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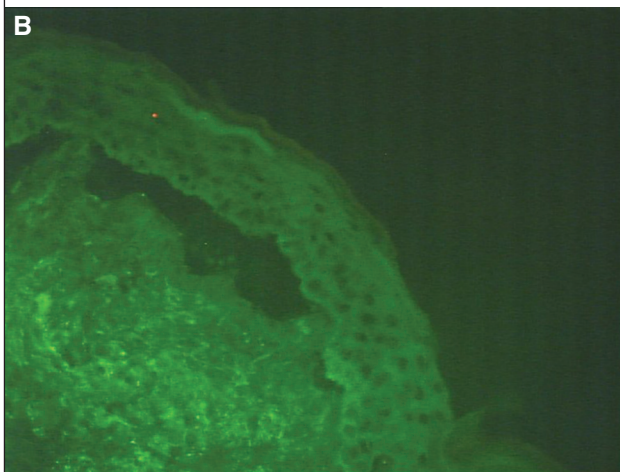
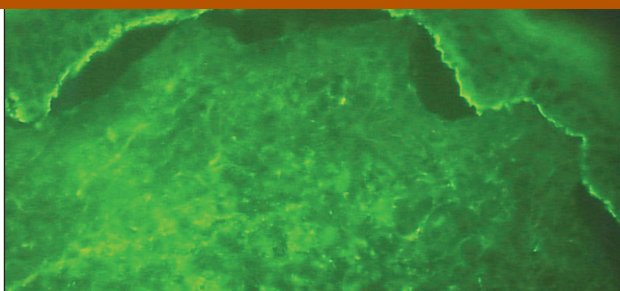


Figure 1. IIF using 1 mol/L NaCl split normal human skin as a substrate.

autoantigen of LABD by immunoblot studies in only a few reports [4, 5].

The major target antigen in LABD is LAD-1, a protein of 120 k-Da, which is a proteolytic fragment of the extracellular domain of BP180 as a result of its cleavage on the surface of keratinocytes by the action of an enzyme. This enzyme belongs to the family of ADAMTS (A disintegrin and A metalloprotease). The other well known antigen is a 97 k-Da protein which is a fragment of the 120 k-Da protein [6]. In our immunoblot analysis, we also found autoantibodies reactive with a 97 k-Da protein, but less reactive.

In this case, we report the novel finding of an IgA directed against a protein of 200 k-Da located on the epidermis. Zone *et al.* also found a band at 200 k-Da in patient's serum with LABD [4]. In fact, this 200 k-Da band, reactive with whole serum, was not present when the purified antibody was used. This suggested that the antibody that bound to this 200 k-Da band was not purified by the immunoaffinity process using human BMZ. These results suggested that the presence of those antibodies may be just an epiphenomenon. However, Fujimoto *et al.* [6] showed that circulating IgA autoantibodies to 200 and 280 k-Da antigens were detected in an LABD patient's serum by immunoblot analysis using extracts from normal human epidermis and human epidermal keratinocytes. These two antibodies also reacted with the epidermal side of 1 mol/L NaCl split skin on IIF

microscopy and bound to hemidesmosomes as determined by immunoperoxidase electronic microscopy. Our results may also suggest the presence of 200 k-Da hemidesmosomal proteins as target antigens in linear IgA disease.

pathogenic, where this 200 k-Da protein is situated and which protein of the BMZ is implicated in such cases. ■

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1. Paul C, Wolkenstein P, Prost C, *et al.* Drug-induced linear IgA disease: target antigens are heterogeneous. *Br J Dermatol* 1997; 136: 406-11.

2. Christophoridis S, Büdinger L, Borradori L, Hunziker T, Merk HF, Hertl M. IgG, IgA and IgE autoantibodies against the ectodomain of BP180 in patients with bullous and cicatricial pemphigoid and linear IgA bullous dermatosis. *Br J Dermatol* 2000; 143: 349-55.

3. Jonkman MF, Groot AC, Slegers TP, Jong MC, Pas HH. Immune diagnosis of pure ocular mucous membrane pemphigoid: indirect immunofluorescence versus immunoblot. *Eur J Dermatol* 2009 ; 456-60.

4. Zone JJ, Taylor TB, Kadunce DP, *et al.* IgA antibodies in chronic bullous disease of childhood react with 97 kDa basement membrane zone protein. *J Invest Dermatol* 1996; 106: 1277-80.

5. Zhou S, Ferguson DJ, Allen J, Wojnarowska F. The localization of target antigens and autoantibodies in linear IgA disease is variable: correlation of immunogold electron microscopy and immunoblotting. *Br J Dermatol* 1998; 139: 591-7.

6. Fujimoto W, Ohtsu T, Toi Y, Nakanishi G, Arata J. Linear IgA disease with IgA antibodies directed against 200- and 280-kDa epidermal antigens. *Br J Dermatol* 2000; 142: 1213-8.

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Sjögren-Larsson syndrome due to a novel mutation in the FALDH gene

Sjögren-Larsson syndrome (SLS) is a rare autosomal recessive disorder characterized by the presence of congenital ichthyosis, spastic diplegia or tetraplegia and mild to moderate mental retardation. SLS is caused by mutations in the ALDH3A2 gene, which encodes for fatty aldehyde dehydrogenase (FALDH) [1-3], an enzyme that catalyzes the oxidation of medium- and long-chain aliphatic aldehydes

[3-5]. The genotype of SLS patients varies considerably, with many private mutations in individual patients [4, 5]. We report a 2.5-year-old girl presenting with a history of generalized extremely dry and scaly skin since birth. Her mental and motor development was also delayed; she had spastic diplegia of the legs and expressed herself only in monosyllables. She had been born at gestational week 35, after a normal pregnancy with no antenatal or perinatal complications. There was no history suggestive of a colloidion membrane at the time of birth. Family history showed consanguinity, the parents were second degree cousins, but there were no other relatives with SLS. Gradually the skin lesions worsened, becoming thicker, especially over the flexures, with some lichenified hyperkeratosis (*figure 1*). The lesions were very pruritic. Scaling and hyperlinearity of palms and soles was present. Ophthalmological and otorhinolaryngological examination was normal.

Magnetic resonance imaging of the brain revealed unmyelinated white matter, and the proton spectroscopy demonstrated an abnormal white matter peak at 1.3 ppm. Histopathologic examination showed orthohyperkeratosis, acanthosis and papillomatosis. Genetic analysis of the patient detected a deletion of thymidine at position 805 of the exon 6 [c.805delT (p.Tyr269fsX5)] of the FALDH gene in a homozygous state. The same mutation was found in heterozygous state in both parents. This mutation has not been previously reported and functional analysis was not performed. However, as it introduces a premature stop codon, it is predicted to cause either a reduced expression of the mRNA or a truncated protein, with consequent reduction of the amount of enzyme produced or loss of enzymatic function, respectively.

SLS is a chronic disabling neurocutaneous disease with autosomal recessive inheritance that was described in detail



Figure 1. Generalized dry and scaly skin.

by two Swedish psychiatrists, Sjögren and Larsson, in 1957 [1]. It is caused by mutations in the ALDH3A2 gene, however the relationship between the genotype and the clinical phenotype of SLS has been difficult to establish [4]. The consequent accumulation of fatty aldehyde precursors, including fatty alcohols, caused by the FALDH deficiency, is postulated to affect the normal formation of multilamellar membranes in the stratum corneum and myelin, and to result in the symptoms [5].

The neurological features of SLS appear in the first years of life and then seem to stabilize [5]. Cerebral proton magnetic resonance spectroscopy in patients with SLS demonstrates an abnormal white matter peak at 1.3 ppm [6], consistent with long-chain fatty alcohol accumulation and coincides with retarded myelination [5].

The diagnosis of SLS is invariably delayed, similarly to other rare genodermatosis. On clinical grounds the diagnosis should be suspected in infants with congenital ichthyosis, especially with emerging neurological features. The ichthyosis is usually the first signal that brings the patient to medical attention, emphasizing the role of the dermatologist in the diagnosis. Unlike most other forms of ichthyosis, it has a disturbing pruritic character and, as in our case, patients tend to be born preterm.

To our knowledge, the mutation of our patient has not been previously reported, supporting the rich mutational heterogeneity associated with this syndrome. ■

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1. De Laurenzi V, Rogers GR, Hamrock DJ, *et al.* Sjögren-Larsson Syndrome is caused by mutations in the fatty aldehyde dehydrogenase gene. *Nat Genet* 1996; 12: 52-7.
2. Rizzo WB. Sjögren-Larsson syndrome. *Semin Dermatol* 1993; 12: 210-8.
3. Auada MP, *et al.* Sjögren-Larsson syndrome: biochemical defects and follow up in three cases. *Eur J Dermatol* 2002; 12: 263-6.
4. Auada MP, Puzzi MB, Cintra ML, *et al.* Sjögren-Larsson Syndrome in Brazil is caused by a common c.1108-1G → C splice-site mutation in the ALDH3A2 gene. *Br J Dermatol* 2006; 154: 770-3.
5. Lassos A, Khoury M, Rizzo W, *et al.* Phenotypic variability among adult siblings with Sjögren-Larsson Syndrome. *Arch Neurol* 2006; 63: 278-80.
6. Mano T, Ono J, Kaminaga T, *et al.* Proton MR Spectroscopy of Sjögren-Larsson Syndrome. *Am J Neuroradiol* 1999; 20: 1671-3.

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