

## Outbreak and control of haemorrhagic pneumonia due to *Streptococcus equi* subspecies *zooepidemicus* in dogs

M. K. KIM, H. JEE, S. W. SHIN, B. C. LEE, B. PAKHRIN, H. S. YOO, J. H. YOON, D. Y. KIM

NECROTISING haemorrhagic pneumonia due to infection with *Streptococcus equi* subspecies *zooepidemicus* has been described in dogs (Garnett and others 1982, Chalker and others 2003). The severity of pulmonary disease in a population of kennelled dogs with endemic infectious respiratory diseases has been correlated with the presence of *S equi* subspecies *zooepidemicus*. The pathogen has also been isolated from horses, cows, pigs, sheep, goats, guinea pigs, dogs and domestic fowl, and interspecies transmission has been suspected (Biberstein and Hirsh 1999, Quinn and others 2003, Greene 2006).

This short communication describes an outbreak and the immediate control of *S equi* subspecies *zooepidemicus*-induced haemorrhagic pneumonia in a research dog colony. Prompt fluid and antibiotic therapies were initiated, and clinical problems were stabilised within two weeks after the initial presentation without further progression.

Twenty-five adult mixed-breed dogs had been kept separated in a metal cage in a research facility at the College of Veterinary Medicine, Seoul National University. Three days before the incident began, four dogs from a private kennel were moved into the colony. Ten dogs, including newly introduced dogs, developed acute signs that included a moist cough, depression, anorexia, fever, dyspnoea and nasal discharge. Seven dogs, including all four newly introduced dogs, died two to 12 hours after first showing clinical signs.

Blood samples taken from three moribund dogs revealed leucopenia ( $1.8$  to  $3.2 \times 10^{12}$  white blood cells) with monocytosis (68 per cent), a strong indication of acute bacterial infection. There were no apparent abnormalities in the serum chemistry profiles. Radiographic examination revealed that the affected dogs had increased pulmonary opacity to the point where almost the entire lung was affected, except the caudodorsal area. The increase in pulmonary opacity was attributable to bilateral alveolar infiltration, which revealed typical air bronchograms and air alveolograms, particularly in the cranial lobes.

Postmortem examination was performed on the seven dogs soon after death to determine the cause of the outbreak. At postmortem examination the lungs were diffusely haemorrhagic, wet and failed to collapse (Fig 1). A large amount of haemorrhagic frothy fluid was present in the trachea and bronchial airways on cut sections. The nasal cavity and thorax were filled with bloody fluid. No other significant gross abnormalities were found in the thoracic and abdominal cavities. For light microscopic examination, samples from the major parenchymal organs, including lungs, were collected, fixed in 10 per cent phosphate-buffered neutral formalin, routinely processed and stained with haematoxylin and eosin for histopathology. Portions of the lungs were obtained aseptically and used for routine aerobic and anaerobic bacterial cultures. The samples



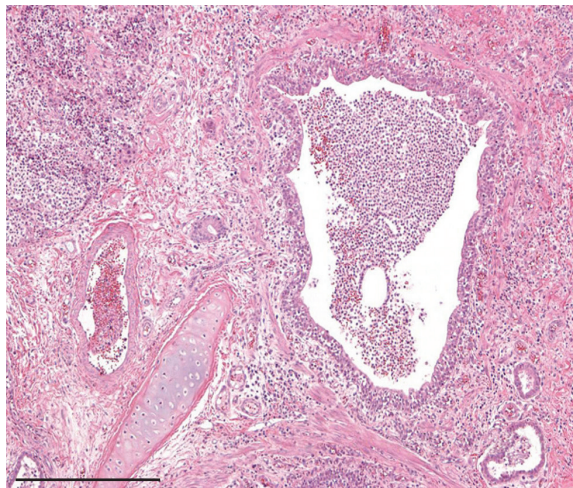
**FIG 1:** Lung from a dog with a history of epistaxis and respiratory distress. All the lobes are haemorrhagic and moist, with consolidation. Scale=cm

were plated on to blood agar and incubated for 24 hours at 37°C; the bacterial colonies were identified using the Vitek system (bioMérieux). Further molecular identification was performed by detection of the gene 23S rRNA, *sodA* (encoding superoxide dismutase A), *seeH* and *seeI* (encoding the superantigenic toxins *seeH* and *seeI*), using PCR, as previously described by Alber and others (2004) and Kawata and others (2004).

Histopathologically, the pulmonary architecture was obscured by haemorrhage and inflammation. The bronchioles, bronchioles and alveolar lumens were diffusely infiltrated with necrotic debris and inflammatory cells, which consisted of predominantly neutrophils and a few lymphocytes, plasma cells and macrophages (Fig 2). Alveolar septa were also hyperaemic. No evidence of septicaemic thrombi was observed in any other organ examined.

Bacteria were isolated in pure culture from the lung and were identified as *Streptococcus equi* using the Vitek system. The isolate was 23S rRNA, *sodA*-positive, and *seeH* and *seeI*-negative (Fig 3). The PCR result was consistent with that of *S equi* subspecies *zooepidemicus*.

An antimicrobial susceptibility test was performed on the isolate using 17 antimicrobial drugs, by the disc diffusion method. A sensitivity test revealed that the isolate was



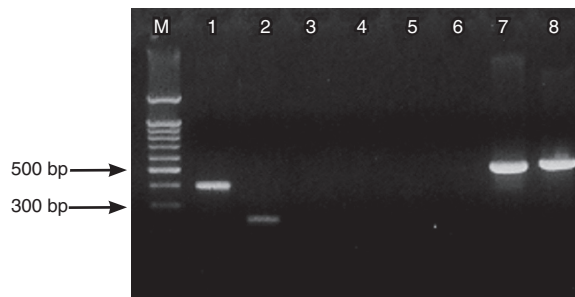
**FIG 2:** Photomicrograph of the lung of a dog. The alveoli and bronchioles are diffusely filled with erythrocytes, necrotic debris and inflammatory cells. Bar=100 µm

*Veterinary Record* (2007) **161**, 528-530

M. K. Kim, DVM, PhD,  
B. C. Lee, DVM, PhD,  
Departments  
of Veterinary  
Therigenology and  
Biotechnology,  
H. Jee, DVM,  
B. Pakhrin, DVM,  
D. Y. Kim, DVM, PhD,  
Department of Veterinary  
Pathology,  
S. W. Shin, DVM,  
H. S. Yoo, DVM, PhD,  
Department of Veterinary  
Infectious Disease,  
J. H. Yoon, DVM, PhD,  
Department of Veterinary  
Radiology, College of  
Veterinary Medicine,  
Seoul National University,  
151-742, Seoul, Korea

Correspondence to  
Dr D. Y. Kim

**FIG 3: Electrophoretic analysis of PCR-amplified products from *Streptococcus equi* isolates and *Streptococcus pyogenes*. Lanes 1, 2, 3, 4 PCR-amplified products from the isolates using primers specific for 23S rDNA, *sodA*, *seeH* and *seel*, respectively; lanes 5, 6, 7, 8 PCR-amplified products from *S pyogenes* using primers specific for 23S rDNA, *sodA*, *seeH* and *seel*, respectively; Lane M DNA size marker (100 base pair [bp] ladder; Intron). The PCR products of the *S equi* isolates have sizes of approximately 399 bp (lane 1) and 320 bp (lane 2); the PCR products of *S pyogenes* have sizes of approximately 500 bp (lane 7) and 520 bp (lane 8). The *S equi* isolates had negative reactions with oligonucleotide primers for *seeH* (lane 3) and *seel* (lane 4); *S pyogenes* had negative reactions with oligonucleotide primers for 23S rDNA (lane 5) and *sodA* (lane 6)**



highly sensitive to penicillin (Penicillin; Becton Dickinson) and enrofloxacin (Baytril; Oxoid). An intravenous infusion of lactated Ringer's solution was given to the three moribund dogs at a maintenance rate of 75 ml/kg/day. A combined preparation of penicillin and streptomycin (consisting of 40,000 iu penicillin and 20 mg/kg streptomycin) (PPS; Dae Sung Microbiological Labs) was also given intramuscularly every 12 hours. This treatment was effective and resolved the dogs' clinical problems promptly.

*S equi* is usually divided into two subspecies, subspecies *zooepidemicus* and subspecies *equi*. Both are a part of the normal bacterial flora in horses (Ruoff 1992). *S equi* acts as an opportunistic pathogen that can cause disease in the upper respiratory tract, uterus, umbilicus and in wounds. Subspecies *zooepidemicus* is associated with several diseases, including lower respiratory tract disorder in horses, pneumonia in ponies and llamas, cervicitis, septicaemia and arthritis in pigs, and mastitis in ruminants (Timoney 1987, Timoney and others 1988, Chanter 1997, Biberstein and Hirsh 1999, Heras and others 2002). Pneumonia and glomerulonephritis due to *S equi* subspecies *zooepidemicus* have also been reported in human beings (Barnham and others 1987, Balter and others 2000).

On the basis of the bacterial isolation together with the histopathological findings, *S equi* subspecies *zooepidemicus* was identified as the aetiological agent of this outbreak. Possible primary viral infections including canine parainfluenza, canine adenovirus type 1 and canine distemper virus were ruled out using appropriate antigen detection methods and histopathology. While this article was being prepared, another *S equi* subspecies *zooepidemicus* epidemic was suspected in the private kennel that had supplied the four dogs involved in the outbreak in the research colony. The authors hypothesise that the outbreak described might have been triggered in the newly acquired dogs by the stress of transportation and joining the colony, possibly associated with an underlying viral infection, since clinical signs were first noticed in these dogs. Tracheal swabs taken from dogs that remained clinically healthy during the outbreak were negative for *S equi*.

In conclusion, *S equi* subspecies *zooepidemicus* should be considered as another definitive causative agent of haemorrhagic pneumonia in dogs, and special caution should be given since interspecies transmission may be possible. To the

authors' knowledge, this is the first outbreak of haemorrhagic pneumonia due to *S equi* subspecies *zooepidemicus* with successful and immediate control by prompt fluid and antibiotic therapy in a research dog colony in Korea.

## ACKNOWLEDGEMENTS

This work was supported by the Brain Korea 21 Program for Veterinary Science and the Korea Research Foundation (KRF-2004-005-E00077).

## References

- ALBER, J., EL-SAYED, A., LAMMLER, C., HASSAN, A. A., WEISS, R. R. & ZSCHOCK, M. (2004) Multiplex polymerase chain reaction for identification and differentiation of *Streptococcus equi* subsp *zooepidemicus* and *Streptococcus equi* subsp *equi*. *Journal of Veterinary Medicine B. Infectious Diseases and Veterinary Public Health* **51**, 455-458
- BALTER, S., BENIN, A., PINTO, S. W. L., TEIXEIRA, L. M., ALVIM, G. G., LUNA, E., JACKSON, D., LACLAIRE, L., ELLIOT, J., FACKLAM, R. & SCHUCHAT, A. (2000) Epidemic nephritis in Nova Serrana, Brazil. *Lancet* **355**, 1776-1780
- BARNHAM, M., LJUNGGREN, A. & MCINTYRE, M. (1987) Human infection with *Streptococcus zooepidemicus* (Lancefield group C); three case reports. *Epidemiology and Infection* **98**, 183-190
- BIBERSTEIN, E. L. & HIRSH, D. C. (1999) Streptococci. In *Veterinary Microbiology*. 1st edn. Eds D. C. Hirsh, Y. C. Zee. Oxford, Blackwell Science. pp 120-126
- CHALKER, V. J., BROOKS, H. W. & BROWNLIE, J. (2003) The association of *Streptococcus equi* subsp *zooepidemicus* with canine infectious respiratory disease. *Veterinary Microbiology* **95**, 149-156
- CHANTER, N. (1997) Streptococci and enterococci as animal pathogens. *Journal of Applied Microbiology* **83** (Suppl 1), 1005-1095
- GARNETT, N. L., EYDELLOTH, R. S., SWINDLE, M. M., VONDERFECHT, S. L., STRANDBERG, J. D. & LUZARRAGA, M. B. (1982) Hemorrhagic streptococcal pneumonia in newly procured research dogs. *Journal of the American Veterinary Medical Association* **181**, 1371-1374
- GREENE, C. E. (2006) Streptococcal and other gram-positive bacterial infections. In *Infectious Diseases of the Dog and Cat*. 3rd edn. Eds C. E. Greene, J. F. Prescott. Philadelphia, Saunders. pp 302-316
- HERAS, L. A., VELA, A. I., FERNANDEZ, E., LEGAZ, E., DOMINGUEZ, L. & FERNANDEZ-GARAYZABAL, J. F. (2002) Unusual outbreak of clinical mastitis in dairy sheep caused by *Streptococcus equi* subsp *zooepidemicus*. *Journal of Clinical Microbiology* **40**, 1106-1108
- KAWATA, K., ANZAI, T., SENNA, K., KIKUCHI, N., EZAWA, A. & TAKAHASHI, T. (2004) Simple and rapid PCR method for identification of streptococcal species relevant to animal infectious based on 23S rDNA sequence. *FEMS Microbiology Letters* **237**, 57-64
- QUINN, P. J., MARKEY, B. K. & MAGUIRE, D. (2003) Streptococci. In *Concise Review of Veterinary Microbiology*. 1st edn. Oxford, Blackwell Publishing. pp 18-19
- RUOFF, K. L. (1992) Group C streptococci: a current view. *Clinical Infectious Diseases* **15**, 175-176
- TIMONEY, J. F. (1987) The Streptococci. In *Pathogenesis of Bacterial Infections in Animals*. 2nd edn. Eds C. L. Gyles, C. O. Theon. Ames, Iowa State University Press. pp 12-13
- TIMONEY, J. F., GILLESPIE, J. H., SCOTT, F. W. & BARLOUGH, J. E. (1988) The genus *Streptococcus*. In *Hagan and Bruner's Microbiology and Infectious Diseases of Domestic Animals*. 8th edn. Ithaca, London, Comstock Publishing Associates. pp 188-192