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1 Influence of aboveground vegetation on seed bank composition and distribution in a

- 2 Great Basin Desert sagebrush community
- 3
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- 16 Abstract
- 17

18 The quantity, composition, and spatial dispersion of seed banks can greatly affect 19 community dynamics. While seed banks of hot deserts have been studied extensively, little is 20 known about seed banks in cold deserts, in particular the relationship between the seed bank and 21 the aboveground vegetation. We investigated the relationship between the seed bank and 22 aboveground vegetation and the effect of microhabitat (shrub interspace or beneath shrub) and 23 aboveground community phase (high or low perennial bunchgrass cover) on the seed bank of a 24 Great Basin Desert sagebrush community. The seed bank and aboveground vegetation differed 25 in their most dominant species, resulting in moderately dissimilar species compositions as 26 determined by Sørensen's similarity index and Bray-Curtis distance. In contrast, comparing the 27 seed bank species composition to the aboveground vegetation structure (functional groups) using 28 non-metric multidimensional scaling (NMDS) revealed a correspondence between the two 29 communities. Shrub seed densities were higher beneath shrubs. Neither microhabitat nor 30 community phase explained variation in total seed density or species richness. Therefore, our 31 measures of the aboveground vegetation did not influence seed density across functional groups 32 or species richness, and the similarity between the seed bank and aboveground vegetation varied 33 depending on the aboveground organizational level used in comparisons. 34 Key words: seed bank, aboveground vegetation, shrub, Great Basin Desert, similarity,

35 microhabitat

36

- **1. Introduction**
- 38

The majority of Great Basin Desert species rely on seeds for propagation; however, seed
banks (or seed pools) of this desert are poorly understood (Kemp, 1989). Seed banks may help

re-establish species that have become locally extinct aboveground. Evaluations of North
American desert seed banks suggest that those of the Great Basin Desert consist of fewer annual
and more perennial species than do hot desert seed banks (Kemp, 1989; Guo et al., 1999).
Although maximum seed densities are generally similar among the North American deserts,
there are some areas of the Great Basin Desert that appear to have very small seed banks (Hassan
and West, 1986).

47 Annual species are more likely than perennial species to form persistent seed banks 48 because they tend to produce dormant seeds (Jurado and Flores, 2005). This strategy allows 49 seeds to wait for proper germination cues which may increase the chance of establishment and 50 survival. However, a long-term seed banking strategy may be difficult for annuals to achieve 51 because of seed reductions caused by continuous germination and granivory (Kigel, 1995). Due 52 to the nature of perennial species, seeds are less likely to be dormant, and therefore, seed banks 53 tend to be transient. Species that form transient seed banks are at risk of becoming locally 54 extinct, especially if seed production is limited (O'Connor, 1991). Seed production can be 55 limited by a number of factors, such as invasive species which may cause native perennial species to produce fewer seeds and die prematurely if the density of the invader is high (Vilà and 56 57 Gimeno, 2007) or by drought years that limit flowering and seed production (St. Clair et al., 58 2009).

59 The invasion of *Bromus tectorum* (cheatgrass) has altered the structure and composition 60 of Great Basin Desert seed banks. Studies examining seed banks of degraded sagebrush 61 communities have shown shifts to greater annual seed abundance with cheatgrass invasion 62 (Young and Evans, 1975; Humphrey and Schupp, 2001). Even in systems that are not 63 considered to be dominated by cheatgrass, introduced species can still account for 20 percent of 64 the total number of seeds in the seed bank (Guo et al., 1999).

65 The relationship between the seed bank and aboveground vegetation is not well understood in Great Basin Desert sagebrush communities. Plant communities dominated by 66 67 perennial species usually have relatively low aboveground-belowground similarities, while 68 annual-dominated communities tend to have a greater correspondence between aboveground 69 vegetation and the seed bank (Thompson and Grime, 1979; Ungar and Woodell, 1993; Milberg, 1995; Bakker et al., 1996; Osem et al., 2006), at least partly because annual-dominated 70 71 communities arise yearly from the available seed bank, which reflects vegetation of the previous 72 year (Osem et al., 2006). 73 When comparing the relationship between the seed bank and aboveground vegetation 74 among forest, grassland (including desert), and wetland seed banks, grassland standing 75 vegetation is most similar to the seed banks in terms of species composition (Hopfensperger, 76 2007). In desert grasslands, extreme environmental conditions may select for species that rely on 77 persistent seed banks, resulting in similar above and belowground communities (Henderson et 78 al., 1988). However, higher similarity between the seed bank and aboveground vegetation in 79 desert grasslands is more likely due to limited dispersal and aggregated seed patterns 80 surrounding parent plants (Shaukat and Siddiqui, 2004). In contrast, some studies have found a 81 lack of correspondence between the seed bank and aboveground vegetation in grasslands which 82 has been attributed to different dominant species in the aboveground and seed bank communities 83 (Eriksson and Eriksson, 1997; Kalamees and Zobel, 1997). For example, the most dominant species in the seed bank may be overrepresented due to high production of small seeds (Eriksson 84 85 and Eriksson, 1997).



The aboveground vegetation not only influences the community composition of the seed

87 bank but also the distribution of the seeds. Although the distribution of seeds within desert seed 88 banks is spatially variable, seeds are frequently more abundant under shrub and tree canopies 89 than in interspaces (Nelson and Chew, 1977; Guo et al., 1998; Marone et al., 2004) and exhibit 90 an aggregated seed pattern due to seeds settling close to the mother plant (Shaukat and Siddiqui, 91 2004). A study investigating spatial patterns of species richness found higher species richness at 92 the mid-point and furthest sampling point from shrubs (2 m and 6 m from shrubs; Li, 2008). In 93 pinyon-juniper woodlands, seed densities and species richness were highest in interspaces and 94 the interface between interspaces and litter under trees (Koniak and Everett, 1983). Shrubs and 95 trees affect the spatial distribution of seeds as they can act as a barrier, altering wind dynamics 96 and subsequent seed deposition patterns. (Guo et al., 1998; Li, 2008; Li et al., 2009). Seeds 97 often accumulate beneath shrubs because they decrease wind velocity and physically trap seeds, 98 leading to deposition close to shrubs (Bullock and Moy, 2004). Seeds can also be redistributed 99 from interspaces to litter beneath shrubs by wind and water (phase II dispersal; Chambers and 100 MacMahon, 1994). 101 This study explores the seed bank and the aboveground vegetation within a Great Basin

Desert plant community and how the aboveground vegetation influences the seed bank community composition and seed distributions. Specific goals were to determine the relationship between the compositions of the seed bank and the aboveground vegetation and the effect of shrubs (microhabitat effects) and perennial bunchgrass cover (community phase effects) on the seed bank community composition, seed density, and seed bank species richness.

107

108 2. Methods

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- 110
- 111 *2.1. Study site*

112	Soil seed bank samples were collected from the Onaqui Sagebrush/Cheatgrass SageSTEP
113	research site in Tooele County, Utah, USA, about 40 km south of Tooele, UT (40°11'53"N
114	112°27'51"W). The Onaqui site is located on the eastern toeslope of the Onaqui mountains at an
115	elevation of 1750-1850 meters (McIver et al., 2010). Based on Utah Climate Center data from
116	the Vernon climate station (Latitude: 40.1125, Longitude: -112.435; Elevation: 1671 m; Period:
117	1953-2010), about 11.3 km south of the study site and in the same valley, mean monthly
118	temperature ranges from -3.1 C in January to 22.0 C in July, while mean monthly precipitation
119	ranges from 17.5 mm in December to 29.5 mm in May, yielding a mean annual precipitation of
120	264.4 mm. Snow can occur in all months except July and August with a peak in January (Utah
121	Climate Center, 2012). Onaqui has fine-loamy soils (McIver et al., 2010).
122	Characteristic vegetation of this site includes Wyoming big sagebrush (Artemisia
123	tridentata ssp. wyomingensis), shadscale saltbush (Atriplex confertifolia), yellow rabbitbrush
124	(Chrysothamnus viscidiflorus), Sandberg bluegrass (Poa secunda), squirreltail (Elymus
125	elymoides), Indian ricegrass (Achnatherum hymenoides), bluebunch wheatgrass
126	(Pseudoroegneria spicata), basin wildrye (Leymus cinereus), and cheatgrass (Bromus tectorum).
127	Seed bank germination assays were conducted at the Utah State University Research
128	Greenhouse Facility in Logan, UT.
129 130	2.2. Experimental design
131	This study presents the results of baseline sampling of the seed bank of the Onaqui
132	sagebrush-cheatgrass site of the SageSTEP network (McIver et al., 2010). The sagebrush-
133	cheatgrass portion of SageSTEP is a large experimental study addressing the effects of
134	restoration treatments (control, prescribed burn, mow, tebuthiuron herbicide, and imazapic

135 [Plateau[®]] pre-emergent herbicide) on sagebrush ecosystems. Burn, mow, and tebuthiuron

treatments were designed to reduce shrub cover, while imazapic is used to reduce the emergence and establishment of exotic annuals. All results presented in this paper are pretreatment and thus do not address the effects of treatments; nonetheless, we describe the experimental design and refer to sampled plots and subplots by their treatment names because treatment effects will be addressed in a subsequent paper. In the present study, the treatment plots should be viewed simply as replicate plots of untreated sagebrush ecosystems.

142 Control, prescribed burn, mow, and tebuthiuron treatments were applied at the whole plot 143 level (75 acres; 30.4 ha), while imazapic was applied at the subplot level (0.1 ha) as a split-plot 144 factor in all whole plots. In the seed bank studies at Onaqui the mow treatment was excluded, 145 leaving three plot-level treatments and one subplot-level treatment.

146 In each sampled plot, subplots with two levels of perennial bunchgrass cover were 147 sampled. Community phases were chosen by dividing the cover of perennial bunchgrasses into 3 148 ranges. Subplots with greater than 19 percent perennial bunchgrass cover were considered phase 149 1 communities, those with 10-19 percent bunchgrass cover were considered phase 2, and those 150 with less than 10 percent perennial bunchgrass cover were considered phase 3 communities. In 151 the present study only phase 1 and 3 communities were sampled in order to examine the effects 152 of high and low native bunchgrass covers on seed banks. Four phase 1 subplots and four phase 3 153 subplots that did not receive Plateau[®] were sampled in the control (CO), prescribed burn (FI), 154 and tebuthiuron (TE) plots, and four phase 1 subplots and four phase 3 subplots that did receive Plateau[®] also were sampled in the control plot for a total of 32 subplots. Although the set of the 155 156 control subplots that received Plateau are not from a true plot, we refer to these subplots as the control-Plateau[®] plot (CP) for convenience. 157

158

159 2.3. Seed bank sampling

160

161	Soil seed bank samples were collected 14-17 and 22-24 August 2006. Within each 0.1-ha
162	(30 x 33 m) subplot, four 28-m transects were laid out so as to not interfere with vegetation
163	surveys. Transects ran north-south and were located at 3, 10, 20, and 27 meters from the
164	northwest corner of the subplot. A composite sample consisting of 5 subsamples from within a
165	25 x 25-cm frame was collected every 3 meters along each of the 4 transects for a total of 10
166	composite samples per transect and 40 per subplot. Collecting many small samples has been
167	shown to increase the precision of estimates of seed numbers in the soil (Bigwood and Inouye,
168	1988). If necessary, sampling locations were shifted slightly in order to assure that all 5
169	subsamples were from the same microhabitat (see below). Subsamples measured 6.1 cm in
170	diameter and were taken to a depth of 4 cm with PVC couplings. Litter and soil layers were
171	collected together. Microhabitat (shrub interspace or beneath shrub) was recorded for each
172	composite sample collected.

173

174 2.4. Vegetation surveys

175 Aboveground vegetation surveys were conducted on transects located at 2, 7, 15, 23, and 176 28 meters from the northwest corner of each subplot. The line-point intercept method was used 177 to measure the cover of each species present along each transect (Herrick et al., 2005). Species 178 intercepted by the pin were recorded every half meter totaling 60 points per transect and 300 179 points per subplot. Subplots were then surveyed to account for any additional species that were 180 not encountered on transects and were assigned a dominance class. Class 1 represents rare 181 species (1-2 plants per subplot); class 2 represents sparse species (<5% cover); class 3 represents 182 common species (5-25% cover); class 4 represents co-dominant species (25-50% cover), and 183 class 5 represents dominant species (>50% cover). Of the additional species found in the subplot

184	survey, none had a higher dominance than 3. Dominance classes were then converted to relative
185	abundance for analysis. Class 1 was converted to 0.0033 (equivalent to a species being hit once
186	during line-point intercept). Class 2 was converted to 0.04 (4% cover), and class 3 was
187	converted to 0.15 (15% cover). Vegetation surveys were conducted in summer 2006.
188	Nomenclature for all plant species followed the USDA NRCS PLANTS Database (2009).
189 190 191 192	2.5. Evaluating the seed bankThe germinable seed bank was evaluated by direct germination in a greenhouse following
193	cold-moist stratification, which has been shown to be an efficient and reliable method for
194	determining species presence in the germinable seed bank (Gross, 1990). Each composite
195	sample was moistened to field capacity and kept in an unlighted refrigerator at 2°C. After 60
196	days of stratification, samples were removed from the refrigerator and spread over a 2-cm layer
197	of sand in planting trays with drainage holes. Planting trays were divided into three 25.4 X 16.9-
198	cm compartments, each containing one soil sample. Spread out soil samples had a depth of ≈ 1.3
199	cm and a volume of \approx 585 cm ³ . Samples were kept moist, and seedlings were identified, counted,
200	and removed as they emerged. Individuals that were not identified in the seedling stage were
201	transplanted and grown until mature.

Due to the high volume of samples collected, not all seed bank samples were evaluated at the same time. Therefore, depending on the availability of greenhouse space, varying numbers of samples were randomly selected from each treatment combination for each germination assay. Eleven of the 40 samples from each treatment combination were germinated and evaluated for each of the first and second germination assay. Six samples from each treatment combination were germinated and evaluated for the third germination assay, and nine samples were germinated and evaluated for the fourth germination assay. Thus, a total of 37 of the 40 samples 209 per subplot were germinated. Each germination assay lasted 150 days. Emergence was initially 210 censused for 115 days, at which point emergence was noticeably reduced. Samples were then 211 dried out for 14 days and mixed, after which watering was resumed and emergence was censused 212 for an additional 21 days.

The first germination assay ran from mid-January to mid-June 2007. The second germination assay ran from mid-June to mid-November 2007. The third germination assay ran from mid-January to mid-June 2008, and the fourth germination assay ran from mid-June to mid-November 2008. First and third germination assays and second and fourth assays ran during the same time of year so as to control for potential seasonal germination effects that could bias results.

219

221

220 2.6. Statistical Analysis

222 Sørensen's similarity index (C_s) and Bray-Curtis distance (BC) were calculated in R 223 version 2.6.2 (R Development Core Team, 2008) to compare the seed community to the 224 aboveground community. These two similarity/distance metrics compare two communities in 225 different ways. Sørensen's similarity is based strictly on presence/absence: $C_s=2w/(2w+A+B)$ 226 where w is the total number of species found in both communities, A is the number of species 227 excusive to above ground, and B is the number of species exclusive to below ground. A C_s of 0 228 represents completely dissimilar communities and C_s of 1 represents identical communities at the 229 level of presence/absence. In contrast to Sørensen's similarity index, Bray-Curtis distance 230 incorporates information on relative abundance (or cover). This metric normalizes relative 231 abundance for communities being compared by dividing the absolute differences by the summation: $BC = \sum_{i}^{n} |x_{ij} - x_{ik}| / \sum_{i}^{n} x_{ij} + x_{ik}$ where x_{ij} is the relative abundance of species i in 232

community *j*, x_{ik} is the relative abundance of species *i* in community *k*, and *n* is the total number of species. A *BC* of 0 represents most similar communities, and a *BC* of 1 represents most different communities. C_s and *BC* were calculated for the entire site and for each plot using relative cover of the aboveground community and relative abundance of the germinable seed bank community to avoid differences in sampling scales.

238 To further compare the community composition of the germinable seed bank to that of 239 the aboveground vegetation, seed bank composition data were ordinated by non-metric 240 multidimensional scaling (NMDS) with a Bray-Curtis distance measure using the metaMDS 241 function in the Vegan package in R version 2.11.1 (Oksanen et al., 2008; R Development Core 242 Team, 2008). Specifically, seed bank composition was compared to the aboveground vegetation 243 structure by assigning one of seven functional groups to all species present aboveground: annual 244 forbs, annual grasses, perennial forbs, perennial grasses, *Poa secunda*, shrubs, and trees. As with 245 the similarity and distance metrics, relative cover of the aboveground community and relative 246 abundance of the germinable seed bank community were used to avoid differences in sampling 247 scales. Functional groups were based on different morphologies and root systems. Poa secunda 248 was considered a different functional group than perennial grasses because *P. secunda* is a 249 shorter grass that has a shallower root system compared to the other perennial grasses. The 250 relative covers of aboveground functional groups were fitted and plotted onto the seed bank 251 species composition ordination solution using the envfit function in the Vegan package in R 252 version 2.11.1 with P < 0.05 to determine significance (Oksanen et al., 2008; R Development 253 Core Team, 2008). NMDS on densities of germinable seed bank species was also used to 254 compare beneath shrub (S) and shrub interspace (I) community compositions and to compare 255 phase 1 and phase 3 community compositions.

256 To determine the number of dimensions for each NMDS, stress values were assessed. 257 Stress is a measure of how much the distances in the reduced ordination space depart from the 258 distances in the original p-dimensional space. High stress values indicate a possibility that sites 259 are randomly being placed without any relation to the original distances. Therefore, ordinations 260 with the lowest possible stress are desirable; values up to 20 are acceptable and can be 261 interpreted ecologically (Clarke, 1993). Regardless of the number of dimensions chosen, all 262 figures are shown in two dimensions because the third dimension did not seem to alter results 263 upon inspection.

264 A mixed-model factorial ANOVA (analysis of variance) was performed to detect 265 differences in total seed density and species richness (total number of species present) between 266 aboveground microhabitat and community phase using the MIXED procedure in SAS version 267 9.1.3 (SAS Institute, 2003) and P < 0.05 to determine significance. Microhabitat and community 268 phase were treated as fixed effects, and plot and subplot were random effects. Because seed 269 bank species composition data were sparse and did not meet the assumptions of normality and 270 heteroscedasticity, seed bank species were grouped into functional groups and analyzed to 271 understand how above ground microhabitat and community phase affect the structure of the seed 272 bank community. The same ANOVA model described above was used to detect seed density 273 differences within each of six functional groups: annual forb, annual grass, perennial forb, 274 perennial grass, *Poa secunda*, and shrub. One tree species (*Tamarix ramosissima*) was found in 275 the germinable seed bank but was not analyzed as a functional group due to very low seed 276 densities and only one species present. Species richness was not analyzed within functional 277 group because sample sizes were too low and assumptions of normality and heteroscedasticity 278 were unable to be met.

279 Total seed density and species richness across functional groups were log transformed to 280 meet the assumptions of normality and homogeneity of variance. Seed densities within the 281 perennial grass, perennial forb, annual forb, Poa secunda, and shrub functional groups were 282 square root transformed. Seed density within the annual grass functional group was log 283 transformed. For significant main effects, least squared means were compared using Tukey's 284 test. Least squared means and standard errors were back-transformed for figures. 285 286 3. Results 287 288 289 3.1. Relationship between the germinable seed bank and aboveground vegetation 290 291 A total of 47 species germinated from the seed bank, and 43 species were recorded 292 aboveground (Appendix A; Appendix B). The germinable seed bank and aboveground 293 vegetation were moderately different in terms of species presence at the overall site scale 294 according to Sørensen's similarity index ($C_s = 0.447$). Results were similar for all individual 295 plots (CO $C_s = 0.421$; FI $C_s = 0.436$; TE $C_s = 0.511$; CP $C_s = 0.433$). When relative abundance 296 was incorporated, Bray-Curtis distance showed a similar trend, where the germinable seed bank 297 and aboveground vegetation were moderately dissimilar at the overall site level and the 298 individual plot levels (Site BC = 0.609; CO BC=0.580; FI BC = 0.587; TE BC = 0.627; CP BC =299 0.638). Sørensen's similarity index concluded that the CO plot had the most dissimilar 300 aboveground-belowground communities while Bray-Curtis distance found that CP had the 301 highest dissimilarity. According to Sørensen's similarity index the aboveground-belowground 302 communities were most similar in the TE plot. However, Bray-Curtis distance identified the CO 303 plot as having the most similar aboveground-belowground communities. 304 The NMDS with two dimensions was an acceptable representation of the original

305	germinable seed bank data (stress = 7.18). Four of the seven above ground structure variables
306	(functional groups) were significantly correlated with the germinable seed bank community, with
307	correlations being highly significant for annual grasses and annual forbs, as expected (Table 1).
308	The two most dominant annual grass species present in the germinable seed bank, Bromus
309	tectorum (brte) and Vulpia octoflora (vuoc), were positively correlated with the cover of the
310	aboveground annual grass functional group (Fig. 1).
311 312 313 314	3.2. Effects of micohabitat and aboveground community phase on germinable seed bank community, seed densities, and species richness
315 316	3.2.1. Germinable seed bank community
317 318	The NMDS plot constructed to compare microhabitats with three dimensions was an
319	acceptable solution (stress=19.88). Interspace and beneath shrub communities did not display
320	distinct community compositions as indicated by the lack of separation in the ordination plot
321	between the two microhabitats (Fig. 2).
322	The NMDS comparing aboveground community phase required three dimensions to
323	achieve an acceptable stress level of 17.38. There was no obvious separation of phase
324	1 and phase 3 community compositions (Fig. 3).
325 326 327	3.2.2. Seed density
328	ANOVA showed that seed density was not significantly affected by microhabitat, phase,
329	or the microhabitat x phase interaction (Table 2). Shrub seed density was the only functional
330	group significantly affected by any of the factors (Table 2). Microhabitat had a significant effect
331	on shrub seed density, with more seeds found beneath shrubs (6.74 ± 3.12) than in interspaces
332	$(3.17 \pm 4.46).$

333

337

334 3.2.3. Species richness

ANOVA showed that species richness of the germinable seed bank was not significantly
 affected by microhabitat, phase, or the microhabitat x phase interaction (Table 2).

338 4. Discussion 339 340 341 4.1. Relationship between the germinable seed bank and aboveground vegetation 342 343 Despite findings that desert grassland germinable seed bank communities may 344 correspond to the aboveground vegetation as a result of limited seed dispersal and clustered 345 seeds surrounding parent plants (Shaukat and Siddiqui, 2004), our results from the similarity and 346 distance metrics did not strongly support this expectation. Both Sørensen's similarity index and 347 Bray-Curtis distance indicate that the germinable seed bank and aboveground community 348 compositions were moderately different at the overall site and individual plot levels. At the plot 349 level, Sørensen's similarity index and Bray-Curtis distance give conflicting results as to which 350 plot had the most and least similar aboveground-belowground communities. This is likely due to 351 Sørensen's similarity index giving greater importance to rare species. The germinable seed bank 352 and aboveground vegetation only shared 19 of the total of 71 species encountered in this study. 353 However, for most species present in both the germinable seed bank and aboveground 354 vegetation, relative abundances were similar except that *Alvssum desertorum* and *Ceratocephala* 355 testiculata were vastly over-represented in the germinable seed bank relative to the aboveground 356 vegetation while Artemisia tridentata was over-represented in the aboveground vegetation 357 relative to the seed bank (Appendix A; Appendix B). 358 Eriksson and Eriksson (1997) have attributed a lack of correspondence between the

359 germinable seed bank and aboveground vegetation to the fact that the dominant species often

360 differ between the two communities. A. tridentata was the most dominant species aboveground 361 at the Onaqui site. Young and Evans (1989) found that no A. tridentata seeds germinated from 362 the seed bank when collected before fall when A. tridentata seeds mature. In contrast, in the 363 present study germinable A. tridentata seeds were found in seed bank samples collected in 364 August, before seed dispersal, but at low densities. Therefore, A. tridentata was over-365 represented aboveground, which decreased the similarity between the germinable seed bank and 366 aboveground vegetation. Conversely, A. desertorum and C. testiculata were abundant in the 367 germinable seed bank but had lower cover aboveground. Annual species such as A. desertorum 368 and C. testiculata that produce small abundant seeds generally may be over-represented in the 369 germinable seed bank (Eriksson and Eriksson, 1997). Although many factors potentially 370 contribute to seed traits, in general annuals are expected to invest more resources in seed 371 production and, all else equal, produce smaller seeds relative to perennials (Silvertown and 372 Charlesworth, 2001). Also, being annuals, A. desertorum and C. testiculata can produce large 373 germinable seed banks and use seed banking as a bet hedging germination strategy (Philippi and 374 Seger, 1989; Gutterman, 2002; Mistro et al., 2005). A. desertorum and C. testiculata may be 375 maintaining dormant seeds to spread the risk of germination over time, allowing seeds to wait for 376 more favorable germination conditions which may increase the chance of establishment and 377 survival. Another possibility for the over-representation of A. desertorum and C. testiculata 378 belowground is the simple fact that these plants were not frequently encountered aboveground 379 during data collection using the line-point intercept method due to their relatively small size (the probability of a pin hitting a smaller plant is lower than the probability of hitting a larger plant) 380 381 and due to primarily actively growing much earlier in the season than when the aboveground 382 sampling occurred.

383 In contrast to the similarity and distance metrics, the NMDS suggested that the 384 germinable seed bank and aboveground vegetation were in fact moderately similar. One reason 385 for this disagreement is the organizational level of the aboveground vegetation used in 386 comparisons. For the similarity and distance metrics, relative abundances were compared at the 387 species-level. However, the NMDS compared the relative abundance of each species in the 388 germinable seed bank to the relative abundance of aboveground vegetation functional groups, i.e. 389 aboveground vegetation structure. Therefore, at the species-level the germinable seed bank and 390 aboveground vegetation communities were only moderately similar, but similarities were 391 considerably greater when comparing germinable seed bank species abundances to the 392 aboveground functional groups. There were a number of species that were only present above or 393 belowground (Tables A.1 and A.2) which decreased similarity between the germinable seed bank 394 and aboveground vegetation. However, the differences between each species present in either 395 community were no longer detected when using functional group as the aboveground 396 organizational level of comparison. 397 The germinable seed bank and aboveground vegetation tend to be more similar in annual

398 communities than in perennial communities (Thompson and Grime, 1979; Ungar and Woodell, 399 1993; Milberg, 1995; Bakker et al., 1996; Osem et al., 2006). NMDS results from the present 400 study did show a significant correlation between the annual germinable seed bank and 401 aboveground structure, but also a significant correlation between the perennial germinable seed 402 bank and aboveground structure. The unexpected correspondence between the perennial 403 germinable seed bank species and aboveground vegetation structure could simply be a function 404 of the comparison between species and functional groups. As displayed by the similarity and 405 distance metrics, the similarity between germinable seed bank and aboveground species

406 compositions was relatively low. However, comparing germinable seed bank species
407 composition to aboveground structure yielded the opposite result. Although the germinable seed
408 bank and aboveground vegetation were not similar at the species level, the germinable seed bank
409 species composition was similar to the aboveground vegetation functional group categories. For
410 example, both *Achnatherum hymenoides* and *Elymus elymoides* were positively correlated with
411 the aboveground perennial grass functional group.

412

413 4.2. Aboveground community phase and microhabitat effects

414 415

416 Shrub was the only functional group whose seed density was significantly affected by 417 microhabitat. The beneath shrub microhabitat contained more shrub seeds than interspaces, 418 which has been observed in other studies (Nelson and Chew, 1977; Guo et al., 1998; Marone et 419 al., 2004). Shrub seed densities tend to be higher under shrub canopies due to seeds falling 420 beneath and adjacent to the parent plant (phase I dispersal; Shaukat and Siddiqui, 2004). Shrubs 421 might have decreased wind velocity, physically trapping seeds beneath shrubs (Bullock and 422 Moy, 2004). Another explanation for higher beneath shrub densities is seeds could have been 423 transported by wind or water from interspaces and trapped in the litter beneath shrubs (phase II 424 dispersal; Chambers and MacMahon, 1994). However, if this was the primary reason we would 425 expect to have higher densities of other seed types beneath shrubs as well.

There are few studies investigating the spatial pattern of species richness of seeds in desert shrub communities, but Li (2008) reported species richness was highest 2 m and 6 m from shrubs. Results from the present research can neither corroborate nor contradict this finding.

429 Species richness was not significantly affected by any of the factors of interest.

430 The invasive grass *Bromus tectorum* was the most dominant annual grass on site.

431 However, annual grass seed density was not affected by aboveground community phase,

432 microhabitat, or the community phase x microhabitat interaction. Although we did not directly 433 examine the effects of Bromus tectorum on the seed bank, the invasion of Bromus tectorum can 434 create shifts to greater annual seed abundance (Young and Evans, 1975; Humphrey and Schupp, 435 2001) which can in turn affect the aboveground vegetation. Regardless of the differences in 436 perennial bunchgrass cover aboveground (community phase), annual grass seed density was not 437 affected—*Bromus tectorum* seed densities were relatively high as was aboveground cover 438 (Appendix A; Appendix B). Even if an invader can be eradicted aboveground, it may be 439 impossible to restore the vegetation community to the composition of an uninvaded community 440 (Vilà and Gimeno, 2007) due to invasive species persisting in the seed bank. Total seed density 441 and species richness, and seed density and species richness for all other functional groups 442 individually, were not significantly affected by aboveground community phase or microhabitat. 443 Seed banks of semiarid deserts can vary extensively spatially (Marone et al., 2004; Coffin and 444 Lauenroth, 1989) which may make it difficult to detect strong effects of these factors on seed 445 density and species richness.

In addition to seed density and species richness, the germinable seed bank species
composition was not strongly affected by community phase or microhabitat, or at least NMDS
did not detect such effects. Due to variability in seed dispersal patterns among species, distinct
germinable seed bank communities as a function of microhabitat and aboveground community
phase may not exist.

451 Prior to our research, the relationship between the seed bank and aboveground vegetation
452 had not yet been examined in cold desert plant communities. Our study found that seed
453 densities, species richness, and the germinable seed bank community composition were not

454 affected by aboveground community phase or, with the exception of shrub functional group seed 455 densities, microhabitat. Species compositions were moderately dissimilar when the germinable 456 seed bank and aboveground vegetation were compared at the species level but were moderately 457 similar when the germinable seed bank was compared to the aboveground vegetation functional 458 groups. These findings provide new insight into seed banks of cold deserts and their potential to 459 influence the aboveground vegetation.

460

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465

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478 Bakker, J.P., Bakker, E.S., Rosen, E., Verweij, G.L., Bekker, R.M., 1996. Soil seed bank

- 479 composition along a gradient from dry alvar grassland to *Juniperus* shrubland. Journal of
 480 Vegetation Science 7, 165-176.
- Bigwood, D.W., Inouye, D.W., 1988. Spatial pattern analysis of seed banks: an improved
 method and optimized sampling. Ecology 69, 497-507.
- Bullock, J. M., Moy, I.L., 2004. Plants as seed traps: inter-specific interference with dispersal.
 Acta Oecologica 25, 35-41.
- Chambers, J.C., MacMahon, J.A., 1994. A day in the life of a seed: movements and fates of
 seeds and their implications for natural and managed systems. Annual Review of Ecology
 and Systematics 25, 263-292.
- 488 Clarke, K.R., 1993. Non-parametric multivariate analyses of changes in community structure.
 489 Australian Journal of Ecology 18, 117-143.
- 490 Coffin, D.P., Lauenroth, W.K., 1989. Spatial and temporal variation in the seed bank of a
 491 semiarid grassland. American Journal of Botany 76, 53-58.
- 492 Eriksson, Å., Eriksson, O., 1997. Seedling recruitment in semi-natural pastures: the effects of
- disturbance, seed size, phenology and seed bank. Nordic Journal of Botany 17, 469-482.
- 494 Gross, K.L., 1990. A comparison of methods for estimating seed numbers in the soil. The
- 495 Journal of Ecology 78, 1079-1093.
- Guo, Q., Rundel, P.W., Goodall, D.W., 1998. Horizontal and vertical distribution of desert seed
 banks: patterns, causes, and implications. Journal of Arid Environments 38, 465-478.
- Guo, Q., Rundel, P.W., Goodall, D.W., 1999. Structure of desert seed banks: comparisons across
 four North American desert sites. Journal of Arid Environments 42, 1-14.
- 500 Gutterman, Y. 2002. Survival strategies of annual desert plants. Adaptations of desert organisms.
- 501 Springer-Verlag, Berlin, Heidelberg, Germany.

- Hassan, M.A., West, N.E., 1986. Dynamics of soil seeds pools in burned and unburned
 sagebrush semi-deserts. Ecology 67, 269-272.
- Henderson, C.B., Petersen, K.E., Redak, R.A., 1988. Spatial and temporal patterns in the seed
 bank and vegetation of a desert grassland community. The Journal of Ecology 76, 717728.
- Herrick, J.E., Van Zee, J.W., Havstad, K.M., Burkett, L.M., Whitford, W.G., 2005. Monitoring
 Manual for Grassland, Shrubland and Savanna Ecosystems. USDA-ARS Jornada
- 509 Experimental Range. The University of Arizona Press, Tucson, Arizona, USA.
- 510 Hopfensperger, K.N., 2007. A review of similarity between seed bank and standing vegetation
- 511 across ecosystems. Oikos 116, 1438-1448.
- Humphrey, D.L., Schupp, E.W., 2001. Seed banks of *Bromus tectorum*-dominated communities
 in the Great Basin. Western North American Naturalist 61, 85-92.
- Jurado, E., Flores, J., 2005. Is seed dormancy under environmental control or bound to plant
 traits? Journal of Vegetation Science 16, 559-564.
- 516 Kalamees, R., Zobel, M., 1997. The seed bank in an estonian calcareous grassland: comparison
- 517 of different successional stages. Folia Geobotanica 32, 1-14.
- 518 Kemp, P.R., 1989. Seed banks and vegetation processes in deserts. In: Leck, M.A., Parker V.T.,
- 519 Simpson, R.L. (Eds.), Ecology of Soil Seed Banks. Academic Press, New York, pp. 257520 282.
- 521 Kigel, J., 1995. Seed germination in arid and semiarid regions. In: Kigel, J., Galili, G. (Eds.),
- 522 Seed Development and Germination. Marcel Dekker, New York, pp. 645-699.
- 523 Koniak, S., Everett, R.L., 1982. Seed reserves in soils of successional stages of pinyon
- 524 woodlands. American Midland Naturalist 108, 295-303.

525	Li, F.R., 2008. Presence of shrubs influences the spatial pattern of soil seed banks in desert
526	herbaceous vegetation. Journal of Vegetation Science 19, 537-548.
527	Li, F.R., Zhao, W.Z., Kang, L.F., Liu, J.L., Huang, Z.G., Wang, Q. 2009. Seed distribution of
528	four co-occurring grasses around Artemisia halodendron shrubs in a sandy habitat. Acta
529	Oecologica 35, 444-451.
530	Marone, L., Cueto, V.R., Milesi, F.A., Lopez de Casenave, J., 2004. Soil seed bank composition
531	over desert microhabitats: patterns and plausible mechanisms. Canadian Journal of
532	Botany 82, 1809-1816.
533	McIver, J.D., Brunson, M., Bunting, S.C., Chambers, J., Devoe, N., Doescher, P., Grace, J.,
534	Johnson, D., Knick, S., Miller, R., Pellant, M., Pierson, F., Pyke, D., Rollins, K., Roundy,
535	B., Schupp, E., Tausch, R., Turner, D. 2010. The Sagebrush Steppe Treatment Evaluation
536	Project (SageSTEP): a test of state-and-transition theory. Gen. Tech. Rep. RMRS-GTR-
537	237.U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station,
538	Fort Collins, CO, 16 pp.
539	Milberg, P., 1995. Soil seed bank after eighteen years of succession from grassland to forest.
540	Oikos 72, 3-13.
541	Mistro, D.C., Rodrigues, L.A.D., Schmid, A.B., 2005. A mathematical model for dispersal of an
542	annual plant population with a seed bank. Ecological Modelling 188, 52-61.
543	Nelson, J.F., Chew, R.M., 1977. Factors affecting seed reserves in the soil of a Mojave desert
544	ecosystem, Rock Valley, Nye County, Nevada. American Midland Naturalist 97, 300-
545	320.
546	O'Connor, T.G., 1991. Local extinction in perennial grasslands: a life-history approach. The
547	American Naturalist 137, 753-773.

548	Oksanen, J., Kindt, R., Legendre, P., O'Hara, B., Simpson, G.L., Stevens, M.H.H. 2008. Vegan:
549	Community Ecology Package. R package version 1.13-1. Available from: http://vegan.r-
550	forge.r-project.org/.
551	Osem, Y., Perevolotsky, A., Kigel, J., 2006. Similarity between seed bank and vegetation in a
552	semi-arid annual plant community: the role of productivity and grazing. Journal of
553	Vegetation Science 17, 29-36.
554	Philippi, T., Seger, J., 1989. Hedging one's evolutionary bets, revisited. Trends in Ecology &
555	Evolution 4, 41-44.
556	R Development Core Team, 2010. R: A language and environment for
557	statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
558	Available from: http://www.R-project.org.
559	SAS Institute. 2003. SAS System for Microsoft Windows. SAS Institute, Inc., Cary, North
560	Carolina, USA.
561	Shaukat, S.S., Siddiqui, I.A., 2004. Spatial pattern analysis of seeds of an arable soil seed bank
562	and its relationship with above-ground vegetation in an arid region. Journal of Arid
563	Environments 57, 311-327.
564	Silvertown, J. and D. Charlesworth. 2001. Introduction to Plant Population Biology. Fourth
565	Edition. Blackwell Science. Oxford, UK.
566	St. Clair, S.B., Sudderth, E.A., Castanha, C., Torn, M.S., Ackerly, D.D. 2009. Plant
567	responsiveness to variation in precipitation and nitrogen is consistent across the
568	compositional diversity of a California annual grassland. Journal of Vegetation Science
569	20, 860-870.
570	Thompson, K., Grime, J.P., 1979. Seasonal variation in the seed banks of herbaceous species in

571	ten contrasting habitats. The Journal of Ecology 67, 893-921.
572	Ungar, I.A., Woodell, S.R.J., 1993. The relationship between the seed bank and species
573	composition of plant communities in two British salt marshes. Journal of Vegetation
574	Science 4, 531-536.
575	USDA, NRCS, 2009. The PLANTS Database. National Plant Data Center, Baton Rouge,
576	Louisiana, USA. Available from: http://plants.usda.gov.
577	Utah Climate Center, 2012. Utah State University. Logan, Utah, USA. Available from:
578	http://climate.usurf.usu.edu/reports/monthly_data_summary.php?stn=429133&unit=SI
579	Accessed 16 December 2011.
580	Vilà, M., Gimeno, I., 2007. Does invasion by an alien plant species affect the soil seed bank?
581	Journal of Vegetation Science 18, 423-430.
582	Young, J.A., Evans, R.A., 1975. Germinability of seed reserves in a big sagebrush community.
583	Weed Science 23, 358-364.

- 584 Young, J.A., Evans, R.A., 1989. Dispersal and germination of big sagebrush (Artemisia
- 585 *tridentata*) seeds. Weed Science 37, 201-206.