



Technological University Dublin  
ARROW@TU Dublin

---

Articles

Radiation and Environmental Science Centre

---

2009-01-01

## Reflection contributions to the dispersion artefact in FTIR spectra of single biological cells.

Paul Bassan

*Technological University Dublin*

Hugh Byrne

*Technological University Dublin, [hugh.byrne@tudublin.ie](mailto:hugh.byrne@tudublin.ie)*

Joe Lee

*University of Manchester*

Franck Bonnier

*Technological University Dublin*

Colin Clarke

*Technological University Dublin, [Colin.Clarke@tudublin.ie](mailto:Colin.Clarke@tudublin.ie)*

Follow this and additional works at: <https://arrow.tudublin.ie/radart>

 [next page for additional authors](#)  
Part of the [Pharmaceutical, Toxicology and Environmental Health Commons](#)

---

### Recommended Citation

Bassan, Paul et al: Reflection contributions to the dispersion artefact in FTIR spectra of single biological cells. *The Analyst*, Vol.134, no. 6, pp.1171-1175, 2009. doi:10.1039/B821349F

This Article is brought to you for free and open access by the Radiation and Environmental Science Centre at ARROW@TU Dublin. It has been accepted for inclusion in Articles by an authorized administrator of ARROW@TU Dublin. For more information, please contact [yvonne.desmond@tudublin.ie](mailto:yvonne.desmond@tudublin.ie), [arrow.admin@tudublin.ie](mailto:arrow.admin@tudublin.ie), [brian.widdis@tudublin.ie](mailto:brian.widdis@tudublin.ie).



This work is licensed under a [Creative Commons Attribution-Noncommercial-Share Alike 3.0 License](https://creativecommons.org/licenses/by-nc-sa/3.0/)



---

**Authors**

Paul Bassan, Hugh Byrne, Joe Lee, Franck Bonnier, Colin Clarke, Paul Dumas, Ehsan Gazi, Michael Brown, Noel Clarke, and Peter Gardner

# Reflection contributions to the dispersion artefact in FTIR spectra of single biological cells

Paul Bassan<sup>a</sup>, Hugh J. Byrne<sup>b\*</sup>, Joe Lee<sup>a</sup>, Franck Bonnier<sup>b</sup>, Colin Clarke<sup>b</sup>, Paul Dumas<sup>c</sup>, Ehsan Gazi<sup>d</sup>, Michael D Brown<sup>d</sup>, Noel W Clarke<sup>d,e,f</sup>, Peter Gardner<sup>a</sup>

<sup>5</sup> Received (in XXX, XXX) Xth XXXXXXXXX 200X, Accepted Xth XXXXXXXXX 200X

First published on the web Xth XXXXXXXXX 200X

DOI: 10.1039/b000000x

Fourier transform infrared spectra of a single cell in transfection geometry are seen to vary significantly with position on the cell, showing a distorted derivative-like lineshape in the region of the optically dense nucleus. A similar behaviour is observable in a model system of the protein albumin doped in a potassium bromide disk. It is demonstrated that the spectrum at any point is a weighted sum of the sample reflection and transmission and that the dominance of the reflection spectrum in optically dense regions can account for some of the spectral distortions previously attributed to dispersion artefacts. Rather than being an artefact, the reflection contribution is ever present in transfection spectra and it is further demonstrated that the reflection characteristics can be used for cellular mapping.

## 15 Introduction

Fourier transform infrared (FTIR) spectroscopy has been demonstrated as a potential tool for disease diagnostics in tissue [1-3] and in its microscopical form, for cellular analysis [4-12]. The use of synchrotron sources and attenuated total reflection imaging have achieved subcellular resolution thus promising a powerful technique for the analysis of the biochemical origin of disease [7, 13, 14]. However, both tissue and cellular material are extremely spatially inhomogeneous on a micrometre scale in terms of physical morphology and chemical content and such inhomogeneity can give rise to elastic scattering of the incident radiation which is the origin of, a broad undulating background to spectra which has been attributed to Mie scattering [15]. In the case of cells, the cytoplasm is relatively sparse whereas the nucleus is rather dense in terms of biochemical content giving rise to a misrepresentation of the relative densities of biochemical constituents [16, 17]. These effects can lead to many ambiguous results depending on the geometry of the measurement. In diffuse reflection infrared Fourier transform (DRIFT) or transfection modes “contamination by contributions of reflection” can lead to further spectral distortions or so-called “dispersion artefacts” [18].

These confounding factors can result in significant frequency shifting of spectral features used to classify biological species for

diagnosis or biochemical analysis. It has been demonstrated that the broad Mie scattering contribution can be modelled [16] and even removed in an automated fashion using Extended Multiplicative Signal Correction algorithm [19]. The reflection “contamination” or dispersion artefact can be corrected for by back-transforming the spectra and extracting the real and imaginary components of the spectra resulting in a significantly reduced dispersive components [15].

In order to progress the potential of FTIR spectroscopy for biomedical applications it is imperative that the true origin of these factors be understood. This is especially true in an interdisciplinary field spanning the physical and life sciences and even the clinical community where terms such as “contamination” and “artefact” mask the true origin of the observed effects.

In this paper it is demonstrated that the spectral distortions observable in FTIR spectra in spatially inhomogeneous samples in a transfection geometry can have origin in a variation of the relative contributions of the reflection and transmission spectra of the material and that at high optical densities the spectrum can be dominated by the reflection component. In cellular spectra, far from being an artefact or contaminant, the pure reflection spectrum provides a biochemical map of the surface, free from artefacts, yielding subcellular information on the nucleus and cytoplasm.

## Experimental

Human metastatic prostate cancer (PC-3) cells were cultured onto calcium fluoride (CaF<sub>2</sub>) windows and fixed according to protocol outlined by Gazi et al [9]. A reflection line map was measured at the SOLEIL synchrotron facility (SMIS beamline) [20] to take advantage of the high brilliance of the source for small spot sizes. The spectra were recorded using the SMIS beamline and end-station which has been described elsewhere [20]. A projected aperture size of 10 × 10 μm<sup>2</sup> was used with a line map step size of 10 μm. 2048 scans were co-added using a spectral resolution of 8 cm<sup>-1</sup> to increase the signal to noise as the reflection signal is inherently weak in magnitude.

As a model sample, a 6% dispersion of the protein albumin in a KBr matrix was produced. 12mg of albumin were mixed

\* Author to whom correspondence should be addressed.

<sup>a</sup> School of Chemical Engineering and Analytical Science, Manchester Interdisciplinary Biocentre, The University of Manchester, 131 Princess Street, Manchester, M1 7DN, UK, E-mail:

<sup>45</sup> Peter.Gardner@Manchester.ac.uk

<sup>b</sup> Focas Research Institute, Dublin Institute of Technology, Kevin Street, Dublin 8, Ireland.

<sup>c</sup> Synchrotron SOLEIL, L'Orme des Merisiers, BP48 - Saint Aubin, 91192 Gif-sur-Yvette Cedex, France.

<sup>50</sup> Genito Urinary Cancer Research Group, School of Cancer and Imaging Sciences Paterson Institute for Cancer Research, University of Manchester, Wilmslow Road, Manchester, UK, M20 4BX, UK

Department of Urology, The Christie NHS Foundation Trust, Manchester, UK, M20 4BX, UK

<sup>55</sup> Department of Urology, Salford Royal NHS Foundation Trust, Salford, UK, M6 8HD, UK

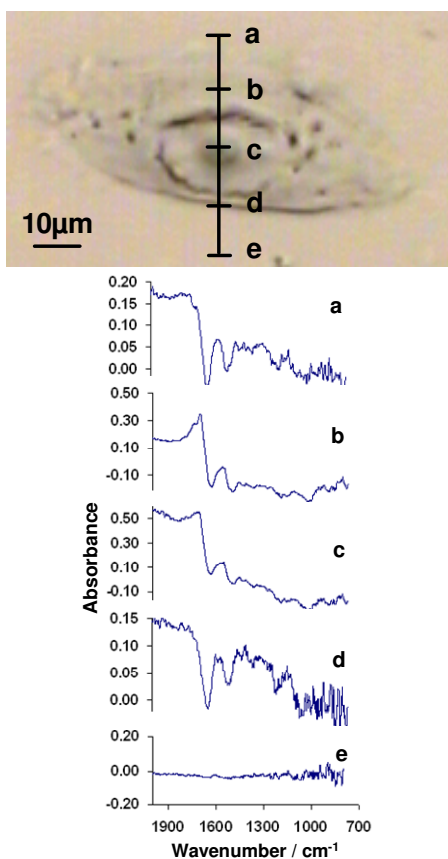


Figure 1: (a) Optical image of PC3 cell on a CaF<sub>2</sub> substrate. (b) Spectra of the cell at the points indicated in (a) expressed as absorbance.

with 188 mg of KBr powder. The two powders were dried in a vacuum oven overnight to decrease as much as possible the contribution of water in the spectra before being pressed into a pellet.

A pellet containing only KBr (200mg) was prepared and used as the reference sample for transmission measurements. A gold substrate was employed as a reference of 100% in reflection mode and transfection mode. While the pellets were freestanding for reflection and transmission measurements, in the case of transfection measurements the pellets were placed on a gold surface to obtain a full reflection of the infrared beam.

In the case of measurements of the model albumin/KBr material, a Perkin Elmer Spotlight 400 was employed. The single point mode was used to record the spectra with an aperture size set at  $100 \times 100 \mu\text{m}^2$ . A map was defined with a step size of  $100 \mu\text{m}$ . 32 scans were co-added using a spectral resolution of  $4 \text{ cm}^{-1}$ .

## Results and discussion

As an example of the complexity of FTIR spectra which can be obtained from FTIR micro-spectroscopy in the transfection geometry, and the variability which can occur as a result of spatial inhomogeneities, Fig. 1 shows a spectral line map of a PC-3 cell taken at  $10 \mu\text{m}$  step intervals. Measurements were taken in transfection mode and are expressed as Absorbance. The background was recorded from a bare CaF<sub>2</sub> disk and thus the spectrum indicates the changes in cellular reflection (specular and/or diffuse) relative to the bare CaF<sub>2</sub>. For all positions, the spectra are relatively noisy as the reflection from the

cell/substrate interface is expected to be weak as the refractive index of the CaF<sub>2</sub> substrate ( $1.387$  at  $1696 \text{ cm}^{-1}$ ) [21] is only slightly larger than that of the cellular material normally considered to be about 1.3 [22]. At position (b), the spot is large enough that it samples some of the cytoplasm and the transfection spectrum shows typical cellular features associated with proteins (Amide I  $\sim 1650 \text{ cm}^{-1}$ , Amide II  $1565 \text{ cm}^{-1}$ ). In the region of the nucleus, however, (c,d), the spectra have become significantly distorted and resemble a first order derivative lineshape. It is these types of distortions which have previously been attributed to dispersion artefacts and interpretation of the spectra for diagnostic applications or biochemical analysis is difficult as changes to physical morphology can mask changes in chemical composition.

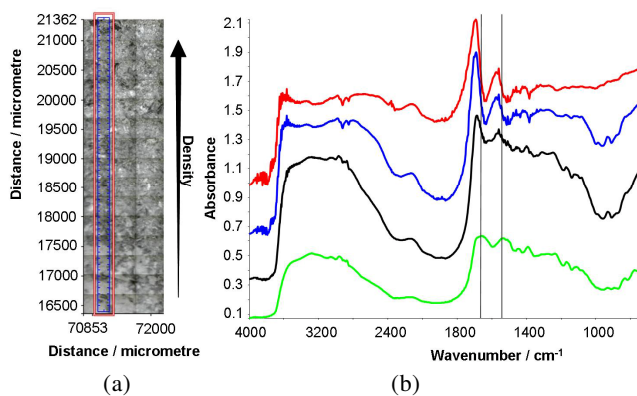


Figure 2: (a) Microscope image of 6% loading albumin/KBr pellet. The red/blue rectangles indicate the regions spectroscopically sampled. (b) Variation in the FTIR transmittance spectrum, expressed as an absorbance as the sampled area varies from the low density to the high density region.

The inhomogeneity of the cellular sample can be mimicked by a dispersion of the protein albumin loaded in a KBr matrix. Figure 2a shows a microscopic image of the sample with 6% loading of albumin in KBr. Within the matrix, the protein is inhomogeneously dispersed, mimicking the spatial inhomogeneity of a cellular material. Figure 2b maps the transfection spectrum, expressed as absorbance, as the measurement position is moved from a region of low albumin density to one of high density. Similar to the case of the cellular line map of figure 1, the spectrum becomes highly distorted in the regions of high density, exhibiting distorted lineshapes in the region of the amide I band characteristic of “dispersion artefacts”. The position of the absorption maxima at, for example  $\sim 1675 \text{ cm}^{-1}$  and  $1560 \text{ cm}^{-1}$ , indicated by the vertical lines in figure 2b, clearly shifts in frequency. In contrast to the case of the cellular spectra, however, the inhomogeneity is purely morphological and there can be no question of spectral changes due to variations in chemical composition.

The variations in the spectral form can be simply understood by considering the relative contributions to the spectrum of the transmitted and surface reflected light. Transfection experiments are commonly performed by placing a sample on a highly reflective substrate, focusing an IR beam onto a region of interest

and collecting the photons that are reflected back into the collection optics. The method is called transflection, because the majority of the signal collected is a transmission signal as the beam passes through the sample, reflects off the substrate, passing through the sample again to the detector as shown in figure 3. As the base substrate is highly reflecting across the spectral region of interest, it is assumed that the spectrum gathered is dominated by the transmission characteristics of the sample under investigation. However, the gathered signal also contains a contribution from the top interface between the air and the sample. The total signal can therefore be approximated by

$$I_{Tf} = 2T + R \quad (1)$$

where  $I_{Tf}$  is the gathered transflection signal,  $T$  the single pass transmission and  $R$  the reflection. The transflection signal therefore consists of a weighted sum of the transmission and reflection characteristics of the sample. In samples where the optical density is low, the weighting is such that the transmission signal dominates and the reflection signal is

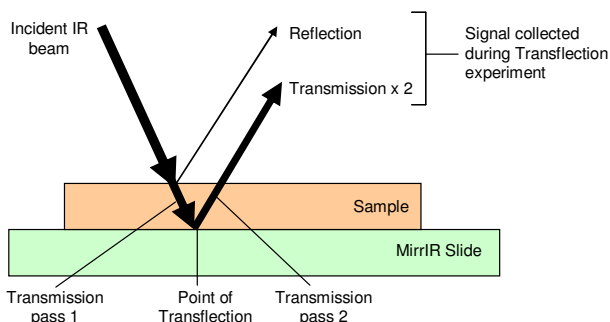


Figure 3. Schematic of a typical transflection experiment. The input beam is drawn at an arbitrary incident angle for clarity.

negligible, producing spectra that closely represent pure transmission data. If the transmission signal is weak or extinct, however, the reflection signal will dominate and a reflectance spectrum is measured.

The optical properties of a material are governed by the complex refractive index [see for example 19]

$$\eta = n + i\kappa = n + i\alpha/2k_0 \quad (2)$$

where  $\alpha$  is the absorption coefficient and  $k_0$  is  $2\pi/\lambda_0$ , such that as a function of driving frequency  $\omega$  for a simple driven electric dipole oscillator of resonance frequency  $\omega_0$ ,

$$n = 1 + \frac{Ne^2}{4\epsilon_0 m \omega_0} \frac{(\omega_0 - \omega)}{(\omega_0 - \omega)^2 + (\gamma/2)^2} \quad (3)$$

$$\alpha = \frac{Ne^2 \pi}{4\epsilon_0 m c} \frac{\gamma/2\pi}{(\omega_0 - \omega)^2 + (\gamma/2)^2} \quad (4)$$

where  $\gamma$  is the usual Lorentz damping factor.

Equations (3) and (4) constitute the Kramers-Kronig dispersion relations between the absorption and dispersion and can be shown graphically as in Figure 4. Equation (3) is calculated for the case of a gaseous media at low pressures such that the off resonant real component approximates to 1. For the sake of the simulation of organic materials, this off resonant value is taken as 1.3 in this simulation.

The reflection from the interface of two materials, in the absence of absorption, is given by

$$R = \left( \frac{n_1 - n_2}{n_1 + n_2} \right)^2 \quad (5)$$

where  $n_1$  is the refractive index of the interface medium (in the case of interest here, air) and  $n_2$  is that of the reflecting medium (cell). Assuming  $n_2$  has a value of 1.3, the nonresonant reflection of an air cell interface can be calculated to be 0.017 or 1.7%.

The sample transmission is given by

$$T = \exp(-\alpha l) \quad (6)$$

where  $l$  is the thickness of the sample. Both the transmission and reflection signals are frequency dependent and the spectra are a manifestation of the electronic states within the material.

The effect of various weightings of the transmission and reflection signals can be simulated from the optical parameters of the idealized simple driven electric dipole oscillator shown in Fig. 4. Fig. 5 shows the variation of such a simulated transflection spectrum, expressed as absorbance (as would commonly be the

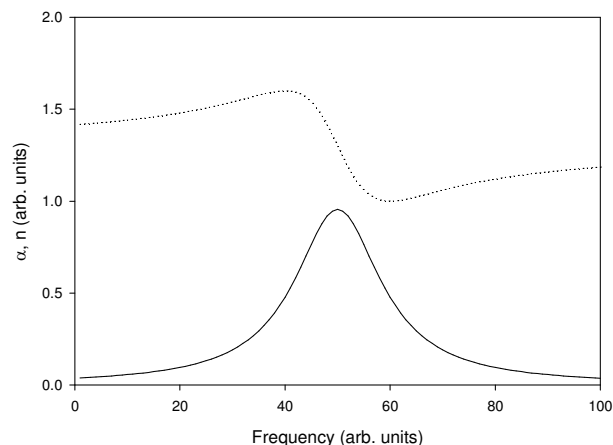


Figure 4 – Simulated absorption  $\alpha(\omega)$  and dispersion  $n(\omega)$ , as a function of frequency  $\omega$ , in the region of a spectroscopic transition of resonance frequency  $\omega_0=50$  (arb. units).

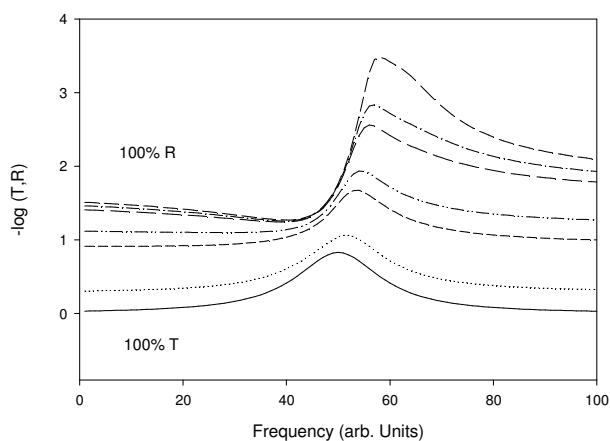


Figure 5. Simulated variation in the spectral profile as a result of weighted sum of the reflection and transmission from a simple harmonic oscillator with resonance frequency  $\omega_0=50$  (arb. units).

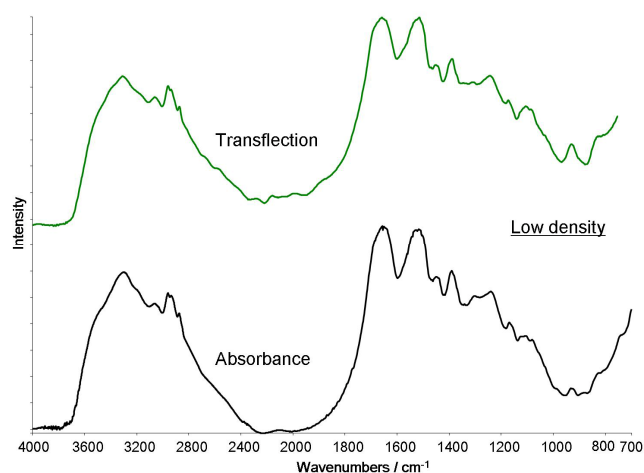


Figure 6: Transflection of albumin/KBr sample as measured in the low density regions of the sample, compared with the absorbance as measured in transmission mode. The spectra are offset for clarity.

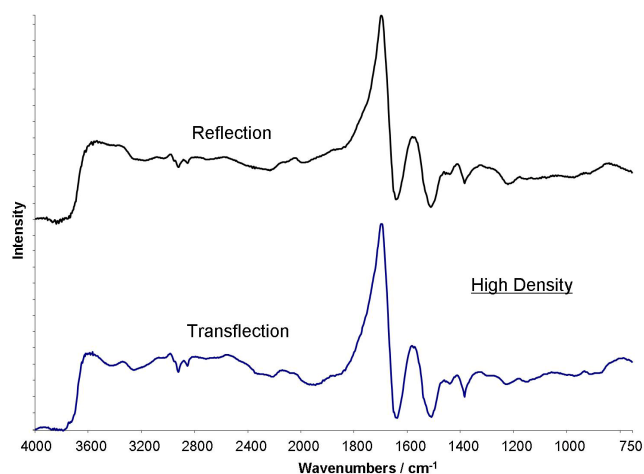


Figure 7: Reflection spectrum of albumin/KBr plate compared with the transflection spectrum in the high density regions. Spectra are offset for clarity.

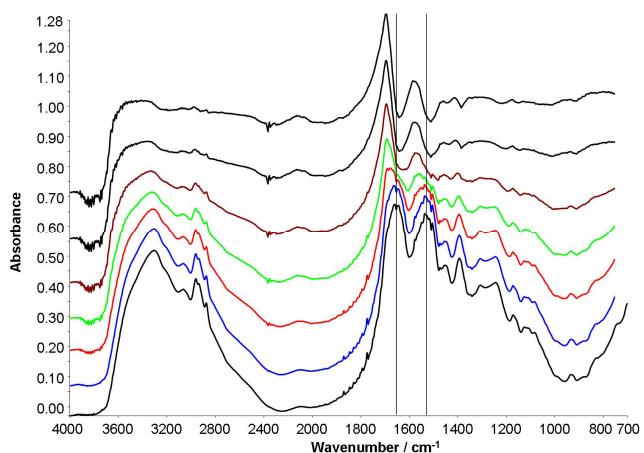


Figure 8 - Simulation of spatial variation of the transflection spectrum of Albumin in KBr using weighted sums of the reflection and transmission spectra, from 100%R (top) to 100%T (bottom).

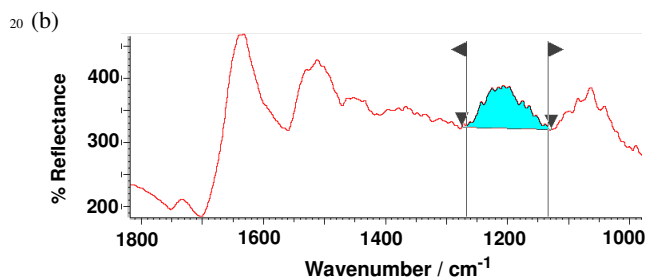
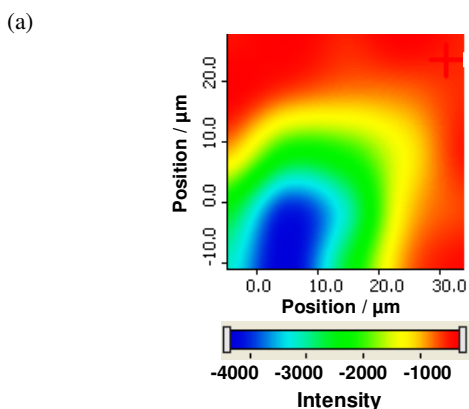


Figure 9 (a) Map of a PC-3 cell on CaF<sub>2</sub> based on the integrated band at 1240cm<sup>-1</sup> shown in (b). Note that blue indicates highest reflectance, red lowest.

case in an FTIR transflection measurement) as the relative weightings  $R\%:T\%$  of the calculated Reflection and Transmissions are varied from 0%:100% (bottom) to 100%:0% (top). It is clear that by effectively changing the optical thickness of the “sample” the spectral lineshape varies considerably, from being initially dominated by the absorption characteristics to being dominated by the reflection. In terms of the distortion of the pure absorbance feature, the increased relative contribution of the reflection component affects spectral changes similar to those termed the dispersion artefact. It is stressed, however, that this is not an artefact, but a contribution of the reflection due to the real component of the refractive index.

In the transflection geometry, therefore, the measured signal is a sum of the sample transmission and the reflection from the top surface. Fig 6 compares the spectrum measured in transmission mode, expressed as absorbance, of the albumin/KBr sample in regions of low density, with the transflection spectrum of the low density region of figure 2. An almost identical spectral match is found. In the high density regions, however, the spectrum becomes dominated by the reflection spectrum, as seen in Fig 7. It should be noted that in such a spatially inhomogeneous system, diffuse reflectance may contribute as much as specular. The transflection spectrum matches almost identically with the spectrum measured in reflection mode. Although the quality of the spectra in terms of signal to noise varies considerably, the spectral shape in reflection mode is found to be independent of albumin density.

As shown in figure 8, in the intermediate regime, the transflection spectrum can be well fitted with a weighted sum of the high density reflection spectrum and the low density reflection spectrum such that

$$I_{Tf} = aT + bR \quad (7)$$

In the examples presented in Figure 1, a low reflectivity substrate, CaF<sub>2</sub>, was employed to minimise the transmission contribution to the transfection spectrum in order to highlight the contribution of the reflection spectrum. Normally, a high reflection (“low e”) substrate such as MirrIR is employed to maximise the transmission component hence minimising the contribution of the reflection spectrum. However, this minimisation leads to the ever present contribution being overlooked or dismissed as a “contamination”. The reflection spectrum has origin in the chemical constituents of the material, however, and is potentially as valuable a characterisation tool as the material absorption.

Fig 9 maps the 1240 cm<sup>-1</sup>, PO<sub>2</sub>, Phosphate DNA backbone, and protein PO<sub>2</sub> groups, which should be dominant in the nucleus of the cell. Although the spectra in the region of the nucleus are dominated by the surface reflection, as indicated by the first derivative type lineshape, it is clear that the map contains information on the nucleus. In a dielectric material, the sampling depth of the pure reflection component is of the order of the wavelength of the incident radiation which in the region of the DNA bands at 1240 cm<sup>-1</sup> corresponds to ~8 microns. Such a penetration depth corresponds well with typical nuclear dimensions. Rather than being a contamination, therefore, the suggestion is that the reflection spectrum of the material is a relatively uncontaminated representation of the biochemical content and variation of the sample under investigation, although it is acknowledged that the signal to noise is considerably compromised and therefore sampling times required would be significantly increased.

## Conclusions

It has been demonstrated that, in a transfection geometry, a Fourier Transform infrared spectrum is a weighted sum of the sample transmission and reflection spectra of the sample. The contribution of the reflection spectrum can become dominant in regions of high optical density and produces spectral distortions characteristic of the so-called “dispersion artefact”. In the transfection geometry, the effect is in reality not dominated by dispersion and is not an artefact, and it has been shown that the reflection spectrum can be used as a biochemical mapping technique in its own right. Recognising that it is omni-present to varying degrees in transfection spectra prompts a move to either reflection or transmission only geometries.

This study has taken an extreme example of a low reflectivity substrate to minimise the transmission contributions for illustrative purposes. In minimising the transmission contributions, transmission dispersive artefacts are also minimised. The origin of the dispersion artefact in purely transmission geometries is currently being explored [23]. Such an elucidation could lead to an evolution of data processing protocols such as EMSC to fully incorporate resonant reflection and scattering contributions such that the real and imaginary components can be deconvolved, yielding a reliable and interpretable absorption spectrum of cellular samples.

## Acknowledgements

This study derived from discussions of the Single Cell Spectroscopy Working Group of the EU FP6 SSA DASIM, Contract Number 005326. The work at DIT is associated with NBIPI, funded by the Irish Government's Programme for Research in Third Level Institutions, Cycle 4, National Development Plan 2007-2013.

## References

- 1 R. Dukor, *Vibrational spectroscopy in the detection of cancer*, Handbook of Vibrational Spectroscopy, Ed J. M. Chalmers and P.R. Griffiths Vol5 2002, Chichester: Wiley 2002
- 2 E. Gazi, M. Baker, J. Dwyer, N. P. Lockyer, P. Gardner, J.H. Shanks, R. S. Reeve, C. Hart, N.W. Clarke M. Brown, *European Urology*, 2006 **50**, 750–761
- 3 D. C. Fernandez, R. Bhargava, S. M. Hewitt, I. W. Levin, *Nature Biotechnol.*, 2005, **23**, 469–474
- 4 N. Jamin, P. Dumas, J. Moncuit, W.H. Fridman, J.L. Teillaud, L.G. Carr, G.P. Williams, *Proc. Natl. Acad. Sci. USA* 1998, **95**, 4837–4840.
- 5 P. Lasch, A. Pacifico, M. Diem, *Biopolymers* 2002, **67**, 335–338.
- 6 P. Lasch, M. Boese, A. Pacifico, M. Diem, *Vibrational Spectrosc.* 2002, **28**, 147–157
- 7 P. Dumas, L. Miller, *Vibrational Spectrosc.* 2003, **32** 3–21.
- 8 P. Dumas, N. Jamin, J.L. Teillaud, L.M. Miller, B. Beccard, *Faraday Discuss.* 2004, **126**, 289–302.
- 9 E. Gazi, J. Dwyer, N.P. Lockyer, P. Gardner, J. Miyan, C.A. Hart, M.D. Brown, J.H. Shanks, N.W. Clarke, *Biopolymers* 2005, **77**, 18–30.
- 10 E. Gazi, J. Dwyer, N.P. Lockyer, J. Miyan, P. Gardner, C.A. Hart, M.D. Brown, N.W. Clarke, *Vibrational Spectrosc.* 2005, **38**, 193–201.
- 11 E. Gazi, P. Gardner, N.P. Lockyer, C.A. Hart, N.W. Clarke, M.D. Brown, *J. Lipid Research* 2007, **48**, 1846
- 12 D. A. Moss, M. Keese, R. Pepperkok, *Vibrational Spectroscopy*, 2005, **38**, 185–191
- 13 P. Dumas, M. J. Tobin, *Spectroscopy Europe*, 2003 **15** 17–23
- 14 P. Dumas, G. D. Sockalingum J. Sulé-Suso, *Trends in Biotechnology*, 2007, **25**, 40–44
- 15 M. Romeo, M. Diem, *Vibrational Spectroscopy*, 2005, **38**, 129–132
- 16 M. Romeo B. Mohlenhoff and M. Diem, *Vibrational Spectroscopy*, 2006, **42**, 19–14
- 17 J. Lee, E. Gazi, J. Dwyer, M. D. Brown, N. W. Clarke, J. Nicholson, P. Gardner, *Analyst*, 2007 **132** 750 - 755
- 18 M. Romeo and M. Diem, *Vibrational Spectroscopy*, 2005, **38**, 115–119
- 19 S.N. Thennadil, H. Martens, A. Kohler, *Applied Spectroscopy*, 2006, **60**, 315–321
- 20 P. Dumas, F. Polack, B. Lagarde, O. Chubar, J. L. Giorgetta, S. Lefrancois, *Infrared Physics & Technology* 2006 **49** 152–160
- 21 CRC Handbook of Chemistry and Physics, 62<sup>nd</sup> Edition 1981 CRC Press Inc, Boca Raton, Florida
- 22 A. Kohler, J. Sulé-Suso, G. D. Sockalingum, M. Tobin, F. Bahrami, Y. Yang, J. Pijanka, P. Dumas, M. Cotte, D. G. van Pittius, G. Parkes, and H. Martens, *Applied Spectroscopy*, 2008 **62**, 259–266
- 23 Paul Bassan, Hugh J. Byrne, Frank Bonnier, Joe Lee, Paul Dumas, Peter Gardner, *The Analyst*, submitted March 2009.