

Supplementary data for article:

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Application of bismuth (III) oxide decorated graphene nanoribbons for enzymatic glucose biosensing

Slađana Đurđić¹, Vesna Vukojević², Filip Vlahović³, Miloš Ognjanović⁴, Ľubomir Švorc⁵, Kurt Kalcher⁶, Jelena Mutić¹, Dalibor M. Stanković⁴

¹Faculty of Chemistry, University of Belgrade, Studentski trg 12-16, 11000 Belgrade, Serbia

²Institute of Chemistry, Technology and Metallurgy, University of Belgrade, Njegoseva 12, 11000 Belgrade, Serbia

³Innovation center of the Faculty of Chemistry, University of Belgrade, POB 51, 118, 11158 Belgrade, Serbia

⁴The Vinca Institute of Nuclear Sciences, University of Belgrade, POB 522, 11001 Belgrade, Serbia

⁵Institute of Analytical Chemistry, Faculty of Chemical and Food Technology, Slovak University of Technology in Bratislava, Radlinského 9, Bratislava, SK-812 37, Slovak Republic

⁶Institute of Chemistry-Analytical Chemistry, Karl-Franzens University Graz, A-8010 Graz, Austria

Supplementary material

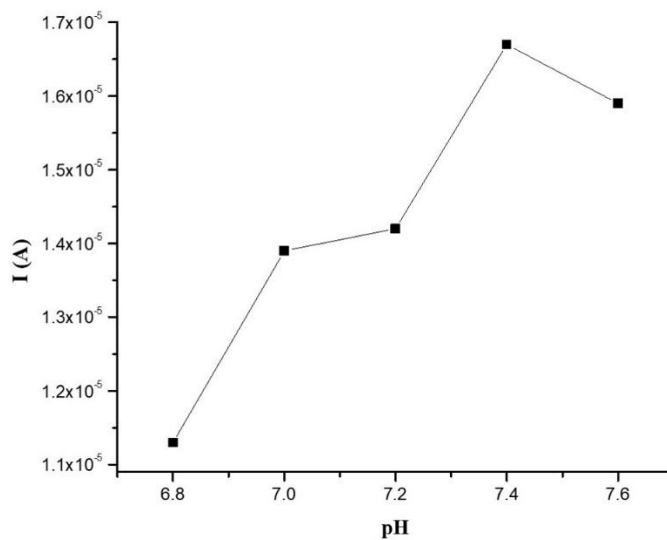


Figure S1. Effect of different pH of 0.1 M phosphate buffer

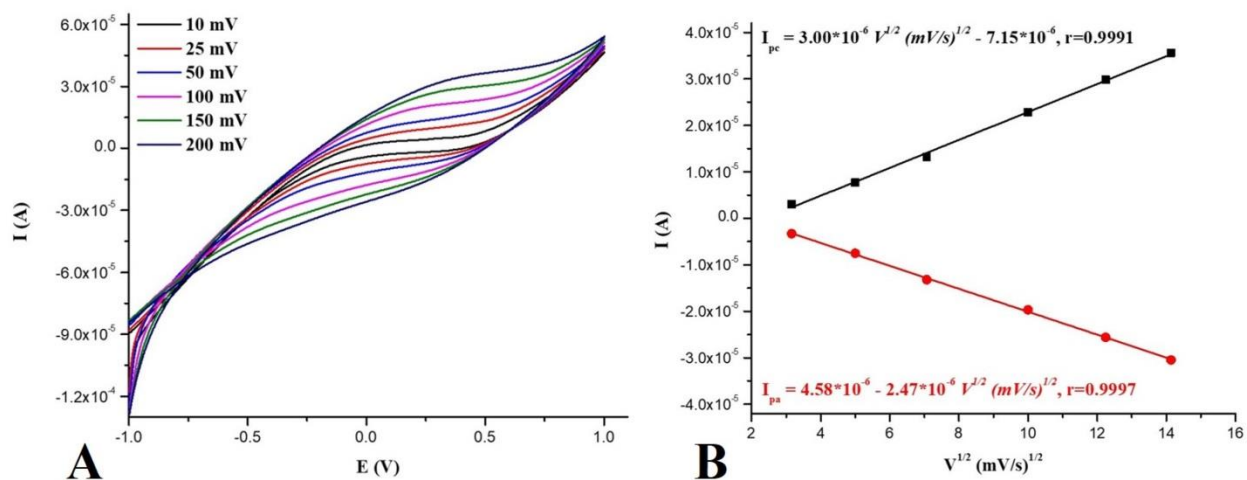


Figure S2. A) Cyclic voltammograms of SPCE/GNR/Bi₂O₃ in 0.1 M PBS (pH=7.40) containing 2.5 mM H₂O₂ at different scan rates (10 mV – 200 mV). B) Plots of the cathodic (I_{pc}) and anodic (I_{pa}) peak current vs. the square root of the scan rate ($V^{1/2}$).

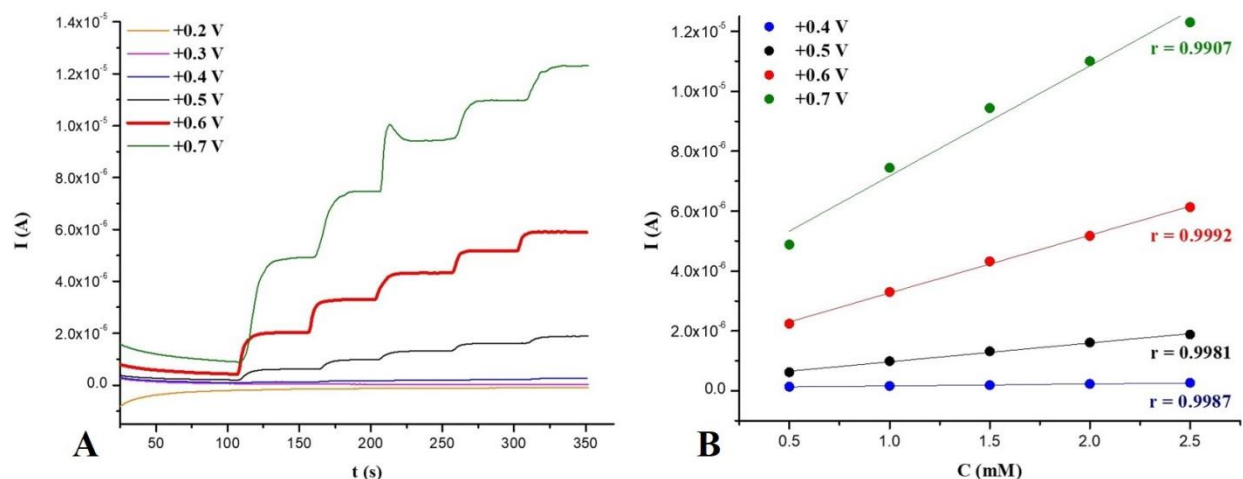


Figure S3. A) Chronoamperometric responses of SPCE/GNR/Bi₂O₃ upon successive addition of H₂O₂ in 0.1 M PBS (pH=7.40) at different working potentials (+0.2 V to +0.7 V). B) Plots of the current intensity vs. the concentration of H₂O₂ for working potentials +0.4 V, +0.5 V, +0.6 V and +0.7 V.

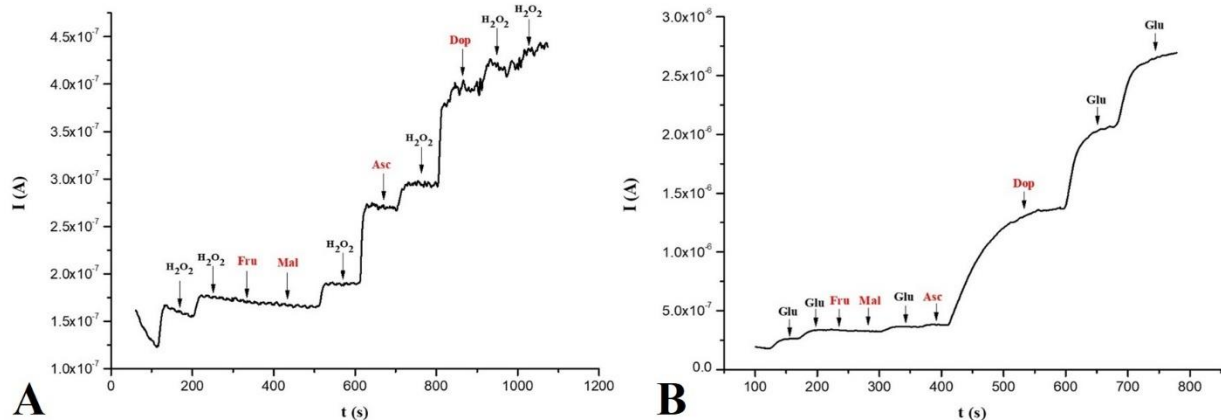


Figure S4. Influence of different interfering compounds during the quantification of H₂O₂ with SPCE/GNR/Bi₂O₃ (A) and glucose with SPCE/GNR/Bi₂O₃/GO_x/Naf/ biosensor (B).

Table S1. Glucose content obtained by analysing the different aliquots of the honey sample

Aliquot (mL)	Declared value (%)	Found value (%)	Recovery (%)
1.0	32.6	32.1	98.5
2.0	32.6	31.0	95.0
3.0	32.6	32.4	99.4
4.0	32.6	32.0	98.2
5.0	32.6	31.9	97.8

Table S2. Glucose concentration (mM) in blood serum and urine samples determined with developed biosensor

	Blood serum samples	Urine samples
Volunteer 1	3.81	<LOD*
Volunteer 2	4.23	<LOD
Volunteer 3	5.76	<LOD

*LOD – limit of detection

Table S3. Comparison of spiked samples and obtained glucose content in urine samples with proposed method

Spiked (mM)	Found (mM)	Recovery (%)
0.32	0.30	93.8
0.48	0.46	95.8
1.04	0.99	95.2
1.68	1.70	98.8