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Draft Genome Sequence of the Phenazine-Producing *Pseudomonas fluorescens* Strain 2-79

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***Pseudomonas fluorescens* strain 2-79, a natural isolate of the rhizosphere of wheat (*Triticum aestivum* L.), possesses antagonistic potential toward several fungal pathogens. We report the draft genome sequence of strain 2-79, which comprises 5,674 protein-coding sequences.**

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The concentration and composition of antibiotic-producing, root-colonizing organisms are important factors that partially determine the suppressiveness of soils toward certain soil-borne diseases (1).

Fluorescent pseudomonads play a major role in suppressing take-all disease of wheat caused by the fungal pathogen *Gaeumannomyces graminis* var. *tritici* (Sacc.) (2). In 1979, Weller and Cook isolated bacteria from roots of wheat plants grown in take-all suppressive soils in Washington state, USA (3). *Pseudomonas fluorescens* 2-79 (NRRL B-15132) was characterized as a strong biological control agent suppressing *G. graminis* in vitro and in planta. Wheat plants infected with *G. graminis* var. *tritici* and additionally treated with *P. fluorescens* 2-79 resulted in taller plants, more heads, and fewer symptoms of root disease compared to the control plants without bacterial treatment. Bacterial treatment could increase the yield up to 147% in soils fumigated with methyl bromide and up to 27% in natural soils (3). *P. fluorescens* 2-79 produces phenazines, which represent a diverse chemical group of nitrogen-containing heterocyclic pigments possessing broadly inhibitory properties toward bacteria and fungi (4). Phenazines undergo redox reactions with NADH/NADPH, leading to an increase of toxic superoxide radicals and hydrogen peroxide in the target cells (5). Mavrodi et al. investigated the biosynthesis pathway of phenazines in *P. fluorescens* 2-79 (6).

Genomic DNA of *P. fluorescens* 2-79 was isolated by using the MasterPure Complete DNA and RNA purification kit (Epicentre, Madison, WI, USA). A shotgun sequencing library was generated employing the Nextera DNA sample preparation kit following the manufacturer's instructions. The whole genome of *P. fluorescens* 2-79 was sequenced with the Genome Analyzer IIx (Illumina, San Diego, CA, USA). In total, 8.5 million paired-end reads of 112 bp were generated. *De novo* assembly of all shotgun reads using SPAdes version 3.0.0 (7) resulted in 143 contigs >3 kb and 123-fold coverage. The draft genome sequence comprises 6.4 Mb and a GC content of 59.83%. Ge-

nome annotation was performed by using Prokka (8). The draft genome harbored 1 rRNA cluster, 47 tRNA genes, 4,286 protein-encoding genes with function prediction, and 1,388 genes coding for hypothetical proteins.

Proteins involved in secondary metabolism were analyzed. The gene *hcnA* (GenBank accession no. 15560558) involved in HCN synthesis and the phenazine operon (GenBank accession no. L48616.1) are present in *P. fluorescens* 2-79. The gene *phlD* (GenBank accession no. 15563828) necessary for the synthesis of 2,4-diacetylphloroglucinol (DAPG) is absent in 2-79.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [JXCQ00000000](https://www.ncbi.nlm.nih.gov/nuccore/JXCQ00000000). The version described in this paper is the first version, [JXCQ01000000](https://www.ncbi.nlm.nih.gov/nuccore/JXCQ01000000).

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REFERENCES

- Haas D, Défago G. 2005. Biological control of soil-borne pathogens by fluorescent pseudomonads. *Nat Rev Microbiol* 3:307–319. <http://dx.doi.org/10.1038/nrmicro1129>.
- Cook RJ, Rovira AD. 1976. The role of bacteria in the biological control of *Gaeumannomyces graminis* by suppressive soils. *Soil Biol Biochem* 8:269–273. [http://dx.doi.org/10.1016/0038-0717\(76\)90056-0](http://dx.doi.org/10.1016/0038-0717(76)90056-0).
- Weller DM, Cook RJ. 1983. Suppression of take-all of wheat by seed treatments with fluorescent pseudomonads. *Phytopathology* 73:463–469. <http://dx.doi.org/10.1094/Phyto-73-463>.
- Mavrodi DV, Blankenfeldt W, Thomashow LS. 2006. Phenazine compounds in fluorescent *Pseudomonas* spp. biosynthesis and regulation. *Annu Rev Phytopathol* 44:417–445. <http://dx.doi.org/10.1146/annurev-phyto.44.013106.145710>.
- Hassett DJ, Woodruff WA, Wozniak DJ, Vasil ML, Cohen MS, Ohman DE. 1993. Cloning and characterization of the *Pseudomonas aeruginosa* *sodA* and *sodB* genes encoding manganese- and iron-cofactored superoxide

- dismutase: demonstration of increased manganese superoxide dismutase activity in alginate-producing bacteria. *J Bacteriol* 175:7658–7665.
6. Mavrodi DV, Peever TL, Mavrodi OV, Parejko JA, Raaijmakers JM, Lemanceau P, Mazurier S, Heide L, Blankenfeldt W, Weller DM, Thomashow LS. 2010. Diversity and evolution of the phenazine biosynthesis pathway. *Appl Environ Microbiol* 76:866–879. <http://dx.doi.org/10.1128/AEM.02009-09>.
 7. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
 8. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068455–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.