

Quantitative 7T MRI does not detect occult brain damage in neuromyelitis optica

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Abstract

Objective

To investigate and compare occult damages in aquaporin-4 (AQP4)-rich periependymal regions in patients with neuromyelitis optica spectrum disorder (NMOSD) vs healthy controls (HCs) and patients with multiple sclerosis (MS) applying quantitative T1 mapping at 7 Tesla (T) in a cross-sectional study.

Methods

Eleven patients with NMOSD (median Expanded Disability Status Scale [EDSS] score 3.5, disease duration 9.3 years, age 43.7 years, and 11 female) seropositive for anti-AQP4 antibodies, 7 patients with MS (median EDSS score 1.5, disease duration 3.6, age 30.2 years, and 4 female), and 10 HCs underwent 7T MRI. The imaging protocol included T2*-weighted (w) imaging and an MP2RAGE sequence yielding 3D T1w images and quantitative T1 maps. We semi-automatically marked the lesion-free periependymal area around the cerebral aqueduct and the lateral, third, and fourth ventricles to finally measure and compare the T1 relaxation time within these areas.

Results

We did not observe any differences in the T1 relaxation time between patients with NMOSD and HCs (all $p > 0.05$). Contrarily, the T1 relaxation time was longer in patients with MS vs patients with NMOSD (lateral ventricle $p = 0.056$, third ventricle $p = 0.173$, fourth ventricle $p = 0.016$, and cerebral aqueduct $p = 0.048$) and vs HCs (third ventricle $p = 0.027$, fourth ventricle $p = 0.013$, lateral ventricle $p = 0.043$, and cerebral aqueduct $p = 0.005$).

Conclusion

Unlike in MS, we did not observe subtle T1 changes in lesion-free periependymal regions in NMOSD, which supports the hypothesis of a rather focal than diffuse brain pathology in NMOSD.

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Glossary

AQP4 = aquaporin-4; **DTI** = diffusion tensor imaging; **EDSS** = Expanded Disability Status Scale; **FLAIR** = fluid-attenuated inversion recovery; **HC** = healthy control; **MRS** = MR spectroscopy; **NAA** = N-acetylaspartic acid; **NAWM** = normal-appearing white matter; **NMOSD** = neuromyelitis optica spectrum disorder; **RRMS** = relapsing-remitting multiple sclerosis; **TE** = echo time; **TI** = inversion time; **TR** = repetition time.

Neuromyelitis optica spectrum disorder (NMOSD) is a severe and often devastating autoimmune and inflammatory CNS disease frequently associated with autoantibodies targeting aquaporin-4 (AQP4) water channels leading to complement activation and focal lesions within AQP4-rich CNS areas such as the spinal cord, the optic nerves, and periependymal regions.^{1,2} In more detail, brain AQP4 water channels are predominantly located within astrocyte foot processes in the glial limiting membrane and in the basolateral cell plasma membrane of ependymal cells.^{3,4}

Clinical,⁵ MRI,⁶ and optical coherence tomography findings mirror the anatomic distribution of AQP4 water channels within the CNS.^{7–9} MRI is used to rule out other disorders and to visualize optic neuritis and signs of myelitis.⁶ On top of that, brain lesion patterns typical for NMOSD have been described including extensive or tumefactive periventricular lesions around the lateral, third, and fourth ventricles and the cerebral aqueduct affecting, e.g., diencephalic structures, the area postrema, the thalamus, the hypothalamus, the corpus callosum, or the periventricular white matter.⁶ Nevertheless, NMOSD-specific brain MRI abnormalities are only detectable within a small proportion of patients,¹⁰ and many patients with NMOSD present with a normal brain MRI,¹¹ which in the past has led to the inclusion of “negative brain MRI at onset” to the 2006 Wingerchuk diagnostic criteria.¹²

Quantitative MRI allows for the quantification of physical variables such as the T1 relaxation time that is sensitive to free-water protons and structural damage to finally compare those variables between tissue regions or participants. When combining quantitative MR techniques with ultra-high-field MRI at 7 Tesla that benefits from an increased signal-to-noise ratio, even subtle degenerative or inflammatory changes that are not obviously present on standard MR images can be assessed.^{13,14}

On this background, we here prospectively performed quantitative T1 relaxometry at 7T to search for occult brain damage within the AQP4-rich, lesion-free and normal-appearing periependymal white or gray matter of patients with NMOSD. We compared our results with those in healthy controls (HCs) and patients with multiple sclerosis (MS).

Methods

Participants

Eleven patients with NMOSD as defined by the 2015 international consensus diagnostic criteria¹ were

prospectively recruited from the outpatient clinic of the department of neurology, Charité—Universitätsmedizin Berlin between January 2014 and December 2015. For comparison, 10 age-matched HCs and 7 patients with relapsing-remitting MS (RRMS), which were best comparable regarding age and sex, were selected from the NeuroCure neuroimaging database as controls. AQP4 antibody serostatus was assessed in patients with NMOSD using one of several established assays.^{15–17} Antibodies against AQP4 were present in all patients with NMOSD. Clinical disability was assessed using the Expanded Disability Status Scale (EDSS) in patients with MS and NMOSD.

Standard protocol approvals, registrations, and patient consents

The Ethics Committee of the Charité–Universitätsmedizin Berlin in conformity with the Declaration of Helsinki approved the study (EA 1/054/09). All participants provided written informed consent.

MRI acquisition

Ultra-high-field MR images were acquired using a 7T Siemens whole-body scanner (Magnetom; Siemens, Erlangen, Germany) and a 24-channel receive head coil (Nova Medical, Wilmington, MA) equipped with a birdcage volume coil for transmission. The imaging protocol included 2-dimensional T2*-weighted fast low angle shot (T2*w FLASH; echo time [TE] = 25.0ms, repetition time [TR] = 1,820 ms; spatial resolution = [0.5 × 0.5 × 2] mm³, supratentorial coverage) and 3-dimensional fluid-attenuated inversion recovery (FLAIR, TE = 90 ms; TR = 16,000 ms; inversion time [TI] = 2,925 ms, spatial resolution = [1.0 × 1.0 × 3.0] mm³).

A 3-dimensional T1-weighted magnetization-prepared rapid gradient-echo sequence with 2 TIs (T1w MP2RAGE, TE = 2.98 ms; TR = 2,300 ms; TI = 900 ms; spatial resolution = (1.0 × 1.0 × 1.0) mm³, whole-brain coverage) was used to generate bias-field corrected T1w images and quantitative T1 maps. Phantom experiments have shown that T1 relaxation times measured using the MP2RAGE approach are closely correlated in a linear fashion with true T1 physical values commonly observed within the brain.¹⁸

Image analysis

All images were analyzed and processed using 3D Slicer (Version 4.6.2 on MacOS 10.11.4, The Slicer Community),

Table 1 Anatomic boundaries for segmentation

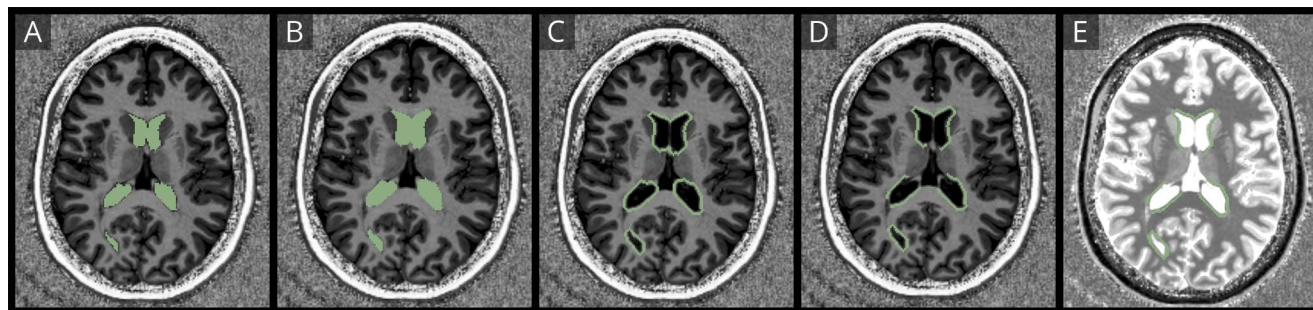
Name of the mask	Anatomic boundaries of the mask					Structures within the mask
	Anterior	Posterior	Superior	Inferior	Medial	
Lateral ventricle	Anterior horn	Posterior horn	Body of the lateral ventricle	End of the anterior horn	Septum pellucidum	Audate nucleus, Genu of corpus callosum
	Corpus callosum			End of the posterior horn	Interventricular foramen	Splenium of corpus callosum
				Inferior horn until collateral trigone		Forceps minor
						Tapetum
						Inferior fronto-occipital fasciculus
Third ventricle	Lamina terminalis	Quadrigeminal cistern	Beginning of the third ventricle	Cerebral aqueduct	Interthalamic adhesion	Thalamus
	Cistern of the lamina terminalis	Posterior commissure				Anterior commissure
	Interventricular foramina	Pineal gland				Posterior commissure
						Fornix
Fourth ventricle	Rhomboid fossa	Cerebellum	Superior medullary velum	Inferior cerebellar	Nodulus	Medial longitudinal fasciculus
	Bottom of the fourth ventricle		Fastigium	Peduncle		Superior cerebellar peduncle
Cerebral aqueduct			Opening of the cerebral aqueduct	Entry in the fourth ventricle		Periaqueductal gray matter
						Inferior colliculus

The table provides an overview on the anatomic boundaries that were used for segmentation of the ventricles.

and `fslmaths` integrated in the FMRIB Software Library (FSL, version 5.0, FMRIB, Oxford, United Kingdom).

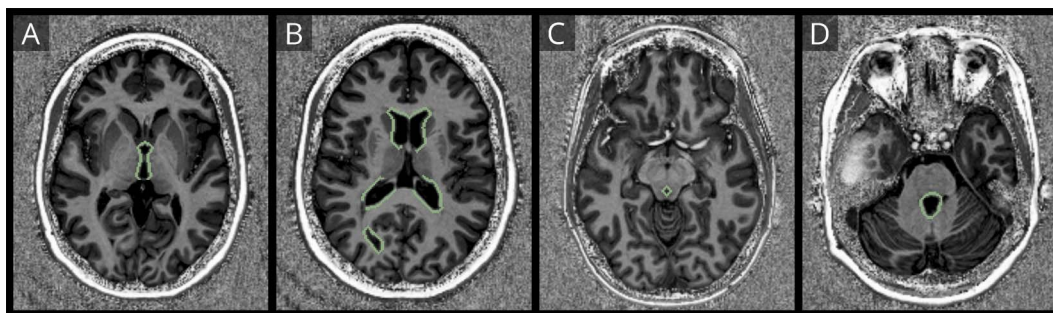
To semiautomatically segment periependymal regions, we defined regions of interest (table 1) and applied a 5-step procedure (figure 1).

First, the lateral, third, and fourth ventricles and the cerebral aqueducts were segmented in consensus reading by a trained and blinded investigator (B.P.) using a semiautomatic threshold-based approach with best visual correction on T1w images. Next, the border of all regions of interest was automatically dilated by 1 mm to avoid partial volume effects of

Figure 1 Segmentation of the normal-appearing periependymal white matter

The figure demonstrates the procedure to segment the normal-appearing periependymal white matter. First, the ventricles were segmented (A) and dilated by 1 mm to avoid partial volume effects and 2 mm to include the periependymal white matter (B). Second, the 1-mm dilated ventricle mask was subtracted from the 1 + 2 mm dilated ventricle mask (C). Finally, the periependymal mask was manually edited for smaller errors, and the lesion mask was subtracted (D) to overlay the final mask with T1 maps (E).

Figure 2 Exemplary masks of periependymal regions



The figure demonstrates exemplary masks of segmented periependymal regions around the third ventricle (A), the lateral ventricles (B), the cerebral aqueduct (C), and the fourth ventricle (D).

the CSF on the final analysis. Third, the border of all regions of interest was automatically dilated by 2 mm, and the 1-mm dilated mask was then subtracted from the 3-mm dilated mask. This step hence creates a 2-mm-thick small rim around the ventricles, which represents the periependymal white matter. Fourth, T1 hypointense and FLAIR/T2*w hyperintense white matter lesions were manually segmented to subtract the lesion map from previously created region-specific periependymal masks. Finally, periependymal masks of the lateral ventricles were split into white and (cortical and deep) gray matter areas by applying T1-threshold-based approach with best visual correction.

The region-specific, CSF-free, lesion-free, and 2-mm-thick periependymal region masks were then used to calculate the mean T1 relaxation time per region and participant using the Label Statistics Module integrated in 3D Slicer (figure 2).

In addition, the lesion count was assessed on T2*w images. Hereby, all T2*w hyperintense lesions larger than 2 mm were counted.

Statistical analysis

All analyses were performed using IBM SPSS Statistics (version 20, IBM, Somers, NY). Normal distribution was assessed visually and by using a Shapiro-Wilk test. T1 measures around the third ventricle, the fourth ventricle, the cerebral aqueduct, and the lateral ventricle (gray matter) were normally distributed. Thus, the Student *t* test was used to assess group differences in mean T1 between patients with NMOSD, MS, and HCs. T1 measures around the lateral ventricle were not normally distributed. Thus, Mann-Whitney *U* test was used to assess group differences in mean T1 (lateral ventricle) between patients with NMOSD, MS, and HCs. Sex differences were assessed using the chi-squared test, and differences in age were assessed using the Student *t* test. *p*-Values <0.05 were considered statistically significant. Given the exploratory nature of the study, no adjustments for multiple comparisons were made.

Data availability

This study was supported by a grant from the Guthy-Jackson Charitable Foundation, which supports the idea of data sharing to facilitate research in the field of NMOSD. Hence, deidentified 7T MRI data of patients with NMOSD included in this study will be shared by the corresponding author with qualified scientific collaborators for research projects on request.

Results

Cohort description

Eleven AQP4 antibody-positive patients with NMOSD and a mean \pm SD age of 43.7 ± 7.12 years (range 22–69 years) were included. Ten HCs (mean \pm SD age 41.6 ± 11.8 years, range 29–67 years) and 7 patients with RRMS (mean \pm SD age 30.2 ± 7.9 years, range 21.4 years) served as controls.

Patients with NMOSD had a total number of 154 (mean \pm SD 14 ± 16.5 , range 0–55) lesions. A total number of 143 lesions (mean \pm SD 20.4 ± 17.4 , range 0–53) were detectable in patients with MS. More clinical details including the EDSS score and sex are presented in table 2.

T1 relaxation time of periependymal regions in healthy controls

Table 3 gives an overview of all results. The mean T1 relaxation time of periependymal regions around the lateral ventricle in HCs was $1,355.6 \pm 49.3$ ms (range 1,296.5–1,472.4 ms). Gray matter areas had a mean T1 relaxation time of $1,772.9 \pm 23.1$ ms (range 1,741.9–1,819.5 ms), whereas white matter areas had a mean T1 relaxation time of $1,220.9 \pm 53.8$ ms (range 1,166.8–1,358.6 ms).

Furthermore, we observed a mean T1 relaxation time of $1,534.7 \pm 47.5$ ms (range 1,470.0–1,597.1 ms) of periependymal regions around the third ventricle, a mean T1 relaxation time of $1,359.2 \pm 25.5$ ms (range 1,322.9–1,409.9 ms) of periependymal regions around the fourth ventricle, and a mean T1 relaxation

Table 2 Cohort overview

	HC	NMOSD	RRMS
n (n, female, <i>p</i> value)	10 (9)	11 (11), <i>p</i> = 0.28	7 (4), <i>p</i> = 0.12
age (years, mean ± SD, range, <i>p</i> value)	41.6 ± 11.8, 29–67	43.7 ± 14.2, 22–69, <i>p</i> = 0.71	30.2 ± 7.9, 21–40, <i>p</i> = 0.04
Disease duration (y, mean ± SD, range)	NA	9.3 ± 8.1, 1–29	3.6 ± 2.1, 0.3–6.6
EDSS (median, range)	NA	3.5, 1.5–5.0	1.5, 0–3.0

Abbreviations: NMOSD = neuromyelitis optica spectrum disorder; RRMS = relapsing-remitting MS; HC = healthy control; EDSS = Expanded Disability Status Scale; NA = not applicable.

Clinical details on studied participants are displayed.

p Values describe differences between NMOSD and RRMS vs HCs.

time of 1,671.0 ± 49.3 ms (range 1,609.5–1753.5 ms) of periependymal regions around the cerebral aqueduct.

T1 relaxation time of periependymal regions in patients with NMOSD

Table 3 and figure 3 give an overview of all results. In comparison to HCs, we observed comparable mean T1 relaxation time of periependymal regions around the lateral (*p* = 0.557), third (*p* = 0.773), and fourth ventricles (*p* = 0.372), as well as around the cerebral aqueduct (*p* = 0.754).

T1 relaxation time of periependymal regions in patients with MS

Table 3 and figure 3 give an overview of all results. In comparison to HCs, we observed longer T1 relaxation times of periependymal regions around the lateral (*p* = 0.043), third

(*p* = 0.027), and fourth ventricles (*p* = 0.013), as well as around the cerebral aqueduct (*p* = 0.005).

In comparison to NMOSD, we observed longer T1 relaxation times of periependymal regions around the lateral (*p* = 0.056), third (*p* = 0.173), and fourth ventricles (*p* = 0.016), as well as around the cerebral aqueduct (*p* = 0.048).

Discussion

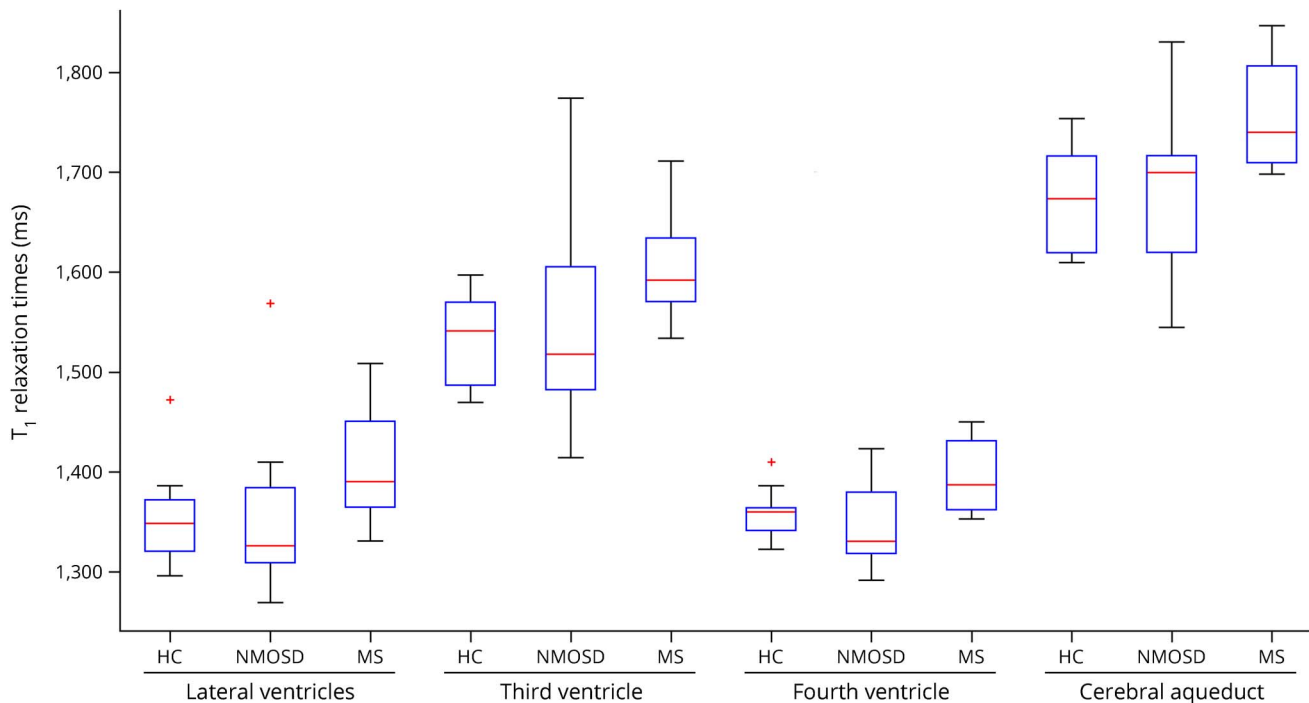
In this study, we aimed to explore occult brain damage in normal-appearing periependymal regions that are characterized by a high expression of AQP4 water channels in patients with AQP4 antibody-positive NMOSD by quantitative MRI with a high signal-to-noise ratio at 7 Tesla. We found that T1

Table 3 T1 of periependymal regions in patients with NMOSD or RRMS and HCs

	HC	NMOSD	MS
Lateral ventricle (T1 in ms, mean ± SD, range, <i>p</i> value)	1,355.6 ± 49.3; 1,296.5–1,472.4	1,354.4 ± 82.1; 1,269.7–1,568.5; <i>p</i> = 0.557	1,405.6 ± 61.9; 1,331.3–1,508.9; <i>p</i> = 0.043
Lateral ventricle GM (T1 in ms, mean ± SD, range, <i>p</i> value)	1772.9 ± 23.1; 1741.9–1819.5	1769.5 ± 45.3; 1,679.9–1831.1; <i>p</i> = 0.838	1816.5 ± 34.4; 1776.8–1877.6; <i>p</i> = 0.007
Lateral ventricle WM (T1 in ms, mean ± SD, range, <i>p</i> value)	1,220.9 ± 53.8; 1,166.8–1,358.6	1,224.8 ± 77.6; 1,162.9–1,441.1; <i>p</i> = 0.809	1,261.5 ± 64.6; 1,210.0–1,393.4; <i>p</i> = 0.070
Third ventricle (T1 in ms, mean ± SD, range, <i>p</i> value)	1,534.7 ± 47.5; 1,470.0–1,597.1	1,545.1 ± 103.8; 1,414.56–1773.81; <i>p</i> = 0.773	1,606.8 ± 58.4; 1,534–1711.2; <i>p</i> = 0.027
Fourth ventricle (T1 in ms, mean ± SD, range, <i>p</i> value)	1,359.2 ± 25.5; 1,322.9–1,409.9	1,345.6 ± 40.3; 1,292.2–1,423.6; <i>p</i> = 0.372	1,397.0 ± 38.2; 1,353.3–1,450.3; <i>p</i> = 0.013
Cerebral aqueduct (T1 in ms, mean ± SD, range, <i>p</i> value)	1,671.0 ± 49.32; 1,609.5–1753.5	1,680.5 ± 82.45; 1,544.8–1830.1; <i>p</i> = 0.754	1757.1 ± 57.1; 1,698.1–1846.3; <i>p</i> = 0.005

Abbreviations: NMOSD = neuromyelitis optica spectrum disorder; RRMS = relapsing-remitting MS; HC = healthy control; GM = gray matter; WM = white matter. T1 relaxation times of periependymal regions in patients with NMOSD or RRMS and HCs are displayed. *p* Values describe differences between NMOSD and RRMS vs HCs. *p* values <0.05 are highlighted in bold.

Figure 3 Box plots



Box plots of mean T1 values within periependymal regions around the lateral ventricles, the third ventricle, the cerebral aqueduct, and the fourth ventricle are displayed.

relaxation times in normal-appearing periependymal regions did not differ between patients with NMOSD and HCs.

The T1 relaxation time is predominantly influenced by structural changes or damage and free protons found in, e.g., inflammatory edema.¹⁹ In other words, both structural damage and edema lead to a prolonged T1 in comparison to healthy brain tissue. By applying quantitative T1 relaxometry at 7 Tesla, one benefits from a substantially increased signal-to-noise ratio and hence increased sensitivity for changes in T1. Thus, T1 relaxation times within the range of those found in HCs argue against the existence of structural damage or edema within the analyzed periependymal regions in patients with AQP4 antibody-positive NMOSD.

Indeed, most previous MRI studies have described no or more focal lesions within the NMOSD brain.^{6,8,9,20} Although NMOSD-specific lesions are often found in AQP4-rich periependymal regions of diencephalic structures, the area postrema, the thalamus, the hypothalamus, the corpus callosum, or the periventricular white matter,⁶ there is only little evidence on a more diffuse or occult damage in those regions.²¹

Several studies have investigated a diffuse or occult damage within the normal-appearing white matter (NAWM) of patients with NMOSD by applying different MRI techniques.

Proton MR spectroscopy (MRS) is a widely applied method used to assess metabolic alterations and the integrity of axonal

and neuronal structures. In NMOSD, normal N-acetylaspartic acid (NAA), creatine, and choline levels were reported within the NAWM, arguing against occult axonal or neuronal damage, inflammation, and gliosis.^{22–24} A recent well-powered MRS study confirmed these findings by reporting normal NAA levels in NMOSD.²⁵

Furthermore, diffusion tensor imaging (DTI)—that is sensitive to structural changes—has been performed in NMOSD. DTI data on the NAWM in NMOSD are inconclusive. On the one hand, no DTI abnormalities were reported within brain regions,²⁶ except for the visual pathway where Wallerian degeneration may occur after optic neuritis.^{27,28} Contrarily, other research groups have observed fractional anisotropy changes—a marker of the structural integrity—within the NAWM of patients with NMOSD.^{29,30} Such DTI abnormalities were, however, rather mild and not as severe as in patients with MS.³¹

Another technique to analyze a more global brain pathology is structural volumetric imaging. Although some groups reported no²⁶ or only mild brain volume changes in NMOSD,^{32,33} others observed white matter volume loss³⁴ but not cortical gray matter volume loss, which contrast atrophy measures in MS.³⁵

Finally, a 7T MRI study on the periventricular venous density in patients with NMOSD did not report changes in venous visibility on highly resolving T2*w images arguing against a widespread hypometabolism in NMOSD.³⁶

All these studies indicate that occult or diffuse brain damage either is absent or only plays a minor role in the pathophysiology of NMOSD,^{21–31} which may result in less brain atrophy in comparison to MS.^{26,32–35} Of note, this assumption is well in line with the clinical presentation of NMOSD.^{2,5} The latter is often characterized by a relapsing-remitting or monophasic disease course. A (secondary) progressive disease course is rare in NMOSD.^{2,5}

In contrast to NMOSD, we clearly observed prolonged T1 relaxation times in patients with MS, especially within the periependymal thalamus and caudate nuclei. Those results may either reflect diffuse normal-appearing white and gray matter damage in MS as indicated by MRS,²⁵ DTI,³¹ volumetric³⁷ or quantitative T1 studies,³⁸ or is caused by small lesions within, e.g., the thalamus that are not obviously seen on conventional MRI.³⁹

Our study is not free of limitations. Although all lesion masks and periependymal regions of interest (ROIs) were best visually corrected with high diligence, we cannot exclude minor misclassifications and partial volume effects. Periependymal ROIs were, however, created with a 1-mm “security” distance around the ventricles. Thus, partial volume effects should not have a relevant effect on this work. In addition, the number of analyzed patients with MS was relatively low.

Our findings of normal T1 relaxation times in normal-appearing lesion-free periependymal regions of patients with NMOSD argue against a severe diffuse or occult brain damage even in AQP4-rich brain regions, which is well in line with the literature, the clinical phenotype of NMOSD, and in contrast to MS. Future work needs to ask, what pathophysiologic processes exactly drive lesion formation in AQP4 antibody-positive human NMOSD.⁴⁰

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Disclosure

B. Pasquier received travel funding from the ECTRIMS. N. Borisow and L. Rasche report no disclosures. J. Bellmann-Strobl received speaker honoraria and travel funding from Bayer, Sanofi-Aventis/Genzyme, Merck, and Teva. K. Ruprecht served on the scientific advisory boards of Sanofi-Aventis/Genzyme, Novartis, and Roche; received travel funding and/or speaker honoraria from Bayer, Biogen, Merck Serono, Sanofi-Aventis/Genzyme, Teva, Novartis, and Guthy-Jackson Charitable Foundation; served as academic editor of *PLoS One*; received publishing royalties from Elsevier; received research support from Novartis, Merck Serono, and German Ministry of Education and Research. T. Niendorf received travel funding from Siemens; served as guest editor for *MR Materials in Physics*,

Biology and Medicine; is founder and CEO of MRI Tools; and received research support from Siemens. T.J. Derfuss served on the scientific advisory boards of Biogen, Novartis, Genzyme, Merck, Bayer, Octapharma, GeNeuro, Roche, Actelion, and Celgene; received travel and speaker honoraria from Bayer, Biogen, Merck, Novartis, Genzyme, and Roche; is a member of the editorial board of *PLoS ONE*; is a member of the steering committee of Mitsubishi Pharma and GeNeuro; is on the speaker bureau of Biogen, Novartis, and Merck; is on the executive board of ECTRIMS; and received research support from Novartis, Biogen, Swiss National Foundation, and Swiss MS Society; his spouse is an employee and holds stock options in Novartis. J. Wuerfel served on the scientific advisory boards of Novartis, Biogen, Genzyme, Teva, and Roche; received travel funding and/or speaker honoraria from Novartis, Bayer, and Biogen; has been employed as CEO at MIAC AG; and received research support from the German Ministry of Education and Research and German Ministry of Economy. F. Paul served on the scientific advisory boards of Novartis and MedImmune; received travel funding and/or speaker honoraria from Bayer, Novartis, Biogen, Teva, Sanofi-Aventis/Genzyme, Merck Serono, Alexion, Chugai, MedImmune, and Shire; is an associate editor of *Neurology: Neuroimmunology & Neuroinflammation*; is an academic editor of *PLoS ONE*; consulted for Sanofi Genzyme, Biogen, MedImmune, Shire, and Alexion; received research support from Bayer, Novartis, Biogen, Teva, Sanofi-Aventis/Genzyme, Alexion, and Merck Serono; and received research support from the German Research Council, Werth Stiftung of the City of Cologne, German Ministry of Education and Research, Arthur Arnstein Stiftung Berlin, EU FP7 Framework Program, Arthur Arnstein Foundation Berlin, Guthy-Jackson Charitable Foundation, and NMSS. T. Sinnecker received travel funding and/or speaker honoraria from Actelion, Roche, and Biogen and has been employed by MIAC. Go to Neurology.org/NN for full disclosures.

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Appendix Author contributions

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Baptiste Pasquier	University Hospital and University of Basel, Basel, Switzerland	Author	Analyzed and interpreted the data, drafted the manuscript, and revised the manuscript
Nadja Borisow, MD	Charité—Universitätsmedizin Berlin, Berlin, Germany	Author	Major role in the acquisition of data and revised the manuscript for intellectual content

Continued

Appendix (continued)

Name	Location	Role	Contribution
Ludwig Rasche, MD	Charité—Universitätsmedizin Berlin, Berlin, Germany	Author	Major role in the acquisition of data and revised the manuscript for intellectual content
Judith Bellmann-Strobl, MD	Charité—Universitätsmedizin Berlin, Berlin, Germany	Author	Major role in the acquisition of data and revised the manuscript for intellectual content
Klemens Ruprecht, MD	Charité—Universitätsmedizin Berlin, Berlin, Germany	Author	Acquisition of data, designed and conceptualized the study, and revised the manuscript for intellectual content
Thoralf Niendorf, MD, PHD	Max Delbrück Center for Molecular Medicine in the Helmholtz Association, Berlin, Germany	Author	Designed and conceptualized the study and revised the manuscript for intellectual content
Tobias J. Derfuss, MD	University Hospital and University of Basel, Basel, Switzerland	Author	Designed and conceptualized the study and revised the manuscript for intellectual content
Jens Wuerfel, MD	Medical Image Analysis Center AG, Basel, Switzerland	Author	Major role in the acquisition of data, interpreted the data, designed and conceptualized the study, and revised the manuscript for intellectual content
Friedemann Paul, MD	Charité—Universitätsmedizin Berlin, Berlin, Germany	Author	Major role in the acquisition of data, interpreted the data, designed and conceptualized the study, and revised the manuscript for intellectual content
Tim Sinnecker, MD	University Hospital and University of Basel, Basel, Switzerland	Author	Major role in the acquisition of data, interpreted the data, designed and conceptualized the study, drafted the manuscript, and revised the manuscript for intellectual content

References

1. Wingerchuk DM, Banwell B, Bennett JL, et al. International consensus diagnostic criteria for neuromyelitis optica spectrum disorders. *Neurology* 2015;85:177–189.
2. Jarius S, Wildemann B, Paul F. Neuromyelitis optica: clinical features, immunopathogenesis and treatment. *Clin Exp Immunol* 2014;176:149–164.
3. Papadopoulos MC, Verkman AS. Aquaporin water channels in the nervous system. *Nat Rev Neurosci* 2013;14:265–277.
4. Jarius S, Paul F, Franciotta D, et al. Mechanisms of disease: aquaporin-4 antibodies in neuromyelitis optica. *Nat Clin Pract Neurol* 2008;4:202–214.
5. Jarius S, Ruprecht K, Wildemann B, et al. Contrasting disease patterns in seropositive and seronegative neuromyelitis optica: a multicentre study of 175 patients. *J Neuroinflammation* 2012;9:14.
6. Kim HJ, Paul F, Lana-Peixoto MA, et al. MRI characteristics of neuromyelitis optica spectrum disorder: an international update. *Neurology* 2015;84:1165–1173.
7. Oertel FC, Kuchling J, Zimmermann H, et al. Microstructural visual system changes in AQP4-antibody-seropositive NMOSD. *Neurol Neuroimmunol Neuroinflamm* 2017;4:e334. doi: 10.1212/NXI.0000000000000541.
8. Popescu BFG, Lennon VA, Parisi JE, et al. Neuromyelitis optica unique area postrema lesions: nausea, vomiting, and pathogenic implications. *Neurology* 2011;76:1229–1237.
9. Pittock SJ, Weinschenker BG, Lucchinetti CF, Wingerchuk DM, Corboy JR, Lennon VA. Neuromyelitis optica brain lesions localized at sites of high aquaporin 4 expression. *Arch Neurol* 2006;63:964–968.

10. Pittock SJ, Lennon VA, Krecke K, Wingerchuk DM, Lucchinetti CF, Weinschenker BG. Brain abnormalities in neuromyelitis optica. *Arch Neurol* 2006;63:390–396.
11. Trebst C, Jarius S, Berthele A, et al. Update on the diagnosis and treatment of neuromyelitis optica: recommendations of the neuromyelitis optica study group (NEMOS). *J Neurol* 2014;261:1–16.
12. Wingerchuk DM, Lennon VA, Pittock SJ, Lucchinetti CF, Weinschenker BG. Revised diagnostic criteria for neuromyelitis optica. *Neurology* 2006;66:1485–1489.
13. Kuchling J, Brandt AU, Paul F, Scheel M. Diffusion tensor imaging for multilevel assessment of the visual pathway: possibilities for personalized outcome prediction in autoimmune disorders of the central nervous system. *EPMA J* 2017;8:279–294.
14. Sinnecker T, Mittelstaedt P, Dörr J, et al. Multiple sclerosis lesions and irreversible brain tissue damage: a comparative ultrahigh-field strength magnetic resonance imaging study. *Arch Neurol* 2012;69:739–745.
15. Kalluri SR, Illes Z, Srivastava R, et al. Quantification and functional characterization of antibodies to native aquaporin 4 in neuromyelitis optica. *Arch Neurol* 2010;67:1201–1208.
16. Jarius S, Probst C, Borowski K, et al. Standardized method for the detection of antibodies to aquaporin-4 based on a highly sensitive immunofluorescence assay employing recombinant target antigen. *J Neurol Sci* 2010;291:52–56.
17. Waters P, Reindl M, Saiz A, et al. Multicentre comparison of a diagnostic assay: aquaporin-4 antibodies in neuromyelitis optica. *J Neurol Neurosurg Psychiatry* 2016;87:1005–1015.
18. Marques JP, Kober T, Krueger G, van der Zwaag W, Van de Moortele PF, Gruetter R. MP2RAGE, a self bias-field corrected sequence for improved segmentation and T1-mapping at high field. *Neuroimage* 2010;49:1271–1281.
19. Sinnecker T, Granziera C, Wuerfel J, Schlaeger R. Future brain and spinal cord volumetric imaging in the clinic for monitoring treatment response in MS. *Curr Treat Options Neurol* 2018;20:17.
20. Jarius S, Ruprecht K, Kleiter I, et al. MOG-IgG in NMO and related disorders: a multicenter study of 50 patients. Part 2: epidemiology, clinical presentation, radiological and laboratory features, treatment responses, and long-term outcome. *J Neuroinflammation* 2016;13:280.
21. Kremer S, Renard F, Achard S, et al. Use of advanced magnetic resonance imaging techniques in neuromyelitis optica spectrum disorder. *JAMA Neurol* 2015;72:815–822.
22. Aboul-Enein F, Krssák M, Höftberger R, Prayer D, Kristoferitsch W. Diffuse white matter damage is absent in neuromyelitis optica. *AJNR Am J Neuroradiol* 2010;31:76–79.
23. Bichueti DB, Rivero RL, de Oliveira EM, et al. White matter spectroscopy in neuromyelitis optica: a case control study. *J Neurol* 2008;255:1895–1899.
24. de Seze J, Blanc F, Kremer S, et al. Magnetic resonance spectroscopy evaluation in patients with neuromyelitis optica. *J Neurol Neurosurg Psychiatry* 2010;81:409–411.
25. Duan Y, Liu Z, Liu Y, et al. Metabolic changes in normal-appearing white matter in patients with neuromyelitis optica and multiple sclerosis: a comparative magnetic resonance spectroscopy study. *Acta Radiol* 2017;58:1132–1137.
26. Finke C, Heine J, Pache F, et al. Normal volumes and microstructural integrity of deep gray matter structures in AQP4+ NMOSD. *Neurol Neuroimmunol Neuroinflamm* 2016;3:e229. doi: 10.1212/NXI.0000000000000541.
27. Pache F, Zimmermann H, Finke C, et al. Brain parenchymal damage in neuromyelitis optica spectrum disorder—a multimodal MRI study. *Eur Radiol* 2016;26:4413–4422.
28. Kuchling J, Backner Y, Oertel FC, et al. Comparison of probabilistic tractography and tract-based spatial statistics for assessing optic radiation damage in patients with autoimmune inflammatory disorders of the central nervous system. *Neuroimage Clin* 2018;19:538–550.
29. Kimura MCG, Doring TM, Rueda FC, Tukamoto G, Gasparetto EL. In vivo assessment of white matter damage in neuromyelitis optica: a diffusion tensor and diffusion kurtosis MR imaging study. *J Neurol Sci* 2014;345:172–175.
30. Liu Y, Duan Y, He Y, et al. A tract-based diffusion study of cerebral white matter in neuromyelitis optica reveals widespread pathological alterations. *Mult Scler* 2012;18:1013–1021.
31. Kim SH, Kwak K, Hyun JW, et al. Diffusion tensor imaging of normal-appearing white matter in patients with neuromyelitis optica spectrum disorder and multiple sclerosis. *Eur J Neurol* 2017;24:966–973.
32. Liu Y, Wang J, Daams M, et al. Differential patterns of spinal cord and brain atrophy in NMO and MS. *Neurology* 2015;84:1465–1472.
33. Hyun JW, Park G, Kwak K, et al. Deep gray matter atrophy in neuromyelitis optica spectrum disorder and multiple sclerosis. *Eur J Neurol* 2017;24:437–445.
34. Blanc F, Noblet V, Jung B, et al. White matter atrophy and cognitive dysfunctions in neuromyelitis optica. *PLoS One* 2012;7:e33878.
35. Chanson JB, Lamy J, Rousseau F, et al. White matter volume is decreased in the brain of patients with neuromyelitis optica. *Eur J Neurol* 2013;20:361–367.
36. Schumacher S, Pache F, Bellmann-Strobl J, et al. Neuromyelitis optica does not impact periventricular venous density versus healthy controls: a 7.0 Tesla MRI clinical study. *MAGMA* 2016;29:535–541.
37. Solomon AJ, Watts R, Dewey BE, Reich DS. MRI evaluation of thalamic volume differentiates MS from common mimics. *Neurol Neuroimmunol Neuroinflamm* 2017;4:e387. doi: 10.1212/NXI.0000000000000541.
38. Vrenken H, Geurts JGG, Knol DL, et al. Whole-brain T1 mapping in multiple sclerosis: global changes of normal-appearing gray and white matter. *Radiology* 2006;240:811–820.
39. Harrison DM, Oh J, Roy S, et al. Thalamic lesions in multiple sclerosis by 7T MRI: clinical implications and relationship to cortical pathology. *Mult Scler* 2015;21:1139–1150.
40. Takeshita Y, Obermeier B, Cotleur AC, et al. Effects of neuromyelitis optica–IgG at the blood–brain barrier in vitro. *Neurol Neuroimmunol Neuroinflamm* 2016;4:e311. doi: 10.1212/NXI.0000000000000541.

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