

1 **ACCEPTED MANUSCRIPT**

2

3 **ANTIFUNGAL ACTIVITY OF *Citrus hystrix* EXTRACT AND ITS APPLICATION AS**
4 **NATURAL FOOD PRESERVATIVE**

5

6 **Nisa K, Herawati ERN, Nurhayati S, Rosyida VT**

7

8 **DOI: -**

9

10 **To appear in : BIOTROPIA Issue**

11

12 **Received date : 06 November 2018**

13 **Accepted date : 18 February 2019**

14

15 **This manuscript has been accepted for publication in BIOTROPIA journal. It is unedited,**
16 **thus, it will undergo the final copyediting and proofreading process before being published in**
17 **its final form.**

18

ACCEPTED MANUSCRIPT

19 ANTIFUNGAL ACTIVITY OF *Citrus hystrix* EXTRACT AND ITS APPLICATION AS
20 NATURAL FOOD PRESERVATIVE

21
22 **Khoirun Nisa***, **Ervika Rahayu Novita Herawati**, **Septi Nurhayati** and **Vita Taufika Rosyida**

23 Research Unit for Natural Products Technology, Indonesian Institute of Sciences, Gunungkidul
24 55861, Yogyakarta, Indonesia

25 *Corresponding author, e-mail: nisa.khoirun@yahoo.com

26
27 Running title: Antifungal activity of *Citrus hystrix* DC as food preservatives

28
29 **ABSTRACT**

30 Traditional ready to eat foods, such as sticky rice cake, are easily contaminated by spoilage
31 pathogens during their storage. Hence, this study aims to evaluate the effect of *Citrus hystrix* extract
32 for reducing spoilage pathogens in sticky rice cake during storage. The tested sticky rice cake was
33 formulated with *Citrus hystrix* extract at varied level of concentrations of 0.65%, 1.26%, and 1.82%
34 (w/w). Treated samples were stored at room temperature for 28 days and evaluated periodically for
35 their microbial activity (total plate count), thiobarbituric acid reactive substances (TBARS), and
36 sensory analysis. The *Citrus hystrix* extract was also examined for its antifungal activity against
37 *Penicillium sp.* and *Aspergillus nidulans* prior to formulation. Results exhibited a significant
38 advantage of the addition of extracts to sticky rice cake. All extract levels effectively eliminated the
39 spoilage microorganism until 28 days storage and significantly lowered the TBARS values.
40 Physico-chemical properties of sticky rice cake including pH, water activity, and moisture content
41 were equal among all formulated samples and slightly different at 1.82% (w/w) extract level.
42 Moreover, the addition of *Citrus hystrix* extract up to 1.82% did not affect the acceptability of
43 sticky rice cake on the sensory attributes as compared to the control ($P>0.05$).

44
45 **Keywords:** Antifungal, *Citrus hystrix*, sticky rice cake, natural preservative

46
47 **INTRODUCTION**

48 In the recent years, food safety issues have become one of the main public health concerns.
49 The spoiled food is unacceptable for human consumption due to alteration in sensory attributes such
50 as taste, colour, appearance, odor, and texture (Holley & Patel 2005). Some methods widely used to
51 protect microbial spoilage of food commodities is the addition of synthetic or natural preservatives,
52 which directly supplemented to the foods or incorporated in the food packages (Brul & Coote
53 1999). Recently, there has been a worldwide effort to minimize the use of chemical preservatives
54 since consumer preferences are inclined towards more natural and healthier products. Consequently,
55 the consumer's inclination for foods deprived of chemical preservatives has led to the discovery of
56 new natural antimicrobial and antioxidant preventing agents (Serra *et al.* 2008).

57 Natural preservatives such as essential oils, flavonoids, phenolic compounds, and microbial
58 metabolites are the chemical agents derived from plants, animals, and microbes that could preserve
59 food by fighting against fungi and food borne bacteria (Prakash *et al.* 2014). They prevent the
60 decomposition of products by inhibit microbial growth, oxidation and certain enzymatic reactions

61 occurring in the foodstuffs (Singh *et al.* 2010). *Citrus hystrix* (family Rutaceae), commonly known
62 as kaffir lime, is a tropical herb distributed in Southeast Asia. The useful parts of *C. hystrix* are the
63 fruit, leaves, and peel. Its leaves are aromatic and used as a spice and for various flavoring
64 purposes. The essential oil of *C. hystrix* was characterized by high contents of terpinen-4-ol
65 (13.0%), α -terpineol (7.6%), 1,8-cineole (6.4%), and citronellol (6.0%). Previous studies reported
66 that *C. hystrix* oil was effective for antioxidant, repellent, and antiviral (Waikedre *et al.* 2010). Its
67 oil has antibacterial activity against 20 serotypes of *Salmonella* and five species of other
68 enterobacteria (Nanasombat & Lohasupthawee 2005).

69 Sticky rice cakes are popular dessert in Asian countries, particularly in Indonesia. Sticky
70 rice cakes can be made in different base matters including sticky or glutinous rice, sugar (palm,
71 brown, and cane), and some other ingredients such as variety of beans, coconut, and sesame seeds.
72 The most popular methods to prepare these foods are by way of steaming, frying, and boiling (Lee
73 *et al.* 2009). Sticky rice cake is one of ready to eat foods which is commonly stored in room
74 temperature. In the state of California, USA, Korean rice cakes are allowed to be sold at room
75 temperature for up to 24 hours after production by listing the date and time of manufacturing
76 (California State Legislature 2016). Most literatures reported that sticky rice cakes can be kept for
77 up to a few days of storage at room temperature. In addition, there have been many investigations
78 on how to maintain the microbiological quality of these products. On this basis, this present study
79 aims to evaluate the potential of *C. hystrix* extract for reducing spoilage pathogens in ready to eat
80 food, sticky rice cake, during storage.

81 82 **MATERIALS AND METHODS**

83 **Preparation of leaves extracts**

84 *C. hystrix* leaves were purchase from Beringharjo market, Yogyakarta, Indonesia. The
85 leaves were washed, dried, and grinded into powder. Then, it was turned into aqueous extract by
86 homogenizing of 250 g of the material with 2500 mL of distilled sterile water, followed by
87 sonication (for 60 min three times). The resulted liquid was filtered and evaporated to get
88 concentrated extract. The extract was further used in formulation for food preservative.

89 90 **Evaluation of antifungal activity**

91 Three food spoilage microorganisms were isolated from sticky rice cake and identified as
92 *Penicillium sp.*, *Aspergillus nidulans*, and *Rhizopus stolonifer*. Those fungi were then isolated and
93 further used as the test organisms. The plates containing three extract concentrations, including
94 control sample were inoculated in the core of the plate by spotting the 8 mm in diameter of fungal
95 species until round inoculums were formed. Inoculation was performed in four replications with

96 two inoculums per plate. After inoculation, Petri plates were closed properly and incubated at 27
97 °C. The evaluation of antifungal activity was carried out by measuring of the radial growth of the
98 mycelium (in diameter) in each plate during 3 days for *Aspergillus nidulans* and *Rhizopus stolonifer*
99 and 7 days for *Penicillium sp* in the presence of extracts. The antifungal activity (AFA) was
100 calculated by the following equation:

$$101 \text{ AFA (\%)} = (GC - GT) / GC \cdot 100$$

102 It is noteworthy that AFA is antifungal activity (%), GC is colony diameter on the control plate
103 (mm) and GT is colony diameter on the test plate (mm) (Mori *et al.* 1997).

104

105 **Preparation of extract granule and sticky rice cake**

106 The concentrated extract of *C. hystrix* leaves was formulated with lactose through a
107 granulation process with a ratio of 1: 4 (w/w). The extracts were homogeneously mixed with lactose
108 fillers to produce a convenient wet mass. The wet mass was passed through a sieve with 30 mesh
109 size. The obtained granules were dried at 60 °C for 36 hours to reach less than 2% of moisture
110 content. Dried granule was then passed back through a sieve with a 30 mesh size and stored until it
111 is used as food preservative.

112 The resulted granule was then applied on the processing of sticky rice cake with varied
113 concentration (0.65%, 1.26%, and 1.82%). Granule was mixed with the sticky rice cake dough in a
114 few minutes before product was completely cooked, as the temperature was getting lower. The
115 product was packed in pieces of small plastic.

116

117 **Physico-chemical properties of sticky rice cake**

118 Moisture content was calculated using moisture balance (AND MX50, Tokyo, Japan). Water
119 activity was determined by Kjeldahl steam distillation (JP Selecta Pro-Nitro S 4002851, Barcelona,
120 ES). The pH value was measured in pH meter (Eutech PC700, NY, US).

121

122 **Microbial analysis of product**

123 Four samples of the tested food product including control were analyzed after 0, 7, 14, 21,
124 and 28 days of storage. Total plate count (TPC) was determined following the incubation at 37 °C
125 for up to 28 days, on Plate Count Agar (PCA, Merck). Twenty five grams of sample were dissolved
126 in 225 ml of 0.85% NaCl solution, and then mashed using a stomacher. The prepared samples were
127 serially diluted tenfold addressed to minimize the number of microbes in the sample solutions. The
128 bacteria were isolated by pour plate methods using 10^{-1} , 10^{-2} , 10^{-3} , and 10^{-4} dilutions. The colonies

129 were counted using colony counter after 24 hrs and 48 hrs incubation and the results were expressed
130 as cfu/ml.

131 Aerobic plate count was determined by the spread plate technique using standard methods
132 with some modification. All yeast and mould counts were done in triplicate, using Potato Dextrose
133 agar (PDA) medium (Merck, Germany), supplemented with 0.01 % chloramphenicol (Merck,
134 Germany). PDA plates were incubated for up to 5 days at 30 °C. The number of visible colonies on
135 Yeast and Mould Count Plate was read after 3 days and 5 days.

136

137 **TBARS determination**

138 The tested food products were analyzed after 0, 14 and 28 days of storage. Lipid stability
139 was evaluated using the thiobarbituric acid reactive substances (TBARS) index according to a
140 method reported by Targladis *et al.* (1960) with slight modification. Ten grams of sticky rice cake
141 sample in the distilled water (50 mL) were minced using a blender (Philips, England). The minced
142 product was filtered and transferred to a distillation system and steam distillation carried out with
143 4N HCl (2.5 mL). The distillate (2.5 mL) was reacted with a 0.02 M thiobarbituric acid solution
144 (2.5 mL), incubated in a boiled water bath for 35 min, and then cooled until it reaches room
145 temperature. Absorbance at 528 nm was measured using a UV Vis Spectrophotometer (Hitachi
146 HALO RB-10). Thiobarbituric acid reactive substances values were calculated from a standard
147 curve and expressed as mg MDA/kg sample.

148

149 **Sensory analysis**

150 The sticky rice cake characteristics was frequently evaluated in terms of odor, tastes, colour,
151 texture, and overall acceptance (Meilgaard *et al.* 1991). The 20 untrained taste panellists assess the
152 sensorial properties every 7 days during products storage. Sensory evaluation scores were
153 determined using a 7-point Hedonic scale ranging from 1 to 7. The same superscript symbols in the
154 sticky rice cake score indicate that sample are not significantly different at a significance level of
155 95%.

156

157 **Statistical analysis**

158 Mean values and standard deviations were calculated from the data obtained from triplicate
159 experiments. One-way Analysis of Variance (ANOVA) with a significant level of 95% test was
160 used to determine the significant differences between variables.

161

162

RESULTS AND DISCUSSION

163 Antifungal activity of *C. hystrix*

164 The antifungal activity (AFA) of *C. hystrix* extract has been evaluated towards the isolated
165 spoilage fungi growth in tested sticky rice cake that was identified as *Penicillium sp* and *Aspergillus*
166 *sp.* The *in vitro* results showed that the selective antifungal activity of *C. hystrix* extract against
167 *Penicillium sp.* and *Aspergillus nidulans* were $33.53 \pm 31.67\%$ and $40.22 \pm 11.08\%$, respectively.
168 Meanwhile, there was no antifungal activity observed against *Rhizopus stolonifer*. Since *C. hystrix*
169 extract exhibited remarkable antifungal activity, thus the extract was further tested in a traditional
170 sticky rice based dessert, sticky rice cake. Warm climate and avoiding refrigeration are the main
171 causes of high level contamination in sticky rice due to spoilage of microorganism. To the best of
172 our knowledge, the effect of *C. hystrix* leaf extracts on the microbiological stability of food
173 products containing sticky rice had not been studied previously.

174

175 Physico-chemical properties of sticky rice cake

176 Abundant nutrients, high water activity, and nearly neutral pH make sticky rice cake a good
177 medium for microbial growth and categorized as products that require temperature control.
178 Therefore, the preservative additive might be necessary because this product is normally stored
179 through un-refrigeration method. Water activity is one of the variables that contribute to the reduced
180 shelf life of a food product. Water can be retained and trapped inside the pores of food products
181 under the activity of water which should be removed. The water activity of all samples was ranged
182 0.68 to 0.78 (see Table 1). This range is relatively lower than the minimum growth requirement for
183 several pathogen bacteria, such as *S. aureus* (0.86), *B. subtilis* (0.95), *E. coli* (0.96), *Salmonella spp.*
184 (0.96), and *Pseudomonas spp.* (0.97) (Molan 1992). According to Filtenborg *et al.* (1996), the
185 spoilage is due to species enduring in a lower water activity (0.95). High lactose concentration
186 (1.82% extract level) led to the reducing of water activity as reported by Mundo *et al.* (2004).

187

188 Table 1 Physico-chemical properties of sticky rice cake formulated with *C.hystrix* extract

Physico-chemical properties	Level of extracts (%)				
	Day	Control ^a	0.65	1.26	1.82
Water activity (a_w)	0	0.78	0.78	0.78	0.76
	14	0.76	0.70	0.70	0.68
	28	0.77	0.77	0.76	0.75
pH	0	6.21	6.07	5.56	6.01
	14	6.18	6.22	6.05	5.88
	28	6.27	6.26	5.93	5.76
Moisture content (%)	0	27.83	25.24	24.86	27.28
	14	20.15	19.14	21.05	20.77
	28	18.27	18.07	18.30	19.14

189 Control : product in the absence of extract.

190
191
192
193

194 Total Plate Count (TPC)

195 Most (sticky) rice-based desserts and snacks are soft and elastic, but during the storage they
196 become hard due to retrogradation in room temperature. Hence, it is not suggested to store them
197 using refrigeration (Morris 1990). They are ready to eat food as they contain some components that
198 are already cooked and do not need further heating prior to consumption, so the microbiological
199 quality of sticky rice cake is very important. Microbiological purity is an important quality criterion
200 in the food products, with a limit for the number of microbes as stipulated in applicable regional
201 law. The average number of microbial colonies in sticky rice cake after 28 days storage is shown in
202 table 2.

203

204 Table 2 Total bacteria of sticky rice cake formulated with *C.hystrix* extract

Storage duration (days)		Bacterial colonies (cfu/ml)			
		Control	A	B	C
0	24 h	0.51×10^4	0.93×10^4	9.75×10^4	1.10×10^4
	48 h	0.36×10^4	0.66×10^4	10.45×10^4	0.71×10^4
7	24 h	7.90×10^4	0.64×10^4	1.02×10^4	1.29×10^4
	48 h	6.45×10^4	0.97×10^4	0.99×10^4	1.02×10^4
14	24 h	1.09×10^4	0.31×10^4	0.19×10^4	1.83×10^4
	48 h	1.04×10^4	0.46×10^4	0.22×10^4	2.89×10^4
21	24 h	0.34×10^4	0.20×10^4	0.10×10^4	0.20×10^4
	48 h	0.43×10^4	0.31×10^4	0.15×10^4	0.30×10^4
28	24 h	0.20×10^4	0.20×10^4	0.15×10^4	0.20×10^4
	48 h	0.41×10^4	0.25×10^4	0.20×10^4	0.60×10^4

205 Control: product in the absence of extract.; A: product formulated with 0.6% extract; B: product
206 formulated with 1.26% extract; C: product formulated with 1.82%.

207

208 Total plate count (TPC) value of sticky rice cakes formulated with *C. hystrix* extract were
209 ranged from 0.1×10^4 to 10.5×10^4 cfu/ml. The addition of uneven extracts during mixing process
210 resulted in a significant increase in the TPC value during storage time. All samples were classified
211 as good or acceptable level of microbiological quality, which are $<10^5$ cfu/g as the guideline for
212 ready to eat foods (NSW Food Authority 2009). Moreover, testing results for products showed that
213 the addition of *C. hystrix* extract as antibacterial agent was effective to reduce the microbial growth
214 on weeks observation compared to the control. However, the better capability was exhibited by the
215 1.26% extract than 1.82% extract. Lee *et al.* (2009) reported that the addition of 1 or 3% green tea
216 or rosemary to rice cakes gave no significantly affect to the number of microbial properties.

217 Conversely, on three days storage, the growth of *B. cereus* and *S. aureus* could be inhibited at room
 218 temperature. The main causes of ready to eat food poisoning are microbial pathogens such as *S.*
 219 *aureus* (Huong *et al.* 2010).

220

221 Yeast and mould counts

222 *C. hystrix* extract improved the microbial quality of sticky rice cake particularly on the level
 223 of 1.82 % extract over 14 days storage compared to the control (see Table 3). However, after 28
 224 days of storage, the sticky rice cake formulated with 1.26% extract gave a better shelf life toward
 225 yeast and mould growth with the number of 26×10^2 cfu/ml compared by control. Mould growth in
 226 this formula is an indication of high moisture content in the product. The higher extract was
 227 formulated in the sticky rice cake, the higher amount of lactose was added. Consequently, it will
 228 easily absorb water, thus allowing the mould to grow. It was shown by the moisture content of
 229 product formulated with 1.82% which is higher than those of other formulas (19.14%).

230

231 Table 3 Total yeast and mould of sticky rice cake formulated with *C. hystrix* extract

Storage duration (days)		Yeast and mould (cfu/ml)			
		Control	A	B	C
0	3 days	2.0×10^2	2.0×10^2	0.4×10^2	0.3×10^2
	5 days	3.0×10^2	4.0×10^2	3.0×10^2	2.0×10^2
7	3 days	3.8×10^2	3.2×10^2	1.4×10^2	1.4×10^2
	5 days	4.1×10^2	4.1×10^2	4.0×10^2	3.0×10^2
14	3 days	10×10^2	1.0×10^2	1.0×10^2	0.9×10^2
	5 days	4.0×10^2	5.0×10^2	2.0×10^2	1.8×10^2
21	3 days	9.0×10^2	8.0×10^2	1.0×10^2	4.0×10^2
	5 days	12×10^2	11×10^2	3.0×10^2	11×10^2
28	3 days	34×10^2	25×10^2	21×10^2	27×10^2
	5 days	39×10^2	28×10^2	26×10^2	32×10^2

232 Control : product in the absence of extract.; A : product formulated with 0.6% extract; B : product
 233 formulated with 1.26% extract; C : product formulated with 1.82%.

234

235 Moulds are filamentous fungi with branching hyphae, multi-cellular, generally aerobic and
 236 grow at a pH range of 3 to 8. The spores can tolerate harsh environmental conditions but sensitive
 237 to heat treatment (Pal 2007). The formulation of sticky rice cake with *C. hystrix* extract slightly
 238 decreased the pH value of product. Moreover, the highest acidity of sticky rice cake was reached
 239 during the 28 days storage on the sticky rice cake formulated with 1.82% extract. The high acidity
 240 favours the growth of moulds (Loureiro & Querol 1999). The addition of extract effectively
 241 eliminated the spoilage microorganism until 28 days storage. After 28 days, the product was fully
 242 contaminated by moulds. As reported by Ji *et al.* (2007), microorganism grows significantly over

243 time in rice cakes and the product would completely spoiled shown by altering the colour, texture,
 244 and flavour after three days of storage at room temperature. Moulds commonly grow on all kinds of
 245 food such as cereals, meat, milk, fruit, vegetables, nuts, and fats. Mycotoxins, the toxic secondary
 246 metabolites toward vertebrate animals in small amounts through natural route, are produced during
 247 growth of moulds on food products. Mycotoxins from *Penicillia* normally grow in cereals kept in
 248 tropical countries with warm climate and high moisture including viridicatumtoxin (*P.*
 249 *aethiopicum*), citrinin (*P.citrinum*), cyclopiazonic acid, patulin and roquefortine C (*P.*
 250 *griseofulcum*) and secalonic acid D (*P. oxalicum*) (Frisvad & Filtenborg 1989).

251

252 TBARS analysis

253 Sticky rice cake, containing coconut milk, renders a major problem associated with the
 254 producing of off-odors due to free radical induced lipid oxidation. The *C. hystrix* extract was added
 255 to a processed sticky rice cake to improve sensory attributes and control oxidation reactions. The
 256 effect of *C. hystrix* extracts after processing and during room temperature storage is shown in Table
 257 4.

258

259 Table 4 Effects of *C. hystrix* extract on oxidative stability (TBARS values) of sticky rice cake

Storage duration (days)	TBARS index (mg MDA/kg)			
	Control	A	B	C
0	0.45	0.22	0.39	0.24
7	0.33	0.39	0.39	0.49
14	0.22	0.16	0.41	0.17
21	0.67	0.47	0.33	0.36
28	0.18	0.19	0.15	0.12

260 Control : product in the absence of extract.; A : product formulated with 0.6% extract; B : product
 261 formulated with 1.26% extract; C : product formulated with 1.82%.

262

263 Adding *C. hystrix* extract to sticky rice cake rather reduced the TBARS values during 28
 264 days storage with no significant difference between the three levels extract formula. Addition of
 265 herbal extracts as antioxidant supplementation in some foods was effective in controlling lipid
 266 oxidation during storage (Formanek *et al.* 2001; Park *et al.* 2016). Moreover, the pH of food
 267 condition gives influence in the activity of oils. At low pH, the hydrophobicity of some essential
 268 oils contained in *C. hystrix* increase and it is subjected to partition in the lipid phase of the food,
 269 thus affecting in reducing oxidative stability.

270

271 Sensory analysis

272 The organoleptic analysis was carried out after a week in storage to evaluate the influence of
273 *C. hystrix* extract on the acceptability of product by customer (see Table 5).

274

275

276 Table 5 Sensory evaluation of sticky rice cake formulated with different levels of *C. hystrix* extract

Level of extracts (%)	Attributes				
	Colour	Odor	Taste	Texture	Overall acceptability
Control (0)	5.89 ^{ab}	6.33 ^b	6.67 ^b	5.94 ^b	6.56 ^b
0.65	5.50 ^a	5.50 ^{ab}	5.61 ^a	4.89 ^a	5.78 ^a
1.26	6.06 ^{ab}	5.39 ^a	5.67 ^{ab}	5.78 ^{ab}	5.94 ^{ab}
1.82	6.44 ^b	5.89 ^{ab}	5.83 ^{ab}	6.33 ^b	6.67 ^b

277 ^{a,b,ab} Mean values in the same column with different letters are significantly different at $p < 0.05$

278

279 The addition of *C. hystrix* extract did not reduce the acceptability of sticky rice cake on the
280 sensory attributes as compared to the control sample, sticky rice cake with no addition of extract.
281 Statistically, there is no significant difference of acceptance among products formulated with
282 0.65%, 1.26%, and 1.82% extracts. However, the sensory test should be performed every week until
283 28 days storage in order to observe the alteration of acceptance level. Most plant extracts do not
284 give the effect on the acceptability of sticky rice cake as long as the appropriate concentration is
285 used. Irradiated and freeze-dried green tea leaf extracts formulated in raw and cooked pork patties
286 were reported giving no influence on the physical and sensory properties (Jo *et al.* 2003). The
287 results of our study indicated that sticky rice cake formulated with *C. hystrix* extracts exhibited
288 sensory stability similar to the control on one-week storage.

289

290

CONCLUSION

291 Addition of *C. hystrix* extract in food formulation could reduce the microbial growth as well
292 as extending the shelf life of sticky rice cake. This suggests that *C. hystrix* enhances the antifungal
293 activity over time. The increasing level of *C. hystrix* extract up to 1.82% of sample (w/w) is still
294 acceptable in overall sensory attributes. Thus, sticky rice cake with *C. hystrix* extract adds potential
295 value to efforts to provide the quality and healthy food options in Indonesia.

296

297

ACKNOWLEDGEMENTS

298 This work was fully supported by Grants-in-Aid for scientific research from the Indonesian
299 Institute of Sciences (LIPI). The authors are grateful to “Program Bantuan Seminar Luar Negeri,
300 Ditjen Penguatan Riset dan Pengembangan, Kemenristekdikti” for providing the conference
301 funding.

302

303

REFERENCES

- 304 Brul S, Coote P. 1999. Preservative agents in foods. Mode of action and microbial resistance
305 mechanisms. *Int J Food Microbiol* 50, 1-17.
- 306 California State Legislature. 2016. Health and Safety Code-HSC. Division 104. Environmental
307 Health. Part 5. Sherman Food, Drug, and Cosmetic Laws. Chapter 5. Food. Article 14.
308 Asian rice noodles, Korean rice cakes and Vietnamese rice cakes.
- 309 Filtenborg O, Frisvad JC, Thrane U. 1996. Moulds in food spoilage. *International Journal of Food*
310 *Microbiology* 33, 85-102.
- 311 Frisvad JC, Filtenborg O. 1989. Terverticillate penicillia: chemotaxonomy and
312 mycotoxinproduction. *Mycologia* 81, 836-861.
- 313 Holley AH, Patel HM. 2005. Improvement in shelf life and safety of perishable food by plant
314 essential oils and smoke antimicrobials. *International Journal of Food Microbiology*, 22,
315 273-292.
- 316 Huong BTM, Mahmud ZH, Neogi SB, Kassu A, Nhien NV. 2010. Toxigenicity and genetic
317 diversity of *Staphylococcus aureus* isolated from Vietnamese ready-to-eat foods. *Food*
318 *Control* 21, 166-171.
- 319 Ji Y, Zhu K, Qian H, Zhou H. 2007. Microbiological characteristics of cake prepared from rice
320 flour and sticky rice flour. *Food Control* 18, 1507-1511.
- 321 Jo C, Son JH, Son CB, Byun MW. 2003. Functional properties of raw and cooked pork patties with
322 added irradiated, freeze-dried green tea leaf extract powder during storage at 4 °C. *Meat Sci.*
323 64, 17-33.
- 324 Lee SY, Gwon SY, Kim SJ, Moon BK. 2009. Inhibitory effect of commercial green tea and
325 rosemary leaf powders on the growth of foodborne pathogens in laboratory media and
326 oriental-style rice cakes. *Journal of Food Protection* 72 (5), 1107-1111.
- 327 Loureiro V, Querol AD. 1999. The prevalence and control of spoilage yeasts in foods and
328 beverages. *Trends in Food Science Technology* 10, 356-365.
- 329 Meilgaard MC, Civille GV Carr BT. 1991. Sensory evaluation techniques. 2nd Edition. Washington
330 DC: CRC Press
- 331 Molan PC. 1992. The antibacterial activity of honey: 1. The nature of the antibacterial activity. *Bee*
332 *World*, 73(1), 5-28.
- 333 Mori M, Aoyama M, Doi S, Kanetoshi A, Hayashi T. 1997. Antifungal activity of bark extract of
334 deciduous trees. *Holz als Roh und Werkstoff* 55: 130-132.
- 335 Morris VJ. 1990. Starch gelation and retrogradation. *Trends in Food Science & Technology* 1, 2-6.
- 336 Mundo MA, Padilla-Zakour OI, Worobo RW. 2004. Growth inhibition of foodborne pathogens and
337 food spoilage organisms by select raw honeys. *International Journal of Food Microbiology*
338 97, 1-8.
- 339 NSW Food Authority. 2009. Microbiological quality guide for ready-to-eat foods.
340 NSW/FA/CP028/0906, 6-7.
- 341 Pal M. 2007. *Veterinary and Medical Mycology*. 1st Edition. Indian Council of Agricultural
342 Research, New Delhi, India.
- 343 Prakash B, Mishra PK, Kedia A, Dubey NK. 2014. Antifungal, antiaflatoxin and antioxidant
344 potential of chemically characterized *Boswellia carterii* Birdw essential oil and its *in vivo*

- 345 practical applicability in preservation of *Piper nigrum* L. fruits. LWT– Food Sci. Technol.
346 56, 240-247.
- 347 Serra AT, Ana AM, Ana VMN, Leitão MC, Brito D, Bronze R, SilvaS, Pires A, Crespo MT, San
348 Romão MV, Duarte CM. 2008. In vitro evaluation of olive- and grape-based natural extracts
349 as potential preservatives for food. Innovative Food Science and Emerging Technologies 9,
350 311-319.
- 351 Singh A, Sharma PK, Garg G. 2010. Natural products as preservatives. International Journal of
352 Pharma and Bio Sciences 1(4), 601-612.
- 353 Targladis BG, Watts BM, Younathan MT, Duggan LR. 1960. A distillation method for the
354 quantitative determination of malonaldehyde in rancid foods. J. Am. Oil Chem. Soc. 37, 44-
355 48.
- 356 Nanasombat S, Lohasupthawee P. 2005. Antibacterial activity of crude ethanolic extracts and
357 essential oils of spices against salmonellae and other enterobacteria. KMITL Sci Tech J. 5,
358 527-538.
- 359 Waikedre J, Dugay A, Barrachina I, Herrenknecht C, Cabalion P, Fournet A. 2010. Chemical
360 composition and antimicrobial activity of the essential oils from new Caledonian *Citrus*
361 *macroptera* and *Citrus hystrix*. Chemistry & Biodiversity 7, 871-877.

ACCEPTED MANUSCRIPT