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1 Linking stocking densities and feeding strategies with social and 2 individual stress responses on gilthead seabream (*Sparus aurata*).

3
4 Pablo Arechavala-Lopez^{1,2}, Joan Nazzaro-Alvarez³, Andrea Jardí-Pons³, Lourdes Reig⁴, Francesca Carella,⁵
5 Maite Carrassón⁶, Ana Roque^{3*}.

6
7 1. Fish Ecology group, Mediterranean Institute of Advanced Studies (IMEDEA-CSIC/UIB), Mallorca, Spain.

8 2. Fish Ethology and Welfare group, Center of Marine Sciences (CCMAR), Faro, Portugal.

9 3. IRTA –SCR , Sant Carles de la Ràpita, Spain

10 4. CREDA- UPC-IRTA, Castelldefels, Spain

11 5. Department of Biology, University of Naples Federico II, Naples, Italy

12 6. Unitat de Zoologia, Facultat de Biociències (UAB), Bellaterra, Barcelona, Spain

13 * Corresponding author: Ana.Roque@irta.cat

14 15 HIGHLIGHTS

- 16
17 1. Different stocking densities did not affect the increment in fish weight.
18 2. High densities might reinforce schooling behavior on seabream juveniles
19 3. Hand-feeding improved fish growth compared to self-demanding systems
20 4. Self-demanding feeding is dependent on particular individuals and social hierarchies
21 5. Individual triggering actions are not correlated with proactive individuals
22 6. Glucose and cortisol levels are not related to behavioral traits

23 24 ABSTRACT

25
26 Intensive aquaculture and poor management practices can cause stress and compromise welfare of
27 farmed fish. This study aimed to assess the potential links between stocking densities and feeding
28 methods with social and individual stress responses on juvenile seabream (*Sparus aurata*) through risk-
29 taking and hypoxia tests. Seabream was first experimentally reared under two different densities: high
30 (HD: 11-65 kg m⁻³) and low (LD: 3-15 kg m⁻³). After 120 days under these conditions, increment in fish
31 weight was not affected by different stocking densities. HD seemed to induce a stronger schooling
32 behavior on seabream juveniles seeking for the group safety during the risk test; while LD increased the
33 mean number of movements per fish recorded and the time of first response. Additionally, HD
34 conditions delayed the time of first response of proactive fish during hypoxia tests. Glucose levels were
35 higher in reactive fish compared to proactive ones, being highly significant in fish reared at HD. In
36 parallel, juvenile seabream was also experimentally reared for 106 days under two different feeding
37 strategies: hand-feeding (HF) and self-demanding feeding (DF), which influenced fish growth and
38 foraging behavior at group and individual level. HF method induced a positive effect on fish weight
39 compared to DF systems. Time of first response during both hypoxia and risk-taking tests was shorter in
40 HF fish than DF fish, and the mean number of movements per fish during risk-taking behavior tests was
41 lower for DF fish compared to HF fish. No differences were found in glucose and cortisol concentrations
42 between behavioral traits (proactive/reactive) and feeding strategies. Triggering actions of seabream in
43 DF systems were also assessed, which seemed to be highly dependent on particular individuals and not
44 related to proactive individuals. DF systems however reinforce the social hierarchy within the fish group,
45 which might lead to a higher competitiveness for resources among fishes, increasing the social
46 hierarchy, and therefore, the stress. The findings of this study provide valuable information to the
47 industry for the management of fish stress and welfare under production conditions at social and
48 individual level.

49
50 **Keywords:** fish individuality, stress copying style, behavior, physiology, welfare, aquaculture.

51 **1. INTRODUCTION**

52

53 Gilthead sea bream (*Sparus aurata*) is a species of great interest for aquaculture, being mostly cultivated
54 in intensive conditions and traditionally throughout the Mediterranean basin (mainly in Greece, Turkey,
55 Italy and Spain). Intensive rearing conditions in aquaculture are associated with a high stocking density,
56 which is considered an aquaculture related chronic stressor, involving many parameters such as water
57 quality, physical space and food availability (Ellis et al. 2002; Hastein et al. 2005). The interest in studying
58 fish stress and welfare has increased to better understanding of potential negative impacts and
59 problems associated with intensive aquaculture production (Huntingford 2006; Ashley 2007). High
60 stocking densities have been shown to produce a wide variety of effects on cultured fish populations,
61 such as alterations in behavior and poor feed utilization, immune suppression leading to increased
62 infections due to associated pathogens, poor growth and even mortality (Tort 2011; Sopinka et al.
63 2016). Higher stocking densities can be used to increase fish production, but the limit beyond which fish
64 welfare is affected is still under discussion. For gilthead seabream, previous studies have demonstrated
65 that high stocking densities or poor management practices (e.g. air exposure, crowding) lead to
66 physiological, biochemical and behavioral stress responses (Arends et al. 1999; Montero et al. 1999;
67 Mancera et al. 2008; Mauri et al. 2011; Sanchez-Muros et al. 2017).

68

69 The gilthead seabream is a schooling species which displays social hierarchies in terms of use of space
70 and competition for food (Goldan et al. 2003; Montero et al. 2009; Arechavala-Lopez et al. 2019;
71 Oikonomidou et al., 2019;). Direct competition for food has been shown to be an important social
72 mechanism in gilthead seabream held in tanks, including the establishment of a dominance hierarchy or
73 increased swimming activity, but there is a direct effect on the size of the group, as well as on the food
74 delivery rate and method (Karplus et al. 2000; Andrew et al. 2003; 2004; Sanchez-Muros et al. 2003;
75 Goldan et al., 2003). Feeding might also affect fish health and growth, feed cost and efficiency, and
76 represents one of the major costs in fish farming (Thorpe et al. 1990; Kentouri et al. 1993; Paspatis et al.
77 1999; Sitjá-Bobadilla et al. 2003). Some studies, however, stated that feeding gilthead sea bream by
78 hand versus automatically, and distributing the daily food ration in two or three equal or unequal-size
79 daily meals, have no effect on the animals growth, nutritional use of the diet or body composition
80 (Velazquez et al. 2006). Hand feeding is one of the main methods used by the industry, but is highly
81 subjective and labour-intensive; automatic feeding has low labour costs but may not be consistent with
82 the feeding needs of fish; and self-demanding feeding has low labour costs and is based on feed
83 demands of the fish but which has been of limited use on an industrial scale (Paspatis et al. 1999).
84 Initially, self-demanding feeders were developed to allow fish to obtain food according to their
85 nutritional needs, but it was shown that feeding activity depends not only on feeding motivation and
86 social organization, but also on individual learning capacity and risk-taking behavior (Attias et al. 2012).

87

88 Different responses to stressors at fish-farms (e.g. stocking densities, feeding strategies) can imply
89 individual behavioral and physiological differences within a population, leading to the concept of stress
90 copying style (SCS), which can be defined as “a coherent set of behavioral and physiological stress
91 responses, which are characteristic to a certain group of individuals” (Koolhaas et al. 1999). In this sense,
92 individual differences are characterized along two axis defined as proactive and reactive individuals.
93 Behaviorally, proactive animals show high aggressiveness towards conspecifics, take risks in the face of
94 potential hazards, are novelty seekers, and present high rates of activity. In contrast, reactive animals
95 are less aggressive with conspecifics; avoid taking risks in unknown environments, show lower rates of
96 activity and passive behaviors such as immobility in response to stressful stimuli (Koolhaas et al. 1999,
97 2007; Coopens et al. 2010). Physiologically, proactive fish present lower production of glucocorticoids
98 (i.e. catecholamines or cortisol) and higher sympathetic activity (i.e. increase noradrenaline and
99 adrenaline) than reactive fish (Øverli et al. 2007). In aquaculture conditions, in which fish densities are
100 usually high and the food sources are regular and predictable, the presence of different SCS within a

101 population can have negative consequences. Individuals with a proactive SCS can monopolize food
102 resources and those with a reactive SCS may not have an adequate amount of food available (Laursen et
103 al. 2011).

104
105 Despite the well-established connection between animal welfare and stress, the implications of these
106 factors on farmed fish need further investigation (Huntingford and Adams 2005). Non-behavioral
107 assessments for the study of coping styles are mainly based on endocrine responses (cortisol)
108 and plasma metabolites such as glucose and lactate (Castanheira et al., 2013a; Laursen et al., 2011),
109 since those parameters are closely related to stress responses (Iwama et al., 2006). The ecological and
110 biological consequences of distinct stress coping styles include potential effects on survival,
111 reproductive success, growth, community organization, and conservation and management of natural
112 resources among others (Mittlebach et al. 2014). Moving into aquaculture, the knowledge of coping
113 styles contribute to improve the sustainability of the aquaculture industry, including welfare and
114 performance of farmed fish, through the establishment of more fine-tuned culture strategies
115 (Castanheira et al. 2017). Despite of the existence of several studies proposing the advantages of
116 characterizing proactive or reactive coping strategies in aquaculture (for a review see Castanheira et al.
117 2017), there is still a lack of knowledge of many cultured fish species, such as gilthead seabream
118 (Castanheira et al. 2013a,b; Herrera et al. 2014). Thus, we hypothesized that both stocking densities and
119 feeding strategies might affect individual and group behavior of seabream subjected under acute stress
120 events. The aim of this study was, therefore, to assess the potential links between different stocking
121 densities and feeding strategies with social and individual stress responses of juvenile seabream through
122 different experiments, in order to shed light on the importance of fish individuality and social
123 hierarchies on fish welfare assessment and aquaculture management.

124
125

126 **2. MATERIAL AND METHODS**

127

128 **2.1. Experimental fish and ethical notes**

129

130 Gilthead seabream juveniles (*S. aurata*) were used as experimental animals. All fish were obtained from
131 a commercial fish farm in Burriana (Spain) in two different periods (experiment 1 in 2017, 1.8 ± 0.4 g
132 body weight at arrival; experiments 2 and 3 in 2018, 1.5 ± 0.4 g body weight at arrival). Upon arrival to
133 the Institute of Agrifood Research and Technology (IRTA) research facilities (Sant Carles de la Ràpita,
134 Spain), two months before the start of each experiment; fish were housed in a stock with standard
135 rearing conditions on fibreglass circular tanks supplied with filtered seawater in a recirculated system
136 (RAS, Recirculation Aquaculture System). Water parameters such as temperature (19-20 °C), oxygen
137 saturation (8-6 mg L⁻¹), pH (~7) and salinity (~36 ‰) were checked daily; ammonia (~0.5 mg L⁻¹) and
138 nitrite (~0.7 mg L⁻¹) were weekly measured ensuring accepted values for seabream. A 12L: 12D
139 photoperiod was maintained with day break set at 8:00 h. Until experiments started, fish were hand fed
140 three times a day (one third of the daily ration) with 5 % of the body weight. This quantity was adjusted
141 every fortnight. All diets were from Skretting and the size of pellet offered according to the fish size and
142 for seabream. All fish experiments were approved by the Ethical Committee of Animal Experimentation
143 and carried out strictly by trained and competent personal, in accordance with the European Directive
144 (2010/63/UE) and Spanish Royal Decree (RD53/2013) to ensure good practices for animal care, health,
145 and welfare.

146

147 **2.2. Experiment 1: Stocking-density**

148

149 The first experiment consisted of studying the potential effects of two different densities on sea bream
150 juveniles regarding individual SCS and stress plasmatic variables. This experiment was conducted in RAS

151 during 120 days (21/03/2017-18/07/2017). A total of 2,511 hatchery-reared sea bream individuals, with
152 initial mean weight of 6.81 ± 0.25 g, was distributed in six 400 L rearing tanks with two different stocking
153 densities: three tanks considered as low densities (LD tanks) holding 180 individuals per tank (initial
154 densities: 3 kg m⁻³; estimated final densities: 15 kg m⁻³); and three tanks considered as high density (HD
155 tanks) with 657 individuals per tank (initial densities: 11 kg m⁻³; estimated final densities: 65 kg m⁻³). All
156 fish was tagged with conventional 12 mm Passive Integrated Transponders tags (PIT-tags, Trovan ID-100
157 A Minitransponder 1.4 x 7 mm cristal made, 10 digits) at day 50 for further individual identification. In
158 order to tag the fish with PIT-tags, fish were fasted overnight and anesthetized with MS-222 at 50 ppm
159 in order to reach surgical anesthesia state (Zahl et al., 2012). PIT-tag was injected on left-hand side of
160 the fish, into the muscle through an IM-200 syringe implanter (Trovan). Fish were recovered in a 60 L
161 PVC tank with the water from the housing tanks and aerated through an airstone connected to the
162 compressed air system at the research facility IRTA.

163

164 During the whole experimental period fish were fed once a day at a rate of 3% of average body mass
165 with a commercial gilthead sea bream diet (Skretting®, Optibream 2 mm; 48.5% crude protein, 18.0%
166 crude fat, 5.9% crude ash, 3.3% crude fibres, 1.0% phosphorus, 0.9% calcium, 0.3% sodium). Fish weight
167 was recorded at the beginning (T_0) and the end (T_{119}) of the experiments, allowing studying the growth
168 rates between stocking densities. All fish individuals were subjected to two different group-based tests
169 (Castanheira et al. 2013a) in order to classify fish individuals regarding their SCS: risk-taking and hypoxia
170 tests (see section 2.4). Every test was repeated twice, first trial at day 70-71 and second trial at days
171 120-121 (50 days between trials). Tests were performed over a two-day period because there were
172 many animals to be tested but animals were tested once in each trial. Additionally, blood samples were
173 taken at the end of the experiment (days 120-121) from selected individuals to determine plasma
174 cortisol and glucose levels (see section 2.5).

175

176 **2.3. Experiment 2: Feeding strategies**

177

178 The second experiment consisted of studying the potential effects of two different feeding methods on
179 sea bream juveniles regarding individual behavioral traits and physiological response to potential stress
180 conditions. This experiment was conducted during 106 days (11/04/2018-26/07/2018). After the
181 acclimation (see section 2.1), a total of 360 fish, with initial mean weight of 10.3 ± 3.2 g were arbitrarily
182 selected, tagged with conventional 12 mm PIT-tags for further individual identification, and randomly
183 distributed in four square 400 L rearing tanks (90 fish per tank) in RAS system. Two tanks were hand-fed
184 twice a day during the whole experimental period, at a rate of 2.4% of average body mass per day with a
185 commercial gilthead sea bream pellet (Optibream 2.5 mm, Skretting, Spain; 48.0% crude protein, 20.0%
186 crude fat, 10.3% crude ash, 1.2% crude cellulose and 1.3% total phosphorus). The other two tanks were
187 supplied with the same food by using self-demand device throughout the experiment, allowing the
188 study of the demand-feeding activity (dominance behavior) of juvenile seabream individuals. Fish weight
189 was recorded at the beginning (T_0) and the end (T_{106}) of the experiments, allowing studying the growth
190 rates between feeding strategies. Fish individuals were subjected to two different group-based tests
191 (Castanheira et al. 2013a) in order to classify fish individuals regarding their SCS: risk-taking and hypoxia
192 tests (see section 2.4). Every test was repeated twice, first trial at day 20-21 and second trial at days 96-
193 97. Additionally, blood samples were taken at the end of the experiment from selected individuals to
194 determine plasma cortisol and glucose levels (see section 2.5).

195

196 In addition, the dominance behavior of two groups of seabream juveniles around a self-feeding system
197 that has to be triggered was separately assessed in order to define the relationship between the
198 individual contribution to the total food demand and behavioral traits (SCS) under stress conditions. To
199 monitor the individual contribution in food demand, PIT-tags were implanted in all individuals. The
200 triggering system consisted of a metal rod with a lead ball at its lower end activated by pushing,

201 submerged 1 cm deep and surrounded by a PIT tag detector antenna (diameter 100/125 x 20mm,
202 Trovan®, Netherlands). The system was based on the fact that fish should activate the food dispenser
203 (ARVO-TEC T Drum 2000®) and PIT-tag registration unit by triggering the lead ball and passing through
204 the PIT-tag antenna, while data were collected on a computer. The food dispenser consists of a 1L
205 hopper that can hold up to 0.7Kg of feed. A roller drum (1 ± 0.2 g /24 cups) inside the device delivered
206 pellets 30 cm away from the trigger and the same amount of food was given each time. This mechanism
207 allowed monitoring two types of variables, the amount of food demanded by the fish during a period of
208 interest and the identification of the fish that activated the mechanism at each moment. Therefore, the
209 relationship between the total food demand and the individual contribution to it was established. The
210 PIT-tag antenna also allowed determining which individuals frequented the self-feeder zone, even
211 though they did not have any contribution in the demand for food. Therefore, depending on their
212 proportional contribution to total number of trigger actuations (%) within the group (triggering activity),
213 fish were classified into three- categories: High triggering (HT, >15% actuations), low triggering (LT, 3-
214 15% actuations) and zero triggering (ZT, 0-3% actuations)(Covès et al. 2006). Feeding-demand behavior
215 was followed over 32 days (from 14/05/18 to 14/06/18). Additionally, these two groups of seabream
216 juveniles were exposed to acute hypoxia stress events, in order to evaluate potential effects on
217 individual stress response during food demanding. The test consisted of inducing an acute stress to the
218 fish by removing the exogenous oxygen supply to the housing tanks, and letting these consume it until
219 reaching values close to 2 mg/L. A first acute stress was carried out one week after behavior monitoring
220 (21/06/18) in which fish were kept in a hypoxia situation for 1 hour and a half (1h30); and a second test
221 was performed six days later (27/06/18), lengthening the hypoxia condition until the first symptoms of
222 loss of consciousness of the individuals and it lasted two hours and a half (2h30). The individual feed
223 demand behavior, as well as the apparent feed consumption of the group, were analysed for a period of
224 one week after the acute stresses.

225

226 **2.4. Stress coping style (SCS) tests**

227

228 Risk-taking test consists in separating the tank in two equal parts, creating safe and risk areas, through a
229 solid plastic wall with a 10 cm diameter hole to let fish pass (Castanheira et al. 2013a). The safe area was
230 shaded and gathered all fish at the beginning of the experiment; the risky zone was naturally lit. Fish
231 individuals were left in the safe area for one hour and then they were allowed to choose between the
232 safe and the risk areas of the tank during one more hour, by allowing passage through an opening in the
233 middle of the divider. A PIT-tag detection antenna was located around the opening of the divider, which
234 allowed monitoring individual passages through the opaque divider. The number of movements
235 between areas and time of response (i.e. first movement) were determined through antenna
236 detections. Risk taking tests were performed in the holding tanks and in all the tanks.

237

238 Hypoxia test consists in reducing oxygen levels in one side of a two-chamber tank and checking escaping
239 behavior from hypoxia to normoxia side (Castanheira et al. 2013a). Both sides were connected with a
240 plastic tube, provided with a removable door, where there was one PIT-tag detection, for monitoring
241 individual passages through the tube. In one side oxygen supply was stopped and nitrogen gas applied
242 to decrease O₂ concentrations for half an hour to achieve values around 2 mg/L (hypoxia conditions),
243 and in the other side oxygen supply was functioning (normoxia). Once hypoxia was achieved the door
244 was opened and fish were allowed to either stay where they were or to move on the unknown normoxic
245 tank. Three rounds of thirty fish from each tank (90 fish per tank, all the tagged fish were tested) were
246 placed in the hypoxia side. Hypoxia test finalised when half of the fish left the hypoxia side. The number
247 of movements between areas and time of response (i.e. first movement) were determined through
248 antenna detections.

249

250 According to previous studies, proactive fishes are behaviorally characterised by high risk taking and
251 exploratory conduct when compared to reactive fishes (Øverli et al. 2006; Mackenzie et al. 2009; Millot
252 et al. 2009; Huntingford et al. 2010; Herrera et al. 2014). Accordingly, fish were classified depending on
253 passed tests. Proactive fishes were considered those passing both runs of hypoxia and both runs of risk-
254 taking tests, while reactive fish were considered those did not pass any of the tests in any session. The
255 remaining individuals were the intermediate ones, corresponding to those that passed only some of the
256 tests. The risk-testing tanks were the same as the housing tanks. Fish were fasted 24 hours prior testing
257 and no feed was given during the tests.

258

259 **2.5. Physiological parameters**

260

261 Additionally, proactive (n=30, experiment 1; n=32, experiment 2) and reactive (n= 45, experiment 1;
262 n=32, experiment 2) fish individuals were selected at the end of the experiment (intermediate fish were
263 not selected); blood samples were obtained from the caudal vein of selected fish, using a 1 ml
264 heparinized insulin syringe. For this step, fish were anesthetized with MS222 at 70 ppm in a separate
265 tank. Plasma was separated by 15-minute centrifugation (4°C, 3000G) and was stored frozen (-80°C)
266 until required for analysis of cortisol and glucose. Finally, all fish were sacrificed with a lethal MS-222 (40
267 ppm) concentration. Plasma cortisol levels were determined by ELISA kit method (“DEMEDIATEC Cortisol
268 ELISA Kit”) with an analytical sensitivity of 3.79 ng/mL and a cross reactivity to the following substances
269 prednisolone (54.3%) and 11-deoxycortisol (35.7%). Other substances to which there is cross reactivity
270 should not be relevant such as prednisone which is a synthetic molecule. Plasma glucose was measured
271 using an endpoint colorimetric method (GLUCOSE MR “Enzymatic Colorimetric Method”), both
272 according to manufacturer instructions.

273

274 **2.6. Data analysis**

275

276 Differences on fish weight between treatments (i.e. stocking densities and feeding strategies) and
277 experimental tanks were assessed through univariate general linear models (uGLM). Levene’s test was
278 applied to analyse data homogeneity. Non-parametric analysis (Mann-Whitney U test) was applied to
279 test for differences between stocking densities and feeding strategies regarding the mean number of
280 fish movements between areas and the minimum time of first response in each SCS test. Pearson
281 correlation test was conducted to assess lineal relationships between the mean number of fish
282 movements between areas and the minimum time of first response according to fish stocking densities
283 and feeding strategies in each SCS test. Univariate general linear models (uGLM) were applied to look
284 for differences in glucose and cortisol concentrations between fish traits (proactive/reactive), stocking
285 densities and feeding strategies.

286

287

288 **3. RESULTS**

289

290 **3.1. Experiment 1: Stocking-density**

291

292 Altogether, mean body weight (BW) at the beginning of the experiment (T_0) was 6.8 ± 1.9 g and there
293 were no differences between stocking densities (uGLM, $p=0.361$) and among rearing tanks (uGLM,
294 $p=0.436$) (Table 1). At the end of the experiment (T_{119}), total mean body mass was 39.6 ± 7.5 g, and
295 similarly, there were no differences between stocking densities (uGLM, $p=0.113$) and among rearing
296 tanks (uGLM, $p=0.112$) (Table 1). The mean number of movements and time of first response were
297 significantly ($p<0.001$) and negatively correlated in both tests and density groups (Table 2, Figure 1). The
298 higher number of movements per fish, the lower is the first response to move. This correlation was
299 higher for the risk-taking tests than for hypoxia tests (Table 2). Regarding the hypoxia test, the number

300 of fish detected and percentage of consistency were higher in LD fish (39.3%) compared to HD fish
301 (27.1%) (Table 2). Non-parametric test revealed significant differences (Mann-Whitney U test; $p=0.001$)
302 between stocking densities during hypoxia tests regarding the first response; first movement of LD fish
303 occurred earlier than HD fish, while no differences were found in the mean number of fish recorded (U
304 test; $p=0.567$) (Table 2, Figure 1a). However, HD fish presented a wider range of time of first response
305 compared to LD fish (Figure 1a). During risk-taking test, percentage of consistency was higher in LD fish
306 (26.7%) compared to HD fish (20.3%), although number of fish detected varied between runs, being
307 lower during second runs in both densities (Table 2). LD fish presented significantly higher values of
308 mean number of fish detected (U test; $p=0.005$) and higher time of first response (U test; $p=0.001$)
309 compared to HD fish (Table 2, Figure1b). HD fish presented a wider range of time of first response
310 compared to LD fish in both tests (Figure 1b). Regarding relationships of plasma metabolites with
311 behavioral traits, glucose mean concentrations of proactive fish were significantly lower (uGLM;
312 $p=0.008$) than concentrations of reactive fish, though no differences were detected between stocking
313 densities (uGLM; $p=0.703$) (Table 3). Similarly, glucose concentrations were significantly lower in
314 proactive fish within HD group compared to reactive fish (uGLM; $p=0.035$), but no differences were
315 detected between reactive/proactive fish within LD group (uGLM; $p=0.098$) (Table 3). No differences
316 were detected on cortisol mean concentrations between stocking densities (uGLM; $p=0.820$) and
317 between proactive/reactive fish (uGLM; $p=0.889$) (Table 3).

318

319 **3.2. Experiment 2: Feeding strategies**

320

321 Altogether, mean body mass (wet weight) at the beginning of the experiment (T_0) was 10.3 ± 0.3 g and
322 there were no differences between feeding methods (uGLM, $p=0.828$) and among rearing tanks (uGLM,
323 $p=0.357$) (Table 1). At the end of the experiment (T_{106}), total mean body mass was 63.9 ± 0.7 g. Fish
324 weight in HF tanks (weight: 67.9 ± 0.9 g) was significantly higher compared to DF tanks (weight: $59.1 \pm$
325 1.1 g)(uGLM, $p= 0.001$); and there were no differences among rearing tanks within treatments (uGLM,
326 $p=0.523$) (Table 1). In addition, mean number of movements and time of first response were
327 significantly ($p<0.001$) and negatively correlated in both tests and feeding strategy groups; the higher
328 number of movements per fish, the lower is the first response to move. This correlation was higher for
329 the risk-taking tests than for hypoxia tests (Table 2; Figure 2). The number of fish detected during
330 hypoxia tests was higher during second run in both feeding groups, and the percentages of consistency
331 were 52.8% and 51.6% for HF and DF fish respectively (Table 2). Non-parametric test revealed significant
332 differences (Mann-Whitney U test; $p=0.012$) between feeding groups during hypoxia tests regarding the
333 time of first response. First detection of HF fish occurred earlier than DF fish, this latter showing a wider
334 range of time (Figure 2a). Though, no differences were found in the mean number of fish detected by
335 the antenna (U test; $p=0.308$) between both fish groups (Table 2), those individuals detected in both
336 runs showed higher number of detections per fish (Figure 2a). The number of fish detected during risk-
337 taking test was higher during first run in both cases, and percentage of consistency was higher for HF
338 fish (59.3%) than for DF fish (37.6%) (Table 2). HF fish presented significantly higher values of mean
339 number of fish detected (U test; $p=0.001$) but lower time of first response (U test; $p=0.001$) compared to
340 DF fish; the range of time of first response was wider for DF fish than for HF fish (Table 2; Figure 2b).
341 Although no significant differences were detected in cortisol mean concentrations between feeding
342 strategies, resulted mean values were higher in HF conditions than in DF (uGLM; $p=0.053$). Regarding
343 individual stress responses, no differences were observed on cortisol levels between proactive and
344 reactive fish (uGLM; $p=0.324$), neither within DF (uGLM; $p=0.703$) or HF (uGLM; $p=0.269$) strategies
345 (Table 3). No differences were detected regarding glucose mean concentrations within feeding
346 strategies (uGLM; $p=0.489$) and within proactive/reactive fish (uGLM; $p=0.147$) (Table 3).

347

348 Social structure by triggering activity in experimental tanks with self-demanding feeders showed that
349 there was only one HT fish in each tank, being responsible of the 71.8% (tank 1) and 46.5% (tank 2) of
350 total detections (TDT); as well as the 30.5% (tank 1) and 32.1% (tank 2) of the total number of triggering
351 actions (TTA), and demanding food the 82% (tank 1) and 95% (tank 2) of the total days (DFD) (Figure 3).
352 HT fish represented the 16.6% (tank 1) and 14.4% (tank 2) of the total population in each tank
353 respectively; LT fish represented 11.1% (tank 1) and 13.3% (tank 2); and ZT fish conformed the
354 remaining 72.3% of the total fish in both experimental tanks (Figure 3). No relationships were observed
355 between those individuals assigned as proactive and resulting individuals triggering levels; indeed, all HT
356 fish were considered reactive individuals. Acute stress tests caused appreciable alterations in the social
357 structure in both tanks under self-feeding demand. The roles of HT fish changed, decreasing its total
358 contribution in food demand (Figure 4). After the acute stresses, LT and ZT fish noticeably increased
359 their individual contribution to the total of triggering actuations, even relieving the position of the HT
360 fish in the case of tank 1 (Figure 4).

361

362

363 4. DISCUSSION

364

365 Farmed fish are typically reared at densities much higher than those observed in the wild, mainly to
366 increase fish production, but to what extent can impact fish welfare and stress is still subject of debate
367 (Champneys et al. 2018). Our findings provide novel insights into the effects of low (LD: 3-15 kg m⁻³) and
368 high (HD: 11-65 kg m⁻³) stocking densities at social and individual level, where the increment of
369 seabream weight and blood parameters (cortisol and glucose) did not differ between treatments.
370 Similarly, previous studies on seabream have shown no effects on growth or weight gain between HD
371 and LD (Montero et al. 1999; Araujo-Luna et al. 2018); while other studies found an increase on weight
372 on seabream reared at LD compared to stocks at HD (Sangiao-Alvarellos et al. 2005; Sanchez-Muros et
373 al. 2017). Contradictory results have been also shown regarding blood parameters on seabream. Some
374 studies reported higher levels of cortisol and glucose on seabream held in HD (Montero et al. 1999;
375 Sangiao-Alvarellos et al. 2005; Mancera et al. 2008; Laiz-Carrion et al. 2009); while most recent studies
376 found no differences among treatments (Sanchez-Muros et al. 2017; Araujo-Luna et al. 2018). However,
377 these later studies showed a high variation on physiological values, which might indicate a wide range
378 stress responses at individual level. According to the concept of SCS, proactive fish present lower
379 production of cortisol and glucose than reactive fish (Øverli et al. 2007; Castanheira et al. 2017). In this
380 sense, resulting glucose levels were higher in reactive fish compared to proactive individuals in the
381 present study, being significant in HD conditions. Regarding cortisol levels, no significant differences
382 were found between individual traits, though proactive fish presented lower levels in LD and higher
383 levels in HD conditions compared to reactive fish. In the present study cortisol levels were high under all
384 circumstances when compared with some previously published work (Papaharisis et al. 2019), and this
385 can be due to either the stress of fishing since fish were sedated outside the holding RAS or
386 alternatively due to the experimental design itself. Carbonara et al (2019) published a study where
387 bigger seabream was grown in different stocking densities and obtained similar levels of cortisol.
388 Cortisol and glucose levels reported in this study were higher than previously reported in the literature
389 for this species (Montero et al. 1999; Sangiao-Alvarellos et al. 2005; Mancera et al. 2008; Laiz-Carrion et
390 al. 2009; Sanchez-Muros et al. 2017; Araujo-Luna et al. 2018); therefore, an indirect effect due to
391 handling on fish stress cannot be ruled out. Stocking densities influenced the time of first response of
392 seabream during SCS tests in this study. It seemed that HD induced a stronger schooling behavior on
393 seabream juveniles, given that proactive fish from HD conditions took longer time to move from a
394 hostile environment during hypoxia test compared to LD fish, probably feeling protected by the group.
395 On the contrary, proactive HD seabream were more explorative moving earlier to a new environment
396 during risk-taking test than LD fish. Sanchez-Muros et al. (2017) studied the individual behavior and
397 social kinetics of seabream held at different stocking densities. They found that seabream showed

398 different shoaling shape and higher cohesion in swimming direction at HD compared to lower densities
399 (LD), which showed no tendency or higher diversification. At individual level, however, fish in HD
400 conditions showed higher exploration and frequency of movements, and lower static movements, than
401 LD fish; but also reported that there was great variation among individuals (Sanchez-Muros et al 2017).
402 In our case, higher individual variations were found in seabream at HD than in LD conditions in terms of
403 time of first response to a stress stimulus. Thus, it can be suggested that individual behavior at HD are
404 more dependent and influenced by the group behavior than at lower densities.

405
406 It is known that juvenile seabream establish dominance relationships during feeding (Montero et al.
407 2009), when most of the aggressive behaviors occur (Goldan et al. 2003). Indeed, direct competition for
408 food is probably one of the major social mechanisms regulating growth in small groups of juveniles of
409 this species when food is limited and defendable (Karplus et al. 2000; Goldan et al. 2003). However, in
410 bigger groups like in rearing conditions might differ depending on individuals, group size and feeding
411 method. The dominance hierarchies in seabream can induce an increase of energy costs related to
412 behavioral strategies, having a direct effect on fish specific growth rate and food consumption (Montero
413 et al. 2009). Those animals able to avoid conflicts could be able to obtain food without a high energy
414 cost, whereas those animals that are not able to avoid conflicts with a fish are not able to obtain enough
415 food to cope with the high energetic cost imposed by the social hierarchy (Montero et al. 2009). It is
416 probable that the amount of food obtained by non-dominant animals can also be directly related to the
417 delivery rate of the food since at high rates of feed delivery, dominant animals could not monopolize all
418 delivered feed, allowing more access by the rest of the animals to the feed (Andrew et al. 2004). Indeed,
419 our results showed that hand-feeding (HF) induced a positive effect on fish weight compared to self-
420 demanding feeding (DF) systems. In agreement, Sanchez-Muros et al. (2003) showed that seabream fed
421 on demand had a significantly lower growth and food conversion rate (FCR) than those fed by hand.
422 Similarly, higher specific growth rate of seabream was observed when fed manually compared to
423 automatic feeding and modulated automatic feeding (Velazquez et al. 2006). A study using underwater
424 cameras showed higher proportions of seabream individuals at feeding during hand-feeding at sea-
425 cages (regular method), and therefore higher intensity, than in fish fed on demand (Andrew et al. 2002).
426 A review of laboratory demand-feeding experiments suggested that self-feeding activities depend not
427 only on feeding motivation and social organization, but also on individual learning capacity and risk-
428 taking behavior (Attia et al. 2012). Our results showed that time of first response during both hypoxia
429 and risk-taking tests was shorter in HF fish than DF fish, and the mean number of movements per fish
430 during risk-taking behavior tests was lower for DF fish compared to HF fish. Therefore, it must be
431 suggested that DF systems seemed to reinforce the social hierarchy within the fish group, which might
432 lead to a higher competitiveness for resources among fishes, increasing the social hierarchy, and
433 therefore, the stress conditions at individual level if feed is not provided in sufficient quantity and
434 quality.

435
436 Social hierarchy has been demonstrated to act as a stressor in seabream in experimental conditions,
437 causing higher stress in subordinate fish, characterized by higher plasma cortisol levels (Montero et al.
438 2009). On the contrary, individuals exhibiting a lower cortisol response to confinement stress perform
439 more aggressive attacks immediately followed by establishment of dominant social status (Øverli et al.,
440 2004). However, dominant fish might also show high basal plasma cortisol levels (Montero et al. 2009)
441 due to the stress that supposes to dominate the food and maintain the social ranking. Therefore, plasma
442 cortisol values and social status are not always well correlated. Our results support this lack of
443 correlation, given that no differences were found in glucose and cortisol concentrations between
444 behavioral traits (proactive/reactive) or feeding strategies. Indeed, individual triggering actions in DF
445 groups do not seem to be related with proactive individuals. Ferrari et al. (2014) characterized the
446 personality of seabass (*Dicentrarchus labrax*) and assessed the link between personality traits and
447 individual triggering activity towards the self-feeder apparatus. They found that triggering activity was

448 negatively correlated with exploratory capacities and boldness, but no differences were observed
449 between triggering categories during the restraint test. Another study on seabass showed that those
450 few high triggering individuals did not exhibit a higher specific growth rate or agonistic behavior as
451 observed by video monitoring (Covès et al. 2006), which suggest a lack of relation between triggering
452 and personality traits. Feeding demand may be very different from one individual to another within the
453 same group subjected to the same conditions. It depends on multiple parameters including density,
454 social organization, genetics, individual learning ability and boldness (Attia et al. 2012). DF systems have
455 low labour costs; they are based on feed demands of the fish, and are nowadays used by the industry,
456 considered a suitable tool which can optimize production performance without compromising fish
457 welfare. However, feed must be provided in sufficient quantity and quality to allow fish expressing their
458 normal feeding behavior (Attia et al. 2012). An optimal food distribution system should address the fish
459 physiological needs, which are in turn dependent upon many variables, including endogenous factors
460 such as biological rhythms, growth stage, species, environmental factors (such as photoperiod, water
461 temperature and salinity, oxygen level, etc.), and external factors such as stress and other disturbances
462 (Velázquez et al. 2004).

463
464 Relationships between number of movements and time of first reponse were negative for both risk-
465 taking and hypoxia tests regardless the densities or feeding strategies. Similarly, a previous study on
466 seabream showed that latency to take risks was negatively correlated to movement, but also to oxygen
467 consumption rates; indicating that risk-avoiders (long latency) were less active and, hence, did not
468 consume so much oxygen as risk-takers (Herrera et al. 2014). Other studies on seabass (*Dicentrarchus*
469 *labrax*) and carp (*Cyprinus carpio*) found a positive correlation between boldness and metabolic rate,
470 suggesting that the risk-takers are associated with high metabolic rates as opposed to risk-avoiders
471 (Huntingford et al. 2010; Killen et al. 2011). Individulas with higher metabolic demand, which means
472 higher energetic requirements, might need to forage more often or take more risks to achive a higher
473 rate of food intake. Hence, the shorter time of response of HF seabream compared to DF fish reinforce
474 the idea of HF as better strategy for meeting the energy demands of seabream in captivity. However,
475 Herrera et al. (2014) found a pronounced individual variation in oxygen consumption rate suggesting
476 that each seabream individual reacted differently when housed in the confinement chambers. On the
477 contrary, they reported higher consistency of individual behavior during the risk-taking tests, but some
478 differences, however, were observed within same individuals after the test repetition. This suggests an
479 habituation of fish to the experimental assays with fish reacting faster during the second run (Martins et
480 al. 2011; Herrera et al. 2014). In this study, a variation of the percentage of consistency was observed
481 during hypoxia (27.1%-52.8%) and risk-taking (20.3%-59.3%) tests, but also varied among treatments
482 (density and feeding strategies) and fish groups (HD, LD, HF, DF), suggesting diverse behavioural
483 reactions under different stress conditions. Experiencing a stress situation does not necessarily lead to
484 negative consequences and can result in an adaptive process, i.e, one fish individual can respond more
485 efficiently to the stressor the second time they are exposed to it (Tort et al. 2011). On the other hand,
486 failure to adapt or overcome the stress situation leads to maladaptation with low performance
487 physiological imbalance and maybe death. This is more common under chronic stress or under
488 combined stressors (Tort et al. 2011).

489
490 In conclusion, this work reports the first data on the links between stocking densities and feeding
491 strategies with social and individual stress responses on gilthead seabream (*Sparus aurata*), providing
492 novel insights into the plasticity of fish behavior under stress conditions. Different stocking densities did
493 not affect the increment in fish weight, although seemed to influence on fish behavior. High densities
494 might reinforce schooling behavior on seabream juveniles while low densities did not show any
495 behavioral effect. Regarding feeding strategies, hand-feeding improved fish growth compared to self-
496 demanding systems, which seems to be more dependent on particular individuals and social hierarchies.
497 Individual triggering actions, however, were not correlated with proactive individuals, suggesting that

498 the divergent copying styles are different from the social organization during feeding. The relationships
499 between behavioral traits and physiological variables were not significant, highlighting the necessity of
500 further studies addressing secondary and tertiary stress effects on the individual physiology and
501 behavior response of sea bream due to stocking densities and feeding strategies, which can be highly
502 informative for future applications to aquaculture.

503
504

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512 **6. REFERENCES**

513

514 Andrew, J. E., Anras, M. B., Kadri, S., Holm, J., & Huntingford, F. A. (2003). Feeding responses of
515 hatchery-reared gilthead sea bream (*Sparus aurata* L.) to a commercial diet and natural prey items.
516 *Marine and Freshwater Behaviour and Physiology*, 36(2), 77-86.

517

518 Andrew, J. E., Holm, J., Kadri, S., & Huntingford, F. A. (2004). The effect of competition on the feeding
519 efficiency and feed handling behaviour in gilthead sea bream (*Sparus aurata* L.) held in tanks.
520 *Aquaculture*, 232(1-4), 317-331.

521

522 Andrew, J. E., Noble, C., Kadri, S., Jewell, H., & Huntingford, F. A. (2002). The effect of demand feeding
523 on swimming speed and feeding responses in Atlantic salmon *Salmo salar* L., gilthead sea bream *Sparus*
524 *aurata* L. and European sea bass *Dicentrarchus labrax* L. in sea cages. *Aquaculture Research*, 33(7), 501-
525 507.

526

527 Araújo-Luna, R., Ribeiro, L., Bergheim, A., & Pousão-Ferreira, P. (2018). The impact of different rearing
528 condition on gilthead seabream welfare: Dissolved oxygen levels and stocking densities. *Aquaculture*
529 *Research*, 49(12), 3845-3855.

530

531 Arechavala-Lopez, P., Diaz-Gil, C., Saraiva, J.L., Moranta, D., Castanheira, M.F., Nuñez-Velázquez, S.,
532 Ledesma-Corvi, S., Mora-Ruiz, M.R., Grau, A. (2019) Effects of structural environmental enrichment on
533 welfare of juvenile seabream (*Sparus aurata*). *Aquaculture Reports*, in press.

534

535

536 Arends, R. J., Mancera, J. M., Munoz, J. L., Bonga, S. W., & Flik, G. (1999). The stress response of the
537 gilthead sea bream (*Sparus aurata* L.) to air exposure and confinement. *Journal of Endocrinology*, 163(1),
538 149.

539

540 Ashley, P. J. (2007). Fish welfare: current issues in aquaculture. *Applied Animal Behaviour Science*, 104(3-
541 4), 199-235.

542

543 Attia, J., Millot, S., Di-Poï, C., Bégout, M. L., Noble, C., Sanchez-Vazquez, F. J., ... & Damsgård, B. (2012).
544 Demand feeding and welfare in farmed fish. *Fish Physiology and Biochemistry*, 38(1), 107-118.

545

546 Carbonara P., Alfonso S., Zuppa W., Manfrin A., Fiochhi E., Pretto T., Spedicato M.T., & Lembo G. (2019).
547 Behavioral and physiological responses to stocking density in sea bream
548 (*Sparus aurata*): Do coping styles matter? *Physiology and Behaviour*, 212: 112698.
549
550 Castanheira, M. F., Conceição, L. E., Millot, S., Rey, S., Bégout, M. L., Damsgaard, B., ... & Martins, C. I.
551 (2017). Coping styles in farmed fish: consequences for aquaculture. *Reviews in Aquaculture*, 9(1), 23-41.
552
553 Castanheira, M. F., Herrera, M., Costas, B., Conceição, L. E., & Martins, C. I. (2013a). Can we predict
554 personality in fish? Searching for consistency over time and across contexts. *PLoS One*, 8(4), e62037.
555
556 Castanheira, M. F., Herrera, M., Costas, B., Conceição, L. E., & Martins, C. I. (2013). Linking cortisol
557 responsiveness and aggressive behaviour in gilthead seabream *Sparus aurata*: Indication of divergent
558 coping styles. *Applied Animal Behaviour Science*, 143(1), 75-81.
559
560 Champneys, T., Castaldo, G., Consuegra, S., & Garcia de Leaniz, C. (2018). Density-dependent changes in
561 neophobia and stress-coping styles in the world's oldest farmed fish. *Royal Society Open Science*, 5(12),
562 181473.
563
564 Coppens, C. M., de Boer, S. F., & Koolhaas, J. M. (2010). Coping styles and behavioural flexibility:
565 towards underlying mechanisms. *Philosophical Transactions of the Royal Society B: Biological Sciences*,
566 365(1560), 4021-4028.
567
568 Covès, D., Beauchaud, M., Attia, J., Dutto, G., Bouchut, C., & Bégout, M. L. (2006). Long-term monitoring
569 of individual fish triggering activity on a self-feeding system: An example using European sea bass
570 (*Dicentrarchus labrax*). *Aquaculture*, 253(1-4), 385-392.
571
572 Ellis, T., North, B., Scott, A. P., Bromage, N. R., Porter, M., & Gadd, D. (2002). The relationships between
573 stocking density and welfare in farmed rainbow trout. *Journal of Fish Biology*, 61(3), 493-531.
574
575 Ferrari, S., Benhaïm, D., Colchen, T., Chatain, B., & Bégout, M. L. (2014). First links between self-feeding
576 behaviour and personality traits in European seabass, *Dicentrarchus labrax*. *Applied Animal Behaviour
577 Science*, 161, 131-141.
578
579 Goldan, O., Popper, D. & Karplus, I. (2003). Food competition in small groups of juvenile gilthead sea
580 bream (*Sparus aurata*). *The Israel Journal of Aquaculture* 55, 94–106.
581
582 Hastein, T., Scarfe, A. D., & Lund, V. L. (2005). Science-based assessment of welfare: aquatic animals.
583 *Revue Scientifique et Technique-Office International des Epizooties*, 24(2), 529.
584
585 Herrera, M., Castanheira, M. F., Conceição, L. E., & Martins, C. I. (2014). Linking risk taking and the
586 behavioral and metabolic responses to confinement stress in gilthead seabream *Sparus aurata*. *Applied
587 Animal Behaviour Science*, 155, 101-108.
588
589 Huntingford, F., & Adams, C. (2005). Behavioural syndromes in farmed fish: implications for production
590 and welfare. *Behaviour*, 142(9), 1213-1228.
591
592 Huntingford, F. A., Adams, C., Braithwaite, V. A., Kadri, S., Pottinger, T. G., Sandøe, P., & Turnbull, J. F.
593 (2006). Current issues in fish welfare. *Journal of Fish Biology*, 68(2), 332-372.
594

595 Huntingford, F. A., Andrew, G., Mackenzie, S., Morera, D., Coyle, S. M., Pilarczyk, M., & Kadri, S. (2010).
596 Coping strategies in a strongly schooling fish, the common carp *Cyprinus carpio*. *Journal of Fish Biology*,
597 76(7), 1576-1591.

598
599 Iwama, G.K., Afonso, L.O.B., Vijayan, M.M., 2006. Stress in fishes. In: Evans,
600 D.H., Clairbone, J.B. (Eds.), *The Physiology of Fishes*. CRC Press, Boca
601 Raton, US, pp. 319–342.

602
603 Karplus, I., Popper, D., & Goldan, O. (2000). The effect of food competition and relative size of group
604 members on growth of juvenile gilthead sea bream, *Sparus aurata*. *Fish Physiology and Biochemistry*,
605 22(2), 119-123.

606
607 Kentouri, M., Divanach, P., Maignot, E. (1993). Comparaison de l'efficacit-cout de trois techniques de
608 rationnement de la daurade *Sparus aurata*, en élevage intensif en bassins. In: Barnabé, G., Kestemont,
609 P., Eds., *Production, Environment and Quality, Bordeaux Aquaculture '92*. Ghent, Belgium, EAS Spec.
610 Publ. No. 18, pp. 273–283.

611
612 Koolhaas, J. M., Korte, S. M., De Boer, S. F., Van Der Vegt, B. J., Van Reenen, C. G., Hopster, H., ... &
613 Blokhuis, H. J. (1999). Coping styles in animals: current status in behavior and stress-physiology.
614 *Neuroscience & Biobehavioral Reviews*, 23(7), 925-935.

615
616 Laiz-Carrión, R., Fuentes, J., Redruello, B., Guzmán, J. M., del Río, M. P. M., Power, D., & Mancera, J. M.
617 (2009). Expression of pituitary prolactin, growth hormone and somatolactin is modified in response to
618 different stressors (salinity, crowding and food-deprivation) in gilthead sea bream *Sparus auratus*.
619 *General and Comparative Endocrinology*, 162(3), 293-300.

620
621 Laursen, D. C., Olsén, H. L., de Lourdes Ruiz-Gomez, M., Winberg, S., & Höglund, E. (2011). Behavioural
622 responses to hypoxia provide a non-invasive method for distinguishing between stress coping styles in
623 fish. *Applied Animal Behaviour Science*, 132(3-4), 211-216.

624
625 MacKenzie, S., Ribas, L., Pilarczyk, M., Capdevila, D. M., Kadri, S., & Huntingford, F. A. (2009). Screening
626 for coping style increases the power of gene expression studies. *PLoS One*, 4(4), e5314.

627
628 Mancera, J. M., Vargas-Chacoff, L., García-López, A., Kleszczyńska, A., Kalamarz, H., Martínez-Rodríguez,
629 G., & Kulczykowska, E. (2008). High density and food deprivation affect arginine vasotocin, isotocin and
630 melatonin in gilthead sea bream (*Sparus auratus*). *Comparative Biochemistry and Physiology Part A:
631 Molecular & Integrative Physiology*, 149(1), 92-97.

632
633 Mauri, I., Romero, A., Acerete, L., MacKenzie, S., Roher, N., Callol, A., ... & Tort, L. (2011). Changes in
634 complement responses in Gilthead seabream (*Sparus aurata*) and European seabass (*Dicentrarchus
635 labrax*) under crowding stress, plus viral and bacterial challenges. *Fish & Shellfish Immunology*, 30(1),
636 182-188.

637
638 Millot, S., Bégout, M. L., & Chatain, B. (2009). Risk-taking behaviour variation over time in sea bass
639 *Dicentrarchus labrax*: effects of day–night alternation, fish phenotypic characteristics and selection for
640 growth. *Journal of Fish Biology*, 75(7), 1733-1749.

641
642 Mittelbach, G. G., Ballew, N. G., & Kjølsvik, M. K. (2014). Fish behavioral types and their ecological
643 consequences. *Canadian Journal of Fisheries and Aquatic Sciences*, 71(6), 927-944.

644

645 Montero, D., Izquierdo, M. S., Tort, L., Robaina, L., & Vergara, J. M. (1999). High stocking density
646 produces crowding stress altering some physiological and biochemical parameters in gilthead seabream,
647 *Sparus aurata*, juveniles. *Fish Physiology and Biochemistry*, 20(1), 53-60.
648

649 Montero, D., Lalumera, G., Izquierdo, M. S., Caballero, M. J., Saroglia, M., & Tort, L. (2009).
650 Establishment of dominance relationships in gilthead sea bream *Sparus aurata* juveniles during feeding:
651 effects on feeding behaviour, feed utilization and fish health. *Journal of Fish Biology*, 74(4), 790-805.
652

653 Oikonomidou, E., Batzina, A., & Karakatsouli, N. (2019). Effects of food quantity and distribution on
654 aggressive behaviour of gilthead seabream and European seabass. *Applied Animal Behaviour Science*,
655 213, 124-130.
656

657 Øverli, Ø., Korzan, W. J., Höglund, E., Winberg, S., Bollig, H., Watt, M., ... & Summers, C. H. (2004). Stress
658 coping style predicts aggression and social dominance in rainbow trout. *Hormones and Behavior*, 45(4),
659 235-241.
660

661 Øverli, Ø., Sørensen, C., & Nilsson, G. E. (2006). Behavioral indicators of stress-coping style in rainbow
662 trout: do males and females react differently to novelty?. *Physiology & Behavior*, 87(3), 506-512.
663

664 Øverli, Ø., Sørensen, C., Pulman, K. G., Pottinger, T. G., Korzan, W., Summers, C. H., & Nilsson, G. E.
665 (2007). Evolutionary background for stress-coping styles: relationships between physiological,
666 behavioral, and cognitive traits in non-mammalian vertebrates. *Neuroscience & Biobehavioral Reviews*,
667 31(3), 396-412.
668

669 Papaharisis L., Tsironi T., Dimitroglou A., Taoukis P., & Pavlidis M. (2019). Stress assessment, quality
670 indicators and shelf life of three aquaculture important marine fish, in relation to harvest practices,
671 water temperature and slaughter method. *Aquaculture Research*, 50 (9), 2608-2620.
672

673 Paspatis, M., Batarías, C., Tiangos, P., & Kentouri, M. (1999). Feeding and growth responses of sea bass
674 (*Dicentrarchus labrax*) reared by four feeding methods. *Aquaculture*, 175(3-4), 293-305.
675

676 Sanchez-Muros, M. J., Corchete, V., Suarez, M. D., Cardenete, G., Gomez-Milan, E., & De la Higuera, M.
677 (2003). Effect of feeding method and protein source on *Sparus aurata* feeding patterns. *Aquaculture*,
678 224(1-4), 89-103.
679

680 Sanchez-Muros, M. J., Sánchez, B., Barroso, F. G., Toniolo, M., Trenzado, C. E., & Rus, A. S. (2017). Effects
681 of rearing conditions on behavioural responses, social kinetics and physiological parameters in gilthead
682 sea bream *Sparus aurata*. *Applied Animal Behaviour Science*, 197, 120-128.
683

684 Sangiao-Alvarellos, S., Guzmán, J. M., Láiz-Carrión, R., Míguez, J. M., Martín Del Río, M. P., Mancera, J.
685 M., & Soengas, J. L. (2005). Interactive effects of high stocking density and food deprivation on
686 carbohydrate metabolism in several tissues of gilthead sea bream *Sparus auratus*. *Journal of*
687 *Experimental Zoology Part A: Comparative Experimental Biology*, 303(9), 761-775.
688

689 Sitjà-Bobadilla, A., Mingarro, M., Pujalte, M. J., Garay, E., Alvarez-Pellitero, P., & Pérez-Sánchez, J.
690 (2003). Immunological and pathological status of gilthead sea bream (*Sparus aurata* L.) under different
691 long-term feeding regimes. *Aquaculture*, 220(1-4), 707-724.
692

693 Sopinka, N. M., Donaldson, M. R., O'Connor, C. M., Suski, C. D., & Cooke, S. J. (2016). Stress indicators in
694 fish. In *Fish Physiology* (Vol. 35, pp. 405-462). Academic Press.

695
696 Thorpe, J.E., Talbot, C., Miles, M.S., Rawlings, C., Keay, D.S. (1990). Food consumption in 24 hours by
697 Atlantic salmon *Salmo salar* L. in sea cage. *Aquaculture* 90, 41–47.
698
699 Tort, L. (2011). Stress and immune modulation in fish. *Developmental & Comparative Immunology*,
700 35(12), 1366-1375.
701
702 Velázquez, M., Zamora, S., & Martínez, F. J. (2004). Influence of environmental conditions on demand-
703 feeding behaviour of gilthead seabream (*Sparus aurata*). *Journal of Applied Ichthyology*, 20(6), 536-541.
704
705 Velázquez, M., Zamora, S., & Martínez, F. J. (2006). Effect of different feeding strategies on gilthead sea
706 bream (*Sparus aurata*) demand-feeding behaviour and nutritional utilization of the diet. *Aquaculture*
707 *Nutrition*, 12(6), 403-409.
708
709 Zahl, I.H., Samuelsen, O., Kiessling, A., 2012. Anaesthesia of farmed fish: implications for welfare. *Fish*
710 *Physiol. Biochem.* 38, 201–218.

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Table 1. Mean weight (g) ±SE of juvenile seabream at the start (T₀) and at the end (T₁₂₀) of the experiments in different tanks, stocking densities (LD: low densities; HD: high densities) and feeding strategies (HF: hand feeding; DF: self-demanding feeding). Values with different letters indicate significant differences between density or feeding strategy groups (p<0.05; uGLM).

	Stocking densities				Feeding strategies			
	Initial Weight (T ₀)		Final Weight (T ₁₂₀)		Initial Weight (T ₀)		Final Weight (T ₁₀₆)	
	LD	HD	LD	HD	HF	DF	HF	DF
Tank 1	6.6±0.7	6.7±0.7	37.3±2.5	38.6±2.2	10.4±0.5	11.4±0.6	66.7±1.3 ^a	60.3±1.5 ^b
Tank 2	7.2±0.6	7.1±0.5	42.8±1.6	41.7±2.9	10.0±0.6	9.4±0.4	69.1±1.4 ^a	59.7±1.4 ^b
Tank 3	6.6±0.4	6.5±0.8	42.9±1.5	33.9±2.3	-	-	-	-
Total	6.9±0.3	6.8±0.4	41.1±1.2	38.1±1.5	10.2±0.4	10.4±3.2	67.9±0.9^a	59.9±1.1^b

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Table 2. Results from the SCS tests (hypoxia response and risk-taking behavior) regarding fish stocking densities (LD: low densities; HD: high densities) and feeding strategies (HF: hand feeding; DF: self-demanding feeding): number of individuals recorded during first and second run of each test; percentage of consistency between both runs within each test; mean number of movements per fish recorded (±SE) for both tests and mean first response (min:sec) per fish (±SE) of each group tested. Values and significance of Pearson's correlation tests between movements and first response are shown for all fish recorded on any run, and for those who past both runs in each test. Asterisks indicate significant correlation (**: p-value<0.01; ***: p-value<0.001); ns: non significant. Different superscript letters in the same test show significant differences between density or feeding strategy groups (p-value<0.01; Mann-Whitney U test).

		N ind.	N ind.	%	Mean ind.	Mean first	Pearson's correlation	
		run 1	run 2	Cons.	movements	response	(sig.)	
DENSITY								
Hypoxia	LD	132	141	39.3%	1.1 ±0.1	08:33 ±00:27 ^a	-0.212 (**)	-0.251 (ns)
	HD	109	74	27.1%	1.2 ±0.1	17:57 ±00:46 ^b	-0.318 (***)	-0.077 (ns)
Risk-Taking	LD	91	37	26.7%	2.5 ±0.3 ^a	39:49 ±01:12 ^a	-0.574 (***)	-0.459 (**)
	HD	135	31	20.3%	1.9 ±0.2 ^b	33:37 ±01:11 ^b	-0.532 (***)	-0.509 (**)
FEEDING								
Hypoxia	HF	74	116	52.8%	33.1 ±7.9	14:07 ±01:24 ^a	-0.295 (***)	-0.367 (*)
	DF	57	81	51.6%	39.6 ±6.8	18:59 ±01:29 ^b	-0.454 (***)	-0.227 (*)
Risk-Taking	HF	109	79	59.3%	13.2 ±1.4 ^a	15:08 ±01:37 ^a	-0.607 (***)	-0.560 (***)
	DF	70	58	37.6%	4.48 ±0.6 ^b	28:03 ±01:38 ^b	-0.543 (***)	-0.488 (**)

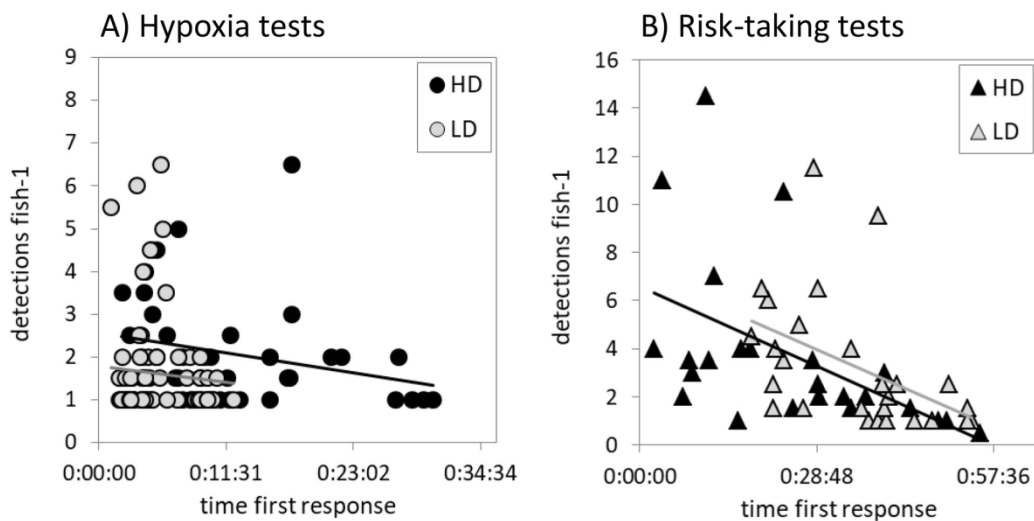
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Table 3. Mean concentrations (\pm SE) of plasma glucose (mmol L^{-1}) and cortisol (ng mL^{-1}) detected in selected fish regarding proactive/reactive traits in two experiments: stocking densities (LD: low densities; HD: high densities) and feeding strategies (HF: hand feeding; DF: demand feeding). Values with different letters indicate significant differences between behavioural traits ($p < 0.05$; uGLM).

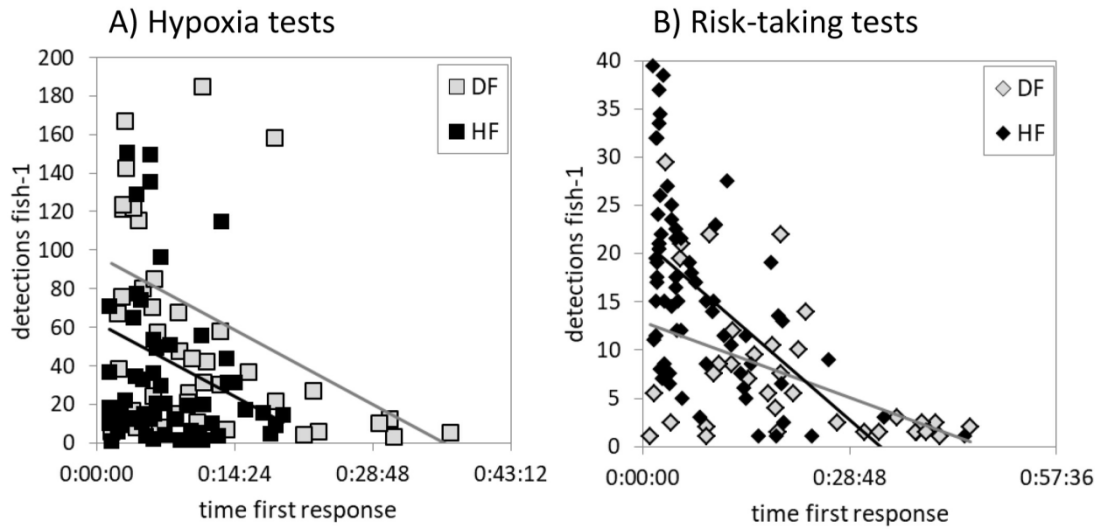
Glucose (mmol L^{-1})		Stocking densities			Feeding strategies			
		LD	HD	Total	HF	DF	Total	
Proactive		6.67 ± 0.35	5.5 $\pm 0.75^a$	6.29 $\pm 0.35^a$	6.25 ± 0.53	5.18 ± 0.35	5.63 ± 0.32	
	Reactive	7.34 ± 0.42	7.98 $\pm 0.79^b$	7.66 $\pm 0.62^b$	4.87 ± 0.32	5.38 ± 0.35	5.1 ± 0.32	
	Total	7.02 ± 0.28	7.23 ± 0.62		5.53 ± 0.34	5.26 ± 0.25		
Cortisol (ng mL^{-1})		LD	HD	Total	HF	DF	Total	
		Proactive	252.6 ± 48.5	308.1 ± 99.1	251.1 ± 42.9	180.6 ± 40.4	98.3 ± 22.2	133.1 ± 22.4
		Reactive	314.1 ± 58.2	227.7 ± 62.7	270.9 ± 42.8	127.1 ± 24.1	93.7 ± 25.9	112.1 ± 17.6
Total	284.1 ± 37.9	250.7 ± 52.5		153.8 ± 23.7	96.5 ± 16.2			

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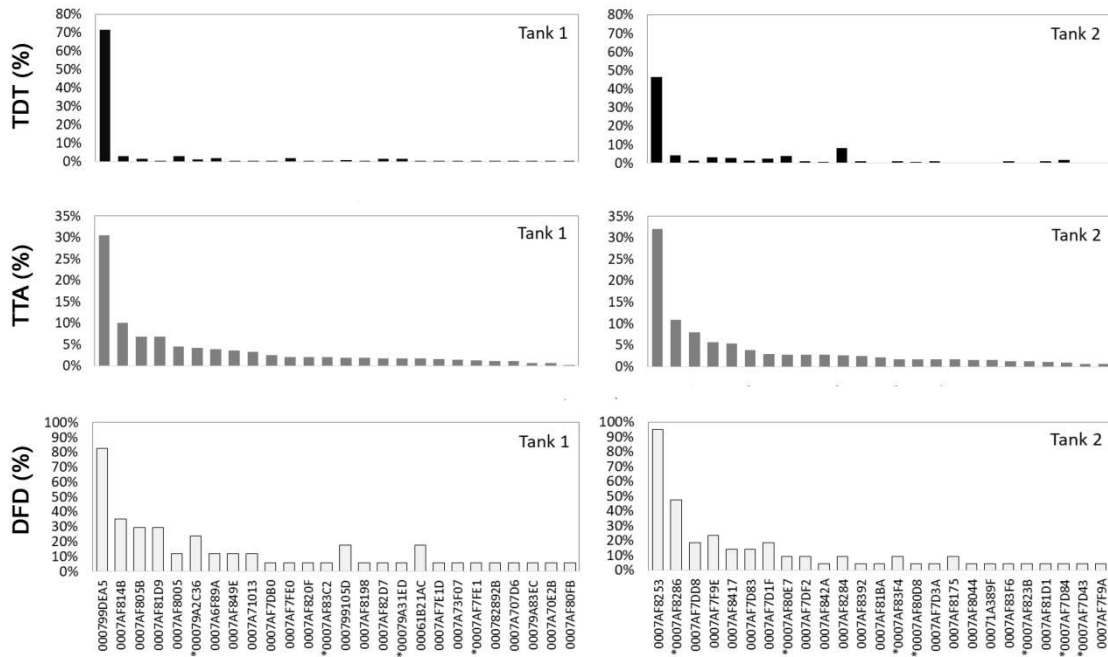
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Figure 1. Scatter-plot and fitted lineal correlation between time of first response and mean number of detections of those fish recorded during both run 1 and 2 within each hypoxia (A) and risk-taking (B) tests, according to fish densities. HD: high density (black symbols and lines); LD: low density (grey symbols and lines). All tests were recorded for 60 minutes. HD tanks in risk taking had around 550 fish and LD tanks had 150 fish. The tanks densities was adjusted bimonthly. Hypoxia tests were performed with groups of 30 fish.



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Figure 2. Scatter-plot and fitted lineal correlation between time of first response and mean number of detections of those fish recorded during both run 1 and 2 within each hypoxia (A) and risk-taking (B) tests, according to feeding strategies. HF: hand feeding (black symbols and lines); DF: self-demanding feeding (grey symbols and lines). All tests were recorded for 60 minutes. All tanks for risk taking tests contained 90 fish. Hypoxia tests were performed with groups of 30 fish.



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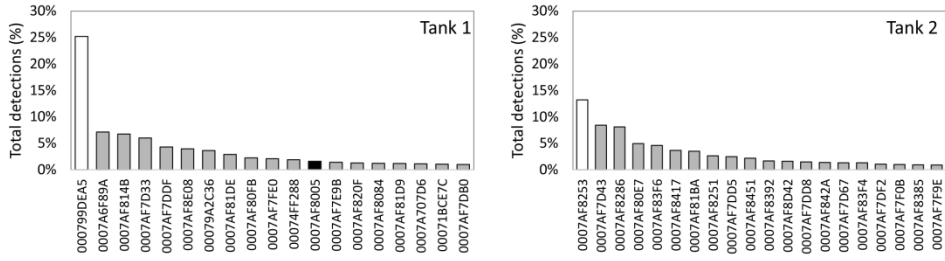
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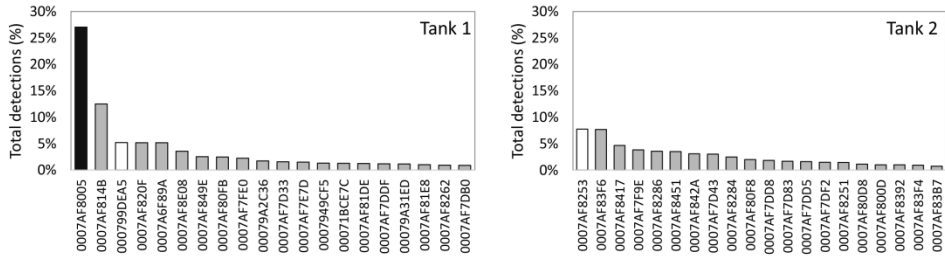
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Figure 3. Percentage of total individual detections (TDT, black bars), percentage of total individual triggering actions (TTA, grey bars), and percentage of days of individual food demand (DFD, white bars), recorded by the PIT-tag antenna around the self-demanding feeders by each juvenile seabream in the experimental tanks. Note: only fish individuals involved in food demand were included in this figure. Asterisks mark individuals considered as proactive.

Post Acute Hypoxia 1



Post Acute Hypoxia 2



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Figure 4. Bar-plots of individual activity (% of total detections) around the self-demanding trigger during the first (5 days; 21/06 - 25/06) and second (7 days; 27/06 - 03/07) post-acute hypoxia periods. White bars highlight the high-triggering (HT) fish during pre-acute hypoxia period. Black bars show the fish individual with the highest proportion of detections during the second post-acute hypoxia period (in tank 1).