



# 6<sup>th</sup> MTERMS 2016

Malaysian Tissue Engineering and Regenerative Medicine Scientific Meeting

in conjunction with

## 2<sup>nd</sup> Malaysian Stem Cell Meeting

*"Ensuring sustainability through innovative regenerative technologies"*



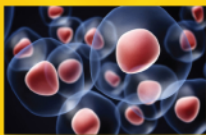
17<sup>th</sup> - 18<sup>th</sup>  
November 2016



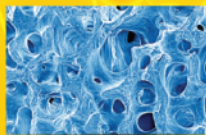
The Light Hotel  
Seberang Jaya, Penang



### Topics



- Reprogramming and pluripotency
- Stem Cell and Cancer



- Biomaterials and Tissue Regeneration
- Transplantation and immunomodulation

- 3D Bioprinting and tissue engineering



- Cell and Gene Therapy
- Imaging and Pre-Clinical Model



### Organised by

Institut Perubatan & Pergigian Ter maju (IPPT), USM and Tissue Engineering & Regenerative Medicine Society of Malaysia (TESMA)

### Co-organised by

Malaysian Society for Stem Cell Research and Therapy (MSCRT)

## P-BTR 2

### **Poly(lactic-co-glycolic acid) and atelocollagen hybrid scaffold seeded with annulus fibrosus cells enhances the formation of cartilaginous tissue engineered construct *in vitro***

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**Purpose:** To evaluate the *in vitro* formation of 3D tissue engineered constructs (TECs) using rabbits' annulus fibrosus (AF) cells seeded on poly(lactic-co-glycolic acid) (PLGA) based scaffolds.

**Methods:** Porous disc-shaped PLGA was fabricated using solvent casting and salt leaching technique. It was crosslinked with atelocollagen to form "PA"scaffold group. Fibrin was added to PLGA and PLGA-atelocollagen composite to form "PF" and "PAF" scaffolds, respectively. The AF cells were seeded into the prefabricated scaffolds ( $1.0 \times 10^5$  cells per scaffold) to form the following TECs groups: AF+PLGA (AFP; *control*), AF+PLGA+atelocollagen (AFPA), AF+PLGA+fibrin (AFPF) and AF+PLGA+atelocollagen+fibrin (AFPAF). The resulting TECs were cultured for three-week and evaluated for cells viability using MTT assay, cellular morphology and attachment using SEM, cartilaginous matrix production using sGAG assay and DNA content using PicoGreen® assay.

**Results:** Significant number of viable cells was observed in the AFPAF group ( $987,985.7 \pm 286,858.9$  cells) when compared to other TECs (AFP:  $373,319.0 \pm 5,456.9$ ; AFPA:  $547,763.4 \pm 66,038.2$ ; AFPF:  $463,763.4 \pm 46,160.8$  cells). Cellular morphology and attachment were comparable in all TECs. The AFPA has the highest sGAG accumulation ( $0.279 \pm 0.117$  mg/ml) but shows no statistical difference when compared to the other TECs (AFP:  $0.083 \pm 0.038$ ; AFPF:  $0.237 \pm 0.131$ ; AFPAF:  $0.181 \pm 0.024$  mg/ml). The AFPF has the highest DNA content ( $1,919.338 \pm 89.050$  ng/ml) but shows no statistical difference when compared to the other TECs (AFP:  $485.659 \pm 27.468$ ; AFPA:  $845.987 \pm 82.134$ ; AFPAF:  $1,575.007 \pm 307.174$  ng/ml). Hence, atelocollagen seemed to provide better environment for cellular attachment and proliferation. This unique collagenous material also promotes sGAG production and DNA content in TECs.

**Conclusion:** The incorporation of atelocollagen into PLGA scaffold enhances the formation of TECs *in vitro*.