

1 **ACCEPTED MANUSCRIPT**

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4 DECIDUOUS FOREST OF PERLIS, PENINSULAR MALAYSIA

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18 **ASSOCIATION OF TREE COMMUNITIES WITH SOIL PROPERTIES IN A SEMI**
19 **DECIDUOUS FOREST OF PERLIS, PENINSULAR MALAYSIA**

20
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30 Running title: Association of tree communities with soil properties
31

32 **ABSTRACT**

33 Distribution of vegetation communities is associated with environmental parameters
34 especially the soil properties of habitats. This study was conducted in a semi-deciduous forest in
35 Perlis State Park (PSP), Perlis to determine the association of tree communities with the soil
36 properties of three distinct forest habitats in this location. Eighteen plots of 40 m × 60 m (0.24 ha
37 each) with sampling area of 1.92 ha (8 plots) in Setul Formation, 0.96 ha (4 plots) in Granite and
38 1.44 ha (6 plots) in Kubang Pasu Formation (totalling 4.32 ha) were established in the PSP. All
39 trees with diameter at the breast height (dbh) of 5.0 cm and above were enumerated, whilst top soil
40 samples were collected from each plot for soil analyses. A total of 412 tree species, 207 genera, and
41 68 families were enumerated in all study plots: 270 tree species from 152 genera and 57 families
42 were found in the Setul forest; 204 tree species of 130 genera and 50 families in the Granite forest;
43 and 109 tree species from 76 genera and 31 families in the Kubang Pasu forest. Euphorbiaceae was
44 the most represented family at Setul, Granite and Kubang Pasu with 36, 19 and 12 species,
45 respectively. Soil analyses showed significant variation in the soil properties of the study sites. The
46 soil at Setul had loam texture, that at Kubang Pasu displayed clay-loam texture, and that at Granite
47 has sandy-loam texture. In summary, the soils were acidic, and low to high concentrations of
48 available nutrients were found in the forest habitats. Ordinations using canonical correspondence
49 analysis indicated that soil factors play an important role in the distribution and diversity of plants
50 in these study sites.
51

52 **Keywords:** canonical correspondence analysis, semi-deciduous forest, Perlis State Park,
53 vegetation–environment relationship
54

55 **INTRODUCTION**

56 Tree species composition varies with the type of habitat in tropical rain forest (Richards
57 1952), and their distributions are often associated with environmental factors (Newbery and Proctor
58 1984; Baillie *et al.* 1987). Many studies conducted in different parts of tropics emphasised the
59 influence of environmental factors, especially soil properties, on plant species distributions
60 (Wentworth 1981; Oliveira-Filho *et al.* 2001; Nizam *et al.* 2013). Relationships between soil
61 variables and plant species distribution have been discovered in various habitats, such as granite and

62 limestone habitats (Nizam *et al.* 2013), habitats of grassland (Amorim and Batalha 2007), savanna
63 communities (Barruch 2005), and tropical rain forest (Silk *et al.* 2010; Sukri *et al.* 2012).

64 Forests of Peninsular Malaysia are predominantly under rainforests, and only a small
65 restricted part of semi-deciduous forests is found in this area. The semi-deciduous forest in
66 Peninsular Malaysia is located near Kra Isthmus, a transitional zone where Malesian and
67 Indochinese floristic regions intersect (van Steenis 1979; Whitmore 1984; Middleton 2003). The
68 State of Perlis, which is situated in the northernmost of the Peninsular Malaysia, is the only area in
69 this country with semi-deciduous forest (Faridah-Hanum 2006). The Perlis State Park (PSP)
70 exhibits a semi-deciduous forest with most areas consisting of limestone hill forests and a small
71 portion composed of granite-based parent material. This study aimed to determine the association
72 between tree communities and their soil properties in different forest habitats with different rock
73 formations. This study is timely because the increasing human pressures on limestone and granite
74 habitats compromise their integrity, diversity and function. Limestone forests are being replaced by
75 plantations, crops and forages to increase their productivity.

76

77

MATERIALS AND METHODS

78 Study site and tree sampling

79 A semi-deciduous forest in PSP, Perlis, Peninsular Malaysia, was selected as our study site
80 (latitude 6° 34' to 6° 43' N, longitude 100° 10' to 100° 13' E) (Figure 1). Tree samplings were
81 conducted in 18 plots (40 m × 60 m each), covering survey area of 4.32 ha. Due to different
82 topography of the selected locations of the forest habitats, thus several plots were selectively
83 established to avoid rocks and huge boulders. As such, the number of plots managed to be
84 established varied between forest habitats; eight plots (1.92 ha) in the Setul, six plots (1.44 ha) in
85 the Kubang Pasu, and four plots (0.96 ha) in the Granite, of different geological formations. All
86 trees with diameter at breast height (dbh) of 5 cm and above were enumerated and identified using
87 published keys (Whitmore 1972; Whitmore 1973; Ng 1978; Ng 1989).

88 Since the number of plots in each forest habitat was unequal as mentioned earlier, hence
89 rarefaction analysis was conducted using EcoSim software program (Gotelli and Entsminger 2003)
90 to produce rarefaction curves. Rarefaction allows observed richness and diversity between sites
91 being compared though sampling efforts are not equal, or samples differ in the total number of
92 individuals (Lee *et al.* 2002). The rarefaction analysis of the study site confirmed that although the
93 sampling efforts between habitats were unequal, nevertheless the species richness was of the same
94 trend with the actual observation in the survey plots (Zakaria *et al.* 2015).

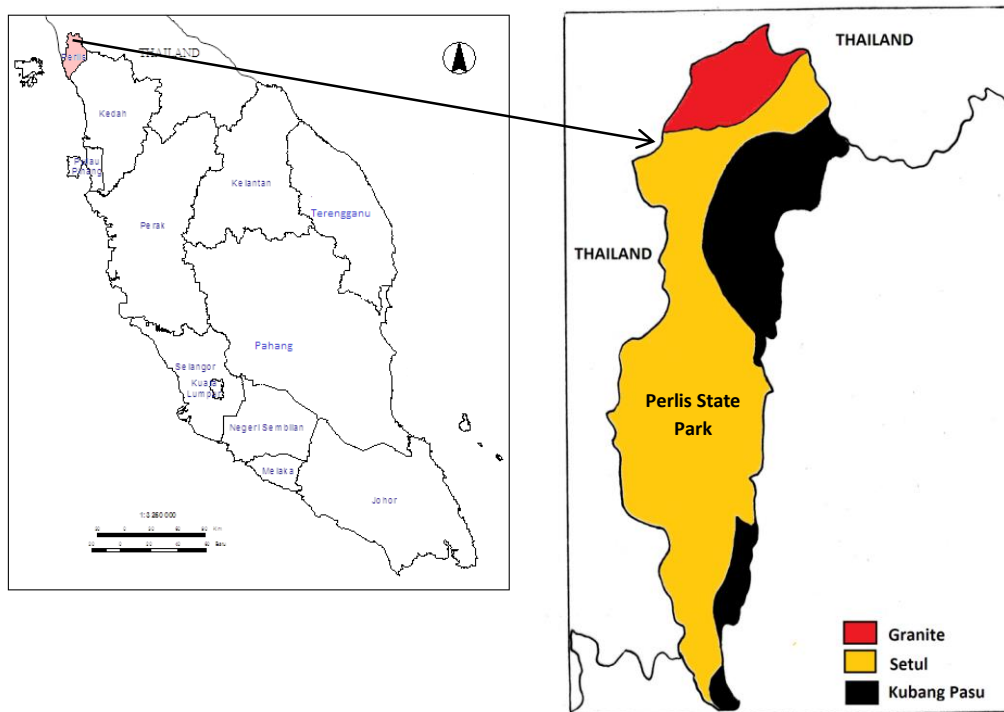


Figure 1. Location of the Perlis State Park in the state of Perlis, Peninsular Malaysia

Soil sampling and analysis

Top soil samples at 0–15 cm depth were collected using soil corer from 18 established plots of various habitats within the PSP. Five replicate samples were obtained from each plot and air dried at room temperature. The replicates were then homogenised together to represent one composite sample from each plot. Root fragments and unwanted materials were removed, and the soils were lightly ground and passed through a 2 mm mesh sieve before analysis.

All soil samples were analysed for physical properties, i.e. particle size distribution and organic matter (OM) content and chemical properties, i.e. exchangeable acid cations (Al^{3+} and H^+), exchangeable base cations (K^+ , Mg^{2+} , Na^+ and Ca^{2+}) and available nutrients (phosphorus (P), potassium (K), and magnesium (Mg)). OM content was measured using loss on ignition techniques, whilst soil pH was recorded based on soil–water ratio of 1:2.5 (McLean 1967). Exchangeable acid cations were investigated in 1 M KCl by titration, whilst exchangeable base cations were analysed in 1 M ammonium acetate extract (Black 1967; Shamshuddin 1981). The extracts were run using the Perkin-Elmer Atomic Absorption Spectrophotometer (AAS) Model 3300 to determine the cation concentrations. Available nutrients (P, K, and Mg) in the soil samples were extracted using 1 M ammonium acetate–acetic acid. The concentration of P in the extract was recorded using Ultraviolet (UV) spectrophotometer, whilst K and Mg concentrations were measured using the AAS.

117 **Data analysis**

118 All enumerated trees in the plots were investigated for overall floristic composition. Values
119 of soil parameters among the study plots were tested for significance using t-test. Association
120 between tree communities and soil variables was analysed using multivariate techniques of
121 canonical correspondence analysis (CCA) (ter Braak 1987; ter Braak and Prentice 1988; ter Braak
122 1992), which is available in CANOCO for Windows 4.56 (ter Braak 2009). Tree species with less
123 than or equal to four occurrences were not included in the analysis because they weaken the
124 uniformity when assigned to the groups (Legendre and Gallagher 2001; Abd Rahman *et al* 2002;
125 Barruch 2005). We selected species with four occurrences or less to limit the number of species to
126 less than 200 and consequently increase the accuracy of the results of CCA ordination diagram.
127 Thus, 161 tree species were selected for the CCA. Detrended Correspondence Analysis (DCA) was
128 first conducted on the species data to confirm their unimodality and thus justify the appropriateness
129 of using CCA (Lepš and Šmilauer 2003). The soil variables selected for the CCA were soil pH,
130 available P, available magnesium (Mg^{2+}), calcium (Ca^{2+}), sodium (Na^+), potassium (K^+) content,
131 ammonium-N (NH_4-N), and nitrate-N (NO_3-N). As recommended by ter Braak (1995), all
132 abundance values were log-transformed prior to matrix processing because their distributions were
133 skewed towards extremely large values. The correlation significance between matrices was tested
134 using Monte-Carlo permutation test based on 499 random trials at a 0.05 significant level (Lepš and
135 Šmilauer 2003). The ordination diagrams were subsequently plotted by CANODRAW 4.14 to
136 illustrate the association patterns of tree communities in relation to soil properties.

137

138

RESULTS AND DISCUSSION

139 **Tree species composition**

140 The tree enumeration was recorded at 4300 trees within the sampling area of 4.32 ha in the
141 PSP. The floristic composition includes 412 tree species, 207 genera and 68 families. The Setul
142 habitat displayed high species richness of 270 species from 152 genera and 57 tree families from
143 1722 trees; the plots in Granite habitat showed 204 tree species, 130 genera and 50 families from
144 1245 trees; and the plots in the Kubang Pasu habitat contained 109 tree species, 76 genera and 31
145 families from 1333 enumerated trees. In all three habitats, Euphorbiaceae was the most represented
146 family with 36 species in Setul, 19 species in Granite; and 12 in Kubang Pasu. On the contrary, 11
147 families were represented by only one species and a single individual in Setul, Granite and Kubang
148 Pasu habitats. These families include Actinidiaceae (*Saurauia pentapetala*), Ancistrocladaceae
149 (*Ancistroclados tectorius*), Anisophylleaceae (*Anisophyllea apetala*), Convolvulaceae (*Erycibe*
150 *albida*), Ctenolophonaceae (*Ctenolophon parvifolius*), Hypericaceae (*Cratoxylum formosum*),
151 Magnoliaceae (*Magnolia elegans*), Oleaceae (*Chionanthus macrocarpus*), Opiliaceae (*Milientha*

152 *suavis*), Oxalidaceae (*Sarcotheca griffithii*), and Rhizophoraceae (*Gynotroches axillaris*). Hence,
 153 these species were not considered in the analysis of species association with soil variables.

154

155 Soil properties

156 The three habitats have different soil properties as follows: the soil in Setul had loam
 157 texture, whereas those in Granite and Kubang Pasu had sandy- and silty-loam textures, respectively.
 158 Granite habitat had the lowest silt and clay contents but the highest sand content at more than 69%
 159 among all study sites (Table 1). The high sand content in Granite is related to sandy parent material;
 160 the quartz sand derived from granitic hill was washed out and deposited a long time ago as
 161 reflecting by the high sandy materials and extremely low clay and silt contents (Osumi 1979;
 162 Zaidey *et al.* 2010). In terms of OM content, the soil of PSP indicates percentages in the range of
 163 2.92% to 18.69% with a mean value of 7.36 ± 1.25 . The soil in Granite showed the significantly
 164 lowest OM content with a mean percentage of 3.12 ± 0.08 ($P < 0.05$), followed by that in Setul with a
 165 mean value of 6.97 ± 1.96 and Kubang Pasu with a mean value of 10.72 ± 2.01 (Table 1). Overall, the
 166 amount of OM in the study sites is considered low. The low OM content in the soil of tropical
 167 rainforests is due to the high decomposition rate of OM (Longman and Jenik 1987).

168

169 Table 1. Summary of soil variables in Setul, Granite and Kubang Pasu habitats at Perlis State Park,
 170 Perlis

Soil parameters	Granite (mean±s.e.)	Setul (mean±s.e.)	Kubang Pasu (mean±s.e.)	p value
Soil pH	4.39±0.12 ^a	5.64±0.35 ^b	5.98±0.30 ^b	p<0.01
Exchangeable cations (meq/100g)				
Ca ²⁺	0.94±0.27 ^a	5.5±2.05 ^b	7.87±2.18 ^b	p<0.05
Mg ²⁺	0.41±0.02 ^a	0.75±0.16 ^b	0.95±0.44 ^a	p<0.05
Na ⁺	0.20±0.04	0.13±0.02	0.13±0.02	NS
K ⁺	0.32±0.03 ^a	0.50±0.06 ^{ab}	0.57±0.12 ^b	p<0.01
Al ³⁺	0.60±0.16	0.19±0.14	0	NS
H ⁺	0.50±0.03	0.35±0.04	0.34±0.02	NS
CEC	2.92±0.2 ^a	7.58±2.09 ^b	9.87±2.54 ^b	p<0.05
Available nutrients (ug/g)				
Phosphorus (P)	9.04±0.31 ^a	7.18±0.69 ^b	7.27±0.37 ^b	p<0.05
Nitrate-N (NO ₃ -N)	27.00±1.5	34.39±6.45	30.96±4.94	NS
Ammonium-N (NH ₄ -N)	3.59±0.46 ^a	18.64±3.57 ^b	30.81±3.57 ^b	p<0.01
Magnesium (Mg)	28.56±0.51	105.35±48.56	139.49±66.78	NS
Potassium (K)	175.02±8.43 ^a	227.27±32.4 ^b	253.80±26.35 ^{ab}	p<0.05

171 Note: mean values with the same letter were not significantly different

172

173 The soil pH showed low mean values of 4.39, 5.64 and 5.98 in Granite, Setul and Kubang
 174 Pasu, respectively, indicating that the soils were strongly to moderately acidic. The soil pH in

175 Granite was significantly lower ($p < 0.01$) than that in Setul and Kubang Pasu (Table 1). Granite soil
176 has the highest acidity probably because it originated from acidic parent rocks (Juo 1981), and the
177 decomposition of OM leads to the acidification of surface soil (Tange *et al.* 1998; Zaidey *et al.*
178 2010). This finding also agrees with a study (Othman and Shamshuddin 1982) which stated that
179 most soils in Peninsular Malaysia tropical rainforests are acidic with pH values between 4.5 and
180 5.5.

181 The cation exchange capacity (CEC) for H^+ , Al^{3+} , Ca^{2+} , Mg^{2+} , Na^+ , and K^+ showed mean
182 value of 7.31 ± 1.35 meq/100g. The CEC in Granite showed the lowest mean value of 2.92 ± 0.20
183 meq/100 g, which was significantly different ($p < 0.05$) from that in Setul (7.58 ± 2.09 meq/100 g) and
184 Kubang Pasu (9.87 ± 2.54 meq/100 g). The CEC varies in size with the clay-humus content of the
185 soil. Clay soils usually have large CEC, whereas sandy soils have low CEC (Cruickshank 1972).
186 The sites showed various concentrations of available macronutrients i.e. magnesium (Mg),
187 potassium (K), phosphorus (P), and soluble nutrients i.e. ammonium-N and nitrate-N (Table 1). The
188 phosphorus concentration in Granite plots was significantly higher than those in Setul and Kubang
189 Pasu with mean concentrations of 9.04 ± 0.31 , 7.18 ± 0.69 and 7.27 ± 0.37 $\mu\text{g/g}$, respectively ($p < 0.05$).
190 The mean concentrations of available magnesium (Mg) in Granite, Setul, and Kubang Pasu were
191 28.56 ± 0.51 , 105.35 ± 48.56 , and 139.49 ± 66.78 $\mu\text{g/g}$, respectively. Nevertheless, these values did not
192 differ significantly.

193 The available inorganic nitrogen in the soil is in the form of nitrate-N ($NO_3\text{-N}$) and
194 ammonium-N ($NH_4\text{-N}$). Overall, the mean values for both $NO_3\text{-N}$ and $NH_4\text{-N}$ of soils at the PSP
195 were 31.60 ± 3.25 and 19.35 ± 3.07 $\mu\text{g/g}$, respectively. The concentrations of these two soluble
196 nutrients showed a consistent trend, wherein Granite habitat displayed lower concentration than
197 Setul and Kubang Pasu habitats did. The mean concentration of ammonium-N was 3.59 ± 0.46 $\mu\text{g/g}$
198 at Granite, 18.64 ± 3.57 $\mu\text{g/g}$ at Setul, and 30.81 ± 3.57 $\mu\text{g/g}$ at Kubang Pasu, and the values were
199 significantly different ($p < 0.01$). Nevertheless, the concentration of nitrate-N was not significantly
200 different among the habitats with values of 27.0 ± 1.50 $\mu\text{g/g}$ at Granite, 34.39 ± 6.45 $\mu\text{g/g}$ at Setul, and
201 30.96 ± 4.94 $\mu\text{g/g}$ at Kubang Pasu (Table 1). The inorganic N in the study sites is characterised by a
202 large $NO_3\text{-N}$, which exceeds the $NH_4\text{-N}$ at all habitats; this might be attributed to the drier
203 conditions in the study sites (Yamashita 2003).

204

205 **Relationships between tree communities and soil variables**

206 DCA confirmed that the data on tree communities were unimodal with the length of gradient
207 of the first axis was 5.758 (Table 2), which is greater than 4 standard deviation (SD) before being
208 analysed by CCA (Lepš and Šmilauer 2003). The eigenvalue produced by the DCA axis 1 (0.758)

209 was high, indicating an environmental gradient where most species vary essentially in their
 210 abundance (ter Braak 1995).

211

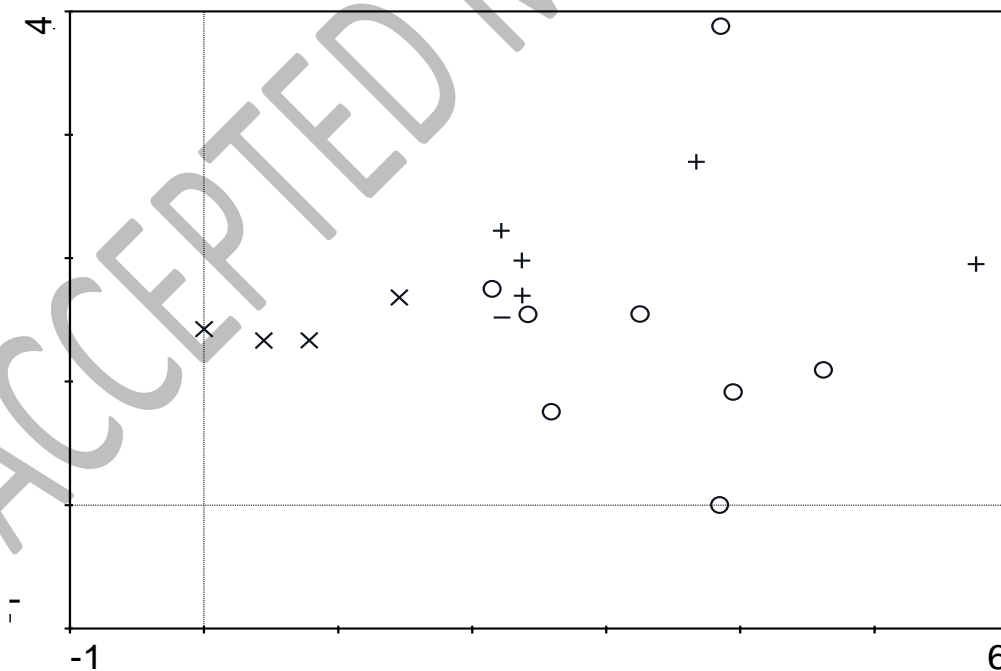
212 Table 2. Summary of the Detrended Correspondence Analysis (DCA) of the vegetation data in 18
 213 plots at Perlis State Park, Perlis

Axes	1	2	3	4	Total inertia
Eigenvalues	0.758	0.464	0.393	0.174	6.527
Lengths of gradient	5.758	3.879	3.012	2.509	
Cumulative percentage variance of species data	11.6	18.7	24.7	27.4	
Sum of all eigenvalues					6.527

214

215 Figure 2 shows the DCA ordination plot indicating the approximate locations of sample
 216 plots and locations. Plots in Granite (symbol x) were clustered together, whilst plots in Setul
 217 (symbol o) and Kubang Pasu (symbol +) exhibited the gradient that occurs from granite to
 218 limestone. The plots clumped together represent the plots with relatively similar floristics, whilst
 219 the separated plots indicate dissimilar floristic composition. The unclearly separated plots in
 220 Kubang Pasu and Setul proved that Kubang Pasu Formation is overlain by Setul Formation (Basir
 221 and Zaiton 2002). Therefore, these habitats shared similar needs for mineral nutrient elements and
 222 other edaphic variables.

223



224

225 Figure 2. DCA ordination diagram of plots based on the abundance of tree species at Perlis State
 226 Park, Perlis. x = plots in Granite; o = plots in Setul; + = plots in Kubang Pasu.

227

228 Based on the CCA analysis, the species–environment correlations were high on the first and
 229 second axes with values of 0.988 and 0.974, respectively (Table 3). The eigenvalue was 0.561 for
 230 the first axis and 0.444 for the second axis. Additionally, the cumulative variation explained by the
 231 first three axes of the species–environment relationship was 55.2%. The Monte-Carlo permutation
 232 test also indicates a significant difference on the eigenvalues among the ordination axes ($p = 0.002$).
 233 Figure 3 shows the CCA ordination plot with the approximate locations of sample plots and
 234 locations, lengths and directions of soil chemical variables. Plots in Granite (1–4) were clustered
 235 together and associated with available phosphorus. Meanwhile, Setul and Kubang Pasu plots did not
 236 cluster into the well-defined group. Their plots were mostly assembled on the bottom right part of
 237 the diagram and were strongly correlated with several soil variables, i.e. magnesium (Mg),
 238 potassium (K), calcium (Ca), pH, available nitrate-N (NO_3), and available ammonium-N (NH_4).
 239

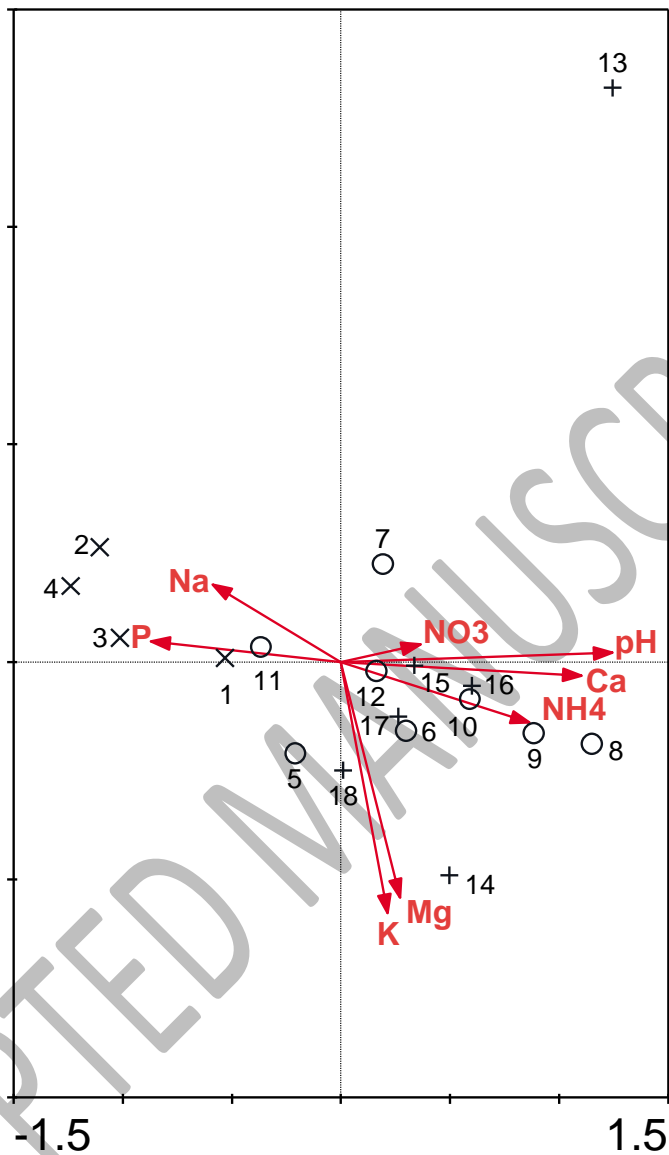
240 Table 3. Summary of the canonical correspondence analysis (CCA) of the vegetation and soil
 241 chemical properties in the 18 plots at Perlis State Park, Perlis

242 Axes		1	2	3	4	Total inertia
243 Eigenvalues	:	0.561	0.444	0.356	0.301	4.238
244 Species–environment correlations	:	0.988	0.974	0.961	0.987	
245 Cumulative percentage variance						
246 of species data	:	13.2	23.7	32.1	39.2	
247 of species–environment relation:		22.7	40.8	55.2	67.5	
248 Sum of all eigenvalues						4.238
249 Sum of all canonical eigenvalues						2.464

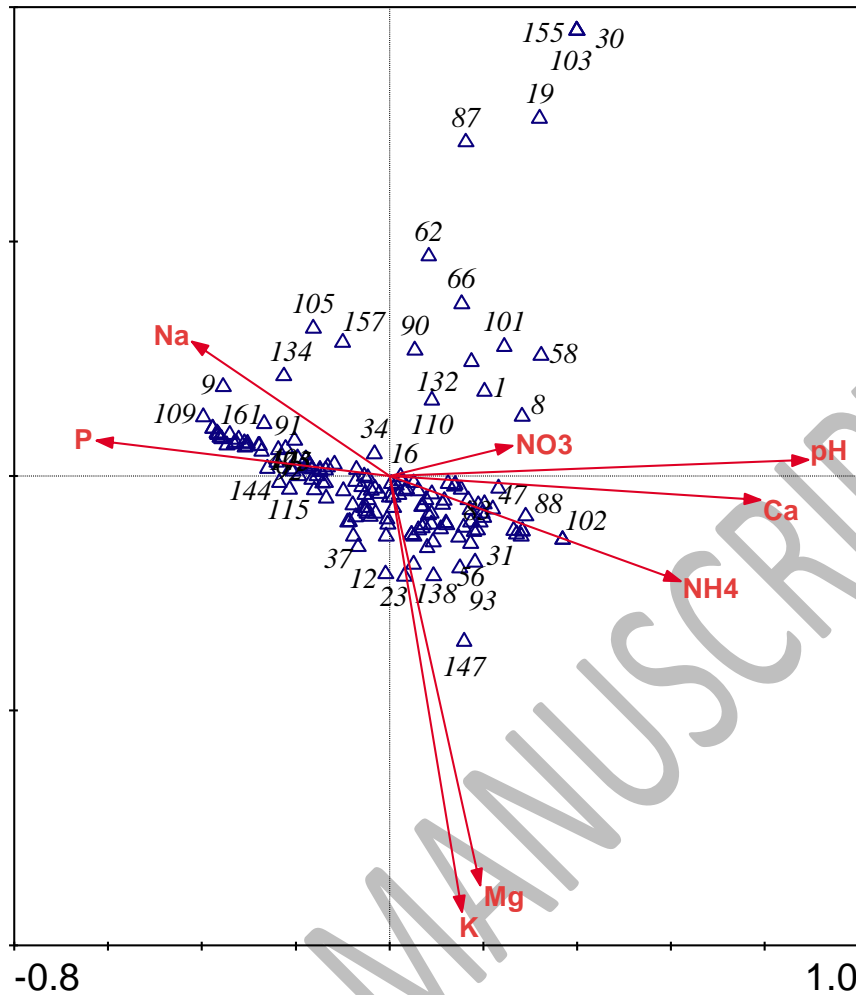
251 The CCA biplot of species and soil variables in Figure 4 shows the species occurrence in
 252 relation to soil variables. Most species were clumped on the centre part of the diagram and
 253 correlated with several soil variables such as available phosphorus (P) and available ammonium-N
 254 (NH_4). Some species such as *Antidesma cuspidatum* (47) were clearly associated with calcium,
 255 whilst *Ficus annulata* (102) and *Duabanga grandiflora* (88) were strongly associated with
 256 ammonium-N. Furthermore, *Hopea ferrea* (34) had strong association with sodium. *Aglaia affinis*
 257 (93) had strong association with magnesium, whilst *Bombax valetonii* (23) was strongly associated
 258 with potassium and magnesium. Based on Figure 4, *Alstonia macrophylla* (19), *Terminalia*
 259 *subspathulata* (30), *Senna timorensis* (87), *Ficus aurata* (103), and *Colona merguensis* (155)
 260 showed a weak association with soil factors when they are farther away from all soil gradients.

261 The variations in species distribution within the study plots were strongly correlated to
 262 edaphic factors and vegetation distribution pattern. Calcium and pH are some factors that strongly
 263 influenced the vegetation pattern in the limestone study area (Setul and Kubang Pasu), whereas
 264 sodium and phosphorus strongly affected the vegetation pattern in Granite area. Soils which
 265 developed on granite are low in calcium and magnesium (Burnham 1974), whereas soils that

266 developed on limestone parent material such as in Setul and Kubang Pasu are mainly high in
267 Calcium and pH (Gauld and Robertson 1985).
268



269
270 Figure 3. CCA ordination plot showing the approximate locations of sample plots and location,
271 length and directions of soil variables. x = plots in Granite (1-4); o = plots in Setul (5-12);
272 + = plots in Kubang Pasu (13-18)
273



274

275 Figure 4. Canonical correspondence analyses of the biplot for tree species and soil variables at the
 276 Perlis State Park. Numbers denote the species in the plots as listed in Table 4
 277

278 Table 4 List of species number denote in diagram of Figure 4

Species code	Species	Species code	Species
1	<i>Alangium kurzii</i>	37	<i>Shorea macroptera</i>
2	<i>Boea oppositifolia</i>	38	<i>Shorea siamense</i>
3	<i>Buchanania arborescens</i>	39	<i>Vatica cinerea</i>
4	<i>Dracontomelon dao</i>	40	<i>Diospyros andamanica</i>
5	<i>Gluta elegans</i>	41	<i>Diospyros buxifolia</i>
6	<i>Gluta velutina</i>	42	<i>Diospyros scortechinii</i>
7	<i>Parishia insignis</i>	43	<i>Diospyros sumatrana</i>
8	<i>Pentaspodon curtisii</i>	44	<i>Diospyros venosa</i>
9	<i>Semecarpus cochinchinensis</i>	45	<i>Elaeocarpus rugosus</i>
10	<i>Semecarpus curtisii</i>	46	<i>Erythroxylum cuneatum</i>
11	<i>Swintonia floribunda</i>	47	<i>Antidesma cuspidatum</i>
12	<i>Alphonsea curtisii</i>	48	<i>Aporosa aurea</i>
13	<i>Goniothalamus tenuifolius</i>	49	<i>Baccaurea griffithii</i>
14	<i>Mitrephora maingayi</i>	50	<i>Chondrostylis kunstleri</i>
15	<i>Orophea cuneiformis</i>	51	<i>Cleistanthus hirsutululus</i>

16	<i>Polyalthia glauca</i>	52	<i>Croton argyratus</i>
17	<i>Xylopi ferruginea</i> var. <i>oxyantha</i>	53	<i>Croton cascarilloides</i>
18	<i>Xylopi magna</i>	54	<i>Croton laevifolius</i>
19	<i>Alstonia macrophylla</i>	55	<i>Dimorphocalyx muricatus</i> var. <i>minor</i>
20	<i>Kibatalia maingayi</i>	56	<i>Drypetes longifolia</i>
21	<i>Ilex cymosa</i>	57	<i>Koilocedrus longifolium</i>
22	<i>Radermachera glandulosa</i>	58	<i>Macaranga andamanica</i>
23	<i>Bombax valetonii</i>	59	<i>Macaranga lowii</i>
24	<i>Durio lowianus</i>	60	<i>Mallotus peltatus</i>
25	<i>Canarium littorale</i> f. <i>rufum</i>	61	<i>Sapium baccatum</i>
26	<i>Dacryodes rubiginosa</i>	62	<i>Sauroupus suberosus</i>
27	<i>Dacryodes rugosa</i>	63	<i>Trigonostemon villosa</i>
28	<i>Lophopetalum javanicum</i>	64	<i>Lithocarpus cantleyanus</i>
29	<i>Parastemon urophyllus</i>	65	<i>Casearia capitellata</i>
30	<i>Terminalia subspathulata</i>	66	<i>Homalium longifolium</i>
31	<i>Tetrameles nudiflora</i>	67	<i>Hydnocarpus castanea</i>
32	<i>Dipterocarpus costatus</i>	68	<i>Hydnocarpus curtisii</i>
33	<i>Dipterocarpus fagineus</i>	69	<i>Hydnocarpus filipes</i>
34	<i>Hopea ferrea</i>	70	<i>Osmelia maingayi</i>
35	<i>Hopea latifolia</i>	71	<i>Garcinia eugeniifolia</i>
36	<i>Parashorea stellata</i>	72	<i>Garcinia hambroniana</i>
73	<i>Garcinia nigroleniata</i>	119	<i>Ardisia pachysandra</i>
74	<i>Garcinia parvifolia</i>	120	<i>Maesa ramentacea</i>
75	<i>Kayea kunstleri</i>	121	<i>Syzygium cerasiforme</i>
76	<i>Mesua ferrea</i>	122	<i>Syzygium glaucum</i>
77	<i>Stemonurus malaccensis</i>	123	<i>Ochna integerrima</i>
78	<i>Cryptocarya ferrea</i>	124	<i>Galearia fulva</i>
79	<i>Cryptocarya rugulosa</i>	125	<i>Galearia maingayi</i>
80	<i>Litsea machilifolia</i>	126	<i>Xanthophyllum affine</i>
83	<i>Leea aequata</i>	127	<i>Xanthophyllum griffithii</i>
84	<i>Callerya artropurpurea</i>	128	<i>Prunus grisea</i>
85	<i>Cynometra malaccensis</i>	129	<i>Aidia densiflora</i>
86	<i>Saraca cauliflora</i>	130	<i>Diplospora malaccensis</i>
87	<i>Senna timoriensis</i>	131	<i>Ixora pendula</i>
88	<i>Duabanga grandiflora</i>	132	<i>Morinda elliptica</i>
89	<i>Lagerstroemia floribunda</i>	133	<i>Psydrax</i> sp. 7
90	<i>Lagerstroemia ovalifolia</i>	134	<i>Psydrax</i> sp. 8
91	<i>Memecylon caeraleum</i>	135	<i>Dimocarpus longan</i> ssp. <i>longan</i>
92	<i>Memecylon minutiflorum</i>	136	<i>Guioa bijuga</i>
93	<i>Aglaia affinis</i>	137	<i>Harpullia cupanioides</i>
94	<i>Aglaia argentea</i>	138	<i>Nephelium costatum</i>
95	<i>Aglaia simplicifolia</i>	139	<i>Nephelium lappaceum</i>
96	<i>Aglaia spectabilis</i>	140	<i>Paranephelium macrophyllum</i>
97	<i>Aphanamixis polystachya</i>	141	<i>Pometia pinnata</i>
98	<i>Chisocheton ceramicus</i>	142	<i>Xerospermum noronhianum</i>
99	<i>Chisocheton patens</i>	143	<i>Chrysophyllum roxburghii</i>
100	<i>Chukrasia tabularis</i>	144	<i>Palaquium hexandrum</i>
101	<i>Artocarpus dadah</i>	145	<i>Palaquium microphyllum</i>

102	<i>Ficus annulata</i>	146	<i>Eurycoma apiculata</i>
103	<i>Ficus aurata</i>	147	<i>Pterocymbium javanicum</i>
104	<i>Ficus fistulosa</i>	148	<i>Pterospermum javanicum</i>
105	<i>Ficus microcarpa</i>	149	<i>Pterospermum pectiniforme</i>
106	<i>Ficus oligodon</i>	150	<i>Pterygota alata</i>
107	<i>Ficus sundaica</i>	151	<i>Sterculia cordata</i>
108	<i>Ficus variegata</i>	152	<i>Sterculia gilva</i>
109	<i>Streblus elongatus</i>	153	<i>Sterculia parviflora</i>
110	<i>Streblus ilicifolius</i>	154	<i>Tetramerista glabra</i>
111	<i>Streblus macrophyllus</i>	155	<i>Colona merguensis</i>
112	<i>Streblus toxoides</i>	156	<i>Grewia viminea</i>
113	<i>Horsfieldia polyspherula</i>	157	<i>Pentace strychnoidea</i>
114	<i>Horsfieldia sucosa</i>	158	<i>Schoutenia accrescens</i>
115	<i>Horsfieldia tomentosa</i>	159	<i>Teijsmanniodendron coriaceum</i>
116	<i>Knema laurina</i>	160	<i>Vitex pinnata</i>
117	<i>Knema patentinervia</i>	161	<i>Vitex siamica</i>
118	<i>Ardisia crassa</i>		

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CONCLUSION

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