



Technological University Dublin
ARROW@TU Dublin

Articles

School of Food Science and Environmental Health

2006

Horseradish and Soybean Peroxidases: Comparable Tools for Alternative Niches?

Barry Ryan

Technological University Dublin, barry.ryan@tudublin.ie

Ciarán Fagan

Dublin City University, ciaran.fagan@dcu.ie

Neil Carolan

Follow this and additional works at: <https://arrow.tudublin.ie/schfsehart>

 Part of the [Life Sciences Commons](#)

Recommended Citation

Ryan, B.J., Carolan, N. and O'Fágáin, C. (2006). Horseradish and soybean peroxidases: comparable tools for alternative niches? *Trends in Biotechnology*, 24, (8), 355-363. <http://dx.doi.org/10.1016/j.tibtech.2006.06.007>

This Article is brought to you for free and open access by the School of Food Science and Environmental Health at ARROW@TU Dublin. It has been accepted for inclusion in Articles by an authorized administrator of ARROW@TU Dublin. For more information, please contact yvonne.desmond@tudublin.ie, arrow.admin@tudublin.ie, brian.widdis@tudublin.ie.



This work is licensed under a [Creative Commons Attribution-NonCommercial-Share Alike 3.0 License](#)



Horseradish and Soybean Peroxidases: Comparable Tools for Alternative Niches?

Barry J. Ryan, Neil Carolan and Ciarán Ó'Fágáin*.

School of Biotechnology and National Centre for Sensors Research, Dublin City University, Dublin 9, Ireland.

Horseradish and soybean peroxidases (HRP and SBP, respectively) are useful biotechnological tools. HRP is often termed the classical plant heme peroxidase, and although it has been studied for decades our understanding has deepened since its cloning and subsequent expression, which has enabled numerous mutational and protein engineering studies. SBP, however, has been neglected until recently; despite offering a real alternative to HRP that actually outperforms it in terms of stability. SBP is now used in numerous biotechnological applications, including biosensors. Review of both is timely. This article summarises and discusses the main insights into the structure and mechanism of HRP, with special emphasis on HRP mutagenesis, and outlines its use in a variety of applications. It also reviews current knowledge and applications to date of SBP, particularly biosensors. The final paragraphs speculate on the future of plant heme-based peroxidases, with probable trends outlined and explored.

Key Words: Horseradish Peroxidase, Soybean Peroxidase, Review, Biosensors, Biocatalysis, Mutagenesis.

[<http://www.dcu.ie/biotechnology/index.shtml>]

[<http://www.dcu.ie/~ncsr/>]

* corresponding author: Ciarán Ó'Fágáin, (Ciaran.Fagan@dcu.ie).

Phone: +353-1-700 5288

Fax: +353-1-700 5412

Abbreviations:

ABTS, 2,2'-Azino-di-(3-ethyl-benzthiazoline-6-sulphonic acid); **Co(bpy)**, Tris-2,2'-dipyridylcobalt(III) perchlorate trihydrate; **CV**, Cyclic voltammogram. **DAB**, 3,3'-diaminobenzidine tetrahydrochloride. **DMPC**, Dimyristoylphosphatidylcholine epoxidised olefins. **DTT**, dithiothreitol; **ELISA**, Enzyme-Linked Immunosorbent Assay. **GOx**, Glucose oxidase; **GP**, Graphite Powder; **H₂O₂**, Hydrogen peroxide; **HQ**, hydroquinone; **IPTG**, iso-propyl-β-thiogalactoside; **MET**, mediated electron transfer; **mm**, millimetre. **PEGDGE**, Poly(ethylene glycol) (400) diglycidylether. **PeIB**, pectate lyase B fragment. **PDB**, Protein Data Bank; **POs-EA**, Os(bpy)₂Cl^{+/2+} poly(4-vinylpyridine) quaternised with 2-bromoethylamine. **Pt**, Platinum. **PVA**, Polyvinyl alcohol. **PVP**, Polyvinyl pyridine. **PVP-Os^{III}**, polyvinyl pyridine-osmium complex. **POCT**, point-of-care testing; **Px**, peroxidase. **Px_{ox}**, oxidised peroxidase. **rHRP**, recombinant HRP; **TTCA**, poly-5,2':5',2"-terthiophene-3'-carboxylic acid.

41 **Introduction.**

42 Peroxidase enzymes span the bioscience and biotechnology spectra, ranging from
43 bioremediation [1] and biocatalysis [2] through diagnostics [3] and biosensors [4] to
44 recombinant protein expression [5], transgenics [6], bioinformatics [7], protein
45 engineering [8] and even to therapeutics [9]. This article contrasts two key heme-
46 containing plant peroxidases, horseradish (HRP) and soybean (SBP), with special
47 emphasis on mutagenesis studies and biosensor applications. HRP is a ‘traditional’
48 enzyme, whereas SBP emerged in the 1990s. A comparison of these key
49 biotechnological tools is timely.

50

51 **Brief biochemistry of horseradish and soybean peroxidase:**

52 All heme peroxidases (E.C. 1.11.1.7) have a ferriprotoporphyrin IX prosthetic group
53 located at the active site [10]. Both HRP and SBP are classified as Class III Classical
54 Secretory plant peroxidases [7 & 11] and as such share common features (Table 1).
55 Their catalytic mechanism involves a two-electron oxidation of the heme moiety to an
56 intermediate known as Compound I. Successive one-electron reductions return the
57 enzyme to its resting state via a second intermediate, Compound II [6].

58 Determining the *in vivo* function of peroxidases is complex owing to the numbers of
59 isoenzymes in[E1] the family [12]. Interestingly, despite the several *in vitro* uses of
60 HRP, its actual *in vivo* role has never been elucidated. Several suggestions have been
61 proposed based on the known roles of other plant peroxidases. Peroxidases are usually
62 found in the cell wall, vacuoles, transport organelles and the rough endoplasmic
63 reticulum, and have noted roles in lignification, wound healing and auxin catabolism
64 [4]. SBP has been isolated from that plant’s seed coat and its presence prevents
65 premature germination[E2] [13]. Plant peroxidases can use lignin and other plant
66 compounds as reducing substrates. Indeed, SBP has been noted to polymerise
67 coniferyl alcohol, indicating that it can efficiently catalyse reactions involving lignin
68 precursors [14]. Therefore, it is possible that peroxidases are involved in the
69 lignification or suberisation processes of plants [15].

70 **Recombinant Peroxidase Expression.**

71 Recombinant hemoprotein expression has been plagued by inclusion body formation,
72 most notably in recombinant HRP expression. Several general methods have been
73 suggested to reduce the formation of inclusion bodies, including reducing cultivation
74 temperature and altering inducer composition and concentration [16]. Other, more
75 peroxidase-specific, methodologies have been cited, including use of specific *E. coli*
76 strains [17], inclusion of chaperones [18] and use of leader sequences [19]. Another
77 major obstacle in the recombinant expression of hemoproteins is the limited
78 availability of heme and iron within a bacterial cell. Bacterial cells each contain 10^5 to
79 10^6 iron ions, which are essential for many metabolic pathways [20]. Culture
80 supplementation with the heme precursor δ -aminolevulinic acid has been suggested
81 [21]; but Goodwin and co-workers have recently developed an elegant co-expression
82 system incorporating a membrane heme receptor, allowing the use of exogenous heme
83 as an iron source [22]. Jung and co-workers [5] noted an increased ratio of holoprotein
84 to apoprotein with less-intense induction conditions, suggesting that slow recombinant
85 hemoprotein production appears to allow easier incorporation of the available heme
86 into the apoprotein [5]. Developments in HRP expression in both prokaryotic and
87 eukaryotic systems are outlined in the supplementary online material.

88

89 **HRP: Cloning and Expression.**

90 The gene coding for the HRP protein was first synthesised by Smith and co-workers
91 [23] based on the protein sequence published by Welinder [24]. This 940 basepairs
92 synthetic gene was designed using commonly used codons in *E. coli* to minimise
93 protein truncation owing to codon bias [25]. Recombinant HRP was over-expressed
94 by induction but this led to the formation of misfolded apoprotein and the requirement
95 to disrupt these aggregates, refold the protein correctly and add the heme centre.
96 Disruption involved addition of EDTA to chelate ions, lysozyme and DNase to
97 reduce viscosity of the bacterial cell lysate, urea to solubilise the protein, and
98 dithiothreitol (DTT) to break disulphide bonds. Refolding required slow exchange of
99 disrupting reagents with folding facilitators such as calcium (for structural integrity),
100 oxidized glutathione (to reform disulphide bridges) and hemin (to provide the
101 prosthetic heme group) [23].

102

103 **SBP: Cloning and Expression.**

104 The gene encoding SBP was first derived from a soybean plant cDNA library
105 screened with a peroxidase-specific probe [26]. The open reading frame for the SBP
106 protein was cloned into the pET-34b (+) expression vector; however, induction of
107 rSBP led to inclusion body formation and *E. coli* cell death. Active SBP was achieved
108 by a refolding strategy similar to that of HRP. Unlike rHRP refolding, however,
109 inclusion of oxidised glutathione in the refolding medium decreased active SBP
110 recovery, possibly due to the formation of mispaired disulphides [26]. Henriksen and
111 co-workers [15] also developed a recombinant SBP for crystallisation studies based
112 on previous cDNA work, in which they successfully refolded SBP from inclusion
113 bodies using a cocktail that included both oxidized and reduced glutathione [27].
114 There have been several other recent examples of recombinantly expressed plant
115 peroxidases including Hushpulia and co-workers' [28] work on tobacco anionic
116 peroxidase.

117 **Mutagenesis of HRP.**

118 Mutagenic studies on HRP began following successful cloning of a synthetic HRP
119 gene [23]. Before the elucidation of the crystal structure [29], most mutations focused
120 on ascertaining the key residues in the active site. Before 1997, researchers based their
121 assumptions on crystal structures of closely related peroxidases, such as cytochrome *c*
122 peroxidase, which suggested positions 38 through 42 as key catalytic residues.
123 Mutations of Arg 38, Phe 41 and His 42 led to dramatic decreases in peroxidase
124 catalytic activity. However, some Phe 41 mutants revealed an augmented thioether
125 sulfoxidation activity owing to increased access channel area [30].

126 Asn 70 was also noted as an important residue in HRP catalysis: although it lies some
127 distance from the heme iron atom, it is hydrogen-bonded to the side chain of the distal
128 His 42. [31] Mutations in this region showed a decrease in HRP activity and a re-
129 orientation of active site residues. Mutation of Phe 221 altered the heme iron of the
130 resting enzyme to a quantum mixed-spin state [32]. Substitutions of Trp 117 revealed
131 this residue's role in protein folding and electron transfer [33]. Mutations within the
132 active site entrance revealed the key role of Phe 142 in binding aromatic molecules
133 [34], whilst mutations within the proximal region (the area below the heme plane)
134 disclosed the parts played by Phe 179 in aromatic molecule binding [35] and by His
135 170 in heme group anchorage [36]. Table 2 lists the various site directed HRP
136 mutants. Recent examples of site directed mutants of HRP have been fewer [37], but
137 Colas and de Montellano [38] identified the key role of carboxylate side chain amino
138 acids in HRP protein-heme interactions.

139 To date, there have been few reports of HRP random mutagenesis. Arnold and co-
140 workers directionally evolved HRP with the aim of increasing activity and stability.
141 Development of a stabilised recombinant HRP is of great importance to increase and
142 consolidate the range of peroxidase applications. Three rounds of random mutagenesis
143 improved expression in yeast, yielding a nine-position HRP mutant displaying an 85-
144 fold increase in activity over the parental molecule. One round of random mutagenesis
145 was also carried out to improve stability, resulting in three mutants more stable than
146 the parent in relation to temperature and H₂O₂ tolerance (supplementary online
147 material) [8, 19, 39, & 40]. Recent publications suggest that targeted, "semi-rational",
148 evolution of enzymes might yield superior mutants in less time [41 and references
149 within]. Mendive and co-workers developed a rapid screening methodology for

150 random mutants displaying increased peroxidase activity, using DAB as substrate.
151 Whereas Arnold and co-workers expressed HRP in *E. coli*, *S. cerevisiae*. and *P.*
152 *pastoris*, Mendive and co-workers utilised a baculovirus expression system [42].
153 No mutagenesis studies on recombinant SBP have been reported to date, and the
154 authors believe that this requires urgent attention. For example, mutagenesis could
155 reveal which residues in SBP endow it with its enhanced stability *vis a vis* HRP [15].
156 Also, similar to HRP, SBP could be subjected to focussed directed evolution to
157 increase the number of substrates accepted.

158

159 **HRP in Biosensors**

160 One of the most common uses of HRP is in biosensors. A biosensor is “*an analytical*
161 *device that brings together an immobilised biological sensing material [often HRP]*
162 *and a transducer to produce an electronic signal that is proportional to the*
163 *concentration of the target chemical substance”* [43]. Although reports of SBP-based
164 biosensors are emerging, HRP biosensor research dominates and has continued to
165 develop through many forms, from the traditional voltammetric- and amperometric-
166 based methods of detection, to nano-sized devices. Real time quantification of
167 hydrogen peroxide continues to be one of the main reasons for sensor development
168 [44], although other diverse applications include the detection of glucose [45], ethanol
169 [46] and tumour markers *in vivo* [47]. Enzyme-based biosensors require rapid and
170 uniform transfer of electrons generated at the enzyme active site to the transducer.
171 The distance between the active site and the transducer can hinder electron transfer;
172 often, posttranslational modifications such as glycosylation increase this distance.
173 Recombinant HRP, devoid of glycans, offers a shorter path for electron transfer and
174 numerous reports of rHRP-based sensors have appeared [48 and references
175 within]. While a detailed review of HRP-based sensors is beyond the scope of this
176 article, we outline some of the emerging trends in HRP-based biosensor development.
177 Biosensors, including immunosensors [49] and electroensors [50], incorporating
178 organic solvents have developed as an expanding area of peroxidase research,
179 primarily due to insolubility of many analytes in aqueous solutions. Recently, Konash
180 and Magner [51] developed a HRP-immobilised, mediated H₂O₂ sensor, which
181 demonstrated good catalytic activity in 2-butanone and ethyl acetate. Organic solvent
182 compatible bi-enzyme peroxidase sensors have also been cited in the literature [52].

183 Size reduction remains a pivotal area in sensor research. The use of nanoparticles
184 offers increased surface area for enzyme immobilisation, whilst simultaneously
185 reducing apparatus size [53]. Currently, HRP-based nano-sensors are at the forefront
186 of biosensor research [54 and Table 3].

187

188 **SBP in biosensors.**

189 Although HRP is the classical heme peroxidase, there is increasing interest in SBP.
190 SBP has advantages over HRP in terms of catalytic activity and stability [55]; these
191 can be exploited in biosensors. Also, unlike HRP, SBP is active in the pH range 2-6,
192 offering a greater range of potential biosensing applications [56]. The first SBP
193 biosensor was reported in 1995 by Vreeke and co-workers [57] as a thermostable
194 wired enzyme electrode using an osmium-based mediator, which aids electron transfer
195 from the active site to the electrode, modified by an epoxide. Kenausis and co-
196 workers [58] also used a poly(4-vinylpyridine) polymer, complexing the pyridine
197 nitrogens to the osmium-based mediator, quaternised with 2-bromoethylamine. Until
198 the use of SBP by Heller and Vreeke [59], no peroxidase-based sensors could be used
199 at 37°C for an extended period (~100 hours). Monitoring of glucose “*in vivo*” for
200 diabetes mellitus, and of lactate for confirmation of hypoxia and ischemia, are vital in
201 patient management; use of thermostable SBP immobilised into a mediator enables
202 this [59].

203 The typical electrochemical reactions of a H₂O₂ sensing, peroxidase based, osmium-
204 mediated electrode system are outlined in Box 1 [60]. In addition to conveying
205 electrons, generated at the active site, to the electrode surface, the pyridin-N-ethylene
206 groups of the osmium-containing mediator also increase the hydration and provide
207 primary amines for cross-linking [58].

208 Table 3 summarizes reports to date of SBP-based biosensors, most of which use a
209 mediator. H₂O₂ can be electrochemically detected by its electrooxidation on a Pt (or
210 other inert Pt group metal) electrode [59]. Utilisation of an enzyme/mediator system
211 produces a multi-step mediated electron-transfer (MET) process, in which each step
212 transports the electron a small distance [61]. However, use of a mediator with a redox
213 enzyme can create its own problems: the mediator-enzyme film can, depending upon
214 its thickness, obstruct substrate diffusion [62]. Chemical modification of redox
215 enzymes with an electron relay moiety can increase multi-step MET by decreasing the

216 electron transfer distance. This leads to improved electrical communication between
217 the enzyme's redox centre and its external environment [61].

218

219 **Peroxidase based Micro- and Nano-Systems**

220 An emerging field in peroxidase research is the use of micro- and nano- sized
221 structures in diagnostic and biosensing fields. Enzyme microreactors, for example,
222 permit chemical and biochemical reactions to be carried out on a microscale. HRP has
223 previously been used as a model microreactor system to monitor HRP catalytic
224 activity and as a diagnostic tool; however, the development of this field will be
225 determined by the ability to immobilise peroxidases onto suitable support structures
226 [63]. Miniaturisation of enzymatic processes is also evident in the diagnostics sector,
227 in "Lab on a Chip" and "Point of Care Testing"(POCT) research. The classical
228 application of HRP in POCT is ClinistixTM; however, recently Cho and co-workers
229 [64] have applied HRP to a portable sequential cross flow immunoanalytical device.
230 Cross flow immunoassays are capable of introducing the antigen-antibody complex
231 to the flow cell whilst sequentially extracting the catalytic signal, thus simplifying the
232 complex traditional ELISA procedure. This device demonstrates many advantages
233 over ELISA-based analytical methodologies including a rapid, sensitive and
234 inexpensive in-situ diagnosis for the presence of Hepatitis B surface antigen in a
235 sample. Miniaturisation of peroxidase-based devices also features in other research
236 fields, including the use of micro-crystals for oxidoreductase-based catalysis in
237 organic solvent [65], nano-immobilisation techniques for peroxidase based
238 wastewater treatment [66], and a more widespread use of nano-structures for
239 peroxidase based sensors [67 & 68]. Recently Yan and co-workers [69] described a
240 microcantilever based biosensor, modified with HRP, for H₂O₂ detection. In this
241 system the enzyme-functionalised microcantilevers deflected irreversibly in response
242 to H₂O₂ concentrations in the nanomolar range. The deflection was caused by
243 conformational change within the HRP molecule as it underwent oxidation by the
244 H₂O₂; the irreversibility was due to the absence of a second, reducing substrate
245 required for reversion of HRP to the resting state. This technique may also provide a
246 sensitive tool for investigating protein structural change. HRP has also been utilised as
247 the functional component of self-assembled three-dimensional (3-D) nano-structures.
248 Rauf and co-workers [70] utilised self-assembly layer-by-layer technology to

249 construct controlled 3-D catalytically active nano-structures. This method of
250 peroxidase immobilisation allows for increased catalytic activity per unit area, and
251 will aid in the miniaturisation of biosensors, biochips and immobilised biocatalysts.
252 With increasing sophistication of support structures on the micro- and nano-scale,
253 miniaturisation of peroxidase-based devices will continue to develop in the future,
254 particularly in the fields of POCT and biosensing.

255

256 **Peroxidase based Biocatalysis.**

257 A major shortcoming of all heme-dependent peroxidases is their low operational
258 stability, owing to oxidative degeneration of the heme group [71]. Operational
259 stability of SBP can be increased by generating H_2O_2 *in situ* from glucose and O_2 .
260 When co-immobilized with glucose oxidase in a polyurethane foam, SBP could act as
261 a peroxygenase to convert thioanisole to its sulphoxide (i.e. by inserting an oxygen
262 atom). Here, SBP uses the H_2O_2 generated *in situ* by glucose oxidase; it formed no
263 sulphoxide with free, exogenous H_2O_2 [71]. Such an arrangement avoids excessive
264 initial H_2O_2 concentrations and, hence, formation of compound III (a reversible dead-
265 end complex formed from compound II in the presence of an excess H_2O_2 , which
266 slowly reverts to native enzyme; [10]) and/or irreversibly inhibited SBP. HRP has
267 been subjected to intense experimentation, including a large body of work focussing
268 on site directed mutants (see above and Table 2). Now that the key catalytic residues
269 are known, researchers have begun to use site directed mutagenesis to alter the
270 function of the HRP molecule [48], e.g. by construction of an improved luminol
271 binding site [72]. Directed evolution stabilised HRP against thermal denaturation [39]
272 and has endowed it with increased H_2O_2 tolerance and increased catalytic activity
273 [40]. Further targeted directed evolution, focussing on the substrate access channel
274 and binding pocket, could allow HRP to accept an increased variety of substrates [73],
275 and promote further diversification of HRP applications in organic synthesis [41].
276 Peroxidase catalysis in organic solvents, both aqueous and anhydrous, offers a huge
277 advantage to organic chemists, as difficult asymmetric oxidation and reduction
278 reactions can take place rapidly and with high specificity [74]. The major problems of
279 substrate solubility and unwanted side reactions promoted by water are also overcome
280 during organic solvent based synthesis. Additionally, in some anhydrous solvents
281 peroxidase (HRP and SBP) activity was actually increased [75], with additional

282 methods, such as salt activation [76] and excipient aided lyophilisation [77] also
283 resulting in increased peroxidase activity. However, in some low water solvents,
284 peroxidases can lose their conformational structure [78]; although recent advances in
285 peroxidase encapsulation in amphiphilic matrices [79], the use of reverse micelles
286 [80] and oil emulsions [81] allow for peroxidase activity in an extended range of
287 anhydrous solvents. Reactions carried out in these solvents include hydroxylations, N-
288 demethylations and sulphoxidations [2]. An interesting recent environmental
289 application of SBP polymerisation in organic solvents is the production of
290 polycardanol in as a potential anti-biofouling agent [82], whilst recently it has been
291 noted that HRP requires a mediator to catalyse the same substrate [83]. The interested
292 reader is directed to a recent review of this area [74].

293

294 **Peroxidase based Bioremediation.**

295 Highly expressed, stabilized, recombinant HRP [39, 40] could be very useful for
296 wastewater cleanup, provided the recombinant enzyme can be produced cost
297 effectively and in sufficient quantities. Phenol cleanup by HRP has been widely
298 reviewed [1] but several drawbacks limit its widespread application, including
299 intolerance of high concentrations of the primary substrate H₂O₂ [84], low enzymatic
300 reusability, and financial costs. Plant heme peroxidase expression in *E.coli* can be
301 frustrating, but advances in peroxidase expression, without formation of inclusion
302 bodies, may pave the way for increased production of recombinant peroxidases (See
303 supplementary online material). SBP has proven itself a worthy alternative
304 peroxidase: it displays superior stability and activity characteristics to the classical
305 HRP. However, research into this enzyme lags far behind HRP. SBP can effectively
306 cleanup phenolic wastewater, yet recent publications cite HRP as being a superior,
307 albeit less stable, catalyst than SBP for phenol cleanup [85]. Development of an
308 enhanced catalytic SBP mutant would provide a powerful tool for wastewater
309 treatment. Bódalo and co-workers [85] noted that the choice of peroxidase for
310 wastewater treatment also depends on effluent characteristics, operational
311 requirements and costs. SBP has been shown to outperform HRP in oxidative dye
312 removal [86]. SBP, possibly owing to its larger substrate access channel, and, hence,
313 greater exposure of the catalytically important delta heme edge, can accept more
314 substrates than HRP [87].

315

316 **HRP: an Unlikely Therapeutic**

317 An exciting application of HRP is as a novel cancer treatment via gene-directed
318 enzyme/prodrug therapy. It has been noted that the non-toxic HRP substrate, indole-3-
319 acetic acid (IAA), forms a radical that is toxic to cancer cells upon HRP catalysis. The
320 exact mechanism of toxicity remains unclear: it is believed to involve lipid
321 peroxidation induced by the free radical formation [88]; however, in human
322 melanoma cells, death receptor-mediated and mitochondrial apoptotic pathways are
323 known to be involved also [89]. Leaving aside the actual reason(s) for toxicity,
324 inactive IAA can be introduced to the body; and then becomes activated by HRP at
325 the region of interest. Localisation of the HRP molecule is achieved via its
326 conjugation to an antibody specific to an extracellular tumour antigen. This approach
327 has become the focus of much research and numerous clinical trials, due to several
328 attractive features: these include the robust nature of the activating enzyme and the
329 low toxicity of the prodrug [90]. HRP has been shown to activate other pro-drugs
330 including ellipticine [91] and halogenated IAA derivatives [92]. The interested reader is
331 directed to the excellent recent review of Dachs et al. [93].

332 **Conclusion and Future Directions:**

333 As outlined, peroxidases are widely studied and very important enzymes, with many
334 applications in the life sciences and beyond. They remain pivotal to advancing
335 biotechnology, and as such, we present two clearly distinct, yet similar members of
336 this classical family. Continued research into the “traditional” HRP, has been
337 accompanied by the slow, but steady progression of SBP. Crude SBP, isolated from
338 waste soybean hulls, offers a cheap bulk peroxidase catalyst for applications such as
339 wastewater treatment and organic synthesis, whilst the more costly peroxidase
340 alternatives (plant HRP and recombinant HRP and SBP) will prove themselves in
341 higher value niches, such as diagnostics and therapeutics. With improved
342 understanding of the catalytic and stability characteristics, the detection of new
343 substrates and the increasing use of implantable devices in the medical field, SBP will
344 rapidly develop its own high value market niche. As noted for HRP, use of
345 recombinant SBP would also benefit the biosensor field by permitting more rapid
346 electron transfer, due to the lack of protein glycosylation. Improvement of these two
347 peroxidases, by rational mutation and “focussed” directed evolution, will widen their
348 applications and expand their roles as key biotechnological tools in the future.

349

350

351

352

353

354

355

356

357 **Acknowledgements**

358 We thank the following for financial support: the Irish Research Council for Science, Engineering and
359 Technology (Embark Initiative postgraduate scholarship, BJR) the National Centre for Sensors
360 Research and School of Biotechnology, Dublin City University (NC) and the Research Advisory Panel
361 of Dublin City University (Albert College Award, C.Ó'F.) (The National Centre for Sensors Research
362 was established under the Programme for Research in Third Level Institutions of the Higher Education
363 Authority)

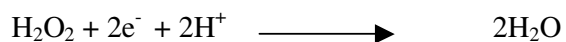
1. Wanger M. and Nicell J.A. Detoxification of phenolic solutions with horseradish peroxidase and hydrogen peroxide. (2002), *Water Research*, **36**, (16), 4041-4052.
2. van de Velde F. et al. (2001), Improving the catalytic performance of peroxidase in organic synthesis. *Trends in Biotechnology*, **19**, (2), 73-80.
3. Dotsikas Y. and Loukas Y.L. (2004). Employment of 4-(1-imidazolyl)phenol as a luminol signal enhancer in a competitive-type chemiluminescence immunoassay and its comparison with the conventional antigen-horseradish peroxidase conjugate-based assay. *Analytica Chimica Acta*, **509**, (1), 103-109.
4. Azevedo A.M. et al. (2003). Horseradish peroxidase: a valuable tool in biotechnology. *Biotechnology Annual Review*, **9**, 199-247.
5. Jung Y. et al. (2001). High-level production of heme-containing holoproteins in *Escherichia coli*. *Applied Microbiological Biotechnology*, **55**, (2), 187-191.
6. Veitch N.C. (2004). Horseradish peroxidase: a modern view of a classic enzyme. *Phytochemistry*, **65**, (3), 249-259.
7. Duroux L. and Welinder K.G. (2003). The Peroxidase Gene Family in Plants: A Phylogenetic Overview. *Journal of Molecular Evolution*, **57**, (4), 397-407.
8. Morawski B. et al. (2000). Functional expression of horseradish peroxidase in *Saccharomyces cerevisiae* and *Pichia pastoris*. *Protein Engineering*, **13**, (5), 377-384.
9. Tupper J. et al. (2004). Use of horseradish peroxidase for gene-directed enzyme prodrug therapy with paracetamol. *British Journal of Cancer*, **90**, (9), 1858-1862.
10. Dunford H.B. (1999). "Heme Peroxidases". Wiley and Sons, New York, pp 1-111[E3].
11. Welinder K.G. (1992). Superfamily of Plant, fungal and bacterial peroxidases. *Current Opinion in Structural Biology*, **2**, 388-393.
12. Hoyle M.C. (1977). High-resolution of peroxidase-indoleacetic acid oxidase isoenzymes from Horseradish by isoelectric-focusing. *Plant Physiology*, **60**, 787-793.
13. dos Santos W.D. et al. (2004). Lignification and related enzymes in Glycine max root growth-inhibition by ferulic acid. *Journal of Chemical Ecology*, **30**, (6), 1203-1212.
14. Schmitz et al. (1997). Characterisation of anionic Soybean Peroxidase (Glycine Max) Seed coat peroxidase. *Canadian Journal of Botany*, **75**, 1336-1341.
15. Henriksen A. et al. (2001). Structure of soybean seed coat peroxidase: A plant peroxidase with unusual stability and haem-apoprotein interactions. *Protein Science*, **10**, (1), 108-115.
16. Hoffmann F. et al. (2004). Minimising inclusion body formation during recombinant protein production in *Escherichia coli* at bench and pilot scale. *Enzyme and Microbial Technology*, **34**, (3-4), 235-241.
17. Teilmann K. et al. (1999). Disulphide bond formation and folding of plant peroxidases expressed as inclusion body protein in *Escherichia coli* thioredoxin reductase negative strains. *Protein Expression and Purification*, **15**, (1), 77-82.
18. Zhang Y. et al. (1998). Expression of eukaryotic proteins in soluble form in *Escherichia coli*. *Protein Expression and Purification*, **12**, (2), 159-165.
19. Lin Z. et al. (1999). Functional Expression of Horseradish Peroxidase in *E. coli* by directed evolution. *Biotechnology Progress*, **15**, 467-471.
20. Wandersman C. and Stojiljkovic I. (2000). Bacterial heme sources: the role of heme, hemoprotein receptors and hemophores. *Current Opinion in Microbiology*, **3**, (2), 215-220.
21. Woodard S.I. and Dailey H.A. (1995). Regulation of Heme Biosynthesis in *Escherichia coli*. *Archives of Biochemistry and Biophysics*, **316**, (1), 110-115
22. Varnado C.L and Goodwin D.C. (2004). System for the expression of recombinant hemoproteins in *Escherichia coli*. *Protein Expression and Purification*, **35**, (1), 76-83.
23. Smith A.T. et al. (1990). Expression of a synthetic gene for horseradish peroxidase in *Escherichia coli* and folding and activation of the recombinant enzyme with Ca²⁺ and Heme. *Journal of Biological Chemistry*, **265**, (22), 13335-13343.
24. Welinder K G. (1979). Amino acid sequence studies of horseradish peroxidase. Amino and carboxy termini, cyanogen bromide and tryptic fragments, the complete sequence, and some structural characteristics of horseradish peroxidase C. *European Journal Biochemistry*, **96**, 483-502.
25. Gustafsson C. et al. (2004). Codon Bias and heterologous protein expression. *Trends in Biotechnology*, **22**, (7), 346-353.
26. Chen H. and Vierling R.A. (2000). Molecular cloning and characterisation of soybean peroxidase gene families. *Plant Science*, **150**, (2), 129-137.
27. Gijzen M. (1997). A deletion mutation at the ep locus causes low seed coat peroxidase activity in soybean. *The Plant Journal*, **12**, (5), 991-998.

28. Hushpulia D.M. (2003). Expression and refolding of tobacco anionic peroxidase from E-coli inclusion bodies. *Biochemistry (Moscow)*, **68**, (11), 1189-1194.
29. Gajhede M. et al. (1997). Crystal structure of horseradish peroxidase C at 2.15 Å resolution. *Nature Structural Biology*, **4**, (12), 1032-1038.
30. Ozaki S. and Ortiz de Montellano P.R. (1995). Molecular engineering of horseradish peroxidase. Thioether sulfoxidation and styrene epoxidation by Phe-41 leucine and threonine mutants. *J.Am.Chem.Soc.*, **117**, (27), 7056-7064.
31. Nagano S, Tanaka M, Ishimori K, Watanabe Y. and Morishima I. (1996). Catalytic roles of the distal site asparagine-histidine couple in peroxidases. *Biochemistry*, **35**, (45), 14251-14258.
32. Howes B.D et al. (2001). Heme-linked interactions in horseradish peroxidase revealed by spectroscopic analysis of the Phe-221 to Met mutant. *Biochem. J.*, **353**, (2), 181-191.
33. Ignatenko O.V. et al. (2000). Catalytic properties of tryptophanless recombinant horseradish peroxidase. *Biochemistry (Moscow)*, **65**, (5), 583-587.
34. Veitch N.C. et al. (1995). Solution characterisation by NMR spectroscopy of two horseradish peroxidase isoenzyme C mutants with alanine replacing either Phe142 or Phe143. *European Journal of Biochemistry*, **233**, (2), 650-658.
35. Veitch N.C. et al. (1997). Identification of a critical phenylalanine residue in horseradish peroxidase, Phe179, by site-directed mutagenesis and H-NMR. Implications for complex formation with aromatic donor molecules. *Biochemistry*, **36**, (48), 14751-14761.
36. Newmyer S.L. et al. (1996). Rescue of the horseradish peroxidase His-170 to Ala mutant activity by imidazole. Importance of proximal ligand tethering. *Biochemistry*, **35**, (39), 12788-12795.
37. Howes B.D. et al. (2005). Spectroscopic and kinetic properties of the horseradish peroxidase mutant T171S. *FEBS Journal*, **272**, 5514-5521
38. Colas C. and de Montellano P. R. O. (2004). Horseradish Peroxidase Mutants That Autocatalytically Modify Their Prosthetic Heme Group. *Journal of Biological Chemistry*, **279**, (23), 24131-24140.
39. Arnold F.H. and Lin, Z. (2000). California Institute Of Technology "Expression of Functional Eukaryotic Proteins". Worldwide Patent: PCT/US99/17127, WO 00/006718.
40. Morawski B. et al. (2001). Functional expression and stabilisation of HRP by directed evolution in *S. cerevisiae*. *Biotechnology and Bioengineering*, **76**, (2), 99-107.
41. Kazlauskas R. (2005). Biological chemistry: Enzymes in focus. *Nature*, **436**, 1096-1097.
42. Mendive F.M. et al. (2003). Construction of a mutant library of horseradish peroxidase gene by directed evolution and development of an *in situ* screening method. *Braz. J. Chem. Eng.*, **20**, (1), 33-38.
43. Luong J.H.T. et al. (1995). Enzyme reactions in the presence of cyclodextrins: biosensors and enzyme assays. *Trends in Biotechnology*, **13**, 457-463.
44. Tønning E. et al. (2005). Chemometric exploration of an amperometric biosensor array for fast determination of wastewater quality. *Biosensors and Bioelectronics*, **21**, (4), 608-617.
45. Alonso Lomillo M. A. et al. (2005). Biosensor based on platinum chips for glucose determination. *Analytica Chimica Acta*, **547**, (2), 209-214.
46. Azevedo A. M. et al. (2005). Ethanol biosensors based on alcohol oxidase. *Biosensors and Bioelectronics*, **21**, (2), 235-247.
47. Lin J. and Ju, H. (2005). Electrochemical and chemiluminescent immunosensors for tumour markers. *Biosensors and Bioelectronics*, **20**, (8), 1461-1470.
48. Ferapontova E. (2004). Direct peroxidase bioelectrocatalysis on a variety of electrode materials. *Electroanalysis*, **16**, (13), 1101-1112.
49. Penalva J. et al. (1999). Analytical properties of Immunosensors working in organic solvents. *Analytical Chemistry*, **71**, 3862-3872.
50. Campanella L. et al. (1999). Analysis of several real matrices using mono-, bi-enzymatic or inhibition organic phase enzyme electrodes. *Analytica Chimica Acta*, **393**, 109-120.
51. Konash, A. and Magner E. (2005). Electrochemically Mediated Reduction of Horseradish Peroxidase by 1,1'-Ferrocene Dimethanol in Nonaqueous Solvents, *Anal. Chem.*, **77**,1647-1654.
52. Castillo J. et al. (2003). Bienzyme biosensors for glucose, ethanol and putrescine built on oxidase and sweet potato peroxidase. *Biosensors and Bioelectronics*, **18**, 705-714.
53. Liu, Y.-L. et al. (2005). Nanosized flower-like ZnO synthesized by a simple hydrothermal method and applied as matrix for horseradish peroxidase immobilization for electro-biosensing. *Journal of Inorganic Biochemistry*, **99**, (10), 2046-2053.
54. Wu L et al. (2006). Electrochemical immunoassay for CA125 based on cellulose acetate stabilized antigen/colloidal gold nanoparticles membrane. *Electrochimica Acta*, **51**, (7), 1208-1214.

55. Kamal J.K. and Behere DV. (2003). Activity, stability and conformational flexibility of seed coat soybean peroxidase. *Journal of Inorganic Biochemistry*, **94**, (3), 236-42.
56. Wang B.Q. et al. (1999). Sol-Gel thin film immobilized soybean peroxidase biosensor for the amperometric determination of hydrogen peroxide in acid medium. *Analytical Chemistry*, **71**, 1935-1939.
57. Vreeke M.S. et al. (1995). A thermostable hydrogen peroxide sensor based on "wiring" of soybean peroxidase. *Analytical Chemistry*, **67**, 4247-4249.
58. Kenausis G. et al. (1997). Electrochemical glucose and lactate sensors based on "wired" thermostable soybean peroxidase operating continuously and stably at 37°C. *Analytical Chemistry*, **69**, 1054-1060.
59. Heller A. and Vreeke M. S. (1997). E. Heller and Company. *Soybean peroxidase electrochemical sensor*. United States Patent. US. 5665222.
60. Li W. et al. (2000). Fabrication of multilayer films containing horseradish peroxidase and polycation-bearing Os complex by means of electrostatic layer-by-layer adsorption and its applications as a hydrogen peroxide sensor. *Analytica Chimica Acta*. **418**, 225-232.
61. Katz, E. (2002). Mediated electron-transfer between redox-enzymes and electrode supports. IN. A. Bard and J. Stratmann. *Encyclopaedia of Electrochemistry, Bioelectrochemistry*, **9**, New York. John Wiley and Sons Inc, 561-626.
62. Sun Y. et al. (1998). Chemically modified electrode via layer-by-layer deposition of glucose oxidase (GOD) and polycation-bearing Os complex. *Thin Solid Films*, **327-329**, 730-733.
63. Verpoorte E. (2002). Microfluidic chips for clinical and forensic analysis. *Electrophoresis*, **23**, (5), 677-712.
64. Cho J-H. et al. (2005). An enzyme immunoanalytical system based on sequential cross-flow chromatography. *Analytical Chemistry*, **77**, 4091-4097.
65. Kreiner M. and Parker M-C. (2005). Protein-coated microcrystals for use in organic solvents: application to oxidoreductases. *Biotechnology Letters*, **27**, 1571-1577.
66. Patel D.S. et al. (2005). Nano-assembly of manganese peroxidase and lignin peroxidase from *P. chrysosporium* for biocatalysis in aqueous and non-aqueous media. *Colloids and Surfaces B*, **45**, 13-19.
67. Zhu J-J. et al. (2005). Reagentless electrochemical biosensor based on the multi-wall carbon nanotubes and nanogold particles composite film. *Frontiers in Bioscience*, **10**, 521-529.
68. Lee Y-M. et al. (2006). Immobilisation of horseradish peroxidase on multi-wall carbon nanotubes and its electrochemical properties. *Biotechnology Letters*, **28**, 39-43.
69. Yan X. et al. (2006). Microcantilevers modified by horseradish peroxidase: intercalated nano-assembly for hydrogen peroxide detection. *Analytical Sciences*, **22**, 205-208.
70. Rauf S. et al. (2006). Building three-dimensional nanostructures with active enzymes by surface templated layer by layer assembly. *Chem. Commun.*, **16**, 1721-1723.
71. van de Velde F. et al. (2000) Improved operational stability of peroxidases by coimmobilization with glucose oxidase. *Biotechnology and Bioengineering*, **69**, (3), 286-291.
72. Tanaka M. et al (1999). Luminol activity of horseradish peroxidase mutants mimicking a proposed binding site for luminol in *Arthromyces ramosus* peroxidase. *Biochemistry*, **38**, (32), 10463-10473.
73. Smith A.T. and Veitch N.C. (1998). Substrate binding and catalysis in heme peroxidases. *Current Opinion in Chemical Biology*, **2**, 269-278.
74. Klivanov A.M. (2003). Asymmetric enzymatic oxidoreductions in organic solvents. *Current Opinion in Biotechnology*, **14**, 427-431.
75. Dai L. and Klivanov A.M. (2000). Peroxidase-catalysed asymmetric sulfoxidation in organic solvents versus water. *Biotechnology and Bioengineering*, **70**, (3), 353-357.
76. Morgan J.A. and Clark D.S. (2004). Salt activation of nonhydrolase enzymes for use in organic solvents. *Biotechnology and Bioengineering*, **85**, (4), 456-459.
77. Dai L. and Klivanov A.M. (1999) Striking activation of oxidative enzymes suspended in non-aqueous media. *Proc. Nat. Acad. Sci. USA*, **96**, 9475-9478.
78. Azevedo A.M. et al. (2001). Stability of free and immobilised peroxidase in aqueous-organic solvents mixtures. *Journal Molecular Catalysis B*, **15**, 147-153.
79. Bruns N. and Tiller J.C. (2005). Amphiphilic Network as nanoreactor for enzymes in organic solvents. *Nano Letters*, **5**, (1), 45-48.
80. Michizoe J. et al. (2003). Control of water content by reverse micellar solutions for peroxidase catalysis in a water-immiscible organic solvent. *Journal of Bioscience and Bioengineering*, **95**, (4), 425-427.

81. Kamiya N. et al. (2000). Catalytic and structural properties of surfactant-Horseradish Peroxidase complex in organic media. *Biotechnology Progress*, **16**, 52-58.
82. Kim Y.H. et al. (2003). Polymerization of cardanol using soybean peroxidase and its potential application as anti-biofilm coating material. *Biotechnology Letters*, **25**, 1521-1524.
83. Won K. et al. (2004). Horseradish peroxidase-catalysed polymerisation of cardanol in the presence of redox mediators. *Biomacromolecules*, **5**, (1), 1-4.
84. Nicell J.A. and Wright H. (1997). A model of peroxidase activity with inhibition by hydrogen peroxide. *Enzyme and Microbial Technology*, **21**, 302-310.
85. Bódalo A. et al. (2005). Comparison of commercial peroxidases for removing phenol from water solutions. *Chemosphere*, **63**, (4), 626-632.
86. Knutson K et al. (2005). Enzymatic biobleaching of two recalcitrant paper dyes with horseradish and soybean peroxidase. *Biotechnology Letters*, **27**, (11), 753-8.
87. Alpeeva I.S. and Sakharov I.Y. (2005). Soybean peroxidase-catalyzed oxidation of luminol by hydrogen peroxide. *Journal of Agriculture and Food Chemistry*, **53**, (14), 5784-8.
88. Folkes L.K. and Wardman P. (2001). Oxidative activation of indole-3-acetic acids to cytotoxic species – A potential new role for plant auxins. *Biochem. Pharmacol.*, **61**, 129-136.
89. Kim D-S. et al. (2004). Oxidation of indole-3-acetic acid by horseradish peroxidase induces apoptosis in G361 human melanoma cells. *Cellular Signalling*, **16**, 81-88.
90. Tafazolli S. and O'Brien, P.J. (2004). Prooxidant Activity and Cytotoxic effects of Indole-3-acetic Acid derivative Radicals. *Chem. Res. Toxicol.*, **17**, 1350-1355.
91. Poljaková J. et al. (2005). Oxidation of an antitumor drug ellipticine by peroxidases. *Biomed. Pap. Med. Fac. Univ. Palacky. Olomouc. Czech Repub.*, **149**, (2), 449-453.
92. Rossiter S. et al. (2002). Halogenated Indole-3-acetic Acids as Oxidatively Activated Prodrugs with potential for targeted Cancer Therapy. *Bioorganic and Medical Chemistry Letters*, **12**, 2523-2526.
93. Dachs G.U. et al. (2005). From bench to bedside for gene-directed enzyme prodrug therapy of cancer. *Anti-Cancer Drugs*. **16**, (4), 349-359.
94. Welinder K.G. and Larsen Y.B. (2004). Covalent structure of soybean seed coat peroxidase. *Biochimica et Biophysica Acta*, **1698**, (1), 121-126.
95. Wuhrer M. et al. (2005). New features of site-specific horseradish peroxidase (HRP) glycosylation uncovered by nano-LC-MS with repeated ion-isolation/fragmentation cycles. *Biochimica et Biophysica Acta.*, **1723**, (1-3), 229-239.
96. Gray J.S.S. and Montgomery R. (2006). Asymmetric glycosylation of soybean seed coat peroxidase. *Carbohydrate Research*, **341**, (2), 198-209.
97. Paul K.G. (1963). "Peroxidases" IN *The Enzymes* (Boyer P.D. ed), Academic Press, **8**, 227-237.
98. Rodriguez-Lopez J.N. et al. (1996). Role of Arginine 38 in horseradish peroxidase a critical residue for substrate binding and catalysis. *Journal of Biological Chemistry*, **271**, (8), 4023-4030.
99. Newmyer S.L. and de Montellano P.R.O. (1996). Rescue of the catalytic activity of an H42A mutant of horseradish peroxidase by exogenous imidazoles. *Journal of Biological Chemistry*, **271**, (25), 14891-14896.
100. Gazaryan I.G. et al. (1994). Effect of single-point mutations Phe-41-His and Phe-143-Glu on folding and catalytic properties of recombinant horseradish peroxidase expressed in E. coli. *FEBS Letters*, **354**, (3), 248-250.
101. Veitch N.C. et al. (1995). Solution characterisation by NMR spectroscopy of two horseradish peroxidase isoenzyme C mutants with alanine replacing either Phe142 or Phe143. *European Journal of Biochemistry*, **233**, (2), 650-658.
102. Howes B.D. et al. (2001). Mutation of residues critical for benzohydroxamic acid binding to horseradish peroxidase isoenzyme C. *Biopolymers*, **62**, (5), 261-267.
103. Mukai M. et al. (1997). Effects of concerted hydrogen bonding of distal histidine on active site structures of horseradish peroxidase. Resonance Raman studies with Asn70 mutants. *J. Am. Chem. Soc.*, **119**, (7), 1758-1766.
104. Gazaryan I.G. et al. (1999). Tryptophanless recombinant horseradish peroxidase. Stability and catalytic properties. *Biochem. Biophys. Res. Commun.*, **262**, (1), 297-301.
105. Morimoto A. et al. (1998). Detection of a tryptophan radical as an intermediate species in the reaction of horseradish peroxidase mutant (Phe-221 to Trp) and hydrogen peroxide. *Journal of Biological Chemistry*, **273**, (24), 14753-14760.
106. Wang B. et al. (2001). Acid-Stable Amperometric Soybean Peroxidase Biosensor Based on a Self-Gelatinizable Grafting Copolymer of Polyvinyl Alcohol and 4-Vinylpyridine. *Electroanalysis*, **13**, (7), 555-558.

107. Zhang Z. et al. (2002). Direct voltammetry and catalysis with *Mycobacterium tuberculosis* catalase-peroxidase, peroxidases, and catalase in lipid films. *Analytical Chemistry*, **74**, (1), 163-70.
108. Song Y. et al. (2006). A novel hydrogen peroxide sensor based on horseradish peroxidase immobilized in DNA films on a gold electrode. *Sensors and Actuators B*, **114**, (2), 1001-1006.
109. Lei C.-X. et al. (2004). An amperometric hydrogen peroxide biosensor based on immobilizing horseradish peroxidase to a nano-Au monolayer supported by sol-gel derived carbon ceramic electrode. *Bioelectrochemistry*, **65**, (1), 33-39.
110. Castilho J.T. et al. (2005). Amperometric biosensor based on horseradish peroxidase for biogenic amine determinations in biological samples. *Journal of Pharmaceutical and Biomedical Analysis*, **37**, (4), 785-791.
111. Li S. and Chen A. (2005). Coadsorption of Horseradish Peroxidase with Thionine on TiO₂ nanotubes for biosensing. *Langmuir*, **21**, 8409-8413.
112. Yuan R. et al. (2005). Electrochemical characteristics of a platinum electrode modified with a matrix of polyvinyl butyral and colloidal Ag containing immobilized horseradish peroxidase. *Anal. Bioanal Chem.*, **381**, 762-768.
113. Schuvailo O.N. et al. (2005). Carbon fibre-based microbiosensors for in vivo measurements of acetylcholine and choline. *Biosensors and Bioelectronics*, **21**, (1), 87-94.
114. Rahman M.A. et al. (2004). A performance comparison of choline biosensors: anodic or cathodic detections of H₂O₂ generated by enzyme immobilized on a conducting polymer. *Biosensors and Bioelectronics*, **19**, (12), 1565-1571.
115. Ferapontova E. and Gorton L. (2002). Effect of pH on direct electron transfer in the system gold electrode-recombinant horseradish peroxidase. *Bioelectrochemistry*, **55**, (1-2), 83-87.
116. Lindgren A. et al. (1999). Direct electron transfer catalysed by recombinant forms of horseradish peroxidase: insight into the mechanism. *Electrochemistry Communications*, **1**, 171-175.
117. Shipovskov S. et al (2004). Recombinant horseradish peroxidase- and cytochrome c-based two-electrode system for detection of superoxide radicals. *Bioelectrochemistry*, **63**, (1-2), 277-280.
118. Ferapontova E. et al. (2001). P-Chip and P-Chip Bienzyme Electrodes Based on Recombinant Forms of Horseradish Peroxidase Immobilized on Gold Electrodes. *Biochemistry (Moscow)*, **66**, (8), 832-839.



Box[E4] 1: Typical biosensor based on a mediated peroxidase (Px) reaction scheme. Upon the addition of H_2O_2 , Px catalyses the reaction forming water; in the process, Px goes through its catalytic cycle. This causes the mediator to go from its resting state of Os^{2+} to Os^{3+} . The osmium species is seen as a one-electron donor, used as the mediator to assist in electron transfer from the active site of Px to the electrode surface. Px_{ox} corresponds to the catalytic intermediate Compound I, formed by a two-electron oxidation. The individual one-electron reduction steps that take place on the enzyme itself (formation of catalytic intermediate Compound II and reversion to resting enzyme, Px) have been omitted from equation 2 for the sake of clarity. Adapted from *Analytica Chimica Acta*. **418**, Li W. et al. (2000). Fabrication of multilayer films containing horseradish peroxidase and polycation-bearing Os complex by means of electrostatic layer-by-layer adsorption and its applications as a hydrogen peroxide sensor. 225-232. [60] Copyright 2000, with permission from Elsevier.

Table 1: Comparison of the biochemical and structural properties of HRP and SBP. Key references are noted.

	Horseradish Peroxidase	Soybean Peroxidase
Species Name	<i>Armoracia rusticana</i> [23]	<i>Glycine max</i> [14]
Number of Amino Acids	306 [23]	306 [94]
Enzyme Classification	1.11.1.7 [29]	1.11.1.7 [15]
PDB Accession Number	1ATJ	1FHF
Molecular Weight	44,100 Da [6]	40,660 Da [15]
Carbohydrate	7,580 Da [6]	7,400 Da [15]
Heme Group	550 Da [6]	550 Da [15]
Calcium Ions	80 Da [6]	80 Da [15]
Glycosylation Sites	Asn: 13, 57, 158, 186, 198, 214, 255, 268, 316 [95]	Asn: 185, 197, 211, 216 [96]
pI	9.0 [24]	4.1 [14]
pH Activity Range	4-8 [97]	2-10 [15]
Secondary Structure	13 α -helices 3 β -sheets [29]	13 α -helices 2 β -sheets [15]
Disulphide Bridges	11-91, 44-49, 97-301, 177-209 [29]	11-91, 44-49, 97-299, 176-208 [15]
<i>In vivo</i> localisation	Roots, cell wall, vacuoles [6]	Hourglass cells, seed coat [14]

Table 2: Summary of HRP site directed mutants[E5]

Mutations		Conclusions
Active Site. [98, 99, & 100]		
R38A	F41T	Arg38 and His42 are key residues in enzyme catalysis.
R38E	F41V	Arg38 and His42 mutations decrease Compound I formation.
R38G	F41W	Mutants affect reactivity towards reducing substrates. NB- Morimoto ref.
R38H	H42A	Arg 38 and His42 are acid base partners. Arg 38 stabilises His 42.
R38K	H42E	Arg38 and His42 operate in concert to distally bind BHA.
R38L	H42L	Arg38 and His42 are dioxygen-heterolytic cleavers
R38S	H42Q	Arg38 plays a role in H ₂ O ₂ binding and cleavage.
F41A	H42R	Hydrophobicity of active site region is critical in enzymatic catalysis.
F41H	H42V	Space creating active site mutants alter substrate specificity.
F41L		Phe41 acts as hydrophobic barrier between Arg38 and His42.
Active Site Entrance. [101]		
S35K	F143E	Phe 142 plays a critical role in aromatic substrate binding.
F142A	F176E	Charged residues are important at the active site entrance.
F143A		Luminol binds to active site via electrostatic interactions in binding area.
Proximal Region. [102 & 36]		
F179A	H170A	Phe 179 is crucial for aromatic substrate binding.
F179H	F172T	His 170 tethers heme moiety in position. Prevents distal His coordination.
F179S		His 170 maintains heme moiety in penta-coordinated state.
Asparagine 70. [103]		
N70V	N70D	Asn70 hydrogen bonds to His42, mutations alter distal heme orientation. Mutant protein displays increased redox potential.
Tryptophan 117. [33 & 104]		
W117F		Mutants displayed increased acid stability. Trp 117 is important in internal electron transfer and protein unfolding.
Threonine 171. [37]		
T171S		Proximal structural alteration, affects proximal pocket hydrogen bonding.
Phenylalanine 221. [105 & 32]		
F221M	F221W	Mutants display decreased stability in alkaline conditions. Trp introduction destabilises protein, due to unfavourable surroundings.

Footnote:

Mutants are grouped into active site, active site entrance, proximal heme region, asparagine 70, tryptophan 117 and Phe 221 mutations. Mutants were expressed in a variety of hosts including *E.coli*, *Trichoplusia ni*, and *Spodoptera frugiperda* cell lines. Key references only are noted for each collection of mutants; further references may be found within these.

Electrode type	Size of Electrode	Enzyme	Method of immobilisation	Mediator	Analyte measured	Method of measurement	Reference
Soybean Peroxidase							
Glassy carbon	4mm	SBP	Adsorption	PVA/PVP	H ₂ O ₂	Amperometry	[106]
Glassy carbon	4mm	SBP	Entrapment	Sol-gel	H ₂ O ₂	Amperometry	[56]
Glassy carbon	3mm	SBP	Entrapment	Pos-EA, PEGDGE	H ₂ O ₂	Amperometry	[59]
Pyrolytic graphite (Rotating disk)	0.2mm	SBP	Entrapment	DMPC	H ₂ O ₂	Amperometry /CV	[107]
Glassy carbon	3mm	GOx/SBP	Entrapment	Pos-EA	Glucose	Amperometry	[58]

Horseradish Peroxidase							
Gold	-	HRP	Adsorption	DNA	H ₂ O ₂	Amperometry	[108]
Glassy Carbon	3mm	HRP	Adsorption	HQ	H ₂ O ₂	Amperometry	[53]
Carbon Ceramic	6mm	HRP	Covalent	Nano Au	H ₂ O ₂	Amperometry	[109]
Carbon Paste	7mm	HRP	Entrapment	GP	Biogenic Amine	Amperometry	[110]
Titanium	0.8mm	HRP	Adsorption	Thionine	H ₂ O ₂	Amperometry	[111]
Platinum	1mm	HRP	Entrapment	Co(bpy)	H ₂ O ₂	Amperometry	[112]
Carbon Fibre	3mm	Cho/HRP	Adsorption	Os- PVP	Choline	Amperometry	[113]
Platinum	7mm	Cho/HRP	Adsorption	TTCA	Choline	Amperometry	114

Recombinant HRP							
Gold	0.15mm	rHRP	Adsorption	Direct	H ₂ O ₂	Amperometry	[115]
Graphite Rotating disk	3mm	rHRP	Adsorption	Direct	H ₂ O ₂	Amperometry	[116]
Gold	0.3mm	CytC / rHRP	Adsorption	Direct	Superoxide Anion radical	Amperometry	[117]
Gold	0.15mm	L-LO / rHRP	Adsorption	Direct	L-lysine	Amperometry	[118]

Table 3: Some SBP, HRP and rHRP biosensors, and their properties, from the literature.