>. Dyslipidaemia is often characterised by low levels of HDL and hypertriacylglycerolaemia. In addition, LDL particles are converted to smaller, more atherogenic lipoproteins<sup>(129)</sup>. A number of factors are likely to be responsible for diabetic dyslipidaemia and these include insulin's effects on liver apoprotein production, dysregulation of lipoprotein lipase (LPL), impaired action of cholesteryl ester transfer protein and the peripheral action of insulin on adipose and muscle<sup>(128)</sup>. Hormone-sensitive lipase and LPL are two enzymes that regulate mobilisation and deposition of fatty acids in adipose tissue in a reciprocal manner<sup>(130)</sup>. Reduced insulin action leads to increased lipolysis in adipocytes with increased NEFA release, which can mediate many adverse metabolic effects, most notably further insulin resistance<sup>(110)</sup> and dyslipidaemia<sup>(128)</sup>. In diabetes greater amounts of fatty acids returning to the liver are reassembled into TAG and secreted in VLDL<sup>(131)</sup>. In the presence of increased concentrations of VLDL in circulation, cholesteryl ester transfer protein will exchange VLDL TAG for cholesteryl ester in the core of HDL, which results in smaller HDL particles, which are more rapidly cleared from plasma<sup>(132)</sup>. This explains the hypertriacylglycerolaemia and reduced HDL commonly observed in type 2 diabetes<sup>(128)</sup>. Cholesteryl ester transfer protein also exchanges VLDL TAG for cholesteryl ester in the core of LDL and the net effect is a decrease in the size and an increase in the density of LDL particles<sup>(128)</sup>. In vitro, small dense LDL can be oxidised more easily, and it binds to LDL receptors less avidly than normal LDL. As a result, diabetic patients are at an increased risk of CVD. Many clinicians measure LDL density and/or size to predict risk of CVD. An increase in waist circumference and waist:hip ratio has been found to be associated with small dense LDL particles<sup>(128,133)</sup>. Therefore, intra-abdominal fat is a critical determinant of an atherogenic lipoprotein profile, including disruptions in the distribution of cholesterol in the different lipoprotein fractions<sup>(105)</sup>.

The characteristic lipid profile in an individual with type 2 diabetes (increased serum VLDL, increased TAG, decreased serum HDL and less commonly, an increase in LDL-cholesterol)(128) may have important implications for MP levels in the eye, as studies have shown that the carotenoids lutein and zeaxanthin are primarily transported by HDL in plasma<sup>(102,103)</sup>. HDL is a known ligand for scavenger receptor B type 1 (SR-B1), and this suggests a 'piggyback' mechanism of lutein and zeaxanthin uptake into the retina via RPE cells which is mediated by SR-B1<sup>(134)</sup>. This hypothesis is supported by evidence from lutein deficiencies observed in the retina of chickens with a genetic HDL defect but not in other organs, implying a major role for HDL in the receptor-mediated transport of lutein to the eye<sup>(102)</sup>. It has been argued that the subspecies of HDL containing apoE supplies lipids and lipid-soluble lutein and zeaxanthin to the retina, and that by increasing HDL, retinal lutein levels may be simultaneously augmented<sup>(100)</sup>.

*n*-3 Long-chain PUFA supplementation may also lead to increases in serum HDL<sup>(135)</sup>. Recent epidemiological studies indicate that increased dietary long-chain-PUFA intakes enhance MPOD<sup>(136)</sup>, and that DHA facilitates the accumulation of lutein in the blood and macula. The mechanism by which DHA increases MPOD may relate to its effect on the transport and uptake of lutein into the macula by mediating changes in lipoprotein levels<sup>(137)</sup>. Putatively, this effect could also be mediated by the anti-inflammatory effects of DHA, but this is conjectural and no direct evidence supports this effect within the retina.

One observational study found that the correlation between MPOD, serum lutein, zeaxanthin and lipids differed depending on the age profile of participants<sup>(138)</sup>. In younger adults, MPOD was found to be influenced by serum lutein, whereas in older adults, circulating lipids were an added determining factor<sup>(138)</sup>. Mares *et al.*<sup>(29)</sup> queried whether any association with serum lipids is truly strong enough to play a meaningful role in the pathogenesis of MPOD depletion. One study which used a diabetic cohort, involving a small group of forty-three participants (fourteen controls, seventeen with diabetes but without retinopathy and twelve with diabetes and with mild non-proliferative retinopathy), found no significant correlation between MPOD and serum lipid levels<sup>(16)</sup>. Our own research group<sup>(28)</sup>, however,

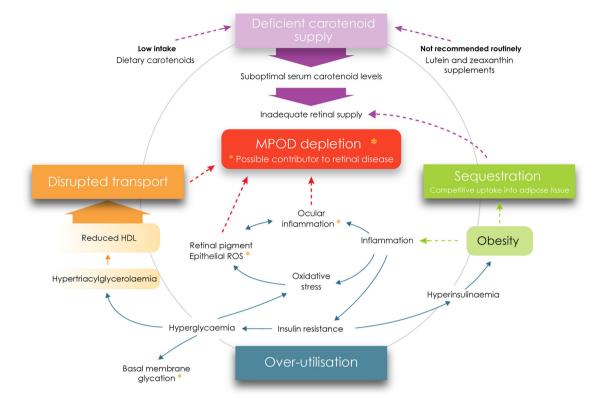


Fig. 2. Metabolic perturbations and supply deficiencies contributing to macular pigment depletion in diabetes. MPOD, macular pigment optical density; ROS, reactive oxygen species. For a colour figure, see the online version of the paper.

reported lower levels of MPOD in type 2 v. type 1 diabetics and noted significantly lower HDL values within the type 2 group, raising the possibility that lower HDL levels may have contributed to this difference. More recently, the DiVFuSS study(31) has demonstrated significant improvements in serum LDL-cholesterol level (9 % mean decrease in the intervention group v. 1 % mean increase in placebo group), HDL-cholesterol (7 % mean increase in the intervention group v.3% decrease in placebo group) and TAG (8.6 % mean decrease in the intervention group v. 2 % mean increase in placebo group), suggesting a positive effect of the study formulation on serum lipids. Whilst this effect was probably attributable to other non-carotenoid components of the supplement, the synchronous increase in MPOD observed in the intervention group may have been partially mediated by these favourable changes in lipid profile and their consequential impact on carotenoid transport to the retina<sup>(102,103,139)</sup>. In the broader context, the evidence from this study is sufficiently robust to prompt further lines of investigation into the role of MP supplementation either in isolation or as a multi-component supplement in diabetes. The research to date, however, is not conclusive and it remains unclear from the limited amount of observational and RCT data whether serum lipids (primarily HDL) play a meaningful role in determining MPOD status. Given the theoretical basis for an association between lipid profile and MP status, and the fact that lipid profile is often altered deleteriously in diabetes, further investigation is warranted. The relationship between dyslipidaemia and lower levels of MP is outlined in Table  $4^{(16,29,31,138-141)}$ .

### Causal mechanisms and metabolic perturbations associated with lower macular pigment in diabetes

The candidate causal mechanisms explored herein as contributors to lower MPOD levels in diabetes relate primarily to type 2 diabetes (or poorly controlled type 1). The complex interrelationship between oxidative stress/inflammation and the metabolic correlates of diabetes (adiposity, dyslipidaemia, insulin resistance, hyperinsulinaemia and hyperglycaemia), and their associations with MPOD, both individually and as a whole, represent a significant scientific challenge. In summary, our analysis indicates that MPOD might be depleted through at least four causal mechanisms previously described, all of which are outlined in Fig. 2.

# Dietary determinants of macular pigment optical density in diabetes

First and foremost, patients with type 2 diabetes may experience lower levels of MP as a result of poor dietary intake of antioxidant nutrients (lutein, zeaxanthin and *meso*-zeaxanthin). Lutein and zeaxanthin are not synthesised *de novo* in humans; therefore, a diet rich in fruit and vegetables is necessary to enhance MP levels in the eye<sup>(4)</sup>. While the exact cause of diabetes is still not fully understood, dietary factors have garnered particular attention over the last number of years. In fact, some research indicates that up to 74 % of type 2 diabetes cases are directly attributable to obesity<sup>(142)</sup>.

Table 5. Dietary factors associated with the metabolic correlates of type 2 c	diabetes mellitus
---	-------------------

Metabolic correlates of type 2 diabetes	(a) Causal dietary factors associated with the metabolic correlates of type 2 diabetes	(b) Preventative dietary factors associated with the metabolic correlates of type 2 diabetes
Oxidative stress	High protein (red meat) <sup>(158)</sup> , high fat (saturated fat) <sup>(161,162)</sup> , high carbohydrate (refined sugar, fructose) <sup>(143,144,163)</sup>	<ul> <li>Whole grains (fibre)<sup>(164)</sup>, nuts (fibre, <i>n</i>-3 PUFA)<sup>(165)</sup>.</li> <li>Fruit and vegetables (antioxidants vitamins C and E, β-carotene, lycopene, quercetin, resveratrol, flavonoids, lutein, zeaxanthin and <i>meso</i>-zeaxanthin)<sup>(4,12,15,30,31,70,152,153,155,166,167)</sup>.</li> <li>Fish (<i>n</i>-3 PUFA)<sup>(168)</sup>, legumes (protein, phytochemicals)<sup>(169)</sup></li> </ul>
Inflammation	High fat (saturated fat, <i>trans</i> -fat) <sup>(170–172)</sup> , high carbohydrate (refined sugar, fructose) <sup>(163,173,174)</sup> , alcohol <sup>(175)</sup>	<ul> <li>Oily fish and <i>n</i>-3 FA<sup>(168)</sup>, folate<sup>(176)</sup>, vitamin D<sup>(177)</sup>.</li> <li>Fruit and vegetables (antioxidants vitamins C and E, β-carotene, lycopene, quercetin, resveratrol, flavonoids, lutein, zeaxanthin and <i>meso</i>-zeaxanthin)<sup>(14,166)</sup></li> </ul>
Obesity	High energy-dense foods <sup>(178)</sup> , high fat (saturated) <sup>(179–182)</sup> , high carbohydrate (refined sugar, fructose) <sup>(143,144)</sup>	Diet and exercise <sup>(183)</sup> , high dietary fibre <sup>(184)</sup> . High fruit and vegetable intake (antioxidants vitamins C and E, β-carotene, lycopene, quercetin, resveratrol, flavonoids, lutein, zeaxanthin and <i>meso</i> -zeaxanthin) <sup>(166)</sup> , high milk intake <sup>(185)</sup>
Dyslipidaemia	High fat (saturated fat, <i>trans</i> -fat) <sup>(186–188)</sup> , high carbohydrate (refined sugar, fructose) <sup>(189,190)</sup> , Na <sup>(189)</sup> , alcohol (especially binges) <sup>(191)</sup> , low physical activity <sup>(192)</sup>	High dietary fibre <sup>(184)</sup> , <i>n</i> -3 FA <sup>(193,194)</sup> , plant stanols/sterols <sup>(195)</sup> , physical activity <sup>(135,192)</sup>

FA, fatty acids.

Many studies have reported a positive association between a high intake of sugars and the development of type 2 diabetes<sup>(143-145)</sup>. Research has also found an association between high fat intakes and type 2 diabetes and impaired glucose tolerance<sup>(146,147)</sup>, with some literature suggesting that high-fat, low-carbohydrate diets are associated with the onset of type 2 diabetes<sup>(146)</sup>. A global change in dietary intake has occurred over recent decades resulting from an increased use of sweeteners such as fructose and sucrose by the food industry<sup>(148)</sup>. Recent evidence suggests a causal link between the high intake of sugar-sweetened soft drinks and obesity and diabetes. This is thought to arise from the large amount of fructose in these soft drinks, which impairs insulin sensitivity and raises blood glucose levels and BMI to dangerous levels(144,149). Fructose consumption increases postprandial TAG concentrations within hours<sup>(150)</sup>, which suggests that postprandial hypertriacylglycerolaemia is one of the earliest metabolic perturbations associated with fructose consumption. The most likely mechanism for this hypertriacylglycerolaemia is increased hepatic de novo lipogenesis, which in turn up-regulates VLDL production and secretion<sup>(151)</sup>, factors which have a knock-on suppressive effect on HDL, a known transporter of carotenoids to the eye<sup>(102,103)</sup>.

Studies have also found an inverse correlation between the intake of green leafy vegetables and risk of developing type 2 diabetes<sup>(152,153)</sup>, and that the consumption of fruits and vegetables may protect against the development of type 2 diabetes, as they are rich in micronutrients, fibre and antioxidants<sup>(153)</sup>, dietary components also known to enhance MPOD. A recent meta-analysis revealed that a higher intake of fruit, especially berries and green leafy vegetables, yellow vegetables and cruciferous vegetables or their fibre, is associated with a lower risk of type 2 diabetes<sup>(152)</sup>. Health benefits have also been observed with the Mediterranean diet<sup>(154,155)</sup>, a diet which contains an abundance of fruit and vegetables. While the beneficial effects of fruits and vegetables on diabetes risk are likely to be multiple, these foods are the primary dietary source of carotenoids

including lutein and zeaxanthin, highlighting their consumption as a key determinant of MPOD. Lower levels of MPOD in diabetes might also be associated with high intakes of fried foods and fat<sup>(156)</sup>, alcohol<sup>(157)</sup>, red and processed meat<sup>(158)</sup>, sugar-sweetened beverages<sup>(143,144)</sup> and high-glycaemic index foods<sup>(159)</sup>, all of which have been linked with higher levels of visceral fat and increased waist circumference. Numerous studies have shown that obesity (in particular, central obesity) can increase the risk of type 2 diabetes<sup>(160)</sup>.

While the body's natural defence against oxidative damage and inflammation is neutralisation by endogenous antioxidants (enzymic and non-enzymic), in association with exogenous antioxidants (vitamin C, vitamin E, carotenoids,  $\beta$ -carotene, lutein, zeaxanthin and *meso*-zeaxanthin)<sup>(50,68)</sup>, these antioxidant defences may be compromised in diabetes. Hence, it is clear that both over-utilisation and under-supply of antioxidant nutrients may contribute to lower levels of MP in the diabetic retina.

Based on the outlined associations between nutritional and dietary factors and the components of the metabolic syndrome which may contribute to MPOD depletion, we outline dietary factors which: (1) may exacerbate the metabolic correlates associated with type 2 diabetes (Table 5 column a); and (2) may help to prevent or resolve the metabolic perturbations associated with type 2 diabetes (Table 5 column b).

Overall, Western dietary habits are a significant factor in the development of the metabolic syndrome. The continuous overprovision of energy via dietary carbohydrate and lipid, when unmatched by physical activity-induced energy expenditure, leads to a state of excess adiposity, chronic low-grade inflammation, and oxidative stress and dyslipidaemia; features associated with type 2 diabetes and which may lead to MPOD depletion.

The importance of diet cannot be overemphasised in diabetes management. Given the variety of mechanisms which may contribute to MPOD depletion and the broader ocular risks associated with such MPOD depletion in diabetes, a comprehensive dietary approach is therefore necessary to optimise ocular health and MPOD status. The generous provision of brightly coloured fruits and vegetables which are rich in a broad range of antioxidants including the target carotenoids lutein, zeaxanthin and meso-zeaxanthin represents an obvious recommendation. This, however, should be complemented by the inclusion of wholegrain-fortified breakfast cereals for their folate, vitamin D and antiobesogenic fibre content; and with plenty of oily fish to provide further vitamin D and n-3 essential fatty acids to attenuate inflammation and correct or prevent hypertriacylglycerolaemia. These foods should be used to displace those which are high in fat, saturated fat, refined sugar and fructose; for example, oily fish should replace red or processed meats at several main meals each week, fruit or raw vegetables should be used to replace snack foods high in fat, saturated fat, trans-fat and refined sugar (for example, baked goods and confectionery), and low-fat fortified milk should replace sugar-sweetened soft drinks. Starchy carbohydrates should be evenly distributed over the day to achieve better glycaemic control, with sterol- or stanol-enriched products used to reduce serum LDL and consequently enhance LDL:HDL ratio. Finally, from a broader lifestyle perspective, these individuals should be advised to moderate their alcohol consumption to no more than two standard drinks per d with several alcohol-free days each week, and to engage in moderate-intensity exercise (for example, walking) on most days of the week to enhance their insulin sensitivity and optimise their blood glucose and lipoprotein profiles. Table 5 (column a) outlines the dietary factors causally associated with the metabolic correlates of type 2 diabetes, and Table 5 (column b) outlines the dietary factors associated with protection against the metabolic perturbations of type 2 diabetes.

### Conclusion

The present review outlines our current understanding of the relationship between diabetes and MPOD. MP appears to be generally lower in diabetic populations compared with healthy controls<sup>(16)</sup>, and this is noted particularly amongst patients with type 2 diabetes<sup>(28)</sup>. Impaired defence against reactive oxidative species (ROS) at the retina may not only be attributable to lower intake of foods rich in carotenoids and other antioxidants (for example, fruit, vegetables, legumes), however, but may also arise from the increased utilisation of these antioxidants in diabetes (via chronic inflammation and oxidative stress). Adiposity, particularly abdominal fatness, is closely linked to type 2 diabetes, and is a significant factor in the condition's pathogenesis. Excess fat tissue may not only compete with the retina for uptake of dietary carotenoids, but may also contribute to increased oxidative stress and inflammation, factors which may have a negative impact on MP levels in the eye. Similarly, the hyperglycaemia which is characteristic of diabetes also contributes to inflammation and oxidative stress, and their depletive effects on MPOD. Dyslipidaemia, more common in poorly controlled diabetes, is a metabolic correlate which may adversely affect the transport and assimilation of carotenes into retinal tissue, by limiting the availability of their principal carrier HDL. The chronic low-grade inflammation and increased oxidative stress observed in patients with type 2 diabetes may also increase the use of these carotenoids at the molecular level, and, in so doing, may elevate our dietary requirement for them.

It is important to note the significant limitations which currently prevail in the reviewed literature. The heterogeneity of studies is a major drawback. Small sample sizes, the merging of type 1 and type 2 diabetic patients in statistical analyses, and the complex interplay of diabetes and its accessory factors (adiposity, dyslipidaemia, oxidative stress and inflammation) with MPOD status are also challenging. While there is some preliminary positive evidence, primarily from animal studies<sup>(19,20)</sup> and the DiVFuss trial<sup>(31)</sup>, that higher MP levels may be protective against retinal damage, there is limited evidence that low MPOD levels are detrimental to ocular health in diabetes, and the actual benefits of higher MPOD in terms of retinal disease development and/or progression in diabetes remain to be elucidated.

In view of the limited evidence available, supplementation with lutein, zeaxanthin and/or meso-zeaxanthin, either with or without other antioxidants, represents a fertile area for future interventional research. It is important to recognise. however, that in order to minimise the risk of MPOD depletion and/or ocular damage in diabetes, it is not sufficient to simply supplement with macular carotenoids. Rather, our review suggests the need for a more holistic evaluation of diet and lifestyle in relation to MPOD in diabetes. In this context, inflammation and oxidative stress, the dual pathogenic driving forces of diabetes which occur independent of hyperglycaemia, may be amenable to dietary intervention and nutritional supplementation. The incorporation of a number of dietary modifications (for example, enhanced antioxidant, vitamin D and n-3 fatty acid intake; and reduced fat, saturated fat, trans-fat and refined sugar consumption), alongside lifestyle changes such as increased physical activity, may not only address the under-supply of macular carotenoids, but may also abrogate the competitive storage, transport, inflammatory and oxidative features of type 2 diabetes which may compromise MPOD status. The safety and low cost of such interventions further commend their potential value as a therapeutic adjunct in diabetic eye disease, and may come to represent an essential element of ocular care for this important and expanding patient group.

#### Acknowledgements

We would like to give special thanks to Kate Loskutova for her help with the graphics in Fig. 2, and to Dr Kate Younger for her review of the draft manuscript.

The present review received no grant from any funding agency, commercial or not-for-profit sectors.

G. S. wrote the paper and has primary responsibility for the final content. J. L. and D. M. helped to draft, evaluate and edit the paper. D. F. helped with the initial drafting and content of the paper.

There are no conflicts of interest.

G. Scanlon et al.

#### References

- 1. Hirsch J & Curcio CA (1989) The spatial resolution capacity of human foveal retina. *Vision Res* **29**, 1095–1101.
- 2. Bone RA, Landrum JT, Hime GW, *et al.* (1993) Stereochemistry of the human macular carotenoids. *Invest Ophthalmol Vis Sci* **34**, 2033–2040.
- 3. Bernstein PS, Li B, Vachali PP, *et al.* (2016) Lutein, zeaxanthin, and *meso*-zeaxanthin: the basic and clinical science underlying carotenoid-based nutritional interventions against ocular disease. *Prog Retin Eye Res* **50**, 34–66.
- Sommerburg O, Keunen JE, Bird AC, *et al.* (1998) Fruits and vegetables that are sources for lutein and zeaxanthin: the macular pigment in human eyes. *Br J Ophthalmol* 82, 907–910.
- Johnson EJ, Neuringer M, Russell RM, et al. (2005) Nutritional manipulation of primate retinas, III: effects of lutein or zeaxanthin supplementation on adipose tissue and retina of xanthophyll-free monkeys. *Invest Ophthalmol Vis Sci* 46, 692–702.
- 6. Nolan J, Meagher K, Kashani S, *et al.* (2013) What is meso-zeaxanthin, and where does it come from? *Eye* **27**, 899–905.
- Snodderly DM, Auran JD & Delori FC (1984) The macular pigment. II. Spatial distribution in primate retinas. *Invest Ophthalmol Vis Sci* 25, 674–685.
- Trieschmann M, Van Kuijk F, Alexander R, *et al.* (2008) Macular pigment in the human retina: histological evaluation of localization and distribution. *Eye (Lond)* 22, 132–134.
- Beatty S, Koh H-H, Phil M, et al. (2000) The role of oxidative stress in the pathogenesis of age-related macular degeneration. Surv Ophthalmol 45, 115–134.
- Loughman J, Davison PA, Nolan JM, *et al.* (2010) Macular pigment and its contribution to visual performance and experience. *J Optom* 3, 74–90.
- Li S-Y, Fung FK, Fu ZJ, *et al.* (2012) Anti-inflammatory effects of lutein in retinal ischemic/hypoxic injury: *in vivo* and *in vitro* studies. *Invest Ophthalmol Vis Sci* 53, 5976–5984.
- Seddon JM, Ajani UA, Sperduto RD, *et al.* (1994) Dietary carotenoids, vitamins A, C, and E, and advanced age-related macular degeneration. *JAMA* 272, 1413–1420.
- 13. Richer S, Stiles W, Statkute L, *et al.* (2004) Double-masked, placebo-controlled, randomized trial of lutein and antioxidant supplementation in the intervention of atrophic age-related macular degeneration: the Veterans LAST study (Lutein Antioxidant Supplementation Trial). *Optometry* **75**, 216–230.
- Izumi-Nagai K, Nagai N, Ohgami K, et al. (2007) Macular pigment lutein is antiinflammatory in preventing choroidal neovascularization. Arterioscler Thromb Vasc Biol 27, 2555–2562.
- Chew EY, Clemons TE, SanGiovanni JP, et al. (2013) Lutein + zeaxanthin and omega-3 fatty acids for age-related macular degeneration: the Age-Related Eye Disease Study 2 (AREDS2) randomized clinical trial. *JAMA* **309**, 2005–2015.
- Lima VC, Rosen RB, Maia M, *et al.* (2010) Macular pigment optical density measured by dual-wavelength autofluorescence imaging in diabetic and nondiabetic patients: a comparative study. *Invest Ophthalmol Vis Sci* 51, 5840–5845.
- 17. Sasaki M, Ozawa Y, Kurihara T, *et al.* (2010) Neurodegenerative influence of oxidative stress in the retina of a murine model of diabetes. *Diabetologia* **53**, 971–979.
- Muriach M, Bosch-Morell F, Alexander G, *et al.* (2006) Lutein effect on retina and hippocampus of diabetic mice. *Free Radic Biol Med* 41, 979–984.
- Kowluru RA, Menon B & Gierhart DL (2008) Beneficial effect of zeaxanthin on retinal metabolic abnormalities in diabetic rats. *Invest Ophthalmol Vis Sci* 49, 1645–1651.
- 20. Kowluru RA, Zhong Q, Santos JM, *et al.* (2014) Beneficial effects of the nutritional supplements on the development of diabetic retinopathy. *Nutr Metab (Lond)* **11**, 8.

- Tang L, Zhang Y, Jiang Y, *et al.* (2011) Dietary wolfberry ameliorates retinal structure abnormalities in db/db mice at the early stage of diabetes. *Exp Biol Med (Maywood)* 236, 1051–1063.
- Yu H, Wark L, Ji H, *et al.* (2013) Dietary wolfberry upregulates carotenoid metabolic genes and enhances mitochondrial biogenesis in the retina of db/db diabetic mice. *Mol Nutr Food Res* 57, 1158–1169.
- Arnal E, Miranda M, Johnsen-Soriano S, *et al.* (2009) Beneficial effect of docosahexanoic acid and lutein on retinal structural, metabolic, and functional abnormalities in diabetic rats. *Curr Eye Res* 34, 928–938.
- Gorusupudi A & Vallikannan B (2012) Glycolipids improve lutein bioavailability and accumulation in eyes in mice. *Eur J Lipid Sci Technol* **114**, 710–717.
- 25. Mamatha BS & Baskaran V (2011) Effect of micellar lipids, dietary fiber and  $\beta$ -carotene on lutein bioavailability in aged rats with lutein deficiency. *Nutrition* **27**, 960–966.
- Davies NP & Morland AB (2002) Color matching in diabetes: optical density of the crystalline lens and macular pigments. *Invest Ophthalmol Vis Sci* 43, 281–289.
- Zagers NP, Pot MC & Van Norren D (2005) Spectral and directional reflectance of the fovea in diabetes mellitus: photoreceptor integrity, macular pigment and lens. *Vision Res* 45, 1745–1753.
- 28. Scanlon G, Connell P, Ratzlaff M, *et al.* (2015) Macular pigment optical density is lower in type 2 diabetes, compared with type 1 diabetes and normal controls. *Retina* **35**, 1808–1816.
- 29. Mares JA, LaRowe TL, Snodderly DM, *et al.* (2006) Predictors of optical density of lutein and zeaxanthin in retinas of older women in the Carotenoids in Age-Related Eye Disease Study, an ancillary study of the Women's Health Initiative. *Am J Clin Nutr* **84**, 1107–1122.
- Hu B-J, Hu Y-N, Lin S, *et al.* (2011) Application of lutein and zeaxanthin in nonproliferative diabetic retinopathy. *Invest Ophthalmol Vis Sci* 4, 303.
- Chous AP, Richer SP, Gerson JD, *et al.* (2015) The Diabetes Visual Function Supplement Study (DiVFuSS). *Br J Ophthalmol* **100**, 227–234.
- Zhou X, Wang S, Ding X, *et al.* (2017) Zeaxanthin improves diabetes-induced cognitive deficit in rats through activiting PI3K/AKT signaling pathway. *Brain Res Bull* 132, 190–198.
- 33. She C-Y, Gu H, Xu J, *et al.* (2016) Association of macular pigment optical density with early stage of non-proliferative diabetic retinopathy in Chinese patients with type 2 diabetes mellitus. *Int J Ophthalmol* **9**, 1433–1438.
- 34. Moschos MM, Dettoraki M, Tsatsos M, *et al.* (2017) Effect of carotenoids dietary supplementation on macular function in diabetic patients. *Eye Vis (Lond)* **4**, 23.
- 35. Rodríguez González-Herrero ME, Ruiz M, Román FJL, et al. (2018) Supplementation with a highly concentrated docosahexaenoic acid plus xanthophyll carotenoid multivitamin in nonproliferative diabetic retinopathy: prospective controlled study of macular function by fundus microperimetry. *Clin Ophthalmol* **12**, 1011–1020.
- 36. Cennamo G, Lanni V, Abbate R, *et al.* (2019) The relationship between macular pigment and vessel density in patients with type 1 diabetes mellitus. *Ophthalmic Res* **61**, 19–25.
- Polonsky KS (2012) The past 200 years in diabetes. N Eng J Med 367, 1332–1340.
- Chen L, Magliano DJ & Zimmet PZ (2012) The worldwide epidemiology of type 2 diabetes mellitus – present and future perspectives. *Nat Rev Endocrinol* 8, 228–236.
- Wilkin T (2001) The accelerator hypothesis: weight gain as the missing link between type I and type II diabetes. *Diabetologia* 44, 914–922.

- Kilhovd BK, Giardino I, Torjesen P, *et al.* (2003) Increased serum levels of the specific AGE-compound methylglyoxalderived hydroimidazolone in patients with type 2 diabetes. *Metabolism* 52, 163–167.
- 41. Kalousova M, Skrha J & Zima T (2002) Advanced glycation end-products and advanced oxidation protein products in patients with diabetes mellitus. *Physiol Res* **51**, 597–604.
- Arnalich F, Hernanz A, Lopez-Maderuelo D, *et al.* (2000) Enhanced acute-phase response and oxidative stress in older adults with type II diabetes. *Horm Metab Res* **32**, 407–412.
- Wright E, Scism-Bacon J & Glass L (2006) Oxidative stress in type 2 diabetes: the role of fasting and postprandial glycaemia. *Int J Clin Pract* 60, 308–314.
- Butkowski EG & Jelinek HF (2017) Hyperglycaemia, oxidative stress and inflammatory markers. *Redox Rep* 22, 257–264.
- Robertson RP, Harmon J, Tran POT, *et al.* (2004) β-Cell glucose toxicity, lipotoxicity, and chronic oxidative stress in type 2 diabetes. *Diabetes* 53, Suppl. 1, S119–S124.
- McCord JM (2000) The evolution of free radicals and oxidative stress. *Am J Med* **108**, 652–659.
- Kowluru RA & Chan P-S (2007) Oxidative stress and diabetic retinopathy. *J Diabetes Res* 2007, 43603.
- 48. Rösen P, Nawroth P, King G, *et al.* (2001) The role of oxidative stress in the onset and progression of diabetes and its complications: a summary of a Congress Series sponsored by UNESCO-MCBN, the American Diabetes Association and the German Diabetes Society. *Diabetes Metab Res Rev* **17**, 189–212.
- Sies H (1991) Oxidative stress: from basic research to clinical application. *Am J Med* **91**, S31–S38.
- Bouayed J & Bohn T (2010) Exogenous antioxidants doubleedged swords in cellular redox state: health beneficial effects at physiologic doses versus deleterious effects at high doses. Oxid Med Cell Longev 3, 228–237.
- Young I & Woodside J (2001) Antioxidants in health and disease. J Clin Pathol 54, 176–186.
- Ceriello A (1997) Acute hyperglycaemia and oxidative stress generation. *Diabet Med* 14, Suppl. 3, S45–S49.
- 53. Schalch W (1992) Carotenoids in the retina a review of their possible role in preventing or limiting damage caused by light and oxygen. *EXS* **62**, 280–298.
- Machlin LJ & Bendich A (1987) Free radical tissue damage: protective role of antioxidant nutrients. *FASEB J* 1, 441–445.
- Borish ET, Pryor WA, Venugopal S, *et al.* (1987) DNA synthesis is blocked by cigarette tar-induced DNA single-strand breaks. *Carcinogenesis* 8, 1517–1520.
- Algvere PV, Marshall J & Seregard S (2006) Age-related maculopathy and the impact of blue light hazard. *Acta Ophthalmol* 84, 4–15.
- 57. Zhang W, Liu H, Rojas M, *et al.* (2011) Anti-inflammatory therapy for diabetic retinopathy. *Immunotherapy* **3**, 609–628.
- Cohen G, Riahi Y, Alpert E, *et al.* (2007) The roles of hyperglycaemia and oxidative stress in the rise and collapse of the natural protective mechanism against vascular endothelial cell dysfunction in diabetes. *Arch Physiol Biochem* **113**, 259–267.
- Esposito K, Nappo F, Marfella R, *et al.* (2002) Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans. *Circulation* **106**, 2067–2072.
- 60. Halliwell B (2006) Oxidative stress and neurodegeneration: where are we now? *J Neurochem* **97**, 1634–1658.
- Brownlee M (2001) Biochemistry and molecular cell biology of diabetic complications. *Nature* **414**, 813–820.
- 62. Kahn BB (1998) Type 2 diabetes: when insulin secretion fails to compensate for insulin resistance. *Cell* **92**, 593–596.
- 63. Ceriello A & Motz E (2004) Is oxidative stress the pathogenic mechanism underlying insulin resistance, diabetes, and

cardiovascular disease? The common soil hypothesis revisited. Arterioscler Thromb Vasc Biol 24, 816–823.

- Brownlee M (2005) The pathobiology of diabetic complications. *Diabetes* 54, 1615–1625.
- 65. Anonymous (1993) UK Prospective Diabetes Study (UKPDS). IX: Relationships of urinary albumin and *N*-acetylglucosaminidase to glycaemia and hypertension at diagnosis of type 2 (non-insulin-dependent) diabetes mellitus and after 3 months diet therapy. *Diabetologia* **36**, 835–842.
- Nishikawa T, Edelstein D, Du XL, *et al.* (2000) Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature* **404**, 787–790.
- Matough FA, Budin SB, Hamid ZA, *et al.* (2012) The role of oxidative stress and antioxidants in diabetic complications. *Sultan Qaboos Univer Med J* 12, 5–18.
- Cabrera MP & Chihuailaf RH (2011) Antioxidants and the integrity of ocular tissues. *Vet Med Int* 2011, 905153.
- Evans P & Halliwell B (2001) Micronutrients: oxidant/antioxidant status. Br J Nutr 85, Suppl. 2, S67–S74.
- 70. Roberts JE & Dennison J (2015) The photobiology of lutein and zeaxanthin in the eye. *J Ophthalmol* **2015**, 687173.
- 71. Sies H, Stahl W & Sundquist AR (1992) Antioxidant functions of vitamins. *Ann N Y Acad Sci* **669**, 7–20.
- 72. Khachik F, Bernstein PS & Garland DL (1997) Identification of lutein and zeaxanthin oxidation products in human and monkey retinas. *Invest Ophthalmol Vis Sci* **38**, 1802–1811.
- 73. Cantrell A, McGarvey D, Truscott TG, *et al.* (2003) Singlet oxygen quenching by dietary carotenoids in a model membrane environment. *Arch Biochem Biophys* **412**, 47–54.
- Bhosale P & Bernstein PS (2005) Microbial xanthophylls. *Appl Microbiol Biotechnol* 68, 445–455.
- Li B, Ahmed F & Bernstein PS (2010) Studies on the singlet oxygen scavenging mechanism of human macular pigment. *Arch Biochem Biophys* **504**, 56–60.
- Akuffo K, Nolan J, Howard A, *et al.* (2015) Sustained supplementation and monitored response with differing carotenoid formulations in early age-related macular degeneration. *Eye* 29, 902–912.
- Hendrychova T, Vytrisalova M, Alwarafi A, *et al.* (2015) Fatand fiber-related diet behavior among type 2 diabetes patients from distinct regions. *Patient Prefer Adherence* 9, 319–325.
- Manna P & Jain SK (2015) Obesity, oxidative stress, adipose tissue dysfunction, and the associated health risks: causes and therapeutic strategies. *Metab Syndr Relat Disord* 13, 423–444.
- 79. Wellen KE & Hotamisligil GS (2005) Inflammation, stress, and diabetes. *J Clin Invest* **115**, 1111–1119.
- Loane E, Kelliher C, Beatty S, *et al.* (2008) The rationale and evidence base for a protective role of macular pigment in age-related maculopathy. *Br J Ophthalmol* **92**, 1163–1168.
- Kowluru RA (2005) Diabetic retinopathy: mitochondrial dysfunction and retinal capillary cell death. *Antioxid Redox Signal* 7, 1581–1587.
- 82. Ishibashi Y, Matsui T, Maeda S, *et al.* (2013) Advanced glycation end products evoke endothelial cell damage by stimulating soluble dipeptidyl peptidase-4 production and its interaction with mannose 6-phosphate/insulin-like growth factor II receptor. *Cardiovasc Diabetol* **12**, 125.
- 83. Semeraro F, Cancarini A, Rezzola S, *et al.* (2015) Diabetic retinopathy: vascular and inflammatory disease. *J Diabetes Res* **2015**, 582060.
- Hollyfield JG, Bonilha VL, Rayborn ME, *et al.* (2008) Oxidative damage-induced inflammation initiates age-related macular degeneration. *Nat Med* 14, 194–198.
- Chen L, Chen R, Wang H, *et al.* (2015) Mechanisms linking inflammation to insulin resistance. *Int J Endocrinol* **2015**, 508409.

16

N Nutrition Research Reviews

- Bhagat N, Grigorian RA, Tutela A, *et al.* (2009) Diabetic macular edema: pathogenesis and treatment. *Surv Ophthalmol* 54, 1–32.
- Zhang X, Zeng H, Bao S, *et al.* (2014) Diabetic macular edema: new concepts in patho-physiology and treatment. *Cell Biosci* 4, 27.
- Lima VC, Rosen RB & Farah M (2016) Macular pigment in retinal health and disease. *Int J Retina Vitreous* 2, 19.
- McArdle MA, Finucane OM, Connaughton RM, *et al.* (2013) Mechanisms of obesity-induced inflammation and insulin resistance: insights into the emerging role of nutritional strategies. *Front Endocrinol (Lausanne)* 4, 52.
- Tangvarasittichai S, Pongthaisong S & Tangvarasittichai O (2016) Tumor necrosis factor-A, interleukin-6, C-reactive protein levels and insulin resistance associated with type 2 diabetes in abdominal obesity women. *Indian J Clin Biochem* **31**, 68–74.
- Phosat C, Panprathip P, Chumpathat N, *et al.* (2017) Elevated C-reactive protein, interleukin 6, tumor necrosis factor α and glycemic load associated with type 2 diabetes mellitus in rural Thais: a cross-sectional study. *BMC Endocr Disord* 17, 44.
- 92. Rodrigues KF, Pietrani NT, Bosco AA, *et al.* (2017) IL-6, TNF-α, and IL-10 levels/polymorphisms and their association with type 2 diabetes mellitus and obesity in Brazilian individuals. *Arch Endocrin Metab* **61**, 438–446.
- Barzilay JI, Abraham L, Heckbert SR, *et al.* (2001) The relation of markers of inflammation to the development of glucose disorders in the elderly. *Diabetes* 50, 2384–2389.
- Freeman DJ, Norrie J, Caslake MJ, *et al.* (2002) C-reactive protein is an independent predictor of risk for the development of diabetes in the West of Scotland Coronary Prevention Study. *Diabetes* 51, 1596–1600.
- Tang J & Kern TS (2011) Inflammation in diabetic retinopathy. Progress Ret Eye Res 30, 343–358.
- 96. Bian Q, Gao S, Zhou J, *et al.* (2012) Lutein and zeaxanthin supplementation reduces photooxidative damage and modulates the expression of inflammation-related genes in retinal pigment epithelial cells. *Free Radic Biol Med* **53**, 1298–1307.
- Chung RW, Leanderson P, Lundberg AK, et al. (2017) Lutein exerts anti-inflammatory effects in patients with coronary artery disease. Atherosclerosis 262, 87–93.
- Johnson EJ, Hammond BR, Yeum K-J, *et al.* (2000) Relation among serum and tissue concentrations of lutein and zeaxanthin and macular pigment density. *Am J Clin Nutr* **71**, 1555–1562.
- Hammond BR, Ciulla TA & Snodderly DM (2002) Macular pigment density is reduced in obese subjects. *Invest Ophthalmol Vis Sci* 43, 47–50.
- Thomson LR, Toyoda Y, Langner A, et al. (2002) Elevated retinal zeaxanthin and prevention of light-induced photoreceptor cell death in quail. *Invest Ophthalmol Vis Sci* 43, 3538–3549.
- Nolan J, O'Donovan O, Kavanagh H, *et al.* (2004) Macular pigment and percentage of body fat. *Invest Ophthalmol Vis Sci* 45, 3940–3950.
- 102. Connor WE, Duell PB, Kean R, et al. (2007) The prime role of HDL to transport lutein into the retina: evidence from HDLdeficient WHAM chicks having a mutant ABCA1 transporter. Invest Ophthalmol Vis Sci 48, 4226–4231.
- Wang W, Connor SL, Johnson EJ, *et al.* (2007) Effect of dietary lutein and zeaxanthin on plasma carotenoids and their transport in lipoproteins in age-related macular degeneration. *Am J Clin Nutr* 85, 762–769.
- Boden G (2008) Obesity and free fatty acids. *Endocrinol* Metab Clin North Am 37, 635–646.
- 105. Nieves DJ, Cnop M, Retzlaff B, *et al.* (2003) The atherogenic lipoprotein profile associated with obesity and insulin

resistance is largely attributable to intra-abdominal fat. *Diabetes* **52**, 172–179.

- 106. Castro A, Macedo-de la Concha L & Pantoja-Meléndez C (2017) Low-grade inflammation and its relation to obesity and chronic degenerative diseases. *Rev Méd Hosp Gen Méx* 80, 101–105.
- Al-Aubaidy HA & Jelinek HF (2011) Oxidative DNA damage and obesity in type 2 diabetes mellitus. *Eur J Endocrinol* 164, 899–904.
- Savini I, Catani MV, Evangelista D, *et al.* (2013) Obesity-associated oxidative stress: strategies finalized to improve redox state. *Int J Mol Sci* 14, 10497–10538.
- Reaven GM, Hollenbeck C, Jeng C-Y, et al. (1988) Measurement of plasma glucose, free fatty acid, lactate, and insulin for 24 h in patients with NIDDM. Diabetes 37, 1020–1024.
- 110. Karpe F, Dickmann JR & Frayn KN (2011) Fatty acids, obesity, and insulin resistance: time for a reevaluation. *Diabetes* **60**, 2441–2449.
- 111. Burke JD, Curran-Celentano J & Wenzel AJ (2005) Diet and serum carotenoid concentrations affect macular pigment optical density in adults 45 years and older. *J Nutr* **135**, 1208–1214.
- 112. Gruber M, Chappell R, Millen A, *et al.* (2004) Correlates of serum lutein + zeaxanthin: findings from the Third National Health and Nutrition Examination Survey. *J Nutr* **134**, 2387–2394.
- 113. Kirby ML, Beatty S, Stack J, *et al.* (2011) Changes in macular pigment optical density and serum concentrations of lutein and zeaxanthin in response to weight loss. *Br J Nutr* **105**, 1036–1046.
- 114. Seddon JM, Cote J, Davis N, *et al.* (2003) Progression of agerelated macular degeneration: association with body mass index, waist circumference, and waist-hip ratio. *Arch Ophthalmol* **121**, 785–792.
- 115. Nolan JM, Feeney J, Kenny RA, et al. (2012) Education is positively associated with macular pigment: The Irish Longitudinal Study on Ageing (TILDA) Education and the TILDA Study. Invest Ophthalmol Vis Sci 53, 7855–7861.
- Bovier ER, Lewis RD & Hammond BR (2013) The relationship between lutein and zeaxanthin status and body fat. *Nutrients* 5, 750–757.
- 117. Blaak E (2001) Gender differences in fat metabolism. *Curr* Opin Clin Nutr Metab Care **4**, 499–502.
- Björntorp P (1991) Adipose tissue distribution and function. Int J Obes 15, 67–81.
- Chung H-Y, Ferreira ALA, Epstein S, *et al.* (2009) Site-specific concentrations of carotenoids in adipose tissue: relations with dietary and serum carotenoid concentrations in healthy adults. *Am J Clin Nutr* **90**, 533–539.
- Gupta A, Raman R, Biswas S, *et al.* (2012) Association between various types of obesity and macular pigment optical density. *Eye* 26, 259–266.
- 121. Fernández-Sánchez A, Madrigal-Santillan E, Bautista M, *et al.* (2011) Inflammation, oxidative stress, and obesity. *Int J Mol Sci* **12**, 3117–3132.
- 122. Bondia-Pons I, Ryan L & Martinez JA (2012) Oxidative stress and inflammation interactions in human obesity. *J Physiol Biochem* **68**, 701–711.
- 123. Kwon H & Pessin J (2013) Adipokines mediate inflammation and insulin resistance. *Front Endocrinol (Lausanne)* **4**, 71.
- 124. Broekmans WM, Berendschot TT, Klopping-Ketelaars IA, *et al.* (2002) Macular pigment density in relation to serum and adipose tissue concentrations of lutein and serum concentrations of zeaxanthin. *Am J Clin Nutr* **76**, 595–603.
- 125. Moeller SM, Voland R, Sarto GE, *et al.* (2009) Women's Health Initiative diet intervention did not increase macular pigment

optical density in an ancillary study of a subsample of the Women's Health Initiative. *J Nutr* **139**, 1692–1699.

- Khan N, Walk A, Edwards C, *et al.* (2018) Macular xanthophylls are related to intellectual ability among adults with overweight and obesity. *Nutrients* 10, E396.
- 127. Edwards CG, Walk AM, Cannavale CN, *et al.* (2019) Macular xanthophylls and event-related brain potentials among adults with overweight and obesity. *Mol Nutr Food Res* **2019**, e1801059.
- 128. Goldberg IJ (2001) Diabetic dyslipidemia: causes and consequences. *J Clin Endocrinol Metab* **86**, 965–971.
- Krauss RM & Siri PW (2004) Metabolic abnormalities: triglyceride and low-density lipoprotein. *Endocrinol Metab Clinics* 33, 405–415.
- Frayn KN, Coppack SW, Fielding BA, *et al.* (1995) Coordinated regulation of hormone-sensitive lipase and lipoprotein lipase in human adipose tissue *in vivo*: implications for the control of fat storage and fat mobilization. *Adv Enzyme Regul* **35**, 163–178.
- Ginsberg HN & Illingworth DR (2001) Postprandial dyslipidemia: an atherogenic disorder common in patients with diabetes mellitus. *Am J Cardiol* 88, 9–15.
- 132. Horowitz BS, Goldberg IJ, Merab J, *et al.* (1993) Increased plasma and renal clearance of an exchangeable pool of apolipoprotein AI in subjects with low levels of high density lipoprotein cholesterol. *J Clin Invest* **91**, 1743–1752.
- 133. Campos H, Willett WC, Peterson RM, *et al.* (1991) Nutrient intake comparisons between Framingham and rural and urban Puriscal, Costa Rica. Associations with lipoproteins, apolipoproteins, and low density lipoprotein particle size. *Arterioscler Thromb Vasc Biol* **11**, 1089–1099.
- 134. During A, Doraiswamy S, Harrison EH, et al. (2008) Xanthophylls are preferentially taken up compared with βcarotene by retinal cells via a SRBI-dependent mechanism. J Lipid Res 49, 1715–1724.
- Thomas TR, Smith BK, Donahue OM, *et al.* (2004) Effects of omega-3 fatty acid supplementation and exercise on low-density lipoprotein and high-density lipoprotein subfractions. *Metabolism* 53, 749–754.
- 136. Delyfer M-N, Buaud B, Korobelnik J-F, et al. (2012) Association of macular pigment density with plasma omega-3 fatty acids: the PIMAVOSA study. *Invest Ophthalmol Vis Sci* 53, 1204–1210.
- 137. Johnson EJ, Chung H-Y, Caldarella SM, *et al.* (2008) The influence of supplemental lutein and docosahexaenoic acid on serum, lipoproteins, and macular pigmentation. *Am J Clin Nutr* 87, 1521–1529.
- 138. Olmedilla-Alonso B, Beltrán-de-Miguel B, Estévez-Santiago R, et al. (2014) Markers of lutein and zeaxanthin status in two age groups of men and women: dietary intake, serum concentrations, lipid profile and macular pigment optical density. Nutr J 13, 52.
- 139. Loane E, Nolan JM & Beatty S (2010) The respective relationships between lipoprotein profile, macular pigment optical density, and serum concentrations of lutein and zeaxanthin. *Invest Ophthalmol Vis Sci* **51**, 5897–5905.
- 140. Renzi LM, Hammond BR, Dengler M, *et al.* (2012) The relation between serum lipids and lutein and zeaxanthin in the serum and retina: results from cross-sectional, case–control and case study designs. *Lipids Health Dis* **11**, 33.
- Nagai N, Izumi-Nagai K, Suzuki M, *et al.* (2015) Association of macular pigment optical density with serum concentration of oxidized low-density lipoprotein in healthy adults. *Retina* 35, 820–826.
- 142. Janssen I (2013) The public health burden of obesity in Canada. *Can J Diabetes* **37**, 90–96.

- 143. Gross LS, Li L, Ford ES, *et al.* (2004) Increased consumption of refined carbohydrates and the epidemic of type 2 diabetes in the United States: an ecologic assessment. *Am J Clin Nutr* **79**, 774–779.
- 144. Bray GA, Nielsen SJ & Popkin BM (2004) Consumption of high-fructose corn syrup in beverages may play a role in the epidemic of obesity. *Am J Clin Nutr* **79**, 537–543.
- 145. Elliott SS, Keim NL, Stern JS, *et al.* (2002) Fructose, weight gain, and the insulin resistance syndrome. *Am J Clin Nutr* **76**, 911–922.
- 146. Marshall JA, Hamman RF & Baxter J (1991) High-fat, lowcarbohydrate diet and the etiology of non-insulin-dependent diabetes mellitus: the San Luis Valley Diabetes Study. *Am J Epidem* **134**, 590–603.
- 147. Saldana TM, Siega-Riz AM & Adair LS (2004) Effect of macronutrient intake on the development of glucose intolerance during pregnancy. *Am J Clin Nutr* **79**, 479–486.
- Nseir W, Nassar F & Assy N (2010) Soft drinks consumption and nonalcoholic fatty liver disease. World J Gastroenterol 16, 2579–2588.
- 149. Raben A, Vasilaras TH, Møller AC, *et al.* (2002) Sucrose compared with artificial sweeteners: different effects on *ad libitum* food intake and body weight after 10 wk of supplementation in overweight subjects. *Am J Clin Nutr* **76**, 721–729.
- 150. Swarbrick MM, Stanhope KL, Elliott SS, *et al.* (2008) Consumption of fructose-sweetened beverages for 10 weeks increases postprandial triacylglycerol and apolipoprotein-B concentrations in overweight and obese women. *Br J Nutr* **100**, 947–952.
- 151. Stanhope KL & Havel PJ (2008) Fructose consumption: potential mechanisms for its effects to increase visceral adiposity and induce dyslipidemia and insulin resistance. *Curr Opin Lipidol* **19**, 16–24.
- 152. Wang PY, Fang JC, Gao ZH, *et al.* (2016) Higher intake of fruits, vegetables or their fiber reduces the risk of type 2 diabetes: a meta-analysis. *J Diabetes Investig* **7**, 56–69.
- 153. Li M, Fan Y, Zhang X, *et al.* (2014) Fruit and vegetable intake and risk of type 2 diabetes mellitus: meta-analysis of prospective cohort studies. *BMJ Open* **4**, e005497.
- 154. Tsoupras A, Lordan R & Zabetakis I (2018) Inflammation, not cholesterol, is a cause of chronic disease. *Nutrients* **10**, E604.
- Romagnolo DF & Selmin OI (2017) Mediterranean diet and prevention of chronic diseases. *Nutr Today* 52, 208–222.
- 156. Guallar-Castillón P, Rodriguez-Artalejo F, Fornes NS, *et al.* (2007) Intake of fried foods is associated with obesity in the cohort of Spanish adults from the European Prospective Investigation into Cancer and Nutrition. *Am J Clin Nutr* **86**, 198–205.
- 157. Traversy G & Chaput J-P (2015) Alcohol consumption and obesity: an update. *Curr Obes Rep* **4**, 122–130.
- Wang Y & Beydoun MA (2009) Meat consumption is associated with obesity and central obesity among US adults. *Int J Obes (Lond)* 33, 621–628.
- 159. Ludwig DS, Majzoub JA, Al-Zahrani A, *et al.* (1999) High glycemic index foods, overeating, and obesity. *Pediatrics* **103**, E26.
- Mokdad AH, Ford ES, Bowman BA, *et al.* (2003) Prevalence of obesity, diabetes, and obesity-related health risk factors, 2001. *JAMA* 289, 76–79.
- 161. Peairs AD, Rankin JW & Lee YW (2011) Effects of acute ingestion of different fats on oxidative stress and inflammation in overweight and obese adults. *Nutr J* **10**, 122.
- 162. Nappo F, Esposito K, Cioffi M, *et al.* (2002) Postprandial endothelial activation in healthy subjects and in type 2 diabetic patients: role of fat and carbohydrate meals. *J Am Coll Cardiol* **39**, 1145–1150.

G. Scanlon et al.

- 163. Castro MC, Massa ML, Arbeláez LG, *et al.* (2015) Fructoseinduced inflammation, insulin resistance and oxidative stress: a liver pathological triad effectively disrupted by lipoic acid. *Life Sci* **137**, 1–6.
- Liu S (2003) Whole-grain foods, dietary fiber, and type 2 diabetes: searching for a kernel of truth. *Am J Clin Nutr* 77, 527–529.
- 165. Kim Y, Keogh JB & Clifton PM (2017) Benefits of nut consumption on insulin resistance and cardiovascular risk factors: multiple potential mechanisms of actions. *Nutrients* 9, E1271.
- 166. Boeing H, Bechthold A, Bub A, *et al.* (2012) Critical review: vegetables and fruit in the prevention of chronic diseases. *Eur J Nutr* **51**, 637–663.
- Prior RL (2003) Fruits and vegetables in the prevention of cellular oxidative damage. *AmJ Clin Nutr* 78, Suppl. 3, S570–S578.
- Calder PC (2012) The role of marine omega-3 (*n*-3) fatty acids in inflammatory processes, atherosclerosis and plaque stability. *Mol Nutr Food Res* 56, 1073–1080.
- Bhathena SJ & Velasquez MT (2002) Beneficial role of dietary phytoestrogens in obesity and diabetes. *Am J Clin Nutr* 76, 1191–1201.
- 170. Fritsche KL (2015) The science of fatty acids and inflammation. *Adv Nutr* **6**, S293–S301.
- Kennedy A, Martinez K, Chuang C-C, *et al.* (2008) Saturated fatty acid-mediated inflammation and insulin resistance in adipose tissue: mechanisms of action and implications. *J Nutr* 139, 1–4.
- Lopez-Garcia E, Schulze MB, Meigs JB, et al. (2005) Consumption of *trans* fatty acids is related to plasma biomarkers of inflammation and endothelial dysfunction. *J Nutr* 135, 562–566.
- 173. Aeberli I, Gerber PA, Hochuli M, *et al.* (2011) Low to moderate sugar-sweetened beverage consumption impairs glucose and lipid metabolism and promotes inflammation in healthy young men: a randomized controlled trial. *Am J Clin Nutr* 94, 479–485.
- Glushakova O, Kosugi T, Roncal C, *et al.* (2008) Fructose induces the inflammatory molecule ICAM-1 in endothelial cells. *J Am Soc Nephrol* **19**, 1712–1720.
- Wang HJ, Zakhari S & Jung MK (2010) Alcohol, inflammation, and gut–liver–brain interactions in tissue damage and disease development. *World J Gastroenterol* 16, 1304–1313.
- Huang X, He Z, Jiang X, *et al.* (2016) Folic acid represses hypoxia-induced inflammation in THP-1 cells through inhibition of the PI3K/Akt/HIF-1α pathway. *PLOS ONE* **11**, e0151553.
- Boucher BJ (2012) Is vitamin D status relevant to metabolic syndrome? *Dermatoendocrinol* 4, 212–224.
- 178. Hill JO & Peters JC (1998) Environmental contributions to the obesity epidemic. *Science* **280**, 1371–1374.

- 179. Golay A & Bobbioni E (1997) The role of dietary fat in obesity. *Int J Obes Relat Metab Disord* **21**, Suppl. 3, S2–S11.
- Bray GA & Popkin BM (1998) Dietary fat intake does affect obesity! *Am J Clin Nutr* 68, 1157–1173.
- Hariri N & Thibault L (2010) High-fat diet-induced obesity in animal models. *Nutr Res Rev* 23, 270–299.
- 182. Raatz SK, Conrad Z, Johnson LK, *et al.* (2017) Relationship of the reported intakes of fat and fatty acids to body weight in US adults. *Nutrients* **9**, E438.
- Huang C-J, Zourdos MC, Jo E, *et al.* (2013) Influence of physical activity and nutrition on obesity-related immune function. *Sci World J* 2013, 752071.
- Smith U (1987) Dietary fibre, diabetes and obesity. Scand J Gastroenterol 22, 151–153.
- Beck AL, Heyman M, Chao C, *et al.* (2017) Full fat milk consumption protects against severe childhood obesity in Latinos. *Prev Med Rep* 8, 1–5.
- Chiu S, Williams PT & Krauss RM (2017) Effects of a very high saturated fat diet on LDL particles in adults with atherogenic dyslipidemia: a randomized controlled trial. *PLOS ONE* 12, e0170664.
- Chen C-L, Tetri LH, Neuschwander-Tetri BA, *et al.* (2011) A mechanism by which dietary *trans* fats cause atherosclerosis. *J Nutr Biochem* 22, 649–655.
- 188. Micha R & Mozaffarian D (2008) *Trans* fatty acids: effects on cardiometabolic health and implications for policy. *Prostaglandins Leukot Essent Fatty Acids* **79**, 147–152.
- Kang YJ, Wang HW, Choen SY, *et al.* (2016) Associations of obesity and dyslipidemia with intake of sodium, fat, and sugar among Koreans: a qualitative systematic review. *Clin Nutr Res* 5, 290–304.
- Basciano H, Federico L & Adeli K (2005) Fructose, insulin resistance, and metabolic dyslipidemia. *Nutr Metab* 2, 5.
- 191. Capurso NA & Petrakis I (2016) Dyslipidemia associated with heavy alcohol use. *Am J Addict* **25**, 188–190.
- 192. Dancy C, Lohsoonthorn V & Williams MA (2008) Risk of dyslipidemia in relation to level of physical activity among Thai professional and office workers. *Southeast Asian J Trop Med Public Health* **39**, 932–941.
- 193. Lepretti M, Martucciello S, Burgos Aceves MA, et al. (2018) Omega-3 fatty acids and insulin resistance: focus on the regulation of mitochondria and endoplasmic reticulum stress. *Nutrients* 10, E350.
- 194. Backes J, Anzalone D, Hilleman D, *et al.* (2016) The clinical relevance of omega-3 fatty acids in the management of hyper-triglyceridemia. *Lipids Health Dis* **15**, 118.
- 195. Gylling H, Plat J, Turley S, *et al.* (2014) Plant sterols and plant stanols in the management of dyslipidaemia and prevention of cardiovascular disease. *Atherosclerosis* **232**, 346–360.