



Technological University Dublin
ARROW@TU Dublin

Articles

School of Physics & Clinical & Optometric
Science

2017

Broadband Acoustic Measurement of the Agar-based Tissue Mimicking Material: a Longitudinal Study

A. Rabell-Montiel
University of Edinburgh

Jacinta Browne
Technological University Dublin, jacinta.browne@tudublin.ie

S. D. Pye
Royal Infirmary of Edinburgh

T. A. Anderson
University of Edinburgh

C. Moran
University of Edinburgh
Follow this and additional works at: <https://arrow.tudublin.ie/scschphyart>

 Part of the [Physics Commons](#)

Recommended Citation

Montiel, A. et al . (2017). Broadband acoustic measurement of the agar-based tissue mimicking material: a longitudinal study. *Ultrasound in Medicine & Biology*, 43(7), pp.1494-1505. doi:10.1016/j.ultrasmedbio.2017.02.019

This Article is brought to you for free and open access by the School of Physics & Clinical & Optometric Science at ARROW@TU Dublin. It has been accepted for inclusion in Articles by an authorized administrator of ARROW@TU Dublin. For more information, please contact yvonne.desmond@tudublin.ie, arrow.admin@tudublin.ie, brian.widdis@tudublin.ie.



This work is licensed under a [Creative Commons Attribution-NonCommercial-Share Alike 3.0 License](#)



1 **Broadband acoustic measurement of the agar-based tissue mimicking**
2 **material – a longitudinal study.**

3 Rabell-Montiel A¹, Browne J. E.², Pye S. D.³, Anderson T. A.¹ and Moran C. M.¹

4 ¹Centre for Cardiovascular Science, University of Edinburgh, Edinburgh, UK

5 ²School of Physics & IEO, FOCAS, Dublin Institute of Technology, Dublin, Ireland

6 ³Medical Physics, NHS Lothian, Royal Infirmary of Edinburgh, Edinburgh, UK

7

8 Corresponding author: Adela Rabell-Montiel

9 Email: adela.rabell@ed.ac.uk

10 Present Address: 47 Little France Crescent, Queen's Medical Research Institute,
11 Centre for Cardiovascular Science, EH16 4TJ, Edinburgh, UK

12 Mobile Phone: +447983126239

13 Office Phone: +441312429219

14 ABSTRACT

15 Commercially available ultrasound quality assurance test phantoms rely upon the long-term
16 acoustic stability of tissue-mimicking-materials (TMMs). The measurement of the acoustic properties
17 can be technically challenging and it is important to ensure its stability. The standard technique is to
18 film-wrap samples of TMM and to measure the acoustic properties in a water bath. In this study, a
19 modified technique is proposed whereby the samples of TMM are measured in a preserving fluid that
20 is intended to maintain their characteristics. The acoustic properties were evaluated using a
21 broadband pulse-echo substitution technique over the frequency range of 4.5 – 50 MHz at 0, 6 and
22 12 months using both techniques. For both techniques, the measured mean values for the speed of
23 sound and the attenuation were very similar and within the IEC recommended value. However, the
24 results obtained using the proposed modified technique demonstrated greater stability over the 1-
25 year period when compared with the results acquired using the standard technique.

26

27 *Key words:* ultrasound, high frequency, tissue mimicking material, speed of sound, attenuation
28 coefficient, long-term.

29 INTRODUCTION

30 Commercially available quality assurance (QA) test phantoms are widely used to test the
31 performance of clinical ultrasound scanners. These phantoms, are manufactured from tissue-
32 mimicking-material (TMM) which is designed to closely match the acoustical properties of the speed
33 of sound (SoS) and the attenuation coefficient from soft tissue. The aim of these phantoms is to
34 provide a reproducible method to assess the performance of ultrasound scanners. However, these
35 phantoms are intended for use with clinical ultrasound scanners at frequencies up to 20 MHz. To the
36 best of our knowledge, there are no commercially available test phantoms to assess the performance
37 of ultrasound scanners employing ultrasound frequencies above 20 MHz.

38 A variety of TMM materials are currently produced both commercially and within
39 laboratories. These include: agar-based TMMs (Teirlinck et al., 1998), condensed milk TMMs
40 (Madsen et al., 1998), gelatine TMMs (Culjat et al., 2010), konjac-carrageenan TMMs (Kenwright et
41 al., 2014; Meagher et al., 2007), urethane rubber TMMs (Culjat et al., 2010), Poly (vinyl alcohol)
42 Cryo-gel (PVA-C) TMMs (Cournane et al., 2010; Culjat et al., 2010; King et al., 2011) and Zerdine™
43 TMMs (CIRS, Inc. Norfolk, VA). The International Electrotechnical Commission (IEC) agar-based TMM
44 has become widely used and popular for clinical and preclinical test objects (Brewin et al., 2008;
45 Browne et al., 2003; Cannon et al., 2011; Culjat et al., 2010; Inglis et al., 2006; Moran et al., 2011;
46 Rajagopal et al., 2014; Sun et al., 2012; Yang et al., 2013). The acoustical properties of this agar-
47 based TMM are designed to comply with the ultrasound acoustical parameters provided by the IEC
48 (IEC, 2001) with the recommended SoS and attenuation over the frequency range 2 – 10 MHz being
49 $1540 \pm 15 \text{ ms}^{-1}$ and $0.5 \pm 0.05 \text{ dB cm}^{-1}$ respectively.

50 Moderately high-frequency ultrasound scanners (up to 20 MHz) have been manufactured for
51 many years and have been utilised clinically in the assessment of skin (Machet et al., 2009), vascular
52 structures (Rhee, 2007) and retinal imaging. In recent years, reliable high (20 – 50 MHz) and very
53 high (>40 MHz) ultrasound scanners have become mainstream technology for the imaging of

54 superficial structures in clinical imaging and for preclinical imaging applications due to improvements
55 in transducer engineering and software technology (Banchhor et al., 2016; Schmitt et al., 2010;
56 Sundholm et al., 2015; Xu et al., 2012).

57 With the increase of high-frequency ultrasound imaging applications, there is a need to
58 develop and to acoustically characterise TMMs suitable for high-frequency ultrasound QA and
59 training phantoms. It has been shown that above 10 MHz the TMMs in the commercial test
60 phantoms, do not have optimum acoustic properties as the attenuation starts to exhibit a nonlinear
61 response with increasing frequency (Browne et al., 2003), whereas the IEC guidelines for TMM
62 properties recommend a linear relationship between attenuation and frequency up to 10 MHz.

63 The agar-based TMM developed under the IEC guidelines and used in this study, has
64 previously been found to have a non-linear response when acoustically investigated at frequencies
65 up to 23 MHz by Brewin et al., (2008), in our own lab and in the National Physical Laboratory at
66 frequencies up to 47 MHz and 60 MHz respectively (Sun et al., 2012 and Rajagopal et al., 2014). In
67 these studies, the use of test cells or TMM samples wrapped with film material (Saran Wrap® or
68 Mylar®) was employed to preserve the samples during acoustic characterisation when degassed,
69 deionised water was used as the reference medium. Moreover, thin slices of TMM ranging in
70 thickness from 2.5 – 30mm were used, enabling higher ultrasound frequencies to propagate through
71 the TMM slices (Brewin et al., 2008; Rajagopal et al., 2014; Sun et al., 2012). The encasing of the
72 TMM in film is important as, without the film, the TMM will degrade rapidly. This degradation is due
73 to leaching of the glycerol from the TMM into the water reference medium, thus altering the acoustic
74 properties of the TMM (Brewin et al., 2008). A reference water test cell, also encapsulated in Saran
75 Wrap® or Mylar® film, was used in the reference measurement in order to account for the effect of
76 the film on measurements (Rajagopal et al., 2014; Sun et al., 2012). However, the production of both
77 the TMM slices wrapped in film and water test cells is time-consuming and technically challenging,
78 especially for thin TMM samples. Therefore, the aim of this study was to compare this well-

79 established technique for the measurement and preservation of an IEC agar-TMM to a technique
80 where TMM is characterised and preserved in a preserving fluid. Furthermore, this method was
81 evaluated over a 1-year period, to determine the longitudinal stability of the acoustic properties.

82 MATERIALS AND METHODS

83 *Acoustical measurements*

84 Data was captured using two different acoustical systems, described briefly here and
85 elsewhere (Sun et al., 2012). Firstly, a Vevo 770® preclinical ultrasound scanner (Visualsonics Inc.,
86 Toronto, Canada) was used at the University of Edinburgh and secondly, a Scanning Acoustic
87 Macroscopy (SAM) system developed in-house at the Dublin Institute of Technology (Cannon et al.,
88 2011). The SAM system was used to provide additional acoustic data and to extend the lower limit of
89 the bandwidth of the measurements to 4.5 MHz.

90 *Manufacture of UTMMs and FTMMs samples.*

91 A batch of the IEC agar-based TMM was manufactured following a widely used standard
92 recipe and method (Brewin et al., 2008; Browne et al., 2003; Cannon et al., 2001; Ramnarine et al.,
93 2001; Teirlinck et al., 1998). This mixture was poured at 42°C onto a pre-warmed metal plate. The
94 plate was pre-warmed to ensure that the mixture spread uniformly. The TMM mixture was then left
95 to cool to room temperature. From this batch of TMM, 22 cylindrical slices of TMM of diameter
96 5.5cm were cut using a thin-walled plastic tube. Due to the short focal lengths associated with high
97 frequency transducers (Table 1), the thickness of the TMM slices was constrained to less than 3.2mm
98 and ranged in value from 1.8mm – 3.2 mm.

99 After being cut, eleven of these cylindrical TMM samples were left uncovered, and placed in a
100 sealed container with TMM preserving fluid. This TMM preserving fluid was manufactured in-house
101 (Brewin et al., 2008; Cannon et al., 2011; Inglis et al., 2006). These samples will be referred to as
102 unwrapped-TMM (UTMM) (Figure 1a).

103 The remaining eleven TMM samples were used to manufacture the samples which were
104 subsequently covered with clear film in the following manner. Initially, a layer (0.015mm thick) of
105 Saran Wrap® film (SC Johnson Inc., Racine, USA) was stretched over an embroidery ring, of 10cm

106 diameter. A fast-hardening epoxy (Araldite Rapid; Huntsman Advanced Materials, Basel, Switzerland)
107 was then applied to one side of a PVC ring (2mm thick, 5.8mm outer diameter) and the stretched
108 Saran Wrap® was lowered onto the PVC ring. This was left to set overnight. The TMM was then
109 manufactured as described above. After setting and cutting, the eleven samples were placed into
110 the PVC rings. Five drops of TMM preserving fluid were added to the surface of the TMM to ensure
111 good acoustic coupling between the film and the TMM, then a second layer of Saran Wrap® was
112 glued to the other side of the ring, similar to that described above, such that the TMM slices were
113 “sandwiched” between the two films and thereby, film-wrapped-TMM (FTMMs). These final film-
114 wrapped samples were left to set overnight. Finally, epoxy was used to seal the edges of the film-
115 ring-film to ensure the FTMMs did not leak. This was re-enforced with insulating tape to ensure that
116 the film would not peel off during the 1-year period of investigation (Figure 1b). These FTMMs were
117 preserved in a box with tissue paper moistened with TMM preserving fluid to create a humid TMM
118 preserving fluid saturated environment. In a similar manner, a water test cell was manufactured
119 whereby the TMM was replaced by degassed deionized water in the manufacturing process.

120 *Experimental set-up of Vevo 770® preclinical ultrasound scanner.*

121 In this study, the radio frequency (RF) data was collected and analysed from 11 FTMMs and
122 11 UTMMs. To measure the acoustic properties, the FTMMs were submerged in a tank filled with
123 degassed, deionised water as the reference medium, while for the UTMM measurements, the tank
124 was filled with TMM preserving fluid. For both measurements, a Polymethylpentene (TPX, Boedeker
125 Plastics, Texas, USA reflector) of 2.5cm diameter and 5mm thickness was located beneath the
126 samples at the focal position of each transducer as illustrated in Figure 2.

127 Measurements were made using 4 transducers (Table 1) at 10% output power, using a high-
128 frequency ultrasound scanner Vevo 770®. This power output was considered sufficient signal
129 magnitude to obtain good signal-to-noise data without the generation of significant nonlinear
130 effects (Sun et al., 2012). The regions of interest (ROI) were located at the upper surface of the TPX

131 reflector with and without the sample in place and from the lower and upper surfaces *of each*
132 *sample. For each measurement the RF data was collected from 10 scan-lines within these pre-*
133 *selected ROIs at 4 different positions on the FTMMs or UTMMs. The data was* analysed off-line using
134 a MatLab script (MatLab R2013a MathWorks, Inc). The calculated angular separation between the RF
135 acquisition lines is approximately 0.15°, so the lines were considered parallel and perpendicular to
136 the TPX reflector. For the FTMMs, identical acoustic measurements were also taken through the
137 water test cell to take into account potential reflections from the Saran Wrap® interfaces and to
138 obtain absolute values of attenuation (Cannon et al., 2011; Sun et al., 2012).

139 The performance of both FTMMs and UTMMs were assessed over a period of one year at
140 *approximately* 0, 6 and 12 month time points.

141 The 3dB bandwidth of each transducer (Table 1) was measured from the frequency spectra
142 taken from the TPX reflector in a degassed, deionized water tank without the sample in place, where
143 the reflector was placed at the focal length of the transducer.

144 *Analysis of speed of sound, thickness and attenuation data of FTMMs and UTMMs samples.*

145 *The analysis of the FTMMs was performed based on a broadband pulse-echo substitution*
146 *technique (AIUM, 2014). The pulse-echo return times from the front and rear surfaces of the FTMMs*
147 *and from the front surface of the TPX reflector were used to determine the thickness and SoS of the*
148 *samples. The magnitude of these pulse-echoes were used to calculate the attenuation.* In addition,
149 the echoes from the TPX reflector with the water test cell was acquired. This data was then used to
150 calculate the SoS, thickness and attenuation of the FTMMs samples in a manner similar to that in
151 Sun et al., (2012).

152 For the SoS and thickness of the UTMMs, the measurement technique and analysis was
153 carried out in a similar manner to the FTMMs. However, since the UTMMs were not wrapped in
154 Saran Wrap®, no water test cell was required and the reference measurements from the TPX were

155 taken through the bath of TMM preserving fluid. The absence of Saran Wrap® meant that the
 156 magnitude of the echoes from the boundaries of the TMM was reduced. This, necessitated manual
 157 selection of the position of the boundaries from the raw RF signals. This was performed by selection
 158 of the largest pulse-echo at each interface of the UTMMs. The criterion for this was that the peak
 159 selected was at least 100% greater than the magnitude of the previous peak within 2 μs time-
 160 window. This was carried out for each of the 10 lines of the raw RF signal inside the ROI of the
 161 UTMMs, enabling the thickness of the sample to be determined at each of these positions. In
 162 addition, the thickness values of the UTMMs were measured manually in five different positions on
 163 each UTMM with a digital calliper (DURATOOL, TM Taiwan, 0 – 150mm). Each sample was placed
 164 between 2 plastic plates of known thickness in order to avoid excessive compression of the UTMMs
 165 (Brewin et al., 2008; Rajaqopal et al., 2014). This was performed at the initial time-point before the
 166 acoustic measurements commenced.

167 The SoS in the UTMMs was calculated using Equation (1).

$$SoS_{TMM} = \left(1 + \frac{T_R - T_{TMMR}}{T_{TMMLo} - T_{TMMUp}} \right) SoS_{TMMfluid} \quad (1)$$

168 Where T_R is the time from the transducer to the TPX in the tank with no sample in the acoustical
 169 path, T_{TMMR} is the time from the transducer to the TPX reflector through the sample, T_{TMMLo} and
 170 T_{TMMUp} are the time from the transducer to the lower and upper surface respectively of the sample
 171 and $SoS_{TMMfluid}$ is the SoS of the TMM preserving fluid in which the samples were immersed.

172 Calculation of the attenuation of the UTMMs was carried out in a similar manner to the
 173 FTMMs, but without the use of the water test cell, to compensate for the interfacial attenuation
 174 loss.

175 Acoustical properties of TMM preserving fluid and degassed, deionized water.

176 The acoustic properties of the TMM preserving fluid were measured by the National Physical
177 Laboratory (Teddington, UK). The SoS was found to be $1538.15 \pm 0.22 \text{ ms}^{-1}$ at $19.3 \pm 0.1^\circ\text{C}$ and a 2nd
178 degree polynomial function was fitted to the attenuation data (α [dB cm⁻¹]) as a function of
179 frequency (f (MHz)): as $\alpha_{TMMfluid} = 0.00309f^2 - 0.004996f$ ($R^2=0.99$) over the frequency range
180 of 1 – 60 MHz.

181 The acoustic properties of the degassed deionized water have previously been measured
182 and found to have an attenuation proportional to f^2 over the range of 7.5 – 67.5 MHz (Duck, 1990;
183 Pinkerton, 1949). Furthermore, the SoS of degassed deionized water varies with temperature
184 (Bilaniuk et al., 1992; Del Grosso et al., 1972). In this study all measurements were undertaken at
185 $22.2 \pm 0.5^\circ\text{C}$ with a SoS of 1488.88 ms^{-1} .

186 *Experimental set-up and data analysis using the SAM system (Dublin Institute of Technology).*

187 The SAM system uses broadband transducers which work as both transmitter and receiver
188 (Olympus NDT Inc., Waltham, MA, USA). The experimental setup was similar to that used with the
189 Vevo 770[®] (Figure 2). Three different transducers were used (Table 1) where the 3dB bandwidth was
190 measured in a similar manner to that of the transducers of the Vevo 770[®].

191 *A pulser-receiver (Model 5052PR; Panametrics, Waltham, MA, USA) was used to transmit*
192 *and receive the pulses. The received reflected pulse was digitised and captured using a data*
193 *acquisition card (PCI-5144: National Instruments, Austin, TX, USA) with the data acquisition*
194 *controlled by a LabView (National Instruments Corporation, TX, USA) program.*

195 The SAM system displayed the RF data in real time during the measurements. Ten lines of
196 data were acquired from each of four different positions on the reflector with and without the
197 sample (FTMMs or UTMMs) in place.

198 The calculation of the SoS and the attenuation coefficient was also based on the broadband
200 pulse-echo substitution technique. However, here the thickness of the sample was inputted into the
201 calculation of SoS and attenuation coefficient. The thickness value input for each FTMMs and
202 UTMMs was the mean of the 10 different measurements at each of 8 locations calculated using the
4 transducers of the Vevo 770[®] ultrasound scanner.

203 At all-time points, measurements taken with the Vevo 770[®] were performed before
204 measurements with the SAM system.

205 *Unpreserved samples, batch to batch variation, and repeatability of the UTMMs.*

206 The acoustic properties of two uncovered TMM samples (UTMM) were measured in an
207 identical manner to the UTMMs described previously using the RMV704 transducer (centre frequency
208 40 MHz, Table 1). Measurements were undertaken initially and then approximately once every 24
209 hours over a 96 hour period. However, in between measurements, the samples were left exposed to
210 the air. These samples will be referred to unpreserved samples.

211 An indication of TMM batch-to-batch variability was assessed by measuring the acoustical
212 properties of 6 UTMMs manufactured from a different batch of TMM. These samples will be referred
213 to as UTMM2. These samples had a mean thickness of 2.01 ± 0.05 mm as measured using the Vevo
214 770[®] scanner. The acoustical properties were measured with the Vevo 770[®] and with the SAM
215 system at the 6 and at 12 month time points.

216 Data analysis was performed in an identical manner to that with the 11 UTMMs described
217 above.

218 To assess the repeatability of the measurement system, the acoustic properties of the 11
219 UTMMs were measured with one transducer RMV710 with the Vevo 770[®] at 5 different times over 1
220 month period. The reference medium was TMM preserving fluid. The temperature for these
221 measurements was $22.0 \pm 0.4^{\circ}\text{C}$.

222 RESULTS

223 The mean thickness of the FTMMs and UTMMs calculated using the RF ultrasound signals
224 measured with the Vevo 770® over all the time points showed a maximum variation of 0.25mm in
225 the case of FTMMs and up to 0.08mm for the UTMMs. The thicknesses measured by the digital
226 calliper from 11 UTMMs showed a maximum variation of 0.03mm.

227 Table 2 shows the mean SoS of FTMMs and UTMMs at each time point. It can be seen that
228 the SoS of the FTMMs exhibited larger variability than the SoS of the UTMMs. Using a Student's t-
229 test it was shown that the mean SoS value of FTMMs and UTMMs samples were not statistically
230 different ($p>0.5$) at 0 time point, but displayed a significant difference ($p<0.05$) at the rest of the
231 time points. The results after 1 year showed that the SoS of the FTMMs decreased 22.1 ms^{-1} when
232 compared with the first measurement at 0 months, whereas the SoS of the UTMMs samples
233 decreased 4.1 ms^{-1} over the same 12 month period. The SoS of UTMM2 samples calculated over a 6
234 month time period showed a decrease from $1558.1 \pm 5.3 \text{ ms}^{-1}$ to $1544.8 \pm 3.3 \text{ ms}^{-1}$, with a difference
235 of 13.3 ms^{-1} .

236 Table 3 shows the mean SoS averaged all time-points for each of the measurement systems.
237 It was found that the SoS measurements using the SAM system showed smaller variability than the
238 SoS measurements using the Vevo 770® for the FTMMs and UTMMs. The mean values over all the
239 samples over all time points, using both measurement systems, were found to be $1538.2 \pm 14.5 \text{ ms}^{-1}$
240 for the FTMMs and $1544.0 \pm 3.5 \text{ ms}^{-1}$ for the UTMMs (Table 4). The mean SoS for the UTMM2 was
241 found to be $1551.4 \pm 6.2 \text{ ms}^{-1}$. Table 4 also shows the SoS results in comparison with those values
242 published for IEC agar TMM (Brewin et al., 2008; Browne et al., 2003; IEC, 2001; Rajagopal et al.,
243 2014; Sun et al., 2012). The mean SoS of FTMMs and UTMMs are within the values specified by the
244 IEC (IEC, 2001).

245 Figure 3 and Figure 4 show the attenuation as a function of frequency for the FTMMs and
246 the UTMMs respectively at 0, 6 and at 12 month time points measured using the seven different
247 transducers. It can be seen that the variation in mean attenuation values for the UTMMs is small in
248 comparison to the variation observed in mean attenuation values measured from the FTMMs.

249 Figure 5 and Figure 6 shows the mean attenuation data over the frequency range 4.5 – 50
250 MHz, averaged over all 11 FTMMs and over all 11 UTMMs and time points, respectively.

251 The batch to batch variation of TMM was assessed by the UTMM2s (2.01 ± 0.05 mm
252 thickness). These samples were measured with both the Vevo 770® and the SAM system (at 6 and 12
253 month time points). The mean SoS of UTMM2s was found to be 0.44% higher (7.4 ms^{-1}) than the
254 mean SoS of the 11 UTMMs. For the attenuation, the UTMM2s were found to have a maximum
255 difference of $\pm 1 \text{ dB cm}^{-1}$ in mean attenuation across the frequency range shown in Figure 7.

256 For the unpreserved samples, after 96 hours of exposure to air, the samples were visibly
257 dehydrated. The mean thickness of the two samples had decreased by 1.22mm and the diameter was
258 decreased by 1.5cm. Additionally, the SoS was shown to increase by 140 ms^{-1} for sample 1 and 180
259 ms^{-1} for sample 2 over the total period of time. The attenuation was found to increase by
260 approximately 10 dBcm^{-1} per day.

261 In the assessment of repeatability, the mean SoS over the five measurements taken from the
262 11 UTMMs was calculated to be 1543.0 ms^{-1} with a range in SoS of $\pm 11.0 \text{ ms}^{-1}$. The mean SoS was
263 found to be smaller by 1 ms^{-1} when compared to the mean SoS calculated using all the transducers
264 at all time points (Table 4) of the UTMMs. The variation in attenuation as a function of frequency
265 was $\pm 1 \text{ dBcm}^{-1}$ over the frequency range of the Vevo770® RMV710B probe.

266 Polynomial functions were calculated for the attenuation as a function of frequency at each
267 time point for the FTMMs and for the UTMMs. The best fit polynomial function was determined over
268 all the attenuation versus frequency data in the range 4.5 – 50 MHz as a combination of Vevo 770®

269 and SAM system data over all time points (Figure 8). The polynomial fit found for FTMM was
270 $0.4649f + 0.007363f^2$ ($R^2=0.80$) and $0.4897f + 0.008366f^2$ ($R^2=0.99$) for UTMM. The goodness
271 of fit (R^2) of the three polynomial fits at each of the three time points for the FTMMs ranged
272 between 0.78 – 0.92 whereas for the UTMMs this value ranged between of 0.96-0.99. In addition, in
273 Figure 8, for comparison, the attenuation data of the IEC agar TMM from studies already published is
274 included.

275 DISCUSSION

276 This aim of this study was to develop a robust and easy-to-use technique for the
277 characterisation and preservation of the IEC agar TMM and to compare the acoustic properties
278 obtained using this modified technique with the standard technique over a period of one year.

279 The thickness used in the calculation of SoS from the SAM system was a mean thickness
280 measured by the Vevo 770[®] ultrasound scanner at eight different locations on the UTMMs (10 lines
281 at each position). Although this mean thickness was used, the SD of the SoS values from the SAM
282 system were smaller than the SD variation calculated using the Vevo 770[®] (Table 4) which would
283 suggest that the use of this mean thickness in the SAM system measurements did not contribute
284 significantly to the experimental error.

285 The acoustic properties of the UTMMs were measured in TMM preserving fluid whose
286 acoustical properties were assessed by NPL at a $19.3 \pm 0.1^\circ\text{C}$, whereas the UTMMs in this study were
287 measured at $22.2 \pm 0.5^\circ\text{C}$. The TMM preserving fluid is composed of the same fluid as used in the
288 TMM manufacture process and Brewin et al (2008) has previously shown a TMM SoS temperature
289 dependence of $2.1 \text{ ms}^{-1} \text{ }^\circ\text{C}^{-1}$. Consequently, there is likely to be a maximum variation of 6 ms^{-1} in the
290 SoS of the TMM preserving fluid which could be attributable to the temperature change. Such a
291 change would result in a potential error of less than 7 ms^{-1} in the measured SoS of the UTMMs.
292 Nevertheless, the SoS values of the UTMM were found to be in good agreement with Rajagopal et
293 al., (2014); Sun et al., (2012) and Brewin et al., (2008). Furthermore, the SoS of the UTMMs was
294 found to decrease by 4.1 ms^{-1} over a 12 month period compared with FTMMs which showered a
295 decrease in the mean SoS of 22.1 ms^{-1} over the same period of time. Additionally the standard
296 deviation of the mean SoS values for FTMM samples was larger than that for UTMMs at all time-
297 points. This increased variation in SoS values for the FTMMs in comparison to UTMMs may be
298 attributable to a number of reasons. Firstly, although a visual inspection was performed on each of
299 the FTMMs before each acoustical measurement and no evidence of leakage was observed,

300 nevertheless, in several of the samples, the epoxy securing the film to the rings showed signs of
301 ageing and the Saran Wrap® film appeared to become less taut over the 1 year period. This could
302 have potentially increased the permeability of the film allowing the glycerol from the TMM to leach
303 into the water medium resulting in a decrease in the measured SoS properties of the FTMM.
304 However, although a decrease in SoS in FTMMs was measured between 0 and 12 months, this did
305 not decrease continuously over the 1-year period which would suggest that the measured variation
306 is not likely to be attributable to glycerol leakage. Secondly, for the FTMMs, the position of the
307 water-film interfaces was selected using Matlab code based on the identification of the position of
308 the maximum rectified RF signal and it was assumed that this signal also marked the TMM interface.
309 Although this is a reasonable assumption, if any of the FTMM samples were subject to shrinkage (by
310 drying out) over the 1-year period, this would represent a potential source of error.

311 The SoS results of both FTMMs and UTMMs in this study were compared with previously
312 published work (Table 4). It can be seen that the UTMM mean SoS values are in good agreement
313 with those published, whereas the mean SoS of FTMM were found to be 5.8 ms^{-1} smaller when
314 compared with Rajagopal et al., 2014 and up to 9.6 ms^{-1} smaller when compared with Sun et al.,
315 2012. In Rajagopal et al., (2014) the manufacture of FTMM was achieved by sandwiching the TMM
316 slice between 2 sheets of Mylar® (~12µm thickness) affixed into Perspex frames, whereas in Sun et
317 al., (2012) the manufacture process of the FTMMs was similar to the method used in this study
318 (referred to as TMM test cells in that study). However, in Rajagopal even though the acoustic
319 measurement was completed relatively quickly (within seconds) the edges of the TMM were not
320 covered, which is likely to have led to some undefined glycerol leakage and potentially affect the
321 measured acoustical properties. Furthermore, in Brewin et al., (2008) the acoustical properties of 2
322 different batches of TMM were measured over a 3 year period with a thickness range from 3mm to
323 12.7mm. In Brewin, the first batch consisted of TMM samples which were not protected by a film
324 and were measured in double degassed, deionized water. As a result of glycerol leaching from the
325 samples in this batch, the SoS was found to decrease by $2.1 \text{ ms}^{-1} \text{ }^{\circ}\text{C}^{-1}$. The second batch consisted of

326 TMM samples protected by Saran Wrap® and were also measured in water. Using this method
327 thinner samples (3mm) displayed the largest SoS variation of 13.4 ms^{-1} . This value is comparable to
328 the SD found in this study for the FTMMs (Table 6) but considerably higher than that measured for
329 the UTMMs.

330 Figure 3 and Figure 4 show the attenuation of FTMMs and UTMMs respectively, at 0, 6 and
331 12 month time points. It can be observed, that there is a much larger variation in the attenuation
332 measurements obtained from the FTMMs compared to the UTMMs. At both the lower (4.5 – 9MHz)
333 and the higher (40 – 50MHz) frequency ranges, the data displayed is obtained from a single
334 transducer. Nevertheless, the attenuation versus frequency for the FTMMs would suggest, that with
335 increasing frequency there is an increasing difference in measured mean attenuation values between
336 the 6 month data and 0 and 12 month data. The maximum difference was of 7 dB cm^{-1} , occurring
337 over the frequency range from 30 – 42MHz and a minimum difference at 15 – 19MHz. For the
338 UTMMs, there is less variation, even in the lower and upper frequency ranges. A maximum variation
339 of 2 dBcm^{-1} was observed across the time points, at a frequency of 47MHz and a minimum variation
340 from 37 to 47MHz.

341 Moreover, the difference in mean attenuation values between the UTMMs and FTMMs
342 would suggest that, despite compensation for the effects of the Saran Wrap®, some additional
343 acoustic effects are introduced which are not fully compensated using the Saran-wrapped reference
344 water test-cell. However, these effects are unlikely to be due to the difference in non-linear effects
345 between water and TMM preserving fluid. It has previously been shown (Sun, 2012) that even in
346 water, at these output powers, the second harmonic component of the ultrasound beam is at least
347 30dB smaller in magnitude than the first harmonic. Since non-linear effects are easier to generate in
348 water than in the TMM preserving fluid, it is unlike that nonlinearities are significantly greater than
349 the experimental errors identified.

350 Figure 5 and Figure 6 show the mean attenuation of the 11 FTMMs and the 11 UTMMs
351 across the 7 different transducers and measured 3 times during the time period of 1 year. The
352 FTMMs (Figure 5) showed larger variability ($\sim 15\text{dB cm}^{-1}$) across samples and transducers. This may
353 be due to inadequate acoustical correction for the interface layers when using the reference water
354 test cell, leading to an increased uncertainty in the attenuation measurements in addition to the
355 factors previously described. The UTMMs (Figure 6) showed good consistency and little variability in
356 the attenuation over the frequency range of 4.5 – 50 MHz.

357 Polynomial fits from FTMM and UTMMs were in good agreement with previous studies in
358 the frequency range of 17 – 23 MHz (Brewin et al., 2008), 10 – 47 MHz (Sun et al., 2012) and 1 – 60
359 MHz (Rajagopal et al., 2014). The polynomial fits were also in good agreement at lower frequencies
360 4.5 to 10 MHz as reported by IEC (IEC, 2001) and in other studies (Brewin et al., 2008; Inglis et al.,
361 2006) as can be seen in Figure 8. The attenuation from FTMMs and UTMMs does not increase
362 linearly with frequency as shown by the quadratic terms of the polynomial fit. This quadratic term
363 was found to be 0.0073 for FTMMs and 0.0083 for UTMMs which is in good agreement with 0.0076
364 reported by Sun et al., 2012 and with 0.0081 reported by Rajagopal et al., 2014.

365 Finally the unpreserved samples displayed significant visual degradation and changes in SoS
366 and attenuation, over the 96 hours. These results are consistent with those of Brewin et al., (2008)
367 who also reported shrinking and hardening of TMM samples which were not preserved.

368 CONCLUSIONS

369 In this study, two different measurement techniques were evaluated for assessing the
370 temporal stability of the acoustic properties of the IEC agar TMM over the frequency range 4.5 – 50
371 MHz. In the first technique thin slices were wrapped and stored in Saran Wrap® and measured in
372 degassed, deionised water. In the second modified technique, thin slices of TMM were preserved and
373 measured in TMM preserving fluid. Measurements were undertaken, over the period of 1 year. The
374 measured SoS of an IEC agar TMM calculated by the Vevo 770® and SAM system was found to be
375 $1538.2 \pm 14.5 \text{ ms}^{-1}$ for the FTMMs and $1544.0 \pm 3.5 \text{ ms}^{-1}$ for the UTMM. For FTMMs the SoS results
376 were less than 10 ms^{-1} lower when compared with those published. The acoustic properties of
377 UTMMs (SoS and attenuation values) were found to be in good agreement with results in earlier
378 studies by Brewin et al., (2008) over the range of 17 – 23 MHz, Sun et al., (2012) over the range of 10
379 – 47 MHz and Rajagopal et al., (2014) over the range of 1 – 60 MHz. Nevertheless, the results for
380 both FTMMs and UTMMs were consistent at low frequencies (Browne et al., 2003; Inglis et al., 2006)
381 and within the range provided by the IEC (IEC, 2001). However, the attenuation coefficient was
382 shown to be nonlinear as a function of frequency. The attenuation was found to increase as
383 $0.4649f + 0.007363f^2$ for FTMMs and as $0.4897f + 0.008366f^2$ for UTMMs with increasing
384 frequency. This second degree polynomial fit was derived based on the data generated in this study
385 using two different measurements systems and was shown to be able to estimate the attenuation of
386 this IEC agar TMM in the frequency range of 4.5 – 50 MHz. The quadratic term was also found to be
387 in good agreement with previous studies.

388 Finally, this study has demonstrated that using unwrapped TMM slices (UTMM), maintained
389 and measured in TMM preserving fluid, results in approximately 4 times smaller SD values for the
390 SOS and up to 5 times smaller variation for the attenuation values when compared with the
391 common method of film-wrapped TMM samples (FTMM) measured in degassed, deionised water.
392 This suggests that, despite compensation within the calculation of the attenuation effects of the

393 Saran Wrap[®], additional acoustic effects are introduced which are not fully compensated using the
394 standard technique (FTMMs). Moreover, this study has also brought into question, the validity and
395 subsequent stability of encasing gel TMM QA phantoms in a sealed film-dry environment.

396 ACKNOWLEDGEMENTS

397 The authors will like to thank Dr. David Kenwright, Mr. Adrian Thomson, Mr. Chris McLeod and
398 Dr. Bakary Diarra for their help during the production of this work and to NPL for the acoustic
399 characterisation of the TMM preserving fluid. This study was funded by a CONACyT (Becas al
400 Extranjero 2014) PhD studentship and Carnegie Trust Small Research Grant.

401 REFERENCES

- 402 AIUM. (2014). *Methods for specifying acoustic properties of Tissue-Mimicking phantoms and objects*.
- 403 Banchhor, S. K., Araki, T., Londhe, N. D., Ikeda, N., Radeva, P., Elbaz, A., Saba, L., Nicolaidis, A.,
404 Shafique, S., Laird, J. L., Suri, J. S. Five multiresolution-based calcium volume measurement
405 techniques from coronary IVUS videos: A comparative approach. *Computer Methods and*
406 *Programs in Biomedicine*, 2016, 134, 237–258. doi:10.1016/j.cmpb.2016.07.009
- 407 Bilaniuk, N., & Wong, G. S. K. Speed of sound in pure water as a function of temperature. *J Acoustic*
408 *Soc*, 1992, 93, 1609–1612.
- 409 Brewin, M., Pike, L. C., Rowland, D. E., & Birch, M. J. The acoustic properties, centered on 20 MHz, of
410 an IEC agar-based tissue-mimicking material and its temperature, frequency and age
411 dependence. *Ultrasound in Medicine & Biology*, 2008, 34(8), 1292–306.
412 doi:10.1016/j.ultrasmedbio.2007.12.017
- 413 Browne, J. E., Ramnarine, K., Watson, A. J., & Hoskins, P. R. Assessment of the acoustic properties of
414 common tissue-mimicking test phantoms. *Ultrasound in Medicine & Biology*, 2003, 29(7),
415 1053–1060. doi:10.1016/S0301-5629(03)00053-X
- 416 Cannon, L. M., Fagan, A. J., & Browne, J. E. Novel tissue mimicking materials for high frequency
417 breast ultrasound phantoms. *Ultrasound in Medicine & Biology*, 2011, 37(1), 122–35.
418 doi:10.1016/j.ultrasmedbio.2010.10.005
- 419 Cournane, S., Cannon, L., Browne, J. E., & Fagan, a J. Assessment of the accuracy of an ultrasound
420 elastography liver scanning system using a PVA-cryogel phantom with optimal acoustic and
421 mechanical properties. *Physics in Medicine and Biology*, 2010, 55(19), 5965–5983.
422 doi:10.1088/0031-9155/55/19/022
- 423 Culjat, M. O., Goldenberg, D., Tewari, P., & Singh, R. S. A review of tissue substitutes for ultrasound
424 imaging. *Ultrasound in Medicine & Biology*, 2010, 36(6), 861–73.

425 doi:10.1016/j.ultrasmedbio.2010.02.012

426 Del Grosso, V. A., & Mader, C. W. Speed of Sound in Pure Water. *J Acoust Soc*, 1972, 52, 1442–1446.

427 Duck, F. A. *Physical Properties of Tissues. A Comprehensive Reference Book*. Bath, England: Academic
428 Press, 1990.

429 Foster, F. S., Pavlin, C. J., Harasiewicz, K. A., Christopher, D. A., & Turnbull, D. H. Advances in
430 ultrasound biomicroscopy. *Ultrasound in Medicine and Biology*, 2000, 26(1), 1–27.
431 doi:10.1016/S0301-5629(99)00096-4

432 He, P. Measurement of acoustic dispersion using both transmitted and reflected pulses. *Acoustical*
433 *Society of America*, 2000, 107(2), 801–807.

434 IEC. European Standard. British Standard. Ultrasonics. Flow measurement systems. Flow test object,
435 2001.

436 Inglis, S., Ramnarine, K., Plevris, J. N., & McDicken, W. N. An anthropomorphic tissue-mimicking
437 phantom of the oesophagus for endoscopic ultrasound. *Ultrasound in Medicine and Biology*,
438 2006, 32(2), 249–59. doi:10.1016/j.ultrasmedbio.2005.10.005

439 Kenwright, D. A., Sadhoo, N., Rajagopal, S., Anderson, T., Moran, C. M., Hadoke, P. W., Gray, G. A.,
440 Zequiri, B., Hoskins, P. R. Acoustic Assessment of a Konjac–Carrageenan Tissue-Mimicking
441 Material at 5–60 MHz. *Ultrasound in Medicine & Biology*, 2014, 40(12), 2895–2902.
442 doi:10.1016/j.ultrasmedbio.2014.07.006

443 King, D. M., Moran, C., McNamara, J. D., Fagan, A. J., & Browne, J. Development of a Vessel-
444 Mimicking Material for use in Anatomically Realistic Doppler Flow Phantoms. *Ultrasound in*
445 *Medicine and Biology*, 2011, 37(5), 813–826. doi:10.1016/j.ultrasmedbio.2011.02.012

446 Machet, L., Belot, V., Naouri, M., Boka, M., Mourtada, Y., Giraudeau, B., Laure, B., Perrinaud, A.,
447 Machet, M. C., Vaillant, L. Preoperative Measurement of Thickness of Cutaneous Melanoma
448 Using High-Resolution 20 MHz Ultrasound Imaging: A Monocenter Prospective Study and

449 Systematic Review of the Literature. *Ultrasound in Medicine and Biology*, 2009, 35(9), 1411–
450 1420. doi:10.1016/j.ultrasmedbio.2009.03.018

451 Madsen, E. L., Frank, G. R., & Dong, F. Liquid or Solid Ultrasonically Tissue-Mimicking Materials with
452 very Low Scatter. *Ultrasound in Medicine and Biology*, 1998, 24(4), 535–542.

453 Meagher, S., Poepping, T. L., Ramnarine, K., Black, R. A., & Hoskins, P. Anatomical flow phantoms of
454 the nonplanar carotid bifurcation, part II: experimental validation with Doppler ultrasound.
455 *Ultrasound in Medicine & Biology*, 2007, 33(2), 303–10.
456 doi:10.1016/j.ultrasmedbio.2006.08.004

457 Moran, C., Ellis, W., Janeczko, A., Bell, D., & Pye, S. The Edinburgh Pipe Phantom: characterising
458 ultrasound scanners beyond 50 MHz. *Journal of Physics: Conference Series*, 2011, 279, 12008.
459 doi:10.1088/1742-6596/279/1/012008

460 Pinkerton, J. M. M. The Absorption of Ultrasonic Waves in Liquids and its Relation to Molecular
461 Constitution. *Proceedings of the Physical Society*, 1949, Section B, 129–141.

462 Rajagopal, S., Sathoo, N., & Zeqiri, B. Reference Characterisation of Sound Speed and Attenuation of
463 the IEC Agar-Based Tissue-Mimicking Material Up to a Frequency of 60 MHz. *Ultrasound in
464 Medicine & Biology*, 2014, 41(1), 317–333. doi:10.1016/j.ultrasmedbio.2014.04.018

465 Ramnarine, K., Anderson, T., & Hoskins, P. Construction and Geometric stability of physiological flow
466 rate wall-less stenosis phantoms. *Ultrasound in Medicine and Biology*, 2001, 27(2), 245–250.

467 Rhee, S. High frequency (IVUS) ultrasound transducer technology - applications and challenges. In
468 *IEEE International Symposium on the Applications of Ferroelectrics*, 2007, (pp. 856–857).

469 Schmitt, C., Hadj Henni, A., & Cloutier, G. Ultrasound dynamic micro-elastography applied to the
470 viscoelastic characterization of soft tissues and arterial walls. *Ultrasound in Medicine & Biology*,
471 2010, 36(9), 1492–503. doi:10.1016/j.ultrasmedbio.2010.06.007

472 Sun, C., Pye, S., Browne, J., Janeczko, A., Ellis, B., Butler, M., Sboros, V., Thomson, A. J. W., Brewin,

473 M. P., Earnshaw, C. H., Moran, C. The Speed of Sound and Attenuation of an IEC Agar-Based
474 Tissue-Mimicking Material for High Frequency Ultrasound Applications. *Ultrasound in Medicine
475 and Biology*, 2012, 38(7), 1262–1270. doi:10.1016/j.ultrasmedbio.2012.02.030

476 Sun, C. *Acoustic characterisation of ultrasound contrast agents at high frequency*. PhD Medical
477 Physics thesis. 2012. University of Edinburgh.

478 Sundholm, J. K. M., Olander, R. F. W., Ojala, T. H., Andersson, S., & Sarkola, T. Feasibility and
479 precision of transcutaneous very-high resolution ultrasound for quantification of arterial
480 structures in human neonates - Comparison with conventional high resolution vascular
481 ultrasound imaging. *Atherosclerosis*, 2015, 239(2), 523–527.
482 doi:10.1016/j.atherosclerosis.2015.02.016

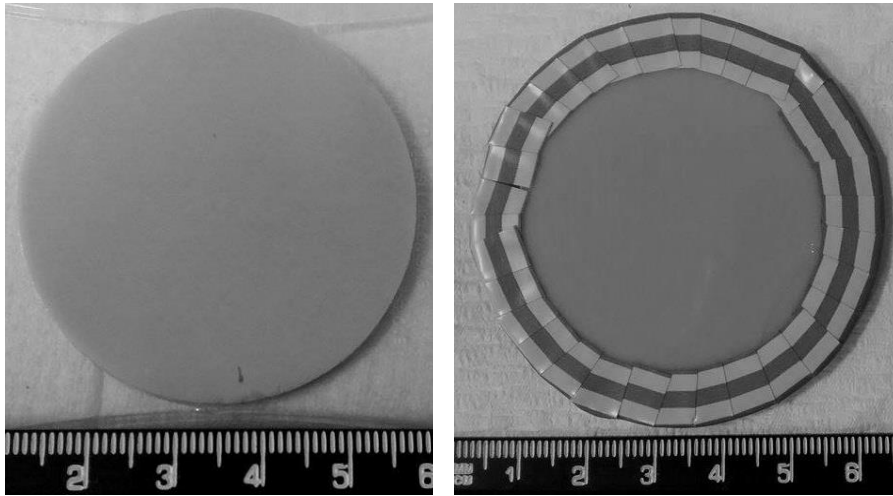
483 Teirlinck, C. J., Bezemer, R. a, Kollmann, C., Lubbers, J., Hoskins, P., Fish, P., Fredfeldt, K. E.,
484 Schaarschmidt, U. G. Development of an example flow test object and comparison of five of
485 these test objects, constructed in various laboratories. *Ultrasonics*, 1998, 36(1–5), 653–60.

486 Xu, J., Tripathy, S., Rubin, J. M., Stidham, R. W., Johnson, L. a, Higgins, P. D. R., & Kim, K. A new
487 nonlinear parameter in the developed strain-to-applied strain of the soft tissues and its
488 application in ultrasound elasticity imaging. *Ultrasound in Medicine & Biology*, 2012, 38(3),
489 511–523. doi:10.1016/j.ultrasmedbio.2011.12.009

490 Yang, X., Sun, C., Anderson, T., Moran, C., Hadoke, P. W. F., Gray, G., & Hoskins, P. Assessment of
491 spectral doppler in preclinical ultrasound using a small-size rotating phantom. *Ultrasound in
492 Medicine and Biology*, 2013, 39(8), 1491–1499. doi:10.1016/j.ultrasmedbio.2013.03.013

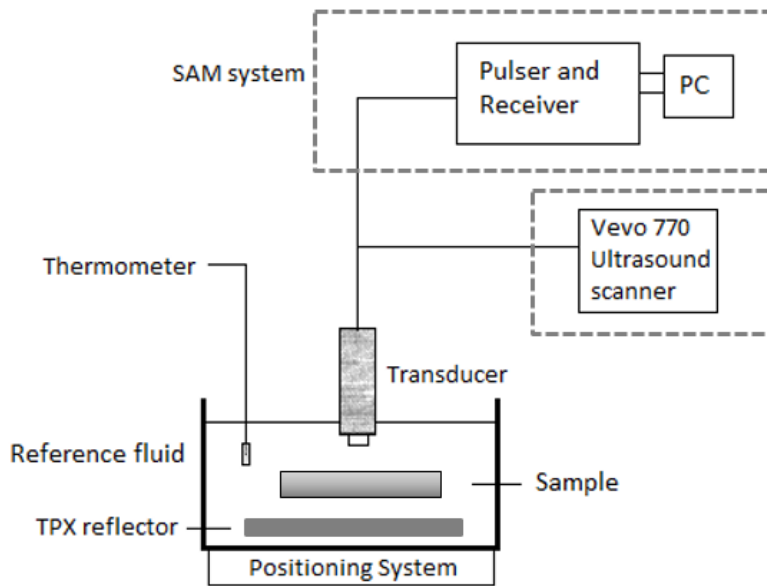
493

495



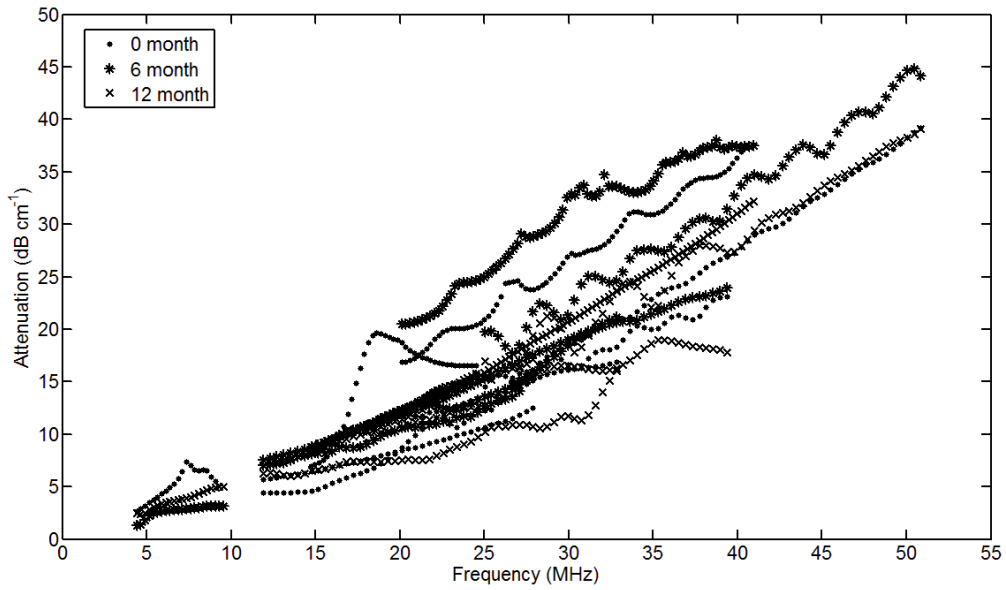
496

497 Figure 1. a) Non-wrapped TMM slices (UTMMs), arrow indicates the identification mark on the
498 sample and b) wrapped TMM slices (FTMMs).



499

500 Figure 2. Experimental set-up using a high frequency ultrasound scanner Vevo 770® used at the
501 University of Edinburgh and a SAM system used at the Dublin Institute of Technology.

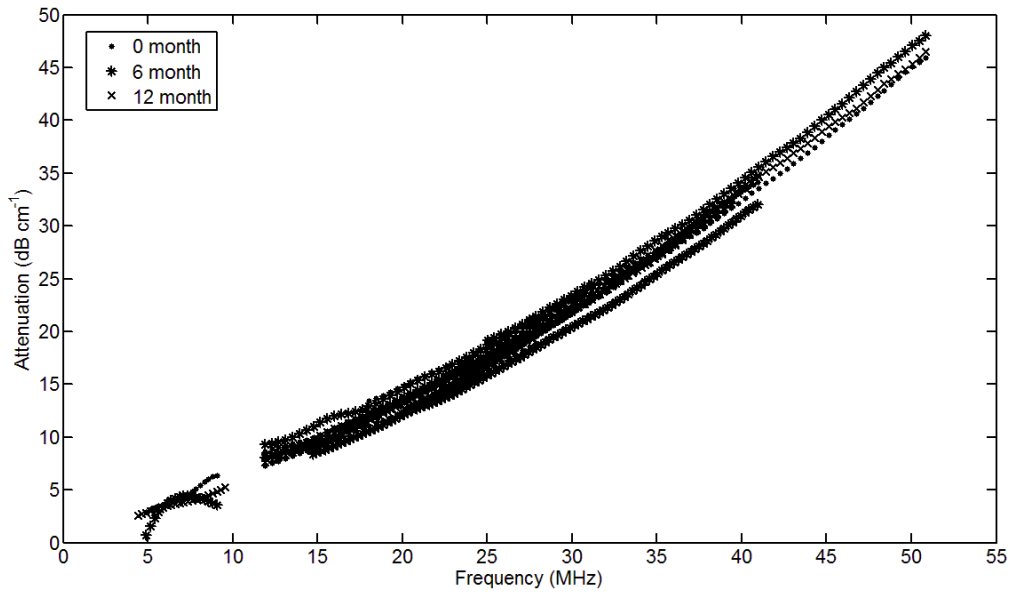


502

503 Figure 3. Measured attenuation with the Vevo 770[®] and SAM system of 11 FTMM at 0, 6 and 12
 504 month time points as a function of frequency. Each line represents one of the seven different
 505 transducers used at each time point [RMV704 (18 – 40 MHz), RMV707B (12 – 32 MHz),
 506 RMV710B (12 – 28 MHz), RMV711 (25 – 50 MHz), V320 (4.5 – 9 MHz), V317 (14 – 25 MHz),
 507 V390 (20 – 40 MHz)].

508

509

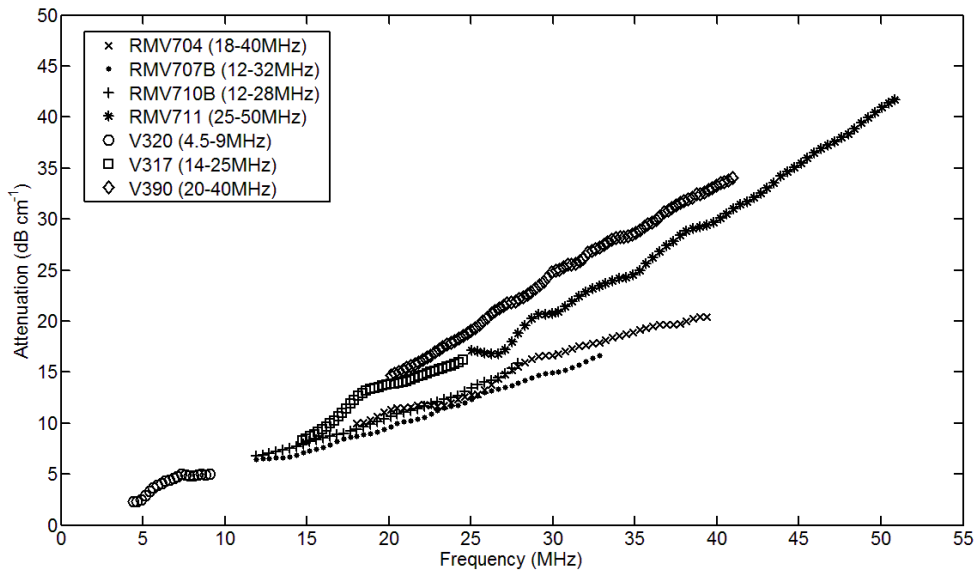


510

511 Figure 4. Measured attenuation with the Vevo 770[®] and SAM system of 11 UTMM at 0, 6 and 12
 512 month time points as a function of frequency. Each line represents one of the seven different
 513 transducers used at each time point [RMV704 (18 – 40 MHz), RMV707B (12 – 32 MHz),
 514 RMV710B (12 – 28 MHz), RMV711 (25 – 50 MHz), V320 (4.5 – 9 MHz), V317 (14 – 25 MHz),
 515 V390 (20 – 40 MHz)].

516

517



518

519 Figure 5. Attenuation data as a function of frequency averaged over all time points (data set: 11

520 FTMM measured in degassed deionized water by the Vevo 770® and SAM system in the

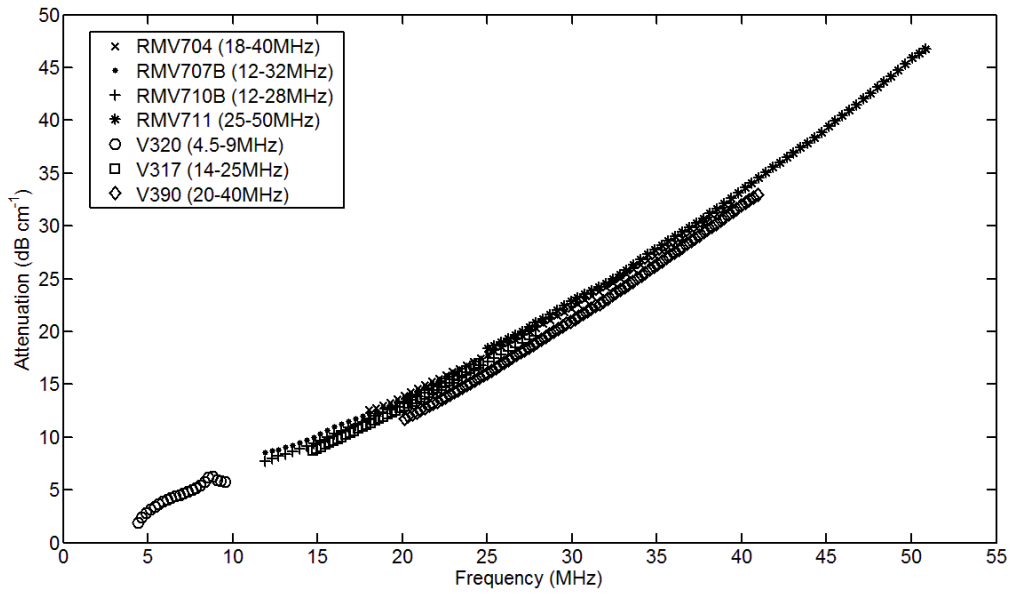
521 frequency range of 4.5 – 50 MHz).

522

523

524

525



526

527 Figure 6. Attenuation data as a function of frequency averaged over all time points (data set: 11

528 UTMM preserved and measured in TMM preserving fluid by the Vevo 770® and SAM system in

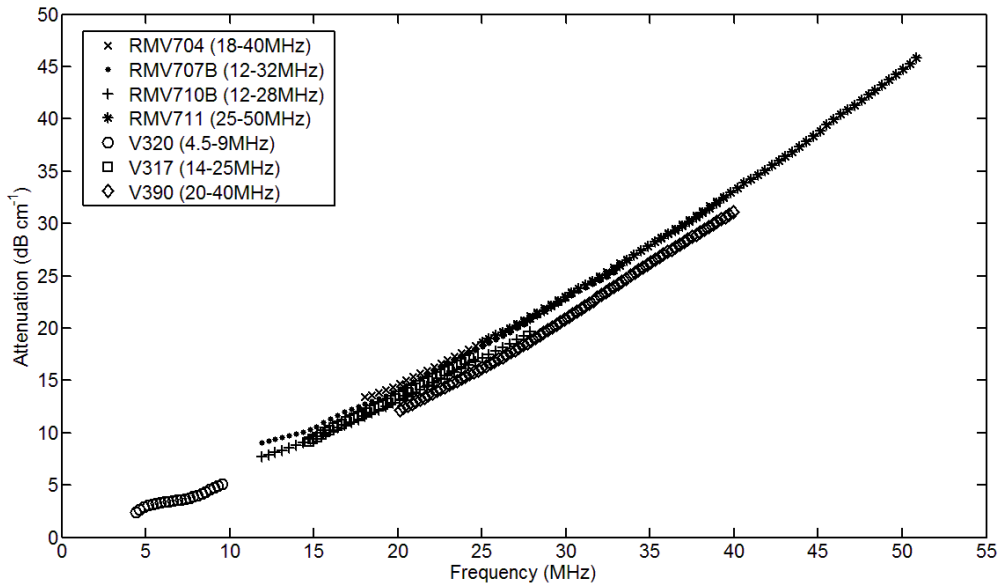
529 the frequency range of 4.5 – 50 MHz).

530

531

532

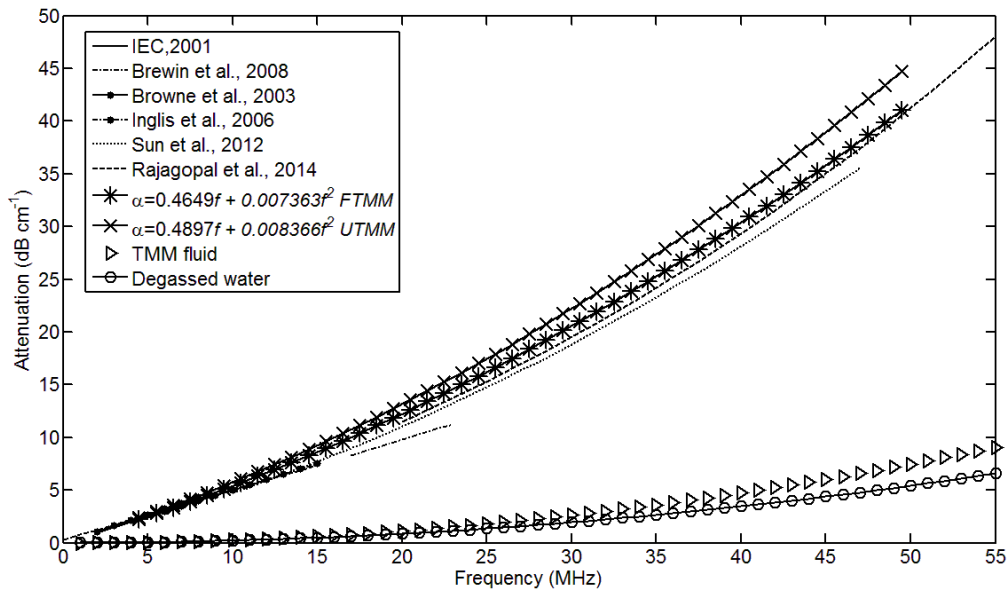
533



534

535 Figure 7. Attenuation data as a function of frequency averaged over all time points from the 6
 536 UTM2 (data set: 6 UTM2 preserved and measured in TMM preserving fluid by the Vevo
 537 770® and SAM system at 6 and 12 month time points in the frequency range of 4.5-50 MHz).

538



539

540 Figure 8. Polynomial curve-fit of all the attenuation data as a function of frequency and the
 541 attenuation (compensated for the attenuation in water) of TMM in 2 – 10 MHz (IEC, 2001), 17

542 – 23 MHz (Brewin et al., 2008), 2.25 – 15 MHz (Browne et al., 2003), 6 – 15 MHz (Inglis et al.,
543 2006), 10 – 47 MHz (Sun et al., 2012) and 1 – 60 MHz (Rajagopal et al., 2014). Also the
544 attenuation as a function of frequency for the TMM preserving fluid and degassed deionised
545 water.

Transducer model and measurement system		Central Frequency (MHz)	Focal Length (mm)	Measured 3dB bandwidth (MHz)	Peak negative pressure (MPa)	Beam width (cm)
704	Vevo 770®	40	6	18 – 40	0.52	0.008
707B		30	12.7	12 – 32	1.05	0.0115
710B		25	15	12 – 28	1.06	0.014
711		55	6	25 – 50	0.23	0.009
V320	SAM system	7.5	95	4.5 – 9	0.05	1.27
V317		20	65	14 – 25	0.021	0.63
V390		50	12	20 – 40	0.022	0.63

547

548 Table 1. Characteristics of the Vevo 770® and SAM system transducers. The central frequency and
 549 focal length are parameters provided by the manufacturer from Vevo 770® (VisualSonics, Inc.,
 550 Toronto, Canada) and SAM system (Olympus Panametrics NDT), the 3dB bandwidth from
 551 measurements and the peak pressure from (Sun, 2012).

552

SoS ± SD (ms ⁻¹)	0 months	6 months	12 months
FTMM	1547.4 ± 19.2	1547.2 ± 21.5	1525.5 ± 16.5
UTMM	1545.9 ± 10.4	1544.2 ± 11.0	1541.8 ± 1.6

553 Table 2. The mean and SD of SoS (ms-1) measured with the Vevo 770® and SAM system at each time
 554 point for the FTMMs and UTMMs samples.

555

556

557

558

559

SoS \pm SD (ms^{-1})	Vevo 770 [®]	SAM system
FTMM	1539.6 \pm 17.1	1536.3 \pm 10.3
UTMM	1542.9 \pm 3.6	1545.3 \pm 3.0

560 Table 3. The mean and SD of SoS (ms^{-1}) over all time points of 11 FTMM and 11 UTMM measured by
561 the four transducers of the Vevo 770[®] and by the three transducers of the SAM system.

562

Sources	Type of samples (covered with film or uncovered)	Mean SoS \pm SD (ms^{-1})	Frequency range (MHz)
IEC, 2001		1540 \pm 15	2 – 10
Browne et al., 2003	TMM uncovered measured in degassed water	1546.5 \pm 3	2.25 – 15
Brewin et al., 2008	TMM uncovered measured in degassed water	1537 \pm 2.6	17 – 23
	TMM covered	1540.9 \pm 8.7	
Sun et al., 2012	TMM covered	1547.8 \pm 3.7	10 – 47
Rajagopal et al., 2014	TMM covered	1544 \pm 3.1	1 – 60
This study	TMM covered (FTMM)	1538.2 \pm 14.5	4.5 – 50
	TMM uncovered (UTMM) measured in TMM fluid	1544.0 \pm 3.5	

563 Table 4. Values of SoS ($\text{ms}^{-1} \pm$ SD) measured in this study using 2 different methods, compared with
564 the published data.